Ahmed glaucoma valve implant: surgical technique and complications

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Abstract: Implantation of Ahmed glaucoma valve is an effective surgical technique to reduce intraocular pressure in patients affected with glaucoma. While in the past, the use of this device was reserved to glaucoma refractory to multiple filtration surgical procedures, up-to-date mounting experience has encouraged its use also as a primary surgery for selected cases. Implantation of Ahmed glaucoma valve can be challenging for the surgeon, especially in patients who already underwent previous multiple surgeries. Several tips have to be acquired by the surgeon, and a long learning curve is always needed. Although the valve mechanism embedded in the Ahmed glaucoma valve decreases the risk of postoperative hypotony-related complications, it does not avoid the need of a careful follow-up. Complications related to this type of surgery include early and late postoperative hypotony, excessive capsule fibrosis around the plate, erosion of the tube or plate edge, and very rarely infection. The aim of this review is to describe surgical technique for Ahmed glaucoma valve implantation and to report related complications.

Keywords: glaucoma, surgical technique, glaucoma drainage devices, Ahmed glaucoma valve, complications

Introduction

Glaucoma, the second leading cause of blindness in the world, is defined as a chronic optic neuropathy, linked to progressive visual field defects. In 2013, the total number of people (aged 40–80 years) with glaucoma worldwide was estimated to be 64.3 million.1 Asia alone accounted for ~60% of the world’s total glaucoma cases (39 million), and Africa had the second highest number of cases with 8.3 million (13%). Europe and North America had 6.77 and 3.36 million cases, respectively.1

Although many risk factors have been described for glaucoma development and progression (intraocular pressure [IOP], age, genetic predisposition, and vascular parameters), lowering IOP is the only scientifically demonstrated method to slow the progression of the disease. The Early Manifest Glaucoma Trial has demonstrated a 10% reduction of the risk of glaucoma progression with each mmHg of IOP decrease from baseline (hazard ratio [HR]=0.90 per mmHg decrease; 95% confidence interval [CI], 0.86–0.94).2 Similarly, in the UK Glaucoma Treatment Study, a 19% decrease of the risk of visual field progression has been found with each mmHg reduction from baseline.10

IOP reduction in glaucoma patients can be achieved with medical, laser, or surgical therapy.11 Medical therapy is generally the first step, due to the low rate of side effects. However, when target IOP is not achievable, laser and/or surgery should be chosen.12 Trabeculectomy, which was first described by Cairns in 1968,13 is considered the surgical “gold standard” for primary open-angle glaucoma and primary angle-closure glaucoma.
Variable success rates for trabeculectomy have been reported in literature. Although success rates are high in the first few years after surgery (70%-92%),
14–16
they tend to decrease with time (42%-90%),
17–20
especially in secondary glaucomas.21–24
Moreover, as trabeculectomy involves anterior subconjunctival space, it is largely influenced by healing processes, and healing modulation with intraoperative and postoperative antimetabolites (ie, mitomycin C [MMC] and 5-fluorouracil [5FU]) is a fundamental part of the procedure.

Aqueous shunts are a reliable alternative to trabeculectomy.25 Conceptually, shunting aqueous humor (AH) to the posterior subconjunctival space may avoid healing issues, especially in patients who have already undergone previous glaucoma surgeries or conjunctival manipulation. The first successful prototype of aqueous shunt was the Molteno implant, followed by Krupin, Ahmed, and Baerveldt implant. While in the past, these devices were usually reserved as a second choice surgery, today mounting experience has encouraged their use also as a primary surgery.26 The aim of this review is to evaluate surgical technique and complications of Ahmed glaucoma valve (AGV) implant.

AGV: device description and technical data
First attempts at developing a glaucoma drainage implant were published in 1906,27 even if the first device currently in use was developed by Molteno et al only in 1976.28 Molteno device offers no resistance to AH outflow and initially was burdened by a high rate of complications, such as hypotony, shallow anterior chamber (AC), choroidal effusion, and choroidal detachment.26 Implants with an embedded valve mechanism were developed as an attempt at overwhelming these complications. In 1976, Krupin designed a pressure-sensitive unidirectional valve to provide filtration restriction.29 Its passive mechanism contemplates a silastic tube, whose distal end is sealed and contains several horizontal and vertical slits. Krupin implant is designed to open when IOP is >11 mmHg.29,30

AGV provides a more complex mechanism to control AH outflow. It was developed by Mateen Ahmed and was approved by the Food and Drug Administration in 1993.31 It consists of 3 parts (Figure 1A): 1) a plate, in medical grade silicone, polypropylene, or porous polyethylene, depending on the model; 2) a drainage tube in medical grade silicone; and 3) a valve mechanism in medical grade silicone. Polypropylene is a rigid plastic, not flexible and highly resistant to torsional forces, whereas silicone is a flexible rubber.

The adult model (S2) of AGV provides 180 mm² of total plate area, whereas the pediatric one (S3) has a total area of 96 mm². Obviously, a smaller plate facilitates positioning in infants and subjects with a small eye. A variant of the device with 2 plates (total filtration area: 360 mm²), and one with a clip for pars plana tube insertion have been also designed (Table 1).

The M4 AGV model, recently introduced, is a modified AGV S2 containing identical valve mechanism, but with a case made of porous high-density polyethylene (Medpor; Porex, Atlanta, GA, USA; subsequently, Stryker Corp., Kalamazoo, MI, USA).32 Total plate area is 160 mm², not including the surface area of pores. The pores may facilitate fibrotic and vascular ingrowth and increase resistance to infection, exposure, extrusion, and mechanical deformation.32–34 In animal models, this new implant behaved as a variable resistor with higher resistance at low flow rates and lower resistance at high flow rates.35 These results support the idea that porous biomaterial may improve hydraulic conductivity of the capsule.

![Figure 1](https://www.dovepress.com/)

**Figure 1** Ahmed glaucoma valve implant.

**Notes:** (A) Ahmed glaucoma valve implant components; (B) Ahmed glaucoma valve mechanism. $V_A$ presented as velocity of fluid flowing in section A. $V_B$ presented as velocity of fluid flowing in section B.
Valve mechanism of AGV consists of thin silicone elastomer membranes, which are 8 mm long and 7 mm wide and create a Venturi-shaped chamber. The membranes are pretensioned to open and close in response to IOP variations, in the range of 8–12 mmHg. After implantation, AH flows slowly and continuously into the trapezoidal chamber of the valve (Figure 1B). As the pressure reaches the preset threshold value, the valve opens, thus decreasing the IOP. As the inlet cross-section of the chamber is wider than the outlet, a pressure differential is created across the chamber. This pressure differential enables the valve to remain open even with a small pressure differential between the AC and the subconjunctival spaces surrounding the device. In order for Bernoulli’s equation to be satisfied (fluid flowing into section A = fluid flowing out of section B, Figure 1B), the fluid’s velocity has to increase as it leaves the chamber through the drainage tube. This increased velocity and the nonobtrusive flow account for better evacuation and smaller valve friction. The tension in the silicone membranes helps to reduce hypotony by closing after the pressure has decreased to normal level again.

**Surgical technique**

AGV is designed to guarantee no manipulation during the surgical procedure (ie, no restrictive methods to limit AH filtration). The implant should be examined for integrity and primed before implantation. Priming is performed by using a 26G cannula, injecting ~1 cc of balanced salt solution (BSS) or sterile water through the drainage tube. Functionality of the implant is demonstrated by BSS flow through the plate.31

Surgical technique for AGV implantation consists of a fornix-based or limbal-based conjunctival incision to create a conjunctival flap between 2 recti muscles, generally in the superotemporal quadrant. As much as possible, Tenon’s capsule is dissected from the episclera and episcleral vessels are gently cauterized. The technique does not require recti muscles isolation.

Body implant is positioned 8–10 mm from the limbus, outside limbal healing space. The plate is then sutured to the sclera with a 9.0 or 10.0 nylon suture. The drainage tube is trimmed to permit a 2–3 mm insertion in the AC and is bevel cut to an angle of 30°, to facilitate AC entering. An AC paracentesis is performed, and viscoelastic substance is injected to increase spaces. The AC is then entered 1–3 mm posteriorly to the corneoscleral limbus with a 22–23G needle. The needle tract is anterior and parallel to the plane of the iris. The tube, which is trimmed so that the bevel faces to the corneal endothelial surface, is inserted into the AC through the needle tract. Care must be taken at this point to ensure that the drainage tube does not contact iris or corneal endothelium after insertion.

An “hangback” technique has been recently proposed to reduce tissue manipulation and facilitate plate insertion during AGV implantation.36 According to this technique, the plate is not sutured to the sclera and is allowed to “hang” from the tube, which is anchored to the sclera 6–7 mm from the limbus, using a 6.0 vicryl suture. Despite initial results for this technique are promising, more extensive research is needed.36

In patients with a previous vitrectomy, implant’s tube can be inserted in vitreous cavity, avoiding complications that can arise from the presence of the tube in the AC. In this case, a pars plana clip is used to secure the tube to the sclera and to give the tube a suitable angle. The clip is anchored to the sclera with a 9.0 or 10.0 nylon suture.

The drainage tube and eventually the pars plana clip are covered with a piece of preserved, donor sclera, pericardium, cornea, or other suitable patch graft material, which is sutured to the sclera. Alternatively, a two-third thickness limbus scleral flap is created, and the tube is inserted into the AC through a needle tract under the flap. The flap should be closely sutured to avoid peritubular leakage of AH.

In the final step, conjunctiva is anchored to the limbus with adsorbable/nonadsorbable sutures. Optionally, a subconjunctival injection of corticosteroids and antibiotics is performed at the end of the procedure. Postoperatively, a regimen of topical corticosteroids and antibiotics is introduced.

A sutureless variant for tube covering has been recently proposed. The tube is covered with human donor scleral graft and kept in place with fibrin glue (Tissue Coll®).37 Tissue Coll is a biologic glue with adhesive properties derived from the formation of fibrin polymers. However, costs, commercial availability, poor uniformity, and quality of donor sclera may not be easily accessible.

### Table 1 Ahmed glaucoma valve implant available models

<table>
<thead>
<tr>
<th>Type</th>
<th>Model</th>
<th>Size (mm²)</th>
<th>Material (plate)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Single plate</td>
<td>S2</td>
<td>184</td>
<td>Polypropylene</td>
</tr>
<tr>
<td>Pediatric size</td>
<td>S3</td>
<td>96</td>
<td>Polypropylene</td>
</tr>
<tr>
<td>Double plate</td>
<td>B1</td>
<td>364</td>
<td>Polypropylene</td>
</tr>
<tr>
<td>Single plate</td>
<td>FP7</td>
<td>184</td>
<td>Silicone</td>
</tr>
<tr>
<td>Pediatric size</td>
<td>FP8</td>
<td>96</td>
<td>Silicone</td>
</tr>
<tr>
<td>Double plate</td>
<td>FX1</td>
<td>364</td>
<td>Silicone</td>
</tr>
<tr>
<td>Single plate</td>
<td>M4</td>
<td>160</td>
<td>Polyethylene</td>
</tr>
<tr>
<td>Pars plana</td>
<td>PS2</td>
<td>184</td>
<td>Polypropylene</td>
</tr>
<tr>
<td>Pars plana (pediatric)</td>
<td>PS3</td>
<td>96</td>
<td>Polypropylene</td>
</tr>
<tr>
<td>Pars plana</td>
<td>PC7</td>
<td>184</td>
<td>Silicone</td>
</tr>
<tr>
<td>Pars plana (pediatric)</td>
<td>PC8</td>
<td>96</td>
<td>Silicone</td>
</tr>
</tbody>
</table>
limit the widespread use of this procedure. The sutureless technique has been further modified with the use of bovine pericardial graft patch (Tuttopatch, Tutogen Medical GmbH, Neunkirken am Brand, Germany) instead of human sclera, with good results in the medium term (Figure 2).38 The relative high costs of fibrin glue are in this way counterbalanced by the low cost of Tuttopatch, when compared with human scleral donor patch. The absence of sutures is advantageous in the postoperative, because sutures can promote inflammation and provide a nidus for infections or neovascularization. Written informed consent was received from the patients for this review including publication of their medical data and images.

Antimetabolite and Ahmed valve surgery

The use of antimetabolites in AGV surgery is debated. In 2004, Costa et al randomized 60 patients affected by neovascular glaucoma to receive intraoperative MMC (0.5 mg/mL for 5 minutes; n=34) or BSS (n=26) during surgery for AGV implantation.39 Kaplan–Meier survival analysis showed a 59% and 61% probability of success at 18 months for the MMC and control groups, respectively, with no statistically significant difference. Similar results were obtained by Kurnaz et al in a study on 48 patients affected with refractory glaucoma.40 Authors found no difference in success rates between the MMC and no-antimetabolite groups at 1 year (86.36% versus 80.76%, respectively), but 3 cases of tube exposure were encountered in the MMC group.

Alvarado et al obtained high success rates at 6-year follow-up in patients who underwent AGV implantation (either alone or in combination with cataract surgery) with both intraoperative MMC and postoperative 5FU injections.41 MMC-soaked sponges were placed subconjunctivally for ~5–8 minutes during surgery (median: 8 minutes, MMC concentration: 0.5 mg/mL). 5FU subconjunctival injections were performed postoperatively, for 4 consecutive weeks, starting at 1 week, with a fifth injection at week 6. Kaplan–Meier estimates of the cumulative probability of implant success at the sixth follow-up year were 0.72 and 0.84 for eyes that underwent AGV implantation (n=88) and AGV implantation + cataract extraction (n=42), respectively. Comparing results at 2- and 4-year follow-up with those from other studies in which antimetabolites were not used, authors concluded that there was a potential benefit associated with the use of MMC and 5FU during the intraoperative and early postoperative period.

The way antimetabolites are administered could be important in determining the efficacy of the surgical procedure. Zhou et al proposed a new technique for MMC administration, in which the valve plate was first encompassed with a thin layer of cotton soaked with MMC and then positioned on the sclera.42 After 2–5 minutes, the cotton and the encompassed AGV were removed and irrigated with 200 mL BSS. In comparison with the traditional MMC administration procedure (ie, sponges soaked with MMC and applied on the sclera), the new technique obtained better results in the short and long terms. The new-technique group had only 1 case of encapsulated cyst over the plate out of 38 surgeries (2.6%), whereas the traditional-technique group had 8 cases of encapsulated cyst out of 41 surgeries (19.5%) ($P=0.030$). However, it should be noted that MMC concentrations in this study were considerably smaller than in other studies (0.25–0.33 mg/mL, 2–5 minutes), and this could have biased the results.

AGV complications

A report from the American Academy of Ophthalmology has reported the major short-term (up to 5 years after surgery)
to medium-term (5–10 years after surgery) complications of aqueous shunt devices. They include immediate and late hypotony after surgery, excessive capsule fibrosis and clinical failure, erosion of the tube or plate edge, and very rarely infection.

Actually, there is no evidence in literature about different rates of complications with 1 AGV model than another. Although an higher IOP reduction with the silicone-plate model than the polyethylene one has been described in the short term, long-term results are not conclusive.

Hypotony
AGV valve mechanism was designed with the aim of preventing postoperative hypotony, allowing for AH drainage when IOP is in the range of 8–12 mmHg. Studies have demonstrated that the mechanism is effective in reducing, but not abolishing, postoperative hypotony, in comparison with other nonvalve implants. The Ahmed versus Baerveldt Study (AVB Study) was a prospective, multicenter, randomized clinical trial comparing AGV (model FP7) and Baerveldt 350 mm² implant in patients affected by refractory glaucoma. In this study, 7 of 114 patients (6.1%) in the Baerveldt group experienced vision-threatening complications related to hypotony at 3-year analysis (3 developed suprachoroidal hemorrhage, 3 had retinal/choroidal detachments, and 1 had refractory hypotony), against 0 of 124 AGV patients. Similarly, in the Ahmed–Baerveldt Comparison (ABC) study, a multicenter clinical trial designed to prospectively compare safety and efficacy of these two commonly implanted devices, 1 eye (2%) in the AGV group experienced failure due to persistent hypotony, against 6 eyes (13%) in the Baerveldt group after 5-year follow-up. In comparison with Molteno implant (both single and double plates), AGV demonstrated lower risk of hypotony-related complications, both in retrospective and prospective studies. Besides these results, postoperative hypotony following the placement of AGV (S2 and FP7) has been reported up to 3% of cases.

The reason of persistent hypotony after AGV implantation is not completely clear. Attention should be taken during surgical procedure to not over-prime the tube and to not excessively manipulate the valve housing, as these actions could damage the valve mechanism embedded in the implant. Importance has been placed on the utilization of a 22 or 23G needle when creation of the sclerostomy is undertaken in order to avoid the egress of AH around the silicone tube in the immediate post-operatory. In addition, ciliary body function may fail or decrease after surgery in complicated eyes in which glaucoma drainage implants are used.

In a study by Prata et al, pressure flow characteristics at physiologic flow rates, in vitro and in vivo rabbits, were evaluated for valve (Ahmed and Krupin) and nonvalve (Baerveldt and Molteno) implants. Authors concluded that none of the implants tested maintained advertised pressure levels during in vitro tests, when immersed and while being perfused at flow rates close to those expected in normal human eyes. AGV and Krupin implants functioned as flow restriction devices or regulators, rather than as valves that truly open and close in response to pressure change after immersion in fluid. In vivo, conjunctival tissue reaction surrounding the explant portion of the device contributed significantly to the outflow resistance, increasing the restriction effect due to the valve mechanism.

Besides persistent and long-term hypotony, transient hypotony in the immediate postoperative has been frequently described after AGV implantation. An “hypotensive” phase was recorded in 13% and 15% of patients in the AVB and ABC studies, respectively. Choroidal effusion may also be present. In these cases, hypotony resolves spontaneously as soon as encapsulation of the plate increases outflow resistance, within days or weeks from the surgery. Observation should be the choice, monitoring AC depth and extension of choroidal detachment. Prompt intervention should be taken if shallow AC, hypotony maculopathy, or near-kissing choroidal detachment is present.

IOP increase and excessive capsule fibrosis
An “hypertensive” phase after glaucoma drainage implantation is quite common and has been frequently described in patients with AGV. Typically, this phase peaks at 1 or 2 months postoperatively and resolves within 6 months. The hypertensive phase could be less frequent in patients who have been implanted with the silicone than with the polypropylene AGV, probably because silicone is less inflammatory than polypropylene. The primary reason for elevated IOP in the postoperative period is from capsular fibrosis. Attempts have been made to modulate the fibrotic reaction around the plate, varying plate size, shape, flexibility, and materials. Initial data showed that a mitigation of the early postoperative hypertensive phase may be achievable with the new M4 AGV model, in comparison with the FP7 and S2 model.

An option in the management of the hypertensive phase is, similar to trabeculectomy, digital massage. The purpose of digital massage is to force AH through the tube, opening the valve mechanism, and reducing scar formation. Caution should be placed in this maneuver in order to avoid repeated tube-corneal endothelial touch.
Late IOP increase (>6 months) is the main cause of long-term failure of AGV surgery. If encapsulation of the plate is evident, a needling revision of the bleb may be attempted, with the aim of reducing outflow resistance. An encapsulated bleb is recognizable as a bleb that has been walled-off by the Tenon’s capsule, resulting in a rigid elevation of the mobile conjunctival tissue. Quaranta et al performed bleb needling with 5FU at the slit lamp in 36 consecutive patients implanted with an S2 AGV, noncontrolled IOP, and encapsulated bleb over the plate.67 Qualified success (IOP ≤18 mmHg with or without medications) was achieved at 1, 3, 6, 12, 18, and 24 months in 100%, 97.8%, 86.1%, 75%, 75%, and 72.2%, respectively. Complications were encountered in 14 eyes (38.8%) and resolved spontaneously in 12 out of 14 cases.

If medical therapy and needling revision are not successful, surgical revision of the implant should be performed. Conjunctiva is dissected over the encapsulated bleb, and the cyst wall is excised. After excision, the conjunctiva is closed with a nylon or vicryl suture. In a retrospective study by Eibschitz-Tsimhoni et al, surgical revision was effective in achieving adequate IOP control in 8 of 11 patients, with or without medications.68 However, in 3 patients, cyst excision was not successful, and further surgical interventions were needed.

**Tube exposure**

Tube exposure is a well-known complication of glaucoma drainage implants (Figure 3). Erosion of the conjunctiva and of the covering patch graft has been described in the late postoperative period in 2%–7% of eyes after implantation of glaucoma devices.62,69–74 With regard to AGV, frequency of tube exposure varies from 5% to 14.3% of cases.83,75,76 In a recently published study on 12 patients, tube exposure has been encountered in up to 30.8% of cases.77 However, these high frequencies are not in accordance with data deriving from multicenter studies.

![Figure 3 Ahmed glaucoma valve: tube exposure due to conjunctival erosion.](image)

Tube exposure can lead to ocular inflammation, hypotony, poor vision, and ptosis. Most of all, tube exposure represents a major risk factor for the development of late endophthalmitis, as the exposed tube provides a way for microorganisms to migrate into the eye from ocular surface and conjunctiva.78,79 Although covering the tube with a patch graft material has markedly reduced the rate of tube exposure,80 no significant difference in patch graft survival has been found when sclera, dura, and pericardium were compared.73

The mechanism responsible for tube exposure is not completely clear. A high grade, immune-mediated process could be responsible for rapid melting (<6 months) of the patch, via cell- or noncell immune-mediated process.73 A mechanical process could be involved in patch erosion as well. If tube is not fixed on the sclera, continuous and minimum movements may produce tube-graft tension, resulting in gradual patch atrophy.73,81 Finally, patch melting could occur as a result of a low grade, possibly immune-mediated, long-term, atrophy process, with consequent gradual patch thinning.73

Chaku et al evaluated risk factors for tube exposure in a comparative, retrospective study of 64 eyes, including 32 eyes with tube exposure and 32 control eyes.82 All patients had a glaucoma drainage device implanted, including 35 AGVs and 29 Baerveldt implants. Patients developed tube exposure at a mean of 1.43±1.5 years from surgery, and no difference was found in the rate of exposure between AGV and Baerveldt implant. In both univariate and multivariate analyses, younger age (P<0.01 and P=0.02) and inflammation prior to tube exposure (P<0.01) were significant risk factors for tube exposure. Diabetes was a significant factor only in the univariate analysis (P=0.02).

In a meta-analysis by Stewart et al, 38 previously published studies describing conjunctival erosion in patients with a glaucoma device (16 AGVs, 12 Baerveldt, and 17 Molteno implants) were evaluated.83 A total of 3,105 patients and 3,255 eyes with an average follow-up of 26.1±3.3 months were included in the analysis. The incidence of tube exposure from these studies was 2.0±2.6% (n=64), with an average exposure rate per month of 0.09±0.14%. No difference was found among AGV, Baerveldt, and Molteno implant. Although the correlation between study length and incidence of exposure was not significant, there appeared to be a little increase in exposure incidence for studies up to 2-year follow-up.

Implant positioning may be important in determining the risk of tube exposure. In a study by Pakravan et al, 58 eyes underwent AGV implantation in the superotemporal
Corneal complications

The presence of the silicone tube in the AC is known to disturb corneal endothelium and may induce corneal decompensation and edema. The exact frequency of corneal issues in patients implanted with AGV is not known, but it has been reported to be 9%–27% in the long term.

Topouzis et al, after a mean follow-up of 30.5 months, found a 27% incidence of corneal decompensation in patients who underwent AGV implantation. However, in this study, 16 of the 60 eyes enrolled had prior or concurrent corneal grafts and 9 of these 16 eyes had corneal graft failure during the follow-up. In the ABC study, persistent corneal edema was recorded in 20.1% of patients in the AGV group, at 5 years from surgery. However, edema was attributed to nonimplant causes in 50% of cases, so that the real percentage of subjects with persistent corneal edema due to the implant was 12%. No difference was found between AGV and Baerveldt implant in terms of persistent corneal edema incidence (20.1% and 20.4%, respectively). In the AVB study, corneal edema affected 7% of patients in the AGV group and 14% of patients in the Baerveldt group at 3-year follow-up (P=0.08). Authors hypothesized that the higher rate of corneal edema in the Baerveldt group was the result of greater IOP variability in the early postoperative phase.

The exact mechanism causing endothelial damage in patients with a drainage implant is unknown. Jet flow around the end of the tube caused by heartbeat, AC inflammation, intermittent tube-corneal touch, tube-uvex touch, and foreign body reaction to the silicon tube are all potential mechanisms of endothelial damage in these patients. However, factors independent of tube existence in the AC may contribute to corneal decompensation. High IOP and long duration of elevated IOP before surgery, toxicity of eye-drop preservatives, duration of surgery, and changes in the composition of AH may influence endothelial cell functions and take a part in corneal decompensation.

Endothelial cell loss in patients implanted with AGV has been prospectively and retrospectively evaluated. Kim et al found a mean percentage decrease in corneal endothelial cell density of 3.5% at 1 month, 7.6% at 6 months, and 10.5% at 12 months from surgery. Lee et al recorded an average decrease in corneal endothelial cell count of 5.8% at 1 month, 11.5% at 6 months, 15.3% at 12 months, 16.6% at 18 months, and 18.6% at 24 months from surgery. In both studies, the superotemporal area, which was closest to the site of the tube, showed the greatest decrease in endothelial cell count, whereas the central area showed the smallest decrease. Besides these data, in the study by Lee et al, Kaplan–Meier analysis showed an alarming 36.6% cumulative rate of failure (ie, patients with a decrease in central corneal endothelial cell density >20%) at 24 months.

In a recent study, medical records of 127 patients who underwent AGV implantation were retrospectively reviewed to evaluate changes in corneal endothelial cell density and rates of corneal decompensation. Mean follow-up was 43.1±20.5 months, with 53% of patients followed-up to 60 months. Corneal decompensation occurred only in 3 cases and the cumulative risk of corneal decompensation was 3.3% at 5 years from surgery. Seventy-two eyes were evaluated for annual change in endothelial cell count and compared with 31 control eyes. Although a more rapid loss of endothelial cell count was found in these 72 eyes compared with controls (−7.0% and −0.1%/year, respectively; P<0.01), the rate of loss decreased over...
time and statistical significance compared with control eyes disappeared after 2 years postoperatively.

Apart from corneal decompensation due to direct tube-corneal touch, tube position could be important in determining corneal complications. Koo et al measured various AH parameters in 39 eyes with previous superotemporal AGV implantation. Parameters measured included tube distance from the cornea, tube angle, and AC tube length. All parameters were obtained by anterior segment optical coherent tomography. In univariate analysis, tube-cornea angle and the closest distance from the tip of the tube to the cornea were statistically significant predictors of superotemporal corneal endothelial cell loss after glaucoma surgery. In multivariate analysis, only the distance from the tip of the tube to the cornea was significant; each millimeter that the tube was closer to the endothelial surface was associated with 353.1 (95% CI, 56.1–650.1; P=0.02) fewer endothelial cells.

In patients with corneal decompensation and AGV, Descemet stripping endothelial keratoplasty (DSAEK) has been attempted, with survival rates at 1 year similar to full-thickness keratoplasty. Surgical procedure can be challenging in these cases. The length and location of the glaucoma drainage tube within the AC may need to be modified for DSAEK to be successful, and eventually the glaucoma implant posteriorly repositioned. Moreover, air management is often difficult because injected air may escape through the implant to the subconjunctival space, thereby making it difficult to obtain a firm intraoperative fill. This is consistent with a potential high rate of early postoperative graft detachment. Finally, a higher rate of endothelial cell loss has been described in eyes with AGV and previous DSAEK, compared with nonglaucomatous eyes. In a study by Kim et al, mean reduction in graft endothelial cell count was 69% over a 20-month follow-up period, whereas Schoenberg et al found an average endothelial cell loss of 40.7% at 12 months from surgery.

Infection and endophthalmitis

Endophthalmitis or infections associated with grafting material covering the tube is a rare complication of glaucoma drainage implants. For this reason, recurrent blebitis after trabeculectomy can be a reasonable indication for shunt implantation, according to a report by the American Academy of Ophthalmology. Several retrospective studies about glaucoma drainage implants included few cases of endophthalmitis, resulting in rates ranging from 0.8% to 6.3% (mean: 2.0%). There appears to be no significant difference in reported rates of endophthalmitis among various glaucoma drainage implants.

In a retrospective study, Al-Torbak et al reviewed records of patients implanted with AGV from 1994 to 2003 in a single eye hospital in Saudi Arabia and found an incidence of endophthalmitis of 1.7% (9 patients on 542 implanted). Five cases of endophthalmitis were detected in pediatric patients (ie, age <18 years) and 4 in adults. Median interval between AGV implantation and diagnosis of endophthalmitis was 260 days and delayed onset endophthalmitis developed in 8 of 9 eyes (88.8%). Conjunctival erosion over the tube was found in 6 of 9 patients (66.6%) who developed endophthalmitis and 4 of 6 eyes (66.6%) had a Seidel-positive leak. Multiple regression analysis revealed younger age (<18 years; P<0.05) and conjunctival erosion over the tube (P<0.01) as significant risk factors for endophthalmitis development.

Conjunctival erosion and tube exposure appear to be a major risk factor for infection development in eyes with glaucoma drainage implants. Several case reports in literature describe episodes of late endophthalmitis in patients with AGV and tube exposure. It has been postulated that the exposed tube may act as a direct channel for the intraocular passage of the conjunctival flora from the ocular surface. Younger age is another major risk factor. Case series of pediatric patients implanted with glaucoma drainage devices have shown an incidence of endophthalmitis ranging from 2.9% to 5%. This could be related to the higher rate of conjunctival erosion and implant exposure in children. Interestingly, younger age has been advocated as a significant risk factor for endophthalmitis also in patients who underwent trabeculectomy, with an incidence of late bleb-related endophthalmitis as high as 8.3% in the pediatric population.

Diplopia and strabismus

Diplopia and strabismus are well-known postsurgical complications of glaucoma drainage devices. The cause of diplopia is likely a restrictive strabismus, either from the plate itself or from the plate impinging on the muscle insertion. Manipulation of the rectus muscles during surgery may induce strabismus as well, which usually resolves spontaneously in weeks or months. A systematic review of literature by Hong et al found an higher incidence of diplopia in patients with Baerveldt implant compared with patients with other glaucoma devices. However, the ABC study found an equal, ~12% cumulative risk of persistent diplopia in both the AGV and Baerveldt groups. Authors suggested that the end-plate fenestrations of the new designed Baerveldt implant (applied in the ABC study) could reduce the height of the bleb and consequently minimize restrictive strabismus. For this reason, an equal risk of diplopia may have been found in both groups of the ABC study.
In a retrospective study, including 159 eyes implanted with AGV, Huang et al found diplopia in 4 patients (2.5%), 3 requiring extraocular muscle surgery and 1 who had removal of the device.63 However, Ayala et al reported an incidence of transient diplopia of 4.7% (4 of 85) in patients who underwent AGV surgery.64 Fifty percent cases of diplopia occurred within 3 months of surgery.

**Conclusion**

AGV implantation is an effective and relatively safe surgical procedure, which allows to manage particular phenotypes of glaucoma (ie, secondary glaucoma) and glaucoma refractory to previous filtration surgeries (ie, second choice surgery). Despite an apparently “user-friendly” technique, many surgical tips are to be acquired by the surgeon, and a long learning curve is always needed. In comparison with other nonvalve glaucoma drainage devices, AGV has the great advantage of an easier postoperative management. Nevertheless, early postoperative hypotony is still a dangerous complication that can affect this type of surgery.

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