Impact of prostaglandin glaucoma drops on platelet-activating factor action: an in vitro study

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Aim: The aim of this study was to investigate the effect of different prostaglandin analogs on platelet-activating factor (PAF) levels.

Methods: Three prostaglandin analogs were selected: bimatoprost 0.3 mg/mL, latanoprost 50 μg/mL, and tafluprost 15 μg/mL. Each drug sample was tested for its ability to cause platelet aggregation, which was measured as PAF-induced aggregation, before and after the addition of various concentrations of the examined sample, creating a linear curve of percentage inhibition (ranging from 0% to 100%) versus different concentrations of the sample. The concentration of the sample that inhibited 50% PAF-induced aggregation was calculated based on this curve, and this value was defined as IC50. In addition, the effect of eye drops on PAF metabolism was examined, through an in vitro analysis on PAF basic metabolic enzymes (PAF-cholinephosphotransferase, PAF-acetyl-CoA:1-O-alkyl-sn-glycero-3-phosphocholine acetyltranseferase, and PAF-acetylhydrolase).

Results: The IC50 values for Lumigan UD® (bimatoprost 0.3 mg/mL), Monoprost® (latanoprost 50 μg/mL), and Saflutan (tafluprost 15 μg/mL) were 8.7, 0.28, and 1.4 μg/mL, respectively.

Discussion: All three prostaglandin analogs suspended PAF, but bimatoprost induced the most potent inhibition, compared to tafluprost and to the weak effect of latanoprost.

Keywords: glaucoma, platelet-activating factor, prostaglandin analogs, treatment, platelet aggregation

Introduction

Glaucoma is estimated to be the second leading cause of severe vision loss and blindness worldwide, affecting ~60.5 million people and accounting for 2.65% of the population aged >40 years in 2010.1,2 The medical treatment for glaucoma includes prostaglandin analogs, which are approved as the first-line treatment for glaucoma in the European Glaucoma Society guidelines, carbonic anhydrase inhibitors, β-receptor antagonists, adrenergic agonists, and parasympathomimetics.3,4 Prostaglandins are a family of lipid compounds that are derived enzymatically from essential fatty acids, with each containing 20 carbon atoms, including a 5-carbon ring.5 In eye, they are synthesized by the trabecular endothelial and the ciliary muscle cells from membrane phospholipids and arachidonic acid, exhibiting anti-inflammatory action.6,7 They seem to alter both the function and structure of the uveoscleral pathways, inducing extracellular matrix remodeling, widening of the intermuscular spaces, and dissolution of collagens types I and III.8 Besides prostaglandin E2, various inflammatory molecules, including cytokines, cyclooxygenase 2, tumor necrosis factor-alpha, interleukins, platelet-activating factor (PAF), vascular endothelial growth factor, and C-reactive protein have been implicated in the mechanisms of glaucoma, highlighting the role of inflammation in the pathogenesis of the disease.9,10

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PAF (1-0-alkyl-2-acetyl-sn-gluceryl-3phosphorylcholine) is a low-molecular phospholipid mediator that is involved in inflammatory process acting on various types of cells through specific receptors. Two metabolic pathways are responsible for PAF secretion: 1) de novo synthesis, where PAF is produced by a lipid intermediate (1-0-alkyl-2-acetyl-sn-glycerol), through the action of the enzyme cholinephosphotransferase (PAF-CPT), converting cytidine diphosphate-choline (CDP-choline) to cytidine monophosphate; and 2) remodeling pathway, which is activated under inflammatory conditions and results in the secretion of PAF from preexisting membrane-bound ether-linked phospholipids through the enzyme acetyl-CoA:1-O-alkyl-sn-glycero-3-phosphocholine acetyltransferase (lyso-PAF-AT), which transfers an acetyl group in the sn-2 position of lyso-PAF. In the remodeling pathway, when cytosolic phospholipase A2 is activated, it becomes membrane-associated and hydrolyzes the fatty acyl moiety esterified at the sn-2 position of membrane phospholipids. The subsequent release of 2-lyso-phospholipids (1-acyrlsn-glycero-3-phosphocholine or lyso-choline-containing membrane phospholipids) serves as substrates for the activation of PAF-acetylhydrolase (PAF-AH). The latter represents a heterogeneous group of PLA2 that removes sn-2 acetyl group, without affecting normal membrane phospholipids; they are responsible for the hydrolysis, the inactivation, and the concentrations of PAF in plasma and tissues.

The activation of PAF requires its coupling with its receptor, which is a 7-transmembrane-G-protein-coupled receptor, located in a great variety of cells, including monocytes, lymphocytes, vascular endothelial and smooth muscle cells, renal and mesangial cells, hepatocytes, keratinocytes, and osteoclasts. The range of cell responses to PAF explains its role in both physiological processes, such as platelet aggregation and secretion, activation of neutrophils and macrophages, and pathological mechanisms, resulting in asthma, systemic or cardiac anaphylaxis, endotoxin-induced shocks, thrombosis, and gastrointestinal ulceration. The detection of PAF in ocular tissues, such as iris, ciliary body, retina, and vascular endothelium, confirms its implication in ocular inflammation and blood flow.

Regarding the inflammatory background of glaucoma and the implication of PAF in the ocular inflammation and in the glaucomatous neuronal damage, the effect of different prostaglandin analogs on PAF levels was investigated.

Materials and methods

Equipment and solutions

The following equipment was used in the present study:

1. Heraeus® Labofuge® 400R for centrifugations
2. Superoxid Bandelin Sonopuls HD 2070 sonicator (Heinrichstraze 3–4, Berlin, Germany) for homogenizations (30% of the power)
3. 1209 Rackbeta (Pharmacia, Wallac, Finland) as liquid scintillation counter
4. Chrono-Log model 400 VS aggregometer (Havertown, PA, USA) to study platelet aggregation.

The eye drops tested in the present study were as follows: Lumigan UD® (bimatoprost 0.1 mg/mL; Allergan Pharmaceuticals Ltd., Westport, Ireland), Monoprost® (latanoprost 50 μg/mL; Laboratoires Thea, Clermont-Ferrand, France), and Saflutan (tafluprost 15 μg/mL; Vianex S.A., Athens, Greece).

Calculation of the inhibitory effects of sample on PAF-induced aggregation

This is an in vitro study, where no human or animal subjects are included, but blood samples was required. The ethics committee of the General Hospital of Athens G. Gennimatas approved the protocol. Because the study was an in vitro one, no patient informed consent was needed. A total of 9 mL of blood was collected from the marginal ear vein of a New Zealand White rabbit and added in 1 mL of an anticoagulant solution of sodium citrate/citric acid. The sample was centrifuged at 630× g for 13 min at 25°C, and the supernatant was subsequently centrifuged at 1,400× g for 20 min at 25°C in order to obtain plasma (rich in platelets). The isolation of plasma and leukocytes from blood samples and the biological assay on washed rabbit platelets were carried out according to the methods described by Moschos et al. The leukocytes were isolated after the sedimentation of the erythrocytes using 3.4 mL of dextran solution (3% dextran in 0.15 M NaCl) for 1 h at room temperature. The homogenates of leukocytes were aliquoted and stored at −80°C after protein determination by using Bradford method.

Briefly, PAF and the examined drug samples were dissolved in 12.5 mg of bovine serum albumin (BSA) per 1 mL of saline. The ability of each selected drug to cause platelet aggregation was estimated by adding various concentrations of each sample into the platelet suspension. The PAF-induced aggregation was calculated at baseline (0% inhibition) and after the addition of the examined samples (in a variety of concentrations), and a linear curve of the percentage inhibition to the concentrations of each sample was created. The concentration of the sample that inhibited 50% of the PAF-induced aggregation was defined as IC50.
Drug was extracted according to the Bligh–Dyer method and was separated by thin-layer chromatography (TLC) on silica gel G-coated plates with a development system consisting of chloroform/methanol/acetic acid/water (100:57:16:8, v/v/v/v). The specific activities of PAF-CPT and lyso-PAF-AT were expressed as pmol of produced PAF/min/mg of sample protein present in each assay.

The homogenate of leukocytes, isolated from New Zealand white rabbits, was used to perform the PAF-CPT and lyso-PAF-AT activity assays, as described previously. The PAF-CPT activity assay was carried out at 37°C for 20 min in a final volume of 200 µL, started by the addition of CDP-choline and stopped by the addition of 0.5 mL of cold methanol after 20 min. The reaction of lyso PAF-AT was carried out at 37°C for 30 min in a final volume of 200 mL, started with the addition of metalloproteinase 2 (MMP-2) and metalloproteinase 3 (MMP-3) than latanoprost and tafluprost, whereas tafluprost and latanoprost increased the expression MMP-1 and MMP-9, respectively, more than bimatoprost. In a previous study, it was found that latanoprost (IC50 = 0.29 µM) was found to be the most potent in inhibiting PAF compared to single travoprost (IC50 = 0 µM) or to the combination of the latter with timolol (IC50 = 0.11 µM).

Jager et al had previously observed that the accumulation of PAF in the aqueous humor of rabbits was implicated by the increase of intraocular pressure. Moreover, they supported that the systemic administration of a PAF-antagonist (BN 52021) prevented the rise of intraocular pressure. It has already been revealed that the binding of PAF with its receptor results in the activation of PLA2 and phospholipases C, D, P, and A2; releasing arachidonic acid; prostaglandin metabolites; and protein kinase C. PAF also stimulates the expression of selective MMPs, whose levels and activity seem to be increased in the iris/ciliary body of mice with pigmentary glaucoma.

The present study concluded that all prostaglandin analogs suspended PAF. However, bimatoprost seems to have a potent inhibitory action against PAF compared to the moderate effect of tafluprost and the weak action of latanoprost. It was also concluded that bimatoprost can efficiently eliminate the inflammatory process that takes place in glaucomatous lesions, eliminating the concentrations of PAF.

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**Author contributions**

All authors contributed toward data analysis, drafting and revising the paper and agree to be accountable for all aspects of the work.
Disclosure
The authors report no conflicts of interest in this work.

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