Expression of Ovol2 is related to epithelial characteristics and shows a favorable clinical outcome in hepatocellular carcinoma

Hui Fu1,2,*
Lisha Qi1,2,3,*
Lu Chen1,2
Yuchao He1,2
Ning Zhang1,2
Hua Guo1,2

1Department of Tumor Cell Biology, National Clinical Research Center for Cancer, Tianjin Medical University Cancer Institute and Hospital, 2The Key Laboratory of Tianjin Cancer Prevention and Treatment, National Clinical Research Center for Cancer, 3Department of Pathology, National Clinical Research Center for Cancer, Tianjin Medical University Cancer Institute and Hospital, Tianjin, People’s Republic of China

*These authors contributed equally to this work

Abstract: Metastasis involves epithelial–mesenchymal transition (EMT) and mesenchymal–epithelial transition. Ovol2 belongs to the Ovo-like family (Ovol) of evolutionarily conserved zinc-finger transcription factors that regulate gene expression in various differentiation processes. Recent studies have demonstrated that Ovol2 affects mesenchymal–epithelial transition by inducing the expression of miR-200 in a range of human cancers. Downregulated Ovol2 expression is involved in the invasion and metastasis of breast and prostate cancers, but little is known about its expression and prognostic value in other cancers, including hepatocellular carcinoma (HCC). This study was designed to explore the clinical and prognostic significance of Ovol2 in patients with HCC. The expression of Ovol2 in tumor samples from patients with HCC and HCC cell lines was examined using Western blotting, real-time polymerase chain reaction, and immunohistochemistry. The expression levels of EMT-related markers, including E-cadherin, N-cadherin, and vimentin, were detected in relation to Ovol2 expression. The prognostic significance of Ovol2 in patients with HCC was statistically analyzed by Kaplan–Meier and Cox regression analyses. Ovol2 expression was significantly lower in HCC tissues than in adjacent noncancerous tissues. Low expression of Ovol2 was detected in HCC tissues with poor histological differentiation, microvascular invasion, and cirrhosis. A significant relationship was observed between Ovol2 and EMT marker expression levels. Kaplan–Meier analysis showed that overall survival was significantly worse in patients with HCC with low Ovol2 expression, indicating that Ovol2 deletion was an independent predictor of unfavorable prognosis in patients with HCC. Elevated Ovol2 expression may suppress HCC cell invasion and metastasis via restricting EMT.

Keywords: Ovol2, hepatocellular carcinoma, prognosis, epithelial–mesenchymal transition, mesenchymal–epithelial transition

Introduction

Globally, hepatocellular carcinoma (HCC) remains the sixth most common cancer and third for cancer-related deaths, with 86% occurring in developing countries (~50% in the People’s Republic of China).1,2 Increasing prevalence rates of HCC have been observed worldwide.3,4 However, most HCCs are generally diagnosed at advanced stages.5 Although some advances in surgical technologies and medical management have been reported, the clinical outcome remains unsatisfactory because of the high incidence of recurrence and metastasis after hepatic resection.6,7 Therefore, identifying new sensitive and reliable biomarkers for the diagnosis, prognosis, and target therapy of HCC is extremely important. Significant efforts have been made to elucidate the molecular mechanism of HCC metastasis.
Epithelial–mesenchymal transition (EMT) is a critical cellular process required for embryonic development, adult wound healing, chronic inflammation, and hypoxia. During EMT, epithelial cells lose their apicobasal polarity, which is maintained by a cytoskeleton structure, apical tight junctions, and cell junction, and become mesenchymal cell types. EMT is characterized by the downregulation of epithelial markers, such as E-cadherin, and upregulation of mesenchymal markers (eg, fibronectin, vimentin, and N-cadherin). Mesenchymal–epithelial transition (MET) describes the reverse process. Cancer cells undergo EMT to enable metastasis and invasion, which can be triggered by secreted growth factors and cytokines. The metastatic cascade is a multifaceted process, in which EMT mediates the initial transformation from benign to invasive carcinoma, and MET is critical for colonization in the later stages of metastasis. Despite the widely recognized implications for EMT in cancer progression, rigorous functional studies on MET in the metastatic cascade are scarce.

One candidate that may induce tumor cells to undergo MET is Ovo-like 2 (Ovol2), a member of the Ovol family of zinc-finger transcription factors (TFs) known to prevent mesenchymal transdifferentiation and maintain epithelial identity. Functional studies in human and selected model organisms (eg, *Caenorhabditis elegans*, *Drosophila*, zebrafish, chick, and mice) have corroborated that Ovol genes are involved in the epithelial development and differentiation of a wide variety of organs from epidermis, hair follicles, and kidney to male germ cells. The Ovol family (including Ovol1, Ovol2, and Ovol3) of zinc-finger TFs functions downstream of key developmental pathways (eg, Wnt, epidermal growth factor, and bone morphogenetic protein/transforming growth factor beta). During mammary morphogenesis, Ovol operates as a “critical molecular brake on MET” by preventing the terminal end bud cells that have gained partial plasticity from undergoing complete EMT. Studies have indicated that Ovols affect MET of prostate cancer (PC) and breast cancer (BC) and regulate cancer progression. Ovol2 suppresses nearly all known EMT-inducing TFs by directly binding to their promoters. Watanabe et al reported that Ovol2 expression levels are closely related to the overall survival (OS) and metastasis-free survival of postoperative patients with BC. However, no studies exist regarding the expression levels and prognostic value of Ovol2 in other cancers, including HCC. In the present study, we detected the expression dynamics of Ovol2 in HCC cells and tissues, analyzed the relationships among Ovol2 expression, clinicopathological features, and EMT-related markers, and evaluated the significance of Ovol2 in the prediction of HCC prognosis.

**Materials and methods**

**Patients and samples**

The study samples included 24 frozen tissues for Western blotting and quantitative real-time polymerase chain reaction (PCR). Furthermore, 74 formalin-fixed, paraffin-embedded tissues (among which there were 33 sets of matched HCC tissues and noncancerous tissues) were collected from patients who underwent hepatectomy for HCC between 2014 and 2015 at the Department of Liver Cancer, Tianjin Medical University Cancer Institute and Hospital. Frozen HCC tissues were collected immediately after resection. The matched noncancerous liver tissue without evidence of liver metastases was acquired at least 10 cm away from the tumor. Tissues from the tumor center (c), the tumor invasion front (e2), and the middle between the tumor center and tumor invasion front (e1) were obtained, frozen in liquid nitrogen, and stored at −80°C for further preparation. Written informed consent was obtained from all patients enrolled in this study. This research is in compliance with the Helsinki Declaration and was approved by the Ethics Committees of Tianjin Medical University.

**Western blotting and quantitative real-time PCR**

Frozen tissues were minced into 1 mm cubes. Tissues or cells were lysed with ice-cold Triton X-100 buffer (40 mM Tris, 120 mM NaCl, 1% Triton X-100, 1 mM NaF, and 1 mM NaVO₄) added with a protease inhibitor cocktail (Hoffman-La Roche Ltd., Basel, Switzerland). The tissues or cell lysates were then centrifuged at 15,000×g at 4°C for 15 minutes, and the tissue concentrations were measured with a bicinchoninic acid assay. Approximately 60 mg of protein was separated by 8% or 10% sodium dodecyl sulfate polyacrylamide gel electrophoresis and transferred to a polyvinylidene difluoride membrane (EMD Millipore, Billerica, MA, USA), which was blocked with 5% nonfat milk. The membrane was then incubated with primary antibodies. Signals were developed with electro-chemi-luminescence (EMD Millipore) after incubation with horseradish peroxidase-conjugated secondary antibody. The antibodies used in Western blotting are listed in Table S1.

Total RNA was extracted from tissues and cells using Trizol (Thermo Fisher Scientific, Waltham, MA, USA) according to the manufacturer’s protocol. Reverse transcription was performed with a kit (TaKaRa, Dalian, People’s Republic of China) according to the manufacturer’s protocol. A forward primer specific for Ovol2 (5′-CAGT...
TGCTCACATTCAACAAAG-3’) and a reverse primer (5’-GAAATACCCTCTCTCTCGTG-3’) were used. Quantitative real-time polymerase chain reactions were performed with 1 μl of cDNA and SYBR Green Real-time PCR Master Mix (TaKaRa). The relative gene expression levels were analyzed using the 2−ΔΔCt method, with glyceraldehyde-3-phosphate dehydrogenase as an endogenous control gene.

Tissue microarrays and immunohistochemistry analysis

Approximately 2 mm-thick formalin-fixed, paraffin-embedded tissue cores were obtained from representative cancerous and surrounding tissues to construct tissue microarray (TMA) slides. TMA slides were deparaffinized and rehydrated according to standard protocols. For antigen retrieval, the sections were boiled under high pressure in citrate buffer (pH 6.0) for 3 minutes. Endogenous peroxidase was quenched with 3% H2O2 for 10 minutes at room temperature. After rinsing the sections with phosphate buffered saline for 3 minutes, the primary antibodies were incubated overnight at 4°C, followed by incubation with horseradish peroxidase-conjugated goat antimouse immunoglobulin G and goat antirabbit immunoglobulin G (dilution 1:200; Zhongshan, Beijing, People’s Republic of China) for 1 hour. Staining was performed by incubating for 1 minute with 3,3′-diaminobenzidine. The slides were counterstained with hematoxylin, followed by dehydration and mounting. The percentages for positive cells were scored as follows: 0 (0%–25%), 1 (26%–50%), 2 (50%–75%), and 3 (76%–100%). Staining intensity was stratified into the following three categories: 0 (negative), 1 (weak), and 2 (strong). The sum of the above two scores was defined as the final immunohistochemistry (IHC) staining score as follows: 0–1, low expression; 2–3, moderate expression; and 4–5, high expression. The antibodies used in IHC analysis are listed in Table S1.

Cell lines

Five HCC cell lines (Huh7, Hep3B, HCC-LM3, MHCC97L, and MHCC97H), purchased from the American Type Culture Collection (Manassas, VA, USA), were maintained in Dulbecco’s Modified Eagle Medium – high glucose (Hyclone Laboratories Inc, Novato, CA, USA) supplemented with 10% fetal bovine serum and 1% glutamine Pen-Strep solution at 37°C and 5% CO2.

Statistical analysis

Statistical analysis was conducted using SPSS 16.0 software (SPSS Inc., Chicago, IL, USA). Wilcoxon matched-pairs test was used to evaluate the significance of Ovol2 expression in HCC and adjacent noncancerous tissues. Chi-square test and Pearson’s test were applied to assess the correlation among Ovol2 expression, clinicopathological characteristics, and EMT biomarker expression. Univariate and multivariate analyses were performed with the Cox proportional hazards regression model. All data were expressed through at least three independent determinations. Statistical significance was initially set at values of P<0.05.

Results

Expression of Ovol2 was downregulated in HCC tissues

To identify the expression levels and role of Ovol2 in HCC, we analyzed the protein (n=7) and messenger RNA (mRNA) (n=10) levels in fresh tissues from patients with HCC using Western blotting and real-time PCR. The protein levels of Ovol2 exhibited a significant decrease in HCC tumor tissues compared with adjacent noncancerous tissues (Figure 1A). Real-time PCR showed low mRNA levels in ten HCC tissues and high mRNA levels in the matched noncancerous tissues (Figure 1B), because EMT occurs predominantly at the invading tumor edge. We also detected Ovol2 expression levels in compartment-specific tissues from the tumor invasion front (e1 and e2) and tumor center (c) (Figure 1C). A significant decrease in the expression of Ovol2 was noted in the tumor invasion front compared with the tumor center. We then examined Ovol2 expression in 33 sets of HCC and adjacent nontumorous tissues by IHC. The expression of Ovol2 by immunoreactivity with the specific monoclonal antibody was generally localized in both the nucleus (pointed with red arrowheads) and cytoplasm of HCC cells (Figure 1D). This was consistent with the previous study, in which, Ito et al33 assessed the immunoreactivity of Ovol2 in hair bulb and in pilomatricoma and yielded the similar results. The distribution pattern of this protein expression may be related with the differentiation of the tumor. Similar findings were also documented in the expression pattern of ZEB2, another EMT TF, in HCC, which displayed both cytoplasmic and nuclear staining.30 The underlying mechanisms still need further to be clarified. Ten (30.3%) HCC tissues were positively stained for Ovol2, whereas 23 (69.7%) matched nontumorous tissues were positively stained for Ovol2 (Figure 1D) (P<0.001; Table 1).

Relationship between Ovol2 expression and clinicopathologic features of HCC

Considering the low Ovol2 mRNA and protein expressions in HCC tissues, we investigated the relationship between Ovol2 expression and clinicopathological features, such as age, sex, tumor size, histological differentiation, microvascular invasion, clinical stage, serum alpha fetal protein,
hepatitis B virus, hepatitis C virus, cirrhosis, and satellite markers (Table 2). As shown in Table 2, 74 patients with HCC were subdivided into the following three groups based on the Ovol2 expression levels: 55 patients with no expression (74.32%, negative), eight patients with weak expression (10.81%, weak), and eleven patients with strong expression (14.86%, strong). Ovol2 protein expression was significantly correlated with histological differentiation ($P=0.016$), microvascular invasion ($P=0.033$), and cirrhosis ($P=0.035$).
No significant association was found with age, sex, tumor size, clinical stage, serum alpha fetoprotein, hepatitis B virus surface antigen positivity, hepatitis C virus surface antigen positivity, and satellite markers.

**Correlation of Ovol2 expression with expression of the EMT indicator proteins**

EMT plays an important role in invasion and metastasis. E-cadherin and N-cadherin, two most widely distributed members of the cadherin family, mediate cell–cell adhesion, tumor cell migration, and tumor invasion.

As a hallmark of EMT, E-cadherin is cleaved at the plasma membrane and subsequently degraded during the destabilization of adherents junctions. Loss of E-cadherin is closely related to the metastasis of lung cancer, liver cancer, PC, and BC. Specifically, the upregulation of N-cadherin balances the decreased expression of E-cadherin in the metastasis of tumor cells. N-cadherin is crucial for the invasion and migration of tumor cells, thereby contributing to tumor cell metastasis and invasion. Vimentin is widely expressed in fibroblasts, endothelial cells, and lymphocytes in interstitial cells but undetectable in normal epithelial cells. As a canonical symbol of EMT, vimentin was also observed in various epithelial tumors and closely related to tumor cell differentiation, invasion, and metastasis.

To detect the relationship between the expression of Ovol2 and the expression of EMT markers, IHC for E-cadherin, N-cadherin, vimentin, and Ovol2 was performed on 74 cases.
of formalin-fixed, paraffin-embedded TMA slides of human HCC tissues. As shown in Figure 2A, positive N-cadherin staining appeared in the cytomembrane or cytoplasm of tumor cells; positive vimentin staining was present in the cytoplasm of mesenchymal cells; positive E-cadherin staining was mainly detected in the cytomembrane of tumor cells; and positive Ovol2 staining was detected in both the nucleus and cytoplasm of tumor tissues. The histological grade of HCC is a significant prognosis factor after surgery. Poorly differentiated HCCs are associated with higher microvascular invasion and worse survival compared with moderately and well-differentiated HCCs.9,28 In the study,
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expression of Ovol2 in hcc
the histological differentiation of three patients were identified as well differentiated (++) , 45 patients were identified as moderately differentiated (+), and 26 patients was identified as poorly differentiated (−). When analyzed in detail, we found that positive N-cadherin staining and vimentin staining were mainly detected in poorly differentiated (−) HCC. Positive E-cadherin and Ovol2 were mainly detected in well-differentiated (++) HCC. As shown in Table 3, the expression of Ovol2 was positively correlated with E-cadherin (P=0.038) and negatively correlated with N-cadherin (P=0.023), whereas no significant difference was found in positive vimentin expression between different positive Ovol2 staining degrees (P>0.05). However, we detected negative Ovol2 expression in poorly differentiated tumor tissues with strong staining for vimentin (8/8) (Figure 2A) (Table 3). Using Western blotting, we found that Ovol2 was highly expressed in epithelial HCC cell lines (Huh7 and Hep3B) compared with mesenchymal HCC cell lines (MHCC97L, MHCC97H, and LM3) (Figure 2B). Thus, we further confirmed that the expression of Ovol2 is related to epithelial characteristics of HCC cells. In four independent clinical samples, patients with higher Ovol2 expression were correlated with higher E-cadherin expression, whereas lower expression levels of N-cadherin, vimentin, and Ovol2 repressed EMT-inducing TFs, including Snail and Slug (Figure 2C).

Correlation of Ovol2 expression with the survival of patients with HCC
Kaplan–Meier survival analyses for the 74 patients were performed to investigate the association between Ovol2 expression and survival. Patients with high Ovol2 levels had better prognosis than those with low Ovol2 levels (P<0.05) (Figure 3). Univariate analyses (Table 4) indicated that OS was significantly correlated with tumor multiplicity (P=0.037) and Ovol2 expression (P=0.036). Multivariate Cox regression analysis demonstrated that Ovol2 (HR = 4.019, 95% CI, 0.871–9.551; P=0.047) expression could be an independent marker for OS. Furthermore, tumor multiplicity was also an independent predictor for OS (Table 4).

Discussion
Evidence from clinical studies suggested that EMT has been increasingly recognized to occur during the progression of various carcinomas.12,29–31 Accumulating evidence suggested that EMT is a key physiological process in HCC recurrence and metastasis. Expression levels of EMT-related markers are associated with HCC recurrence and metastasis.8 Multiple pro-EMT TFs, including Twist and Snail, are expressed in ~40%–70% of HCC and associated with poor prognosis.25,32 In this situation, an urgent need exists to

Table 3 Correlation between expression of Ovol2 and epithelial–mesenchymal transition-associated proteins

<table>
<thead>
<tr>
<th>Variable</th>
<th>Total, %</th>
<th>Ovol2 expression, n (%)</th>
<th>χ²</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Negative</td>
<td>Weak</td>
<td>Strong</td>
</tr>
<tr>
<td>E-cadherin expression</td>
<td></td>
<td>61</td>
<td>49 (80.3)</td>
<td>5 (8.2)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>13</td>
<td>6 (46.2)</td>
<td>3 (23.1)</td>
</tr>
<tr>
<td>Vimentin expression</td>
<td></td>
<td>66</td>
<td>47 (71.2)</td>
<td>8 (12.1)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>8</td>
<td>8 (100.0)</td>
<td>0 (0.0)</td>
</tr>
<tr>
<td>N-cadherin expression</td>
<td></td>
<td>47</td>
<td>31 (66.0)</td>
<td>5 (10.6)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>27</td>
<td>24 (88.9)</td>
<td>3 (11.1)</td>
</tr>
</tbody>
</table>

Note: *P-value < 0.05 was considered to be significant.

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Discussion
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identify treatment strategies that reduce the ability of tumor cells to undergo EMT.

Metastasis is believed to begin when some cancer cells of the primary tumor undergo EMT and migrate to distant organs, where they undergo MET and grow into secondary tumors. Understanding the basic principles of the interconnection between EMT/MET is crucial in preventing cancer metastasis. During EMT, epithelial cells downregulate the expression of E-cadherin, occludin, tight junction protein 1, or cytokeratin but upregulate the expression of N-cadherin, vimentin, zinc-finger domain proteins (SNAI1/SAIL, SNAI2/SLUG, and ZEB2/SIP1), matrix metalloproteinases, and the basic helix-loop-helix domain protein Twist1.

Several oncogenic signaling pathways (ie, peptide growth factors, Ras, integrin, Wnt/β-catenin, and Notch) were confirmed to induce MET. However, the mechanisms mediating MET are rarely understood. During MET, the epithelium-associated genes are unregulated, and the mesenchymal-associated genes are downregulated. C-met has been reported to induce MET in cancer cells. Vincan et al demonstrated that Frizzled-7 (FZD7) is necessary for MET of colorectal cancer cells, because loss of FZD7 results in the persistence of a mesenchymal state (increased SNAI2/decreased E-cadherin). The role of DNA methylation in MET induction has been recently highlighted as well. Snail plays an important role in the DNA methylation of E-cadherin, and stable RNA interference of Snail expression results in complete MET in carcinoma cell lines.

Current studies described Ovol2 as active gatekeepers acting both as a driver of MET and a brake holder of EMT. Roca et al reported that Ovol2 influence MET by interacting with AP1, STAT1, STAT3, and nuclear factor kappa B TFs in BC and PC; furthermore, the AP1/MYC TF pair plays an important role in regulating gene expression in MET. Roca et al recently revealed the novel role of Ovols as important regulators of MET in BC and PC cells by repressing ZEB1 and inducing ESRP1 and miR-200. Inhibition of Ovols can induce complete MET, and their overexpression can lead to complete MET. Ovol2, a well-studied regulator in various differentiation processes, can drive MET through the suppression of nearly all known EMT TFs by directly binding to their promoters in multiple cancers. Ovol2-null mice die during midgestation, whereas Ovol2-deficient mammary stem/progenitor cells are prone to undergo EMT and exhibit reduced morphogenic and regenerative capacity.

A previous study has shown that the expression of Ovol2 is positively correlated with the OS and metastasis-free survival of postoperative patients with BC. In this study, we present novel evidence significant for understanding the correlations between the expression levels of Ovol2 and EMT-associated markers in HCC. Our results suggested that Ovol2 may be used as a prognostic factor for HCC.

We first investigated Ovol2 expression in patients with HCC and further analyzed the relationship between the expression of Ovol2 and the expression of EMT indicator proteins. Furthermore, the relationship between the expression of these proteins and clinicopathological variables was determined. We found that Ovol2 expression was lower in HCC tissues than in adjacent noncancerous tissues (Figure 1). Notably, the expression levels of Ovol2 were much higher in the tumor centers

<table>
<thead>
<tr>
<th>Prognostic factors</th>
<th>Univariate analysis</th>
<th>Multivariate analysis</th>
</tr>
</thead>
<tbody>
<tr>
<td>HR (95% CI)</td>
<td>P-value</td>
<td>HR (95% CI)</td>
</tr>
<tr>
<td>Age (&lt;60/&gt;= 60, years)</td>
<td>0.777 (0.442–1.364)</td>
<td>0.379</td>
</tr>
<tr>
<td>Sex</td>
<td>1.448 (0.522–4.017)</td>
<td>0.478</td>
</tr>
<tr>
<td>Tumor size</td>
<td>1.139 (0.665–1.951)</td>
<td>0.636</td>
</tr>
<tr>
<td>Microvascular invasion</td>
<td>1.233 (0.696–2.187)</td>
<td>0.473</td>
</tr>
<tr>
<td>Clinical stage</td>
<td>1.291 (0.646–2.579)</td>
<td>0.470</td>
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<tr>
<td>Cirrhosis</td>
<td>0.931 (0.546–1.588)</td>
<td>0.793</td>
</tr>
<tr>
<td>Differentiation</td>
<td>0.688 (0.200–2.366)</td>
<td>0.553</td>
</tr>
<tr>
<td>AFP</td>
<td>1.020 (0.570–1.825)</td>
<td>0.948</td>
</tr>
<tr>
<td>HBV</td>
<td>0.871 (0.192–3.959)</td>
<td>0.859</td>
</tr>
<tr>
<td>Satellite</td>
<td>1.378 (0.721–2.632)</td>
<td>0.332</td>
</tr>
<tr>
<td>Tumor multiplicity</td>
<td>1.765 (1.036–3.008)</td>
<td>0.037*</td>
</tr>
<tr>
<td>Ovol2</td>
<td>0.231 (0.049–1.087)</td>
<td>0.036*</td>
</tr>
<tr>
<td>E-cadherin</td>
<td>0.327 (0.042–2.549)</td>
<td>0.286</td>
</tr>
<tr>
<td>N-cadherin</td>
<td>1.038 (0.345–3.125)</td>
<td>0.947</td>
</tr>
<tr>
<td>Vimentin</td>
<td>2.484 (0.734–8.399)</td>
<td>0.143</td>
</tr>
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Notes: *P-value <0.05 was considered to be significant. Univariate and multivariate analyses: Cox proportional hazards regression model.

Abbreviations: AFP, alpha fetal protein; HBV, hepatitis B virus; HR, hazard ratio; CI, confidence interval.
than those at the edges. Clinically, patients with HCC with high Ovol2 expression had a better prognosis than those with low expression. These results indicated that Ovol2 expression may be correlated with HCC progression.

The loss of E-cadherin (CDH1) and the upregulation of vimentin and N-cadherin are widely considered the hallmarks of EMT.12,25,42,43 Moreover, TFs that are central to promote EMT programs, such as Snail, Slug, Twist, and ZEB1, are regarded as important markers of EMT.44 Our study showed that Ovol2 expression levels in HCC were positively correlated with the expression of the epithelial marker E-cadherin and inversely correlated with the expression of the mesenchymal marker N-cadherin. Vimentin expression in invasive cancer cells was observed, particularly in poorly differentiated tissues.23,27,45 Thus, further studies require a large sample size, especially for cases with poor differentiation, to confirm the relationship between the expression levels of Ovol2 and vimentin in HCC. Consistent with the findings of Western blotting of clinical HCC tissues, we observed that N-cadherin, vimentin, Snail, Slug, Twist, and ZEB1 were downregulated, whereas E-cadherin was upregulated in tissues with higher expression levels of Ovol2. Altogether, these data demonstrated that Ovol2 inhibited EMT of HCC cells. Tumor multiplicity and Ovol2 expression were valuable predictors for OS by univariate analysis. When adjusted by multivariate analysis using Cox proportional hazard model, Ovol2 expression and tumor multiplicity were independent predictors for OS.

Conclusion
We demonstrated for the first time the expression patterns of Ovol2 in HCC cell lines and tissues. Ovol2 downregulation was correlated with upregulated mesenchymal markers and could predict an unfavorable prognosis. Although the precise mechanisms of Ovol2 function in tumor development require further investigation, our results confirmed the value of Ovol2 as a novel and reliable marker and a potential therapeutic target for HCC.

Acknowledgments
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Disclosure
The authors report no conflicts of interest in this work.

References


Supplementary material

Table S1 Primary antibodies for Western blotting and immunohistochemistry

<table>
<thead>
<tr>
<th>Antibody</th>
<th>WB</th>
<th>IHC</th>
<th>Company</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ovol2</td>
<td>1:1,000</td>
<td>–</td>
<td>Santa Cruz Biotechnology</td>
</tr>
<tr>
<td>Ovol2</td>
<td>–</td>
<td>1:50</td>
<td>Novus Biologicals</td>
</tr>
<tr>
<td>GAPDH</td>
<td>1:3,000</td>
<td>–</td>
<td>Santa Cruz Biotechnology</td>
</tr>
<tr>
<td>E-cadherin</td>
<td>1:2,000</td>
<td>1:100</td>
<td>BD Biosciences</td>
</tr>
<tr>
<td>Vimentin</td>
<td>1:6,000</td>
<td>1:300</td>
<td>BD Biosciences</td>
</tr>
<tr>
<td>Slug</td>
<td>1:200</td>
<td>–</td>
<td>Cell Signaling Technology</td>
</tr>
<tr>
<td>Snail</td>
<td>1:200</td>
<td>–</td>
<td>Cell Signaling Technology</td>
</tr>
<tr>
<td>Twist</td>
<td>1:4,000</td>
<td>–</td>
<td>Abcam</td>
</tr>
<tr>
<td>ZEB1</td>
<td>1:2,000</td>
<td>–</td>
<td>Santa Cruz Biotechnology</td>
</tr>
</tbody>
</table>

Abbreviations: IHC, immunohistochemistry; WB, Western blotting.