NH₄HCO₃ gas-generating liposomal nanoparticle for photoacoustic imaging in breast cancer

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Abstract: In this study, we have developed a biodegradable nanomaterial for photoacoustic imaging (PAI). Its biodegradation products can be fully eliminated from a living organism. It is a gas-generating nanoparticle of liposome-encapsulating ammonium bicarbonate (NH₄HCO₃) solution, which is safe, effective, inexpensive, and free of side effects. When lasers irradiate these nanoparticles, NH₄HCO₃ decomposes to produce CO₂, which can absorb much of the light energy under laser irradiation with a specific wavelength, and then expand under heat to generate a thermal acoustic wave. An acoustic detector can detect this wave and show it as a photoacoustic signal on a display screen. The intensity of the photoacoustic signal is enhanced corresponding to an increase in time, concentration, and temperature. During in vivo testing, nanoparticles were injected into tumor-bearing nude mice through the caudal vein, and photoacoustic signals were detected from the tumor, reaching a peak in 4 h, and then gradually disappearing. There was no damage to the skin or subcutaneous tissue from laser radiation. Our developed gas-generating nanomaterial, NH₄HCO₃, is feasible, effective, safe, and inexpensive. Therefore, it is a promising material to be used in clinical PAI.

Keywords: Photoacoustic tomography, CO₂, NH₄HCO₃, contrast agent, cancer

Introduction

Photoacoustic imaging (PAI) has developed rapidly in recent years. It is a non-destructive biophoton imaging method based on optical absorption differences in biological tissue and mediated by laser irradiation and ultrasound. It has the advantages of the high contrast of pure optical imagery and the high permeability of pure ultrasound; therefore, it is an important method for evaluating morphology, physiological features, metabolic functions, and pathological features of biological tissue.¹⁻² PAI has promising applications in biomedical clinical diagnosis and has become a “hot topic” in recent studies. Its principle is as follows: when irradiated by a pulsed laser, a biological tissue absorbs the laser energy, causing a local temperature rise and thermoelastic expansion that generates an ultrasound signal, which can be detected by an ultrasonic detector near the tissue.³ The photoacoustic signal is then analyzed to reconstruct an optical absorption distribution image.

However, because the biological tissue has a strong scattering effect on lasers, the light intensity and the photoacoustic signal-to-noise ratio exhibit exponential decay corresponding to the increase in tissue depth. Thus, the research direction is to develop a contrast agent to enhance the PAI effect.⁴⁻⁶ At present, major materials used for PAI contrast agents include gold nanomaterials, dyes, and carbon nanomaterials.⁷⁻¹² However, the use of such materials is controversial in PAI. As a heavy metal, gold cannot be excreted by a living organism and, thus, is hazardous to health. Gold is also very...
expensive. Indocyanine green is approved by the China Food and Drug Administration as a safe dye, and while a large portion of it can be eliminated, it is costly and takes a long time to be completely cleared from the body. Carbon nanotubes are not pure carbon and have a long residence time within a living organism. Only a tiny portion of them is eliminated through urine or feces, and they can cause eye discomfort, skin allergy, lung cancer, and pneumoconiosis. Because the above-mentioned contrast agents have such disadvantages as the inability to be fully eliminated by living organisms, have side effects, and are costly, they cannot be widely applied in clinical practice. Therefore, it is necessary to find a safe and inexpensive substance without side effects to serve as a contrast-enhancing material for PAI.

In our previous study, we found that not only could an \( \text{NH}_4\text{HCO}_3 \) solution emit photoacoustic signals but that all of its decomposition products are normal metabolites of the human body, can be completely eliminated from living organisms, and are nontoxic to humans. Based on the results of this study, we propose a nanoscale contrast agent made of a liposome-encapsulating \( \text{NH}_4\text{HCO}_3 \) solution that can be applied in PAI. We have evaluated its feasibility and effectiveness through in vitro qualitative and quantitative tests, explored its imaging capacity, and tested its safety and imaging effects through animal tests to lay a solid foundation for its application in PAI (Figure 1).

**Materials and methods**

**Preparation of the nanoparticles**

The thin-film hydration method plus the liposome extrusion method used in preparation was as follows: 1) At a mass ratio of 3:1:1, cholesterol-free distearoyl phosphatidylcholine, distearoylphosphatidylethanolamine-polyethylene glycol 2000 (Avanti Polar Lipids, Inc., Alabaster, AL, USA), and cholesterol (Sangon Biotech Co., Ltd, Shanghai, People’s Republic of China) were added to 10 mL of chloroform (Chongqing Chuandong Chemical Group Co., Ltd, Chongqing, People’s Republic of China) and mixed thoroughly. 2) On a rotavapor (Shanghai Yarong Biochemical Instruments Factory, Shanghai, People’s Republic of China), the above-mentioned solution was rotated at 50°C at a rotational speed of 60 rpm for ~60 min. 3) After the rotation, 5 mL of \( \text{NH}_4\text{HCO}_3 \) (Chengdu Kelong Chemical Reagent Factory, Chengdu, People’s Republic of China) saturated solution was added, and the resulting solution was placed in an ultrasound cleaner (Ningbo Scientz Biotechnology Co., Ltd, Ningbo, People’s Republic of China) to be shaken for a few seconds. 4) The above-mentioned solution was passed through extruder filter films (Whatman Plc, Kent, UK) with apertures of 800, 400, and 200 nm, respectively. 5) The extruded solution was dissolved in a 5% glucose solution (Chengdu Kelong Chemical Reagent Factory) for dialysis.

![Figure 1 The generation of photoacoustic signal.](image)

**Notes:** When a laser irradiates a nanoparticle, it is decomposed into \( \text{CO}_2, \text{H}_2\text{O}, \) and ammonia. \( \text{CO}_2 \) absorbs much of the light energy at a specific wavelength and expands under heat to produce a thermal acoustic wave, which can be detected by the acoustic detector and then displayed on a screen as a photoacoustic signal.

**Abbreviations:** \( \text{HCO}_3^- \), bicarbonate; \( \text{NH}_4^+ \), ammonium.
Optical microscope and electron microscopy
Nanoparticle morphology and distribution were observed using a BX 54 optical microscope (Olympus Optical, Tokyo, Japan) and a S-3000N field emission scanning electron microscope (Hitachi, Tokyo, Japan); nanoparticle size and zeta potential were measured using a laser particle size analyzer (Malvern Instruments Ltd, Shanghai, People’s Republic of China).

Detection of NH₄HCO₃ solution encapsulation
A CaCl₂ solution was added into the NH₄HCO₃ solution, double-distilled H₂O (dd H₂O)-encapsulating nanoparticles (DDHN), NH₄HCO₃ encapsulating nanoparticles (ABN), broken DDHN, and broken ABN. The changes in them were observed.

Infrared spectrometer
Absorption spectra of NH₄HCO₃ solution, CO₂, dd H₂O, and ammonia solution were determined using the Spectrum 100 infrared spectrometer (PerkinElmer, Inc., Melville, New York, NY, USA).

In vitro PAI test
The NH₄HCO₃ solution was irradiated (excitation wavelength: 680–970 nm) by the VevoLAZR PAI system (VisualSonics, Inc., Toronto, ON, Canada) to determine the optimal irradiation wavelength.

Photoacoustic signal images of NH₄HCO₃ solution, CO₂, dd H₂O, ammonia solution, ABN, and DDHN were determined using the PAI system.

The ABN and DDHN samples were irradiated using the PAI system. The changes in the two samples of photoacoustic and ultrasonic models were observed. The changes of ABN in the photoacoustic image corresponding with time, concentration, and temperature were observed.

In vivo PAI test
Animals and experimental grouping
Twelve female nude mice aged 4–6 weeks were raised and supplied by the Laboratory Animal Center of Chongqing Medical University (Chongqing, People’s Republic of China). All animal experiments were carried out in accordance with the guidelines of the Ethics Committee of Chongqing Medical University, which approved the study. Twelve tumor-bearing nude mice were taken and randomly divided into two groups: 1) treatment group: 1.0 mL of ABN was injected through the caudal vein and 2) control group: 1.0 mL of water-encapsulating nanoparticles was injected through the caudal vein.

Tumor cells culture and animal model
MDA-MB-231 tumor cells are tumor cell lines subcultured by the Institute of Ultrasound Imaging of Chongqing Medical University. These cells were obtained from the Basic Medical Research Institute of Chongqing Medical University. These cells were cultured in culture flasks loaded with 10% fetal bovine serum (Gibco, Shanghai, People’s Republic of China) and Dulbecco’s Modified Eagle’s Medium (Gibco), and incubated at 37°C with 5% CO₂ and saturated humidity. When cell concentration was ~6×10⁷/mL, 0.2 mL of the cell suspension was subcutaneously injected into the nude mice on their right rear legs using a sterile syringe. In ~3 weeks, subcutaneous tumors of 8–9 mm developed.

Photoacoustic imaging
The subcutaneous tumors of tumor-bearing nude mice in each group were irradiated by the PAI system, and photoacoustic images of each group at various time points before and after injection were observed.

Observation with naked eyes
Skin integrity and damage were observed with naked eyes before and after using the PAI system.

Pathological examination
After image acquisition, the tumor-bearing nude mice were sacrificed by neck-breaking, and the tumors with adjacent skin tissues were then taken and fixed with 4% paraformaldehyde. After being paraffin embedded, sectioned, and hematoxylin and eosin (H&E) stained, the tissues were observed under an optical microscope for changes in the cell structure of skin and tumor tissue.

Statistics
The data were analyzed using SPSS 13.0 (SPSS Inc., Chicago, IL, USA). All measurement data were expressed as mean ± standard deviation (SD). The means between groups were compared using one-factor analysis of variance (ANOVA), whereas pair-wise comparisons were made using Fisher’s least significant difference method. P<0.05 was considered statistically significant.

Results
General physical properties test
Observed under an optical microscope, nanoparticles were spherical with a regular shape and uniform size and distribution. There was no obvious aggregation with a concentration of ~2.63×10⁹/mL counted manually (Figure 2).
Observation of a single nanoparticle under a transmission electron microscope (TEM) showed a two-layer phospholipid membrane on the outer compartment of the nanoparticle and \(\text{NH}_4\text{HCO}_3\) solution filling the core (Figure 3).

Measured by a laser particle size analyzer, the mean particle size of nanoparticles was 230.9 ± 54.58 nm, with uniform distribution and dispersion. The zeta potential was \(-22.8\pm5.75\) mV.

**Detection of \(\text{NH}_4\text{HCO}_3\) solution encapsulation**

The \(\text{NH}_4\text{HCO}_3\) solution appeared to have a large amount of white precipitate. The broken ABN sample changed from clear to white. There was no other color change observed before or after \(\text{CaCl}_2\) solution was added into DDHN, broken DDHN, and ABN (Figure 4).

**Study on PAI mechanism by the ABN**

The spectra of the \(\text{NH}_4\text{HCO}_3\) solution, the ammonia solution, \(\text{dd H}_2\text{O}\), and \(\text{CO}_2\) were acquired using the infrared spectrometer. The results showed that absorption peak of the ammonium bicarbonate solution was at \(-1,300\) nm, ammonia solution at \(-1,100\) nm, \(\text{dd H}_2\text{O}\) at \(-720\) nm, and \(\text{CO}_2\) at \(-700\) nm (Figure 5).

**The optimal laser excitation wavelength**

The \(\text{NH}_4\text{HCO}_3\) solution was irradiated with the PAI system at laser excitation wavelengths ranging from 680 to 970 nm; the results showed that the photoacoustic signal intensity was optimal at an excitation wavelength of \(-700\) nm (Figure 6).

Gel models loaded with the \(\text{NH}_4\text{HCO}_3\) solution, \(\text{dd H}_2\text{O}\), and ammonia solution were irradiated with the PAI system. The results showed that the \(\text{NH}_4\text{HCO}_3\) solution emitted a photoacoustic signal, whereas the \(\text{dd H}_2\text{O}\) and ammonia solution emitted no photoacoustic signal (Figure 7).

There was no photoacoustic signal recorded before the infusion of pure \(\text{CO}_2\) into the sealing device, whereas there was a photoacoustic signal recorded after the infusion of pure \(\text{CO}_2\) (Figure 8).

We could see that the ABN had a photoacoustic signal and ultrasonic echo enhancement, but the DDHN did not (Figure 9).

**In vitro PAI experiment**

**Changes in the photoacoustic signal of ABN and \(\text{NH}_4\text{HCO}_3\) solution corresponding with time**

ABN samples were irradiated with the PAI system, and the results showed that the photoacoustic signal intensity gradually increased with time to reach a peak and then gradually decreased with time. The \(\text{NH}_4\text{HCO}_3\) solution reached a peak in a short period of time and decreased soon after (Figure 10).

**Changes in the photoacoustic signal of ABN at different concentrations**

For serial 2-fold diluted nanoparticle samples irradiated with the PAI system, the photoacoustic signal intensity serially decreased with decreasing concentration (Figures 11 and 12).

**Changes in the photoacoustic signal of ABN at different temperatures**

The samples irradiated with the PAI system at different ambient temperatures (25°C, 37°C, and 42°C) showed the
photoacoustic signal intensities of 0.56±0.11, 0.9±0.21, and 0.91±0.15, respectively. Compared to the samples at 37°C and 42°C, the sample at 25°C had the weakest photoacoustic signal intensity. There was no statistical difference found in the photoacoustic signal intensities between the samples at 42°C and 37°C (P>0.05; Figures 13 and 14).

**In vivo PAI experiment**

**Intratumoral PAI**

Irradiating subcutaneous tumors with the PAI system showed that the treatment group recorded no photoacoustic signal before injection, and only a slight photoacoustic signal was detected 30 min after injection. The signal gradually

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**Figure 4** Change of CaCl$_2$ added into the NH$_4$HCO$_3$ solution, DDHN, broken DDHN, ABN, and broken ABN. 
**Abbreviations:** ABN, ammonium bicarbonate-encapsulating nanoparticles; DDHN, double-distilled H$_2$O-encapsulating nanoparticles.

**Figure 5** Infrared absorption spectra of various substances.
**Notes:** The absorption peaks of the NH$_4$HCO$_3$ solution were at ~1,300 nm, of the ammonia solution at ~1,100 nm, of dd H$_2$O at ~720 nm, and of CO$_2$ at ~700 nm. 
**Abbreviations:** dd H$_2$O, double-distilled H$_2$O; NH$_4$HCO$_3$, ammonium bicarbonate.
increased 1 h after injection; the increased signal intensity, which reached its peak 4 h after injection, gradually decreased and then completely disappeared 24 h after injection. No photoacoustic signal was observed in the control group before or after injection (Figures 15 and 16).

Tumors were observed with naked eyes
After laser irradiation, the skin had no redness, but was not swollen, and had injury (Figure 17).

Pathological examination
After H&E staining, tissues from skin and subcutaneous tumors were observed under an optical microscope. There had been no marked difference observed in the cell morphology, cell nucleus, and cytomembrane. There was no obvious necrosis and the structure of tumor tissue was not damaged (Figure 18).

Discussion
NH$_4$HCO$_3$ is a weak alkaline compound with a molecular weight of 79.06. It is a white powder and a food additive that was approved by the Food and Drug Administration of the United States in 1985. It is safe, hazard-free, and inexpensive.

NH$_4$HCO$_3$ decomposes into water, ammonia, and carbon dioxide without leaving any residue; all the decomposition products are normal metabolites of the human body, which are completely eliminated through the respiratory, digestive, and urinary systems.

Our preliminary research showed that a saturated NH$_4$HCO$_3$ solution could emit photoacoustic signals. To investigate the cause of the photoacoustic signals, we analyzed the NH$_4$HCO$_3$ solution, the ammonia solution, dd H$_2$O, and CO$_2$ using an infrared spectrometer. The results showed that the absorption peak of CO$_2$ is at ~700 nm and that of dd H$_2$O is at ~720 nm. In this study, the laser excitation wavelength ranged between 680 and 970 nm, suggesting that dd H$_2$O and CO$_2$ could absorb light energy within this range. We then tested all the samples using the PAI system. First, we needed to confirm the excitation wavelength. When the NH$_4$HCO$_3$ solution was irradiated by the laser at a wavelength of 680–970 nm from the PAI system, it was...
found that the optimal photoacoustic signal was at the 700 nm wavelength. This wavelength is consistent with that of the strongest absorption peak of CO$_2$ detected by the infrared spectrometer, indicating that the excitation wavelength is the optimal wavelength of CO$_2$, and that the photoacoustic signal arises from CO$_2$. Therefore, 700 nm was selected as the excitation wavelength in this study. Our results showed that the NH$_4$HCO$_3$ solution and CO$_2$ produced photoacoustic signals, whereas the other samples did not. Therefore, we propose the following explanation: the NH$_4$HCO$_3$ solution is irradiated by a laser, and the water absorbs the light energy and converts it into heat energy, which causes NH$_4$HCO$_3$ to decompose into CO$_2$, ammonia, and water. Ammonia is extremely highly soluble in water, whereas CO$_2$ has poor water solubility, with most of it remaining in gas form. After being irradiated by a laser at a specific wavelength, the CO$_2$ absorbs much of the light energy at a specific wavelength and expands under heat to produce a thermal acoustic wave, which can be detected by the acoustic detector and then displayed on a screen as a photoacoustic signal.

In our previous study, we also found that the photoacoustic signal only lasted for a short time when the NH$_4$HCO$_3$ solution was irradiated by the PAI system. As we know, NH$_4$HCO$_3$ exists in the solution as HCO$_3^-$ and NH$_4^+$. HCO$_3^-$ tends to be neutralized by the acidic buffer pair in the blood, and NH$_4^+$ tends to transform into urea in the liver and is excreted in the urine. Therefore, it is very difficult to achieve PAI at the target area. It is necessary to encapsulate NH$_4$HCO$_3$ with a shell membrane to prevent it from being neutralized by the buffer pair in the blood,
to reduce its transformation in the liver, and to prolong the time of the photoacoustic signal. As an enveloping membrane, a liposome has characteristics such as good biocompatibility, stability, elasticity, and thinness and has been successfully applied in clinical practice. Therefore, we employed a thin-film hydration method plus an extrusion method to encapsulate the saturated \( \text{NH}_4\text{HCO}_3 \) solution in a spherical liposomal vesicle and prepared nanoparticles of phospholipid membrane encapsulating \( \text{NH}_4\text{HCO}_3 \). After testing with an optical microscope, an electron microscope, a laser particle size analyzer, and a zeta potential analyzer, results showed that the nanoparticles in the sample appeared to be spherical with a regular shape, a relatively uniform size of \( \sim 230 \) nm, and with a potential of \( \sim -23 \) mV. The nanoparticles are smaller than the intercellular space of newly formed tumor endothelial cells, which allows the nanoparticles to cross the intercellular space and move into the tumor tissues.\(^{11}\)

To verify that the \( \text{NH}_4\text{HCO}_3 \) solution was packaged in a liposome, we used a \( \text{CaCl}_2 \) solution. From the changes in each sample, we could see that the membranes of ABN

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**Figure 14** Temperature of ammonium bicarbonate-encapsulating nanoparticles versus photoacoustic signal intensity.

**Notes:** There is no significant difference between 37°C and 42°C. But there is a significant difference between 25°C and 37°C and/or 42°C. *Statistical significance.

**Figure 15** Photoacoustic images of subcutaneous tumor in each group.

**Notes:** Treatment group recorded photoacoustic signal with time. Control group recorded no photoacoustic signal with time. (A1–A10) Treatment group (A1: before injection, A2–A10: 30 min, 1, 2, 3, 4, 5, 6, 12, and 24 h after injection, respectively). (B1–B10) Control group (B1: before injection, B2–B10: 30 min, 1, 2, 3, 4, 5, 6, 12, and 24 h after injection, respectively). The positive result is represented within the dotted lines.
were broken and the NH₄HCO₃ solution overflowed. When the CaCl₂ solution was mixed into the NH₄HCO₃ solution, the NH₄HCO₃ solution changed from clear to muddy (white precipitation/white precipitate). By contrast, the membrane of ABN had no sign of destruction, the NH₄HCO₃ solution was still enveloped in the liposome, and the CaCl₂ solution did not react with the NH₄HCO₃ solution. There was no change in color (from clear to muddy) indicating that DDHN and broken DDHN had not enveloped the NH₄HCO₃ solution. From the above-described phenomenon, we could deduce that the NH₄HCO₃ solution was enveloped in a liposome.

At the same time, we tested ABN and DDHN using the PAI system. The photoacoustic model showed that ABN had a photoacoustic signal, while the DDHN did not. And the ultrasonic model showed an enhanced echo of ABN, while the DDHN did not. Several studies have reported the enhanced echo of ABN caused by CO₂.¹⁴–¹⁶ Our study also found that ABN had echo and photoacoustic signals, which further demonstrated that photoacoustic signals are caused by CO₂.

We then investigated the changes in the photoacoustic signal of the ABN and NH₄HCO₃ solutions with regard to time duration. Our results showed that the photoacoustic signal intensity of ABN gradually increased, and then slowly decreased as the irradiation time was progressively lengthened. The photoacoustic signal of the NH₄HCO₃ solution quickly reached a peak value and decreased soon after. This phenomenon might be related to the rate and amount of CO₂ generated by NH₄HCO₃. After being irradiated by a laser, the ABN generate CO₂, which is inside the liposomal vesicles. Owing to its small molecular weight, CO₂ continuously diffuses across the thin phospholipid membranes out of the spherical vesicles, and then expands under laser irradiation to emit photoacoustic signals that can be detected by the detector. When the generated CO₂ decreases, the photoacoustic signal slowly decreases accordingly. The likely cause of the change in the photoacoustic signals of ABN over time is the fact that under heat, NH₄HCO₃ decomposes into CO₂, which is inside the liposomal vesicles, and continuously diffuses out of the vesicles and gradually decreases. Conversely, there were no membranes to encapsulate the NH₄HCO₃ solution, which was not enveloped in liposome. The decomposed CO₂ was entirely released quickly, producing a photoacoustic signal in a short period of time.

In addition, we investigated the changes in the photoacoustic signals of ABN at different concentrations of ABN (~2.63×10⁷/mL) in vitro. For serial 2-fold diluted ABN samples irradiated with the PAI system, we found that the photoacoustic signal intensity decreased corresponding with a decreased concentration. When the concentrations were higher, the photoacoustic signal was stronger; and when the concentrations were lower, the photoacoustic signal was weaker. The above-described phenomenon is presumably related to the ABN volume. As the ABN volume decreased, the generated CO₂ and the photoacoustic signals also decreased. Furthermore, we investigated the changes in the photoacoustic signals of ABN at different temperatures (25°C, 37°C, and 42°C) in vitro. At ambient temperature...
Figure 18: Hematoxylin and eosin-stained subcutaneous tumor tissue. There was no marked difference in the cell morphology, cell nucleus, and cytomembrane. There was no obvious necrosis, and the structure of tumor tissue was not damaged.

Notes: (A) Laser-irradiated skin (400-fold). (B) Laser-irradiated tumor (400-fold). (C) Laser non-irradiated skin (400-fold). (D) Laser non-irradiated tumor (400-fold).

(25°C), photoacoustic signal intensity of the sample was the weakest, and at 37°C, it increased. At 42°C, the photoacoustic signal intensity of the sample was not significantly different from that at 37°C, suggesting that the generation of photoacoustic signals of the ABN is related to temperature. Most likely, NH$_4$HCO$_3$ tends to decompose at elevated temperatures. The more CO$_2$ generated, the greater and stronger the photoacoustic signal. The temperature of the human body is 37°C, and at this temperature, there were stronger photoacoustic signals recorded, which provided theoretical basis for further tests in vivo.

During in vivo testing, 1.0 mL of ABN was injected through the caudal vein into tumor-bearing mice. Photoacoustic signals from subcutaneous tumors began to appear after 30 min and gradually increased to reach their peak after 4 h, when the scope of the photoacoustic signals was the largest. The signals then gradually decreased and completely disappeared after 24 h. These results indicate that the intravenously injected ABN could enter the local tumor and emit photoacoustic signals. The skin of the tumor area showed no damage and appeared intact when observed by the naked eye. Pathological examination also showed that the structure of the skin and tissue cells around the tumor were intact without obvious damage, indicating that laser energy was safe and did not cause damage to the skin or subcutaneous tumor tissues and cells.

This study used a PAI system to conduct feasible, effective, and safe in vitro and in vivo tests of liposome-derived, gas-generating nanoparticles, and elaborated the mechanism of generating photoacoustic signals. However, the following limitations remain in this study: 1) The stability of ABN is poor, 2) the potential toxicity of ABN to normal tissue or cells was not studied, and 3) there was no direct method to prove that NH$_4$HCO$_3$ was wrapped in liposomes. Therefore, further experimental studies need to be conducted to improve the stability and safety of ABN and to verify NH$_4$HCO$_3$ that was wrapped in liposomes.

Conclusion

In summary, we have succeeded in preparing a biodegradable material (NH$_4$HCO$_3$ nanoparticles) as a PAI contrast agent whose metabolic products can be completely eliminated from a living organism. The mechanism involved may be CO$_2$, which swells under specific optical laser wavelengths. This material, which is a liposome-encapsulating NH$_4$HCO$_3$ solution, is feasible, effective, safe, and inexpensive. Therefore, NH$_4$HCO$_3$ nanoparticles are a promising material to be used in clinical PAI for breast or superficial cancers.
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Disclosure
The authors report no conflicts of interest in this work.

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