Magnetic and fluorescent \( \text{Gd}_2\text{O}_3 : \text{Yb}^{3+}/\text{Ln}^{3+} \) nanoparticles for simultaneous upconversion luminescence/MR dual modal imaging and NIR-induced photodynamic therapy

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Abstract: The development of upconversion nanoparticles (UCNs) for theranostics application is a new strategy toward the accurate diagnosis and efficient treatment of cancer. Here, magnetic and fluorescent lanthanide-doped gadolinium oxide \( \text{Gd}_2\text{O}_3 \) UCNs with bright upconversion luminescence (UCL) and high longitudinal relaxivity \( (r_2) \) are used for simultaneous magnetic resonance imaging (MRI)/UCL dual-modal imaging and photodynamic therapy (PDT). In vitro and in vivo MRI studies show that these products can serve as good MRI contrast agents. The bright upconversion luminescence of the products allows their use as fluorescence nanoprobes for live cells imaging. We also utilized the luminescence-emission capability of the UCNs for the activation of a photosensitizer to achieve significant PDT results. To the best of our knowledge, this study is the first use of lanthanide-doped \( \text{Gd}_2\text{O}_3 \) UCNs in a theranostics application. This investigation provides a useful platform for the development of \( \text{Gd}_2\text{O}_3 \)-based UCNs for clinical diagnosis, treatment, and imaging-guided therapy of cancer.

Keywords: upconversion nanoparticles, upconversion luminescence imaging, MR imaging, photodynamic therapy, singlet oxygen

Introduction

Cancer is still one of the most devastating human diseases, causing millions of deaths every year. Accurate diagnosis and efficient treatment of cancer are crucially important to increase the survival rate of patients. However, current diagnostic and therapeutic techniques are still far from satisfaction. Photodynamic therapy (PDT) is considered to be an effective technique for cancer treatment because it is cost-effective, highly localized, and has fewer side effects compared to radiation therapy and chemotherapy.1–3 It involves local or systemic administration of a photosensitizer (PS), followed by irradiation of the target lesion with light of a specific wavelength. This triggers oxidative photodamage by the generation of reactive oxygen species, especially singlet oxygen, subsequently leading to tumor cell killing.4 However, most of the currently available PSs are activated by visible light (400–700 nm), which has a limited penetration depth in biological tissues. In contrast, the near-infrared (NIR) within the “optical transparency window” (700–1,100 nm) of biological tissues not only results in low photodamage but also possesses high tissue penetration capability.5–7 A novel strategy of combining the PS with lanthanide-doped upconversion nanoparticles (UCNs) can overcome the previous limitations and has recently attracted interest in the field of malignant tumor
therapy. In this novel system, UCNs activate the PS by visible light emission generated from sequential multiphoton NIR excitation via an anti-Stokes process.8–10

In 2007, Zhang et al first reported the use of NaYF₃:Yb³⁺/Er³⁺ UCNs for PDT application.11 Since then, the UCNs have been used for PDT by many groups.12–14 Both the dopants and host of UCNs are important to achieve efficient PDT. There are two types of dopants ions in UCNs: a sensitizer to absorb the NIR light and an activator to emit photons and further activate the PS. The Yb³⁺ ion is the most commonly used sensitizer ion because its 2F⁵/₂ → 2F⁷/₂ transition has a large absorption cross-section around 980 nm. The Er³⁺ ion is a highly efficient activator because many of its f-f transitions resonant well with the 2F⁵/₂ → 2F⁷/₂ transition of Yb³⁺. For effective PDT, the choice of the UCNs host is critical for obtaining good upconversion efficiency. A popular host material is NaYF₃, which exhibits high upconversion efficiency due to the low phonon cutoff energy,15 thus attracting much interest in UCNs-based PDT and upconversion luminescence (UCL) imaging.16–19 Other fluorides containing Yb³⁺ (NaYbF₄) and Gd³⁺ (NaGdF₄) ions, which also function as computed tomography (CT) and magnetic resonance imaging (MRI) contrast agents,20–24 have been utilized to achieve simultaneous dual- or multimodal bioimaging with high sensitivity and high spatial resolution. These multifunctional nanoparticles can satisfy the high efficiency and accuracy requirements of clinical cancer theranostics.

Oxides, such as Gd₂O₃, ZrO₂, and Y₂O₃, are also generally used as luminescence host materials owing to their desirable chemical durability, thermal stability, and lower phonon energy.25–27 Among these oxides, the Gd₂O₃ nanoparticles also exhibit higher relaxivity than clinical Gd-DTPA, and have been considered as effective T₂ MR contrast agents.28–30 Thus, the lanthanide-doped Gd₂O₃ nanoparticles have attracted attention in the dual-modal imaging. Zhou et al used the lanthanide-doped Gd₂O₃ UCNs with the size of 10–270 nm as dual-modal probes for MR/UCL imaging.31 Liu et al reported the application of Gd₂O₃:Yb³⁺/Er³⁺ nanorods (length of 90–150 nm and diameter of 10–25 nm) for CT/MRI/UCL multimodal imaging.32 Our recent work has focused on the synthesis and development of ultra-small (sub-10 nm) lanthanide-doped Gd₂O₃ nanoparticles for MR/fluorescence dual-modal imaging.33–36 However, to the best of our knowledge, the ability of lanthanide-doped Gd₂O₃ UCNs for theranostics application, which combines the diagnosis and treatment of cancer, has not previously been reported. In this contribution, the aim was to synthesis magnetic and fluorescent lanthanide-doped Gd₂O₃ UCNs and simultaneously achieve MR/UCL dual-modal imaging and PDT. This study is part of a long term to improve the accuracy or possibilities to detect (or monitor) and treat cancer with MR/UCL dual-modal imaging and PDT by utilizing the magnetic and fluorescent properties of lanthanide-doped Gd₂O₃ UCNs.

**Experimental section**

**Preparation and characterization of the UCNs**

The UCNs were prepared by a two-step method, as previously reported.35 Analytical grade powders of Gd₂O₃, Yb₂O₃, and Ln₂O₃ (Ln = Tm, Ho, Er) in the stoichiometric ratio of 83:15:2 were used as the raw materials to fabricate targets via a solid-state reaction technique. The UCNs were prepared utilizing the laser ablation in liquid (LAL) technique, “top-down” strategy, which generates higher purity products efficiently, and has been applied to synthesizing a variety of nanomaterials.37 A schematic diagram of the formation mechanism of UCNs by LAL is shown in Scheme 1. The process starts with the penetration of laser pulse in the liquid and arrival to the Gd₂O₃:Yb³⁺/Ln³⁺ target surface (Scheme 1B), then followed by a high-temperature and high-pressure plasma plume containing the ablated material produced at the target/liquid interface, accompanied by the emission of a shockwave (Scheme 1C). As there is continual absorption of laser pulse by the target, the plasma plume subsequent ultrasonic adiabatic expands into surrounding liquid (Scheme 1D).37 During the expansion, the plasma plume quickly cools down and releases energy to the deionized water, resulting in the nucleation of the Gd₂O₃:Yb³⁺/Ln³⁺ UCNs and growth of nuclei (Scheme 1E). Meanwhile, the energy release of plasma plume to the liquid induces the generation of a cavitation bubble, which expands into the liquid.38 During its expansion, the temperature and pressure inside the bubble decreased rapidly and cause the collapse of the bubble. During the collapse, the pressure and temperature recover to the original values, and the high energy is released by emission of a second shockwave,38 finally the Gd₂O₃:Yb³⁺/Ln³⁺ UCNs are formed and ejected into surrounding liquid (Scheme 1F). The ablated colloids were allowed to stand for 24 h, before the upper liquid was collected for further measurement.

The UC fluorescence spectrum of the collected upper liquid was measured at room temperature using an Edinburgh spectrofluorophotometer (FLS920, Edinburgh Instrument Ltd., Edinburgh, UK) equipped with a 980 nm NIR laser. The structure, morphology, and component of the products were characterized using an X-ray diffractometer (XRD, D-MAX2200 VPC, Tokyo, Japan), a transmission electron microscope (TEM, FEI Tecnai G2 Spirit, FEI Company,
In vitro MRI measurements
In vitro T₁-weighted MRI was performed by a 3.0T Siemens Trio MRI scanner (Siemens Medical Solutions, Erlangen, Germany). The TSE T₁ axial sequence was used with the following parameters: 5% dist. factor, FOV = 64 mm, slice thickness = 2.0 mm, TR = 600 ms, TE = 12 ms, no of averages = 6. In this study, various samples with Gd concentrations in the average 0–0.1 mM were measured. The signal intensities were analyzed using a picture archiving and communications system.

In vivo MRI measurements
BALB/c nude mice of 16±2 g weight were purchased from the Animal Experiment Centre of the Medical College, Sun Yat-Sen University (People’s Republic of China) and housed in a pathogen-free animal facility. The study protocol was approved by the Care and Use of Laboratory Animals of Sun Yat-sen University (Permit Numbers: SCXK (Guangdong)
We strictly followed the Guide for the Care and Use of Laboratory Animals of Sun Yat-sen University during the study. The BALB/c nude mice were subcutaneously injected with $5 \times 10^6$ nasopharyngeal carcinoma (NPC) CNE-2 cells in 100 μL phosphate-buffered saline (PBS). Ten days after tumor cell inoculation, mice with xenografted tumor of ~60 mm$^3$ were anesthetized by intraperitoneal injection of 1% mebumalnatrium (10 μL/g), then injected with the Gd$_3$O$_7$-Yb$^{3+}$/Ln$^{3+}$ UCNs (Gd$^{3+}$, 15 μmol/kg) in 100 μL of PBS (1X buffer) via the tail vein, and scanned with the 3.0-T MRI system using a 3 inch in diameter surface coil constructed specifically for small animals. To prevent bias toward abnormally enhanced regions, normalized histograms of signal intensity were generated for the entire tumor.

**Fluorescence imaging of cells**

Cells from a murine macrophage cells line (RAW 264.7 cells) were cultivated under 5% CO$_2$ atmosphere at 37°C in Dulbecco’s Modified Eagle’s Medium (DMEM) containing 10% fetal bovine serum, penicillin (100 units/mL), and streptomycin (100 mg/mL). The cells were incubated with the Gd$_3$O$_7$-Yb$^{3+}$/Ln$^{3+}$ UCNs (20 μg/mL) for 2 h. After co-incubation, the cells were washed with PBS to remove the remaining particles and dead cells and then observed under a laser scanning confocal microscope (LSM 710, Carl Zeiss, Jena, Germany), operating at an excitation wavelength of 980 nm.

**Cytotoxicity assay**

The human NPC cell lines (CNE2 cells, purchased from the Cell bank of Laboratory Animal Center, the Sun Yat-sen University Hospital, Guangzhou, People’s Republic of China, with the permit number SCYK [粤] 2016-0029) were incubated with UCNs concentrations of 20, 30, 40, and 50 μg/mL in 96-well plates for 4, 6, and 8 h and compared to culture media (DMEM) as the negative control and lipopolysaccharides as the positive control. After co-incubation, 20 μL of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) was added for another 4 h of incubation. The culture medium was removed and 100 μL dimethyl sulfoxide added to dissolve the formazan crystals for 10 min. The absorbance at 490 nm was measured by a microplate reader (Bio-Rad, Hercules, CA, USA).

**Singlet oxygen assay**

Singlet oxygen is important in PDT as the singlet oxygen kills cancer cells. The photochemical method with 1,3-diphenylisobenzofuran (DPBF) is generally used to detect singlet oxygen. The singlet oxygen decomposes DPBF, which has an absorption wavelength of ~410 nm, thus the decreases in DPBF absorbance indicate the generation of singlet oxygen. The UCNs (Gd$_3$O$_7$-Yb$^{3+}$/Er$^{3+}$, 3.5 μg/mL, 5 μL), PS (ZnPc, 0.75 μM, 5 μL), and DPBF (10 mM, 15 μL) in solvent were irradiated with NIR light (laser diode, wavelength 980 nm, continuous wave, 0.2 W/cm$^2$). The DPBF absorbance was measured using a Scandop spectrophotometer (Nanodrop 2000, Thermo Fisher Scientific, Waltham, MA, USA).

**In vitro PDT**

To understand the effects of PDT, the cell viability of CNE2 cells containing the UCNs and the PS was measured under NIR irradiation. The cancer cells (2×10$^3$ cells/100 μL) were seeded into 96-well cell culture plates. Following cultivation under dark conditions in DMEM, the cells were treated with UCNs (Gd$_3$O$_7$-Yb$^{3+}$/Er$^{3+}$, 40 μg/mL) and the PS (ZnPc, 0.75, 1.5, and 3 μM), and compared to UCNs and ZnPc control groups. All groups were cultivated under the same conditions for 4 h. The cells were washed three times with PBS and replaced into fresh culture medium. The cells were then irradiated with NIR light (laser diode, wavelength 980 nm, continuous wave, 0.2 W/cm$^2$). After the 30 min cultivation, the cell viability was measured by MTT assay.

**Results and discussion**

**Upconversion fluorescence properties of the Gd$_3$O$_7$-Yb$^{3+}$/Ln$^{3+}$ UCNs**

The sensitizer (Yb$^{3+}$ ions) has a much larger absorption cross-section at 980 nm and a much higher concentration than the activator (Ln$^{3+}$ ions), thus energy transfer upconversion is the main pathway in the combined Yb$^{3+}$/Ln$^{3+}$ system. Under NIR excitation at 980 nm, the Yb$^{3+}$ ions continuously absorbed 980 nm photons and transfer the energy to the activator, resulting in photons emission. Figure 1A shows the upconversion fluorescence photograph of the collected UCNs colloids under excitation at 980 nm, as obtained by a digital camera without using any optical filters. The upconversion fluorescence colors obtained from the Yb$^{3+}$/Tm$^{3+}$, Yb$^{3+}$/Ho$^{3+}$, and Yb$^{3+}$/Er$^{3+}$ combinations are blue, green, and red, respectively, which are naked-eye compatible and reflect the entire spectrum. Figure 1B shows their corresponding fluorescence spectra in the range of 400–750 nm under excitation at 980 nm. The Gd$_3$O$_7$-Yb$^{3+}$/Tm$^{3+}$ UCNs exhibit three main upconversion emission bands in the visible spectrum, including a strong blue emission band at around 476 nm and two relatively weak red emission peaks centered at 653 and 695 nm. The Gd$_3$O$_7$-Yb$^{3+}$/Ho$^{3+}$ UCNs have a strong green emission at 543 nm and a weak red emission at 667 nm, while the Gd$_3$O$_7$-Yb$^{3+}$/Er$^{3+}$ UCNs exhibit a strong red emission at around 672 nm and two quite weak green emissions at ~528 and 547 nm. Interestingly, the red emission of Gd$_3$O$_7$-Yb$^{3+}$/
Er\textsuperscript{3+} UCNs centered at 672 nm have a large red shift (over 10 nm) compared to the reported cubic counterpart.\textsuperscript{32,42}

Tm\textsuperscript{3+}, Ho\textsuperscript{3+}, and Er\textsuperscript{3+} ions are the most common activators in UCNs and possess their own ladder-like energy levels, as shown in Figure S1. The intense blue and weak red emissions of Gd\textsubscript{2}O\textsubscript{3}:Yb\textsuperscript{3+}/Tm\textsuperscript{3+} UCNPs are assigned to the \( ^{1}G_{4} \rightarrow ^{3}H_{6} \) (476 nm), \( ^{3}G_{4} \rightarrow ^{3}H_{6} \) (653 nm), and \( ^{3}F_{2} \rightarrow ^{3}H_{6} \) (695 nm) transitions of Tm\textsuperscript{3+}. For Gd\textsubscript{2}O\textsubscript{3}:Yb\textsuperscript{3+}/Ho\textsuperscript{3+}, the peak near 543 nm is assigned to the \( ^{3}F_{4} \rightarrow ^{5}S_{2} \) transition of Ho\textsuperscript{3+}, and a weak emission in red region comes from \( ^{5}F_{5} \rightarrow ^{5}I_{6} \) (667 nm) transition. The intense red emission and two quite weak emissions in the green region of Gd\textsubscript{2}O\textsubscript{3}:Yb\textsuperscript{3+}/Er\textsuperscript{3+} UCNPs are assigned to the \( ^{3}F_{2} \rightarrow ^{4}I_{15/2} \) (672 nm), \( ^{4}S_{2} \rightarrow ^{4}I_{15/2} \) (547 nm) and \( ^{3}H_{11/2} \rightarrow ^{2}I_{13/2} \) (528 nm) transitions of Er\textsuperscript{3+}, respectively. Thus, the different color fluorescence of the Gd\textsubscript{2}O\textsubscript{3}:Yb\textsuperscript{3+}/Ln\textsuperscript{3+} nanoparticles indicates that the Yb\textsuperscript{3+}, Tm\textsuperscript{3+}, Ho\textsuperscript{3+}, and Er\textsuperscript{3+} ions are successfully doped into the Gd\textsubscript{2}O\textsubscript{3} matrix.

**Structure, morphology, and component of Gd\textsubscript{2}O\textsubscript{3}:Yb\textsuperscript{3+}/Ln\textsuperscript{3+} UCNs**

The crystal structures of the Gd\textsubscript{2}O\textsubscript{3}:Yb\textsuperscript{3+}/Ln\textsuperscript{3+} UCNs were characterized by XRD measurement. The collected UCNs colloids are dropped onto silicon substrates and evaporated for measurement. Figure 2 shows the XRD patterns of Gd\textsubscript{2}O\textsubscript{3}:Yb\textsuperscript{3+}/Ln\textsuperscript{3+} UCNs. The peaks of the UCNs match well to standard monoclinic Gd\textsubscript{2}O\textsubscript{3} (PDF#42-1465). The strong narrow peaks are indicative of the high-quality crystallinity of the UCNs.

The typical TEM image, SAED pattern, HRTEM image, and size distribution histogram of the Gd\textsubscript{2}O\textsubscript{3}:Yb\textsuperscript{3+}/Er\textsuperscript{3+} UCNs are shown in Figure 3A–D. The results show that the as-prepared UCNs possess a nearly spherical shape with a wide-sized distribution. The corresponding size distribution histogram of the nanoparticles, which is fitted by the Gauss function, shows an average diameter of \( \sim 7.8 \) nm. However, the size calculated from XRD data by Scherrer equation is \( \sim 17.43 \) nm. Figure 3C shows the high resolution TEM (HRTEM) of UCNs. Clearly, small and large particles coexist and possess high crystallinity. In general, small nanoparticles usually make the peaks of XRD become wider, and even no peak can be detected for the ultrafine nanocrystals. We deduce that these high crystalline large nanoparticles may have much effect to enhance the signal of XRD measurement or large particles maybe picked in the XRD.
measurement. The presence of large rings in the selected area electron diffraction pattern (Figure 3B) is further evidence of a polycrystalline structure. The ring spacings matched well with the crystallographic planes of monoclinic Gd$_2$O$_3$ (PDF#42-1465). Taking the Gd$_2$O$_3$:Yb$^{3+}$/Er$^{3+}$ UCNs as an example, XPS was employed to confirm the components of the products. As shown in Figure S2, peaks at 1,220.4, 1,187.0, 184.0, and 169.5 eV denote Gd3d$^{3/2}$, Gd3d$^{5/2}$, Yb4d, and Er4d, respectively. The structure and component analyses demonstrate that Yb$^{3+}$/Er$^{3+}$ ions are successfully co-doped into the Gd$_2$O$_3$ host matrix.

In vitro and in vivo MRI study

Samples were fed into the tubes in various concentrations between 0 and 0.1 mM for measurement. Figure 4A shows the T$_1$-weighted MR images of the Gd$_2$O$_3$:Yb$^{3+}$/Ln$^{3+}$ UCNs and Gd-DTPA (clinical MRI contrast agent). The results reveal an enhanced brightness of the MRI signal with increasing UCNs concentration. For the same Gd concentration, the MRI using the UCNs as contrast agent is much brighter than those using Gd-DTPA, indicating that the Gd$_2$O$_3$:Yb$^{3+}$/Ln$^{3+}$ nanoparticles possess better contrast enhancement than Gd-DTPA. The longitudinal relaxivity ($r_1$) value was measured to understand its effectiveness from a quantitative perspective. As shown in Figure 4B, $R_1$ (1/T$_1$) vs Gd concentration curves were plotted, and the slopes of the curves or $r_1$ were obtained for each sample. The $r_1$ values of Gd$_2$O$_3$:Yb$^{3+}$/Tm$^{3+}$, Gd$_2$O$_3$:Yb$^{3+}$/Ho$^{3+}$, and Gd$_2$O$_3$:Yb$^{3+}$/Er$^{3+}$ UCNs are 13.23, 16.56, and 14.52 s$^{-1}$mM$^{-1}$, respectively. Note
that the $r_1$ values of Gd$_2$O$_3$:Yb$^{3+}$/Ln$^{3+}$ UCNs are over three times higher than those of Gd-DTPA (4.16 s$^{-1}$mM$^{-1}$), which is consistent with the above contrast enhancement result.

Figure 4C illustrates the interaction between the contrast agents (Gd-DTPA and Gd$_2$O$_3$:Yb$^{3+}$/Ln$^{3+}$ UCNs) and water. The $r_1$ value of the contrast agent is proportional to the water hydration number which directly corresponds to the unpaired electrons of Gd$^{3+}$ ions. The Gd$^{3+}$ ions of Gd-DTPA can only offer one hydrate position since the other six unpaired electrons are coordinated by the chelates. However, a Gd$^{3+}$ ion on the surface of the UCNs can offer all its seven unpaired electrons for water hydration, resulting in a higher $r_1$ value than Gd-DTPA. The $T_1$-weighted MR images of the NPC CNE-2 xenografted tumor (white arrow) in BALB/c nude mice are shown in Figure 5. $T_1$-weighted images in the axial orientations were obtained at 0, 5, 10, 15, 30, 45, 60, and 90 min after intravenous administration of the Gd$_2$O$_3$:Yb$^{3+}$/Er$^{3+}$ nanoparticles and clearly show a high contrast enhancement of the tumor 30 min after injecting the Gd$_2$O$_3$:Yb$^{3+}$/Er$^{3+}$ nanoparticles. Therefore, both the in vitro and in vivo MR imaging investigations indicate that the Gd$_2$O$_3$:Yb$^{3+}$/Er$^{3+}$ nanoparticles may be a promising $T_1$-weighted MRI contrast agent for biomedical applications.

Confocal fluorescence imaging of live cells

The use of NIR light for the excitation of nanoparticles for molecular, cellular, and tissue imaging is very attractive in bioimaging applications owing to the lack of tissue damage and deep tissue penetrability. To explore the feasibility of using the UCNs as biological fluorescence probes, we conducted in vitro biological experiments using a murine macrophage cells line (RAW 264.7 cells) incubated with Gd$_2$O$_3$:Yb$^{3+}$/Ln$^{3+}$ UCNs. The cells were washed after incubation and imaged using a confocal laser fluorescence microscope operating at an excitation wavelength of 980 nm. As shown in Figures 6 and S3, blue, green, and red fluorescence can be observed from the cells incubated with the
Figure 5 In vivo MRI of a NPC CNE-2 xenografted tumor after intravenous administration of the Gd$_2$O$_3$:Yb$^{3+}$/Er$^{3+}$ UCNs (Gd$^{3+}$, 15 μmol/kg) at 0, 5, 10, 15, 30, 45, 60, and 90 min, respectively.

Abbreviations: MRI, magnetic resonance imaging; NPC, nasopharyngeal carcinoma; UCNs, upconversion nanoparticles; min, minutes.

Figure 6 Confocal fluorescence images of RAW267.4 cells incubated with Gd$_2$O$_3$:Yb$^{3+}$/Ln$^{3+}$ UCNs under excitation at 980 nm.

Abbreviation: UCNs, upconversion nanoparticles.
Magnetic and fluorescent Gd$_2$O$_3$:Yb$^{3+}$/Ln$^{3+}$ nanoparticles, respectively, with no autofluorescence found. The positions of the cells in the bright-field and the dark-field fluorescence imaging correlated well, and the fluorescence products were primarily located in the cell membrane and cytoplasm. The cells show good viability and morphology, suggesting that the products cause no significant damage to the cells. We also assessed the cells’ viability after incubation with Gd$_2$O$_3$:Yb$^{3+}$/Ln$^{3+}$ (Ln = Tm, Ho, and Er) UCNs (Figure S4). Clearly, there is no significant difference between the three kinds of nanoparticles and the negative control (PBS); the data showed that the cell toxicity of the three kinds of nanoparticles is satisfactory. This result indicates that the upconversion fluorescence emission of the UCNs is strong enough for live cell imaging and can be applied as potential fluorescence nanoprobes for in vivo imaging.

In vitro PDT

Cytotoxic singlet oxygen can result in cell death. Therefore, the ability to generate singlet oxygen is one of the crucial factors for effective PDT. Singlet oxygen production is generally monitored using DPBF, which reacts irreversibly with singlet oxygen causing a decrease of its characteristic absorption at ~410 nm.$^{39-41}$ As shown in Figure 7A, the combination of the UCNs (Gd$_2$O$_3$:Yb$^{3+}$/Er$^{3+}$) with PS (ZnPc) resulted in the highest rate of decomposition of the DPBF under NIR irradiation, and thus absorption of DPBF decreased exponentially.
with increasing NIR irradiation time, confirming the efficient generation of singlet oxygen.

To understand the effects of PDT, the viability of cells containing the combination of UCNs and PS under NIR irradiation was studied. First, the viability of CNE2 cells after incubation with UCNs concentrations of 20, 30, 40, and 50 μg/mL for 4, 6, and 8 h was determined by MTT assay. As shown in Figure 7B, the UCNs have no significant intrinsic toxicity effect on the cells’ survival. Then, the viability of CNE2 cells containing Gd-based UCNs (Gd2O3:Yb3+/Er3+, 40 μg/mL) and the ZnPc (0.75, 1.5, and 3 μM) under NIR irradiation was measured and compared to the control experiment. The UCNs-based PDT treatment is schematically illustrated in Figure 7C. The NIR irradiation of the UCNs emitted red light (~672 nm), excited the ZnPc (maximum absorption peak at ~670 nm), which then generated singlet oxygen to kill the cancer cells. The results shown in Figure 7D demonstrate that the cells’ viabilities of samples containing combinations of UCNs and ZnPc are lower than their control groups, meaning that this NIR-induced PDT can inhibit the proliferation of cancer cells.

Conclusion
In summary, magnetic and fluorescent lanthanide-doped Gd2O3 UCNs imaging have been fabricated and can simultaneously achieve MR/UCL dual-modal imaging and PDT. As an MRI contrast agent, the products show a high longitudinal relaxivity (r1) value and T1-weighted contrast enhancement of xenografted tumors in mice. As UCL imaging nanoprobes, the products exhibit bright luminescence under NIR irradiation. Moreover, upon NIR excitation, PDT can be achieved by the combination of the UCNs and the PS ZnPc. We anticipate that the magnetic and fluorescent lanthanide-doped Gd2O3 UCNs will be a highly attractive addition to the theranostic platform, and promote the development of “detect-to-treat” strategies or imaging-guided therapy of cancer.

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Author contributions
JL, LH, XT, XC, YS, and FX: experimental work. DC and LL: project planning. All authors contributed toward data analysis, drafting and revising the paper and agree to be accountable for all aspects of the work.

Disclosure
The authors report no conflicts of interest in this work.

References


Supplementary materials

Figure S1 Schematic illustration of the UC emission process (A), and the proposed UC mechanism schemes (B, C) of the Gd₂O₃:Yb³⁺/Ln⁺ (Ln = Tm, Ho, and Er) UCNs.

Abbreviations: NIR, near-infrared; UC, upconversion; ET, energy transfer; Ex, excitation; Em, emission.
Figure S2 Gd3d, Yb4d, and Er4d XPS spectra of Gd2O3:Yb3+/Er3+ UCNs.
Figure S3 Confocal fluorescence images of Raw267.4 cells incubated with Gd₂O₃:Yb³⁺/Ln³⁺ UCNs.

Figure S4 Cells viability of Raw267.4 cells incubated with Gd₂O₃:Yb³⁺/Ln³⁺ (Ln = Tm, Ho, and Er) UCNs (20μg/mL at 2 hours).

Abbreviations: LPS, lipopolysaccharides; PBS, phosphate-buffered saline.