Effect of inserted spacer in hepatic cell-penetrating multifunctional peptide component on the DNA intracellular delivery of quaternary complexes based on modular design

Luchen Zhang1
Zhenbo Li1
Fangli Sun2
Yuhong Xu1
Zixiu Du1

1School of Pharmacy, Shanghai Jiao Tong University, Shanghai, 2School of Engineering, Zhejiang Agriculture and Forestry University, Lin’an, Zhejiang Province, People’s Republic of China

Abstract: A safe and efficient quaternary gene delivery system (named Q-complexes) was constructed based on self-assembly of molecules through noncovalent bonds. This system was formulated through the cooperation and competing interactions of cationic liposomes, multifunctional peptides, and DNA, followed by coating hyaluronic acid on the surface of the ternary complexes. The multifunctional peptide was composed of two functional domains: penetrating hepatic tumor-targeted cell moiety (KRPTMRFRYTWNPMD) and a wrapping gene sequence (polynarginine 16). The effect of spacer insertion between the two domains of multifunctional peptide on the intracellular transfection of Q-complexes was further studied. Experimental results showed that the formulations assembled with various peptides in the spacer elements possessed different intercellular pathways and transfection efficiencies. The Q-complexes containing peptide in the absence of spacer element (Pc) showed the highest gene expression among all samples. The Q-complexes containing peptides with a noncleavable spacer GA (Pd) had no ability of intracellular nucleic acid delivery, whereas those with a cleavable spacer RVRR (Pv) showed moderate transfection activity. These results demonstrated that the different spacers inserted in the multifunctional peptide played an important role in in vitro DNA transfection efficiency. Atomic force microscopy images showed that the morphologies of ternary complexes (LPD) and Q-complexes (HL, PD) were crystal lamellas, whereas those of other nanocomplexes were spheres. Circular dichroism showed the changed configuration of peptide with spacer GA in nanocomplexes compared with that of its free state, whereas the Pd configuration without spacer in nanocomplexes was consistent with that of its free state. The present study contributed to the structural understanding of Q-complexes, and further effective modification is in progress.

Keywords: Q-complexes, spacer elements, multifunctional peptides, lipid component, intracellular transfection

Introduction

Gene delivery can increase or decrease the expression of any protein in a cell theoretically,1 and thus drugs based on gene have been regarded a promising treatment for many serious diseases, such as cancer and genetic disorders.2,3 Effective gene delivery to the target sites remains a challenge because of the difficulty of transporting these negatively charged macromolecules to overcome various physical barriers without degradation.1,4 A safe and efficient delivery system is thus needed for a successful gene therapy.1,4,5 Since 1960s, nonviral vectors based on cationic polymers6–9 and lipids10
have been preferred as the carriers to wrap and transport genes. Numerous efforts have been made to optimize the designing strategies of nonviral vectors to overcome the barriers during delivery process. The conventional method of designing targeted drug carriers is via chemical conjugation of functional moieties; the conjugation of various ligands requires chemical reactions and may destroy the conformation of biomolecules, thereby resulting in inactivation and cytotoxicity. The self-assembly of molecules via noncovalent bonds to form an organized structure is most helpful in drug delivery system, maintains the nature of each component, and enhances the delivery activity of carriers through the synergies of all functional moieties.

On the basis of self-assembly of molecules via noncovalent bonds, we rationally constructed a highly effective and biologically responsive delivery system (Q-complexes) using nontoxic lipids (1,2-dioleoyl-3-trimethylammonium-propane [DOTAP] and 1,2-dioleoylsn-glycero-3-phosphatidylethanolamine [DOPE]), peptide, and hyaluronic acid (HA). Briefly, the formulation is first composed of a cationic liposome, multifunctional peptide, and DNA at optimized ratios, which can finally self-assemble into ternary complexes (LPD). The ternary complex was associated with negatively charged HA and formulated quaternary complexes (HLPD, also named as Q-complexes). Specifically, we applied a hepatic tumor-specific cell-penetrating peptide (KRPTMRFRYTWNPMK) to polyarginine 16 (R
16) in the absence or presence of spacer to create a multifunctional peptide, which could specifically permeate hepatic tumor cells and tightly condense nucleic acids. The HA coated at the surface of ternary complexes is in charge of shielding the negative charge of the nanoparticle surface, as well as the oriented binding to the tumor tissues and cells because HA actively targets receptors (such as CD44) distributed on the surfaces of the tumor cells. From the similar reported formulation, we could conclude that some of KRPTMRFRYTWNPMK in the multifunctional peptide protruded at the surface of LPD, which possibly induces the nanoparticles to penetrate into the tumor cells, and the lipid component destroys the endosomal membranes through membrane fusion function of DOPE.

In the present study, we found that the transfection efficiency of HLPD is closely related to the inserted spacer of the multifunctional peptides, between KRPTMRFRYTWNPMK and R
16. Thus, we investigated the effect of spacer element on the in vitro DNA transfection of HLPD and mainly explored the characteristics and internalization mechanism of HLPD containing the peptides with different spacers between KRPTMRFRYTWNPMK and R
16. The cell uptake and gene expression of Q-complexes after incubation with different inhibitors were analyzed to validate the successful intracellular gene delivery pathway. In this text, H represents HA, L represents cationic liposome that was composed of DOTAP/DOPE at the weight ratio of 1:1, P represents peptide, and D represents DNA. For example, HLP
D refers to HA, cationic liposome, P
s, and DNA at the weight ratio of 14:1:4:1.

Materials and methods

Materials

DOTAP and DOPE were purchased from Sigma-Aldrich Co. (St Louis, MO, USA). The peptide sequence R
16KMPNWTRYFRMTPRK (P
a), R
16 (P
b), and peptide with the spacers GA and RVRR based on P
s (named as P
c and P
d) were purchased from China Peptides (Shanghai, People’s Republic of China) and dissolved in 10 ng/mL of nuclease-free water (Thermo Fisher Scientific, Waltham, MA, USA). The plasmid pCI-Luc comprising the luciferase gene of pGL3 was purchased from Invitrogen. The plasmids were amplified in DH5α strain of Escherichia coli and prepared using a GoldHi EndoFree Plasmid Maxi Kit (CWBiotech, Beijing, People’s Republic of China). HA (Mw =34 kDa) was purchased from Shandong Freda Biotechnology Co., Ltd. (Shandong, People’s Republic of China). In addition, fluorescein isothiocyanate-labeled HA (FITC-HA) was synthesized as reported.

Preparation of cationic liposome

Liposome preparation was described previously. Briefly, the required amounts of DOTAP and DOPE were first dissolved in chloroform at the weight ratio of 1:1, and a thin lipid film was produced via slowly evaporating the chloroform overnight with a rotary evaporator under vacuum conditions. The lipid film was subsequently hydrated with double-distilled water at room temperature to produce a crude suspension of 1 mg/mL. The crude suspension was sonicated for 10 min to obtain the final product.

Formation of LPD and HLPD

Ternary complexes (LPD) were prepared by mixing the cationic liposome with different peptides and DNA solution at the weight ratio of 1:4:1 (ie, charge ratio of 0.3:4:4:1 for P
s, P
c, and P
d and charge ratio of 0.3:8.4:1 for P
b). These complexes were incubated for 30 min to formulate LPD at room temperature. The thoroughly mixed suspensions were mixed with HA solution at a precise ratio of 14:1 HA/DNA (ie, 5.9:1 charge ratio) and maintained for 15 min to produce Q-complexes (HLPD). For physicochemical studies of HLPD, the complexes were prepared and diluted with
double-distilled water. For in vitro transfection, Opti-MEM was used as diluent.

Hydrodynamic size and zeta potential measurements
The hydrodynamic size and zeta potential of LPD and HLPD complexes were measured via dynamic light scattering using a Zetasizer Nano ZS (Malvern Instruments, Malvern, UK) at 25°C±0.1°C. The amount of DNA used in each formulation was 2 µg. The values for each formulation are the mean values of three measurements.

Gel electrophoresis assay
Approximately 1 µg of DNA and 10 µL of complex suspensions were used in the gel electrophoresis assay. All samples, except for 1 kb DNA ladder (Foregene, Chengdu, People’s Republic of China), were added to 2 µL of 6× loading buffer (Thermo Fisher Scientific) before loading the samples onto a 1% w/v agarose gel in Tris-acetate-EDTA buffer, which contained 3 µL of GelRed. The samples were electrophoresed at 90 V for 30 min and visualized under ultraviolet (UV) light.

Atomic force microscope
A drop of LPD and HLPD nanocomplexes was applied onto the mica. After a few seconds, the drop was dried through blotting with the filter paper. The total component concentration of each sample was 0.3 mg/mL. The morphology of LPD and HLPD was examined using an atomic force microscope (Nanoscope IIIa; Digital Instruments/Veeco, Inc., Santa Barbara, CA, USA).

Circular dichroism
Circular dichroism measurements were performed with aqueous preparations of LPD and HLPD complexes, as well as peptide and HA components at the same concentrations similar to those for the LPD and HLPD complexes. The individual components were used as controls to determine their native structures. All spectra were obtained in a JASCO J-815 spectropolarimeter using a 0.1 cm rectangular quartz cuvette cell, at a scan rate of 50 nm/min, data pitch of 1 nm, response time of 1 s, and bandwidth of 1 nm.

Cell culture
SMMC-7721 (human hepatoma) cells were obtained from Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences (Shanghai, People’s Republic of China). Cell culture media and reagents were purchased from Thermo Fisher Scientific. SMMC-7721 cells were cultured in RPMI-1640 medium. The medium used for cell culture was supplemented with 10% v/v fetal bovine serum and 1% v/v penicillin/streptomycin solution. All cell lines were incubated at 37°C in a humidified atmosphere with 5% CO₂.

Flow cytometry and confocal laser scanning microscopy
DiI-dyed lipid component was used to monitor the trafficking of nanocomplexes. The cell uptake of the complexes was quantitatively measured via flow cytometry (FACS Calibur; BD, Worcestershire, NJ, USA). Cells were seeded in 24-well plates at the density of 1×10⁵ cells per well and incubated at 37°C in a humidified atmosphere with 5% CO₂ for 24 h before transfection. The culture medium was subsequently removed, and the cells were rinsed with 500 µL of 1× phosphate buffered saline (PBS). Afterward, the complexes containing 2 µg of pGL3 plasmid in 500 µL of Opti-MEM were added to each well. The cells were incubated at 37°C with 5% CO₂ for 4 h, after which the complex suspensions were removed. The cells were collected and prepared for flow cytometry assay. For confocal microscopy, fluorescent-labeled Q-complex formulations were prepared with DOTAP and DOPE at 1:1 weight ratio containing DiI at 1.0% weight ratio of total lipids. In this experiment, the cells were incubated for 4 h. The cells were mixed with 4% paraformaldehyde, permeabilized with 0.3% triton in PBS for 10 min, and incubated for 5 min at room temperature with DAPI (0.2 mg/mL; Sigma-Aldrich). The cells were washed with PBS and sealed in mounting media (Invitrogen). Subsequently, visualization was carried out on a Leica TCS SP8 laser scanning microscope system (Leica Microsystems, Wetzlar, Germany).

Expression of green fluorescent protein (GFP) reporter gene
SMMC-7721 cells were seeded in 24-well plates at the density of 1×10⁵ cells per well and incubated at 37°C in a humidified atmosphere with 5% CO₂ for 24 h before transfection. The cells were first rinsed with 500 µL of 1× PBS, and the complexes containing 1 µg of GFP plasmid in 500 µL of Opti-MEM were added to each well. The cells were incubated at 37°C with 5% CO₂ for 4 h. The suspensions were subsequently replaced with fresh medium, and the cells were incubated for additional 20 h. The cells were observed under a fluorescence microscope.

Intracellular internalization mechanism assay of HLPD
Cells were seeded in 96-well plates at the density of 2×10⁴ cells per well and incubated at 37°C in a humidified atmosphere with
Amino acid sequences:

<table>
<thead>
<tr>
<th>Peptide</th>
<th>Amino acid sequences</th>
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<tbody>
<tr>
<td>P₁</td>
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| P₂      | RRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRRR
an unchanged α-helix configuration in a free state, which was the same as in LP$_D$ and HLP$_D$. Furthermore, the $\lambda_{\text{max}}$ at ~208 nm appeared in free P$_c$, but moved to 210 nm in LP$_D$ and HLP$_D$, and also appeared in a positive peak at ~195 nm in LP$_D$ and HLP$_D$. The result indicated that the secondary structure of P$_c$ had changed from α-helix toward β-sheet (Figure 4B). The negative $\lambda_{\text{max}}$ of peptide in LP$_D$ and free P$_c$ at 200 nm indicated the random coil, and the $\lambda_{\text{max}}$ of peptide in HLP$_D$ was at 218 nm (Figure 4C), which showed that the secondary structure of P$_c$ in HLP$_D$ was the transition state from random coil into β-sheet.

**Cell uptake**

The Q-complexes containing P$_d$, P$_b$, and P$_c$, respectively, were effectively internalized after 4 h of transfection (Figure 5). The equivalent mean fluorescence intensity of the cells after incubation with Q-complexes and the corresponding ternary complexes could be observed from the cell uptake transfected to SMMC-7721 cells. Such observation indicated that the internalization efficiency of nucleic acids did not drop after coating the surface of ternary complexes with HA. However, the Q-complexes containing P$_c$ were not easily endocytosed by the tumor cells.
Figure 3 Morphologies of LPD and HLPD wrapping pGL3 plasmid observed via an atomic force microscope.

Notes: (A) LP\textsubscript{a}D; (B) HLP\textsubscript{a}D; (C) LP\textsubscript{b}D; (D) HLP\textsubscript{b}D; (E) LP\textsubscript{c}D; (F) HLP\textsubscript{c}D; (G) LP\textsubscript{d}D; and (H) HLP\textsubscript{d}D. LPD: a cationic liposome, multifunctional peptide, and DNA at optimized ratios; HLPD: H represents hyaluronic acid, L represents cationic liposome that was composed of DOTAP/DOPe at a 1:1 weight ratio, P represents peptide (P\textsubscript{a}–P\textsubscript{d} refers to the different peptide used), and D represents DNA.

Abbreviations: DOTAP, 1,2-dioleoyl-3-trimethylammonium-propane; DOPE, 1,2-dioleoylsn-glycero-3-phosphatidyl-ethanolamine.
EGFP expression

Fluorescence microscope was used to observe the GFP expression in varying intensities after incubation with SMMC-7721 cells for 24 h. The HLP_D sample showed the highest transfection activity (Figure 6) in all the samples containing P_a–P_d. Specifically, the GFP expression efficiency of complexes was in the order of samples containing the following peptides: P_a > P_b > P_d > P_c. The GFP expression results indirectly reflected the DNA delivery trend of Q-complexes with different multifunctional peptides.

Luciferase transfection and internalization pathways of ternary and Q-complexes

SMMC-7721 cells were transfected with LPD and HLPD complexes for the investigation of both transfection efficiency and internalization pathways of Q-complexes. Figure 7 illustrates that the intercellular activity of the ternary and Q-complexes containing P_a showed the highest transfection efficiency among all the samples in the absence of inhibitors, and the complexes containing P_c barely delivered DNA into the cells. The samples containing P_d showed higher DNA transfection efficiency than those complexes containing P_c.

The unchangeable transfection efficiencies of all complexes could be observed after incubation with CD44 antibody (Figure 7A), which indicated the minimal contribution of CD44-mediated internalization to the intercellular transfection.
Figure 6 Enhanced green fluorescent protein transfection of LPD (LPD [A], LPD [C], LPD [E], and LPD [G]) and HLPD (HLPD [B], HLPD [D], HLPD [F], and HLPD [H]) in SMMC-7721 cells incubated for 24 h.

Notes: LPD: a cationic liposome, multifunctional peptide, and DNA at optimized ratios; HLPD: H represents hyaluronic acid, L represents cationic liposome that was composed of DOTAP/DOPe at a 1:1 weight ratio, P represents peptide (P$_a$–P$_d$ refers to the different peptide used), and D represents DNA. Magnification ×100.

Abbreviations: DOTAP, 1,2-dioleoyl-3-trimethylammonium-propane; DOPe, 1,2-dioleoylsn-glycero-3-phosphatidyl-ethanolamine.

Figure 7 Luciferase transfection of LPD (LPD, LPD, LPD, and LPD) and HLPD (HLPD, HLPD, HLPD, and HLPD) in SMMC-7721 cells incubated for 24 h.

Notes: In the absence (normal group) and presence (dynasore group) of (A) anti-cD44 antibody (50 µg/mL) pretreated for 1 h, (B) dynamin inhibitor I with 0.2% DMsO in the solution (DI; dynasore, 80 µM) pretreated for 1 h, (C) chlorpromazine (cPZ; 20 µM) pretreated for 1 h, and (D) 5-(N-ethyl-N-isopropyl) amiloride (eIPa; 100 µM) with 0.1% DMsO in the solution pretreated for 30 min. Values are the means of four replicates ±sD. * represents an alpha value of $P<0.05$, ** represents an alpha value of $P<0.01$, and *** represents an alpha value of $P<0.001$. LPD: a cationic liposome, multifunctional peptide, and DNA at optimized ratios; HLPD: H represents hyaluronic acid, L represents cationic liposome that was composed of DOTAP/DOPe at a 1:1 weight ratio, P represents peptide (P$_a$–P$_d$ refers to the different peptide used), and D represents DNA.

Abbreviations: RLU/mg protein, relative light units per milligram of protein; DOTAP, 1,2-dioleoyl-3-trimethylammonium-propane; DOPE, 1,2-dioleoylsn-glycero-3-phosphatidyl-ethanolamine; ns, not significant.
activity of the complexes. In addition, a significant drop in the luciferase expression of the samples containing Pa was observed after pretreatment with dynasore (Figure 7B). This result showed that the internalization pathway of these nanoparticles was mainly dynamin-dependent, thereby reflecting a strong penetrating ability of the peptide component in the nanoparticles.13 The internalization mechanism of most LPD and HLPD was also partly both clathrin-dependent20 (Figure 7C) and macropinocytosis-dependent11 (Figure 7D) pathways.

We further investigated the intracellular paths of Q-complexes observed through confocal microscopy (Figure 8). FITC-HA and Dil-labeled lipid component (red) were used to monitor the trafficking of HLPD inside SMMC-7721 cells. Nuclei were stained with DAPI (blue). After incubation for 4 h, the HLPaD and HLPbD inside the SMMC-7721 cells were observed through Dil-labeled lipid component, but the dots of FITC-HA were not found. This result implied that HA component had separated from the remaining components of Q-complexes before termination of incubation. Similarly, it also indicated that the FITC-HA of Q-complexes could only be efficiently endocytosed through CD44-mediated pathway (Figure 9). We also found that anti-CD44 antibody impeded the fluorescence intensity of FITC-HA, and dynasore could not prevent the cell uptake of FITC-HA. Therefore, HLPaD could not be endocytosed by the SMMC-7721 cells.

Discussion

Multifunctional peptide consists of both nearly neutral (KRPTMRFRYTNPMK) and cationic peptide (R16) sequences. Thus, the effects of incorporating peptide between these two peptide sequences are important for rationally designing a delivery system. A large gap of transfection efficiency between HLPaD and HLPcD inspired us to investigate the role of the incorporating spacer in the peptide component of Q-complexes. We selected two representative peptide spacers (GA and RVRR) as different spacer elements based on the original peptide of Pa (Table 1) and verified whether these changes would affect the formulation and transfection efficiency of Q-complexes. In addition, we inhibited the intercellular uptake pathways, mainly including the CD44 mediating and dynamin-dependent pathways that HA and the tumor cell-penetrating peptide (KMPNWTYRFRMPRK) specifically depended on,13 respectively. Q-complexes
were hypothesized to achieve intracellular delivery through multifunctional peptide completely wrapping nucleic acids and strongly penetrating specific tumor cell lines. Consequently, we employed DOPE lipid component to disrupt the endosomal membrane, in addition to the negatively charged HA coating at the surface of ternary complexes to shield the positive surface charges and highly effective targeting tumor. The experimental results showed that the nanocomplexes containing different peptide components possessed similar sizes (Figure 1), and the zeta potential of LP_D was negative at approximately −19 mV; other LPD formulations had positive surface charge. The electrophoretic mobility results (Figure 2) indicated that LP_D and HLP_D could not tightly wrap DNA. In contrast to other spherical nanoparticles, the morphology and structure of LP_D and HLP_D obviously changed to crystal lamellas, which perhaps implied the reason why the samples containing P_c had bare DNA transfection efficiency. The circular dichroism result (Figure 4) further demonstrated that folding or conformational changes in P_c integrated into LP_D and HLP_D caused the minimal cell uptake and almost failure in transport DNA ability of the delivery system. Taking the results of cell uptake (Figure 5) and transfection efficiencies (Figures 6 and 7) into account, we thought that the reason why complexes with P_c showed low efficiency might be that undesirable formation could not be changed immediately in the transport process as GA is a noncleavable spacer. Additionally, when RVRR instead of GA was used as a spacer in P_c, the morphologies of nanocomplexes were still spherical, and the conformational change of P_c occurred in HLP_D but not in LP_D. Therefore, different inserted spacer elements of peptides could result in different morphologies and transfection efficiencies of LPD and HLPD formations. The enhanced cellular uptake and transfection efficiency of LP_D and HLP_D demonstrated that the negative effect of RVRR on the intracellular delivery efficiency was lower than that of GA. Thus, the crystal morphologies of complexes and some conformational changes in multifunctional peptides might shield their penetrating function or weaken their penetrating ability. However, when the multifunctional peptide was formed without spacer, the strong cell-penetrating function of KRPTMRFRYTNPMK and tightly wrapping nucleic acid capability of R_N could be fully manifested in LP_D and HLP_D. Moreover, the LP_D and HLP_D formulations showed unified spherical morphology, and the P_c in the ternary and Q-complexes maintained unchangeable configuration. The function of HA was mainly in shielding the positive charge and pushing the nanoparticles to bind to the tumor cells. LP_D and HLP_D transport DNA mainly through dynamin-dependent and CD44-mediated independent pathways (Figure 6), which is completely different from the traditional drug delivery system utilizing HA for both targeting CD44 receptor and entering the cell through CD44-mediated pathway.15,22,23

Conclusion

We rationally designed a stable delivery platform for in vivo nucleic acid delivery based on modular design, which integrates a specific tumor lineage-homing cell-penetrating peptide and gene-condensing peptide into a multifunctional peptide that can both envelope nucleic acids and penetrate the tumor cell membrane. This study has proven that spacers in the peptide could decrease and prevent the intracellular
transfection efficiency through reversing the complexes morphology from sphere to crystal lamellae and changing the peptide configuration. These findings will be important in understanding the formulation of Q-complexes and beneficial in designing an improved gene delivery system in the future.

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**Disclosure**

The authors report no conflicts of interest in this work.

**References**

Supplementary materials

Figure S1 (A–D) The results of hydrodynamic size and zeta potential measurements of ternary complexes and Q-complexes wrapping pGL3 plasmid.

Notes: The data is the weight ratio of each component in Q-complexes. For example, “HLP-D-1:8:1” refers to hyaluronic acid, cationic liposome, P, and DNA at 1:8:1 weight ratios. LP: a cationic liposome, multifunctional peptide, and DNA at optimized ratios; HLP: H represents hyaluronic acid, L represents cationic lipidosome that was composed of DOTAP/DOPC at a 1:1 weight ratio, P represents peptide (P4–P4 refers to the different peptide used), and D represents DNA.

Abbreviation: PDI, polydispersity index.

Figure S2 Electrophoretic mobility in 1% agarose gel for ternary complexes and Q-complexes wrapping pGL3 plasmid.
Effect of inserted spacer in hepatic cell-penetrating peptide