Cataract induction by administration of nitroglycerin in cardiac patients through imbalance in redox status

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Purpose: The objective of this study was to evaluate the role of nitroglycerin in the pathogenesis of cataract.

Design: Prospective study.

Patient and methods: This study was performed in adults from tertiary Saudi Arabian hospitals (34 males and 26 females in each group, aged from 40 to 60 years), who were divided into four groups with an equal number of subjects (control group, cardiac group, idiopathic cataract group, and a group of cardiac patients using nitroglycerin and with cataracts). Fasting glucose concentrations, blood glycated hemoglobin levels, lipid profiles, and levels of nitrite, conjugated dienes (CD), thiobarbituric acid reactive substances (TBARS), superoxide dismutase (SOD), and reduced glutathione (GSH) were determined.

Results: Treatment of cardiac patients with nitroglycerin produced an imbalance in their systemic redox status, leading to the development of cataracts, which was reflected by a significant increase in the levels of nitrite, CD, and TBARS and a significant decrease in SOD activity and GSH, compared with idiopathic cataract patients. The results of correlation studies and multiple regression analysis revealed a significant positive correlation between different biochemical parameters (GSH, SOD, TBARS, CD, and nitrite) in the blood and lens in both idiopathic cataract patients and cardiac patients treated with nitroglycerin.

Conclusion: The study points to the relative and predictive effects of nitric oxide derived from nitroglycerin in the development of cataract in the presence of the oxidative stress induced by nitroglycerin treatment.

Keywords: nitroglycerin, oxidative stress, cataract, nitrite, lens

Introduction
Nitroglycerin (glicyeryl trinitrate, GTN) has been used to treat angina pectoris, myocardial infarction, and heart failure for a significant period of time, and still acts as one of the main therapies for these conditions.1

Several metabolic pathways have been identified through which GTN is enzymatically reduced to produce nitric oxide (NO) or NO precursors.2,3 These enzymes include xanthine oxidase,4 glutathione S-transferase, and, more recently, mitochondrial aldehyde dehydrogenase (ALDH-2).2 Indeed, the main action of ALDH-2 in the mitochondrial electron transport chain is that it mediates the conversion of GTN into nitrite (NO2−) in the mitochondria.5 Under normal conditions, GTN is bioactivated by ALDH-2 to yield 1,2-glyceryl dinitrate and nitrite, which are then converted to NO, S-nitrosothiol, or a related species by the mitochondrial respiratory chain or acidic disproportionation, to induce vasodilatation. In addition, GTN induces vasodilatation through...
activation of the soluble guanylyl cyclase/cyclic guanosine monophosphate (cGMP)/cGMP-dependent protein kinase I pathway without production of free radicals. Both enzymatic and nonenzymatic mechanisms for GTN metabolism and biotransformation have been reported. Many enzymes are involved in GTN metabolism and biotransformation, including ALDH-2, the cytochrome p450 enzymes, xanthine oxidoreductase, and glutathione S-transferases. The reaction of GTN with thiols may also produce vasodilator S-nitrosothiols (SNOs). NO and SNOs activate soluble guanylyl cyclase, causing an elevation in the levels of second messenger cGMP in tissues. cGMP-dependent protein kinase I mediates vasodilation by phosphorylating proteins which regulate cellular Ca2+ levels. GTN can also induce vasodilation through a cGMP-independent pathway.

During bioactivation of GTN through oxidation of essential thiol groups to disulfide, ALDH-2 is inactivated. Restoration of ALDH-2 reductase activity is induced by reducing lipoic acid. Subsequently, the reduced glutathione (GSH)–glutathione reductase system reduces the oxidized form.

Chronic therapeutic use of GTN produces mitochondrial reactive oxygen and reactive nitrogen species (ROS/RNS). The major source of ROS formation is the mitochondria in response to short- and long-term GTN challenges. The diffusion-limited reaction of NO and superoxide produces peroxynitrite. GTN and/or NO byproducts in the mitochondria may lead to uncoupling of respiration and mitochondrial damage. Critical thiols, such as the active-site thiols in ALDH-2, are oxidized by increased production of superoxide and other ROS.

GTN bioactivation is inhibited by oxidative stress through inactivation of ALDH-2 or inhibition of the ALDH-2 repair system, which includes lipoic acid as well as the GSH/GSH reductase system. Moreover, the bioavailability of the vasodilator produced from GTN bioactivation is decreased by superoxide. A characteristic feature of prolonged GTN administration is the formation of GTN tolerance. A possible explanation is either a decrease in GTN bioconversion into NO or a decrease in the bioavailability of GTN-derived NO as a result of enhanced superoxide formation. Both mechanism-based tolerance and cross-tolerance are activated by mitochondrial production of ROS. The contribution of ALDH-2 to GTN tolerance and cross-tolerance remains to be clarified.

Suprapharmacological GTN concentrations cause a significant increase in vascular NO formation, while pharmacological GTN concentrations are not associated with any elevation in NO levels. Increased NO production is possibly related to the “second” GTN pathway, which is not affected by GTN tolerance state. Increased production of RNS causes nitrosylation reactions, which can cause alterations in the structure of proteins and, thus, impair their normal function. Thiol, amine, and heme nitrosylation generates S-nitrosothiols, nitrosyl heme, and N-nitroso compounds, respectively. These reactions suggest new signaling pathways through which NO affects the cell physiology by covalently modifying tissue biomolecules.

Paik and Dillon found modifications of protein structure, which are similar to those found in age-related nuclear cataract, when they incubated nitrite, an end product of NO, with lens α-crystallin. Aminoguanidine, an inhibitor of inducible nitric oxide synthase (iNOS), has been demonstrated to suppress cataractogenesis in animal studies. The objectives of this study were to evaluate the role of NO (as a metabolite of GTN) in the pathogenesis of cataract and to estimate the levels of an NO metabolite (nitrite), lipid peroxidation, superoxide dismutase (SOD), and GSH as an indicator of oxidative stress.

**Methods**

This study was performed in adults from tertiary Saudi Arabian hospitals. Three hundred cardiac patients were considered for this study, of whom 60 patients had not received nitroglycerin and had cataract (2%). One hundred and twenty cardiac patients were selected in accordance with the inclusion and exclusion criteria. They were divided into two groups (cardiac patients who had not received nitroglycerin and cardiac patients who had received nitroglycerin). Other groups included in the current study were healthy control and idiopathic cataract groups. This study was approved by the hospital committee of the tertiary Riyadh Hospital, Kingdom of Saudi Arabia, and written informed consent was obtained from all the patients included in the study. A preliminary examination of all participants consisted of a brief medical history including details of systemic disease, drug administration, cigarette smoking, trauma, and exposure to ultraviolet radiation. A complete ophthalmic history and examination included an assessment of the morphologic type of any cataract present. All participants in this study had similar dietary habits (including low dietary fat, number of daily meals, and a low salt intake). All participants underwent a full clinical examination.

**Inclusion criteria**

Cardiac patients who had been administered nitroglycerin for at least 1 year.
Exclusion criteria
Cardiac patients who had been administered nitroglycerin, but were more than 60 years old; prolonged exposure to sunlight (10 months or more); high blood pressure; obesity; previous eye injury or inflammation; previous eye surgery; antioxidants had not been administered in the previous 6 months; smoking; medication or diseases, which induced oxidative stress in the last 2 years (including prolonged use of corticosteroid medications, diabetes, cancer, asthma, chronic bronchitis, osteoarthritis, and hepatitis).

Sample collection
Approximately 10 mL of venous blood was collected from the antecubital vein of each subject after overnight fasting and was separated into two aliquots: 5 mL in a vacutainer tube without any anticoagulant to obtain the serum and the other 5 mL in a plain vial containing heparin to obtain the plasma. The lenses of affected patients (nucleus plus cortex) were obtained after extracapsular cataract extraction operations and were immediately stored at −70°C until analyzed.

Biochemical analysis
Fasting glucose levels were measured colorimetrically using the glucose oxidase method reported by Trinder. Blood glycated hemoglobin was determined according to the method reported by Little et al using the Helena GLYCO-Tek affinity column method (Helena Laboratories, Beaumont, TX, USA). Nonesterified fatty acid concentrations were analyzed colorimetrically according to the method described by Schuster and Pilz. Serum cholesterol concentrations were estimated according to the method reported by Deeg and Ziegenhorm using a commercial reagent kit (Reactivos Spinreact Company, Girona, Spain). Serum triglyceride concentrations were determined according to the method reported by Fossati and Prencipe using a commercial reagent kit purchased from Reactivos Spinreact Company, Girona, Spain). Serum high-density lipoprotein (HDL) cholesterol concentrations were calculated according to the method reported by Norbert. Serum low-density lipoprotein (LDL) cholesterol levels were calculated using the Friedewald formula (LDL-cholesterol = total cholesterol – triglycerides/5 – HDL-cholesterol).

Nitrite assay
Nitrite was measured using the Griess reaction. Briefly, serum samples were diluted fourfold with distilled water and deproteinized using zinc sulfate (300 g/L) to give a final concentration of 15 g/L. After centrifugation, 100 µL of the supernatant was applied to a microtiter plate well, followed by addition of 100 µL of Griess reagent which was composed of 1 g/L sulfanilamide, 25 g/L phosphoric acid, and 0.1 g/L N-1-naphthylethylenediamine. After 10 minutes of color development at room temperature, the absorbance was measured at a wavelength of 540 nm using a microplate reader (Titertek Multiskan MCC/340; Flow Lab, McLean, VA, USA).

Assay of lipid peroxidation as a measurement of oxidative status
Conjugated dienes assay
Serum conjugated dienes (CD) levels were determined using the method described by Recknagel and Glende. Lipids were extracted using 2:1 (by volume) chloroform–methanol; the extract was evaporated until dry under a stream of nitrogen and then redissolved in cyclohexane. The cyclohexane solution absorbance was measured at 234 nm. The results were expressed as µmol/L.

Thiobarbituric acid reacting substance (TBARS) assay
Serum TBARS concentrations were measured as a lipid peroxidation product using the Kei Satoh method. Serum proteins were precipitated using trichloroacetic acid and the mixture was heated for 30 minutes in a boiling water bath in the presence of thiobarbituric acid in 2 M sodium sulfate. n-Butyl alcohol was used to extract the resulting chromogen and the absorbance of the organic phase was determined at a wavelength of 530 nm.

Enzymatic antioxidant assay – SOD
Estimation of plasma SOD was performed using the method reported by Kakkar et al. A mixture of 1.35 mL of double-distilled water, 0.1 mL of phenazine methosulfate, 50 µL of plasma, 1.2 mL of sodium pyrophosphate buffer (pH 8.3), and 0.3 mL of nitroblue tetrazolium was prepared. The reaction was initiated with 0.2 mL of NADH solution. After incubation at 39°C for 90 seconds, the reaction was terminated using 1 mL of glacial acetic acid. After the addition of 4 mL of n-butanol, the mixture was centrifuged at 4,000 rpm for 10 minutes. The absorbance of the upper butanol layer was recorded at 560 nm.

Nonenzymatic antioxidant assay – reduced GSH
The concentration of GSH was determined using a fluorometric method described by Hissin and Hilf. Briefly, serum samples were deproteinized using an equal volume of 5% trichloroacetic acid. Then, the samples were centrifuged at
10,000× g for 10 minutes. The GSH levels were determined in µmol/L. GSH standard solution or supernatant (200 µL) was added to 2.6 mL of phosphate-ethylenediaminetetraacetic acid buffer (5 mM ethylenediaminetetraacetic acid, 0.1 M sodium phosphate, pH =8) and 200 µL of 1 mg/mL O-phthalaldehyde. The mixture was incubated at room temperature for 45 minutes. Then, fluorescence detection was performed with excitation and emission at 350 and 418 nm, respectively.

**Preparation of lens extract**
Homogenization of the lens was performed on ice using a cold buffer (0.2 M potassium phosphate, 60 mM sodium dodecyl sulfate, 137 mM potassium chloride, pH 7.2). Lipid auto-oxidation was prevented using ethanolic butylated hydroxytoluene solution (2%, v/v). Centrifugation (12,000× g; 40°C; 20 minutes) was performed to remove any insoluble material. Biochemical measurements were carried out on an ultraviolet/visible spectrophotometer (SAFAS 2, Monaco, Monaco).

**Determination of lens nitrite levels**
Nitrite levels were measured using the following spectrophotometric method. Lens samples were homogenized in bidistilled water in a Heidolph glass homogenizer at a neutral pH and centrifuged at 10,000× g for 10 minutes, after measuring the net weight of each lens. These supernatants were incubated in a water bath for 15 minutes at 80°C. Insoluble proteins were separated by centrifugation for 30 minutes at 13,000 rpm. The supernatants were analyzed for nitrite using a modified Griess reaction (1% sulfanilamide solution in 2.5% phosphoric acid and 0.1% naphthylethlenediamine). The reaction product (nitrite) was measured at 541 nm using a spectrophotometer after incubation for 10 minutes at room temperature. The nitrite level was expressed in nanomoles per 100 mg of lens tissue.

**Measurement of lens lipid peroxidation markers**
CD concentrations were assessed in a liposoluble lens extract. In brief, 50 µL of water-insoluble lens material was suspended by vortex-mixing in 1.5 mL of a chloroform–methanol mixture (2:1, v/v) and centrifugation for 5 minutes at 2,000× g (4°C). The lower chloroform layer was removed and evaporated until dry under a stream of nitrogen. The absorbance was measured in a quartz-glass cuvette at λ=234 nm after redissolving the lipid residue in 1 mL of concentrated cyclohexane. The level of TBARS was determined in a hydrosoluble lens extract. The absorbance of a pinkish trimethin complex formed during the reaction of Malondialdehyde (MDA) with 2-thiobarbituric acid (70 mM; NaOH, 50 mM) was determined at λ=532 nm.

**Determination of lens SOD activity**
SOD activity was determined by measuring the rate of inhibition of adrenaline auto-oxidation to adrenochrome. This was continuously monitored at 25°C for 3 minutes at λ=480 nm.

**Determination of lens GSH levels**
Tripeptide GSH is the major intracellular nonenzymatic antioxidant and reducing agent. Hydrosoluble lens extract was deproteinized using 10% metaphosphoric acid. Then, the concentration of GSH was determined using Ellman’s reagent.

**Statistical analysis**
Findings are expressed as mean ± standard deviation (SD) and data were analyzed using the Student’s t-test. One-way analysis of variance was performed followed by a Tukey post hoc test to determine statistically significant differences between groups. A P-value of <0.05 was considered to indicate significant differences. Correlation studies were performed to analyze the relative and predictive effects of NO levels on the development of nitroglycerin-induced cataract in the presence of oxidative stress. The correlations between the studied parameters were assessed to analyze the relative and predictive effects of serum GSH and plasma SOD activity in the development of cataracts in the presence of the oxidative stress induced by nitroglycerin.

**Results**
The basic demographic characteristics of the control participants, idiopathic cataract patients, and cardiac patients who received or did not receive nitroglycerin are presented in Table 1.

The cardiac patients were somewhat younger (<55 years) and had a slightly higher body mass index, on average, than the healthy controls and idiopathic cataract patients.

In the present study, 57% of the participants per group were male (n=34) and 43% were female (n=26). The mean age, sex, educational level, and body mass index were suitably matched between the experimental groups.

The frequency distribution of lifestyle confounding factors was not significantly different between the experimental groups. Twenty percent of the control group (n=12)
patients were highly educated, whereas 80% possessed a low level of education. Twenty-five percent of the cardiac patients were highly educated, whereas 75% possessed a low level of education. Only 17% of the idiopathic cataract patients were highly educated, whereas 83% possessed a low level of education. Twenty-two percent of cardiac patients treated with nitroglycerin were highly educated, whereas 78% were not (Table 1).

Table 2 shows that there were no significant changes among certain biochemical variables between the experimental groups, including fasting blood glucose levels, glycated hemoglobin A, total cholesterol, HDL, LDL, and triglycerides.

In the present study, serum CD levels and TBARS, which are markers for oxidative stress, were found to be significantly increased in the cardiac patients not treated with nitroglycerin and in the idiopathic cataract patients compared with the control group (51% and 53%, and 22% and 15%, respectively). Cardiac patients treated with nitroglycerin showed a significant decrease in the levels of SOD, compared with the cardiac patients not treated with nitroglycerin and the idiopathic cataract patients (10% and 12%, and 62% and 73%, respectively) and the control group (70% and 98%, respectively) (Table 3).

However, the levels of plasma SOD, a marker of enzymatic antioxidant activity, were found to be significantly decreased in the cardiac patients not treated with nitroglycerin and in the idiopathic cataract patients compared with the control group (20% and 22%, respectively). Cardiac patients treated with nitroglycerin showed a significant decrease in the levels of SOD, compared with the cardiac patients not treated with nitroglycerin and the idiopathic cataract patients (37% and 34%, respectively) and the control group (49%) (Table 3).

In addition, the levels of serum GSH, a marker of nonenzymatic antioxidant activity, were found to be significantly decreased in the cardiac patients not treated with nitroglycerin and in the idiopathic cataract patients compared with the control group (10% and 12%, respectively). Cardiac patients treated with nitroglycerin showed a significant decrease in serum GSH, compared with the cardiac patients not treated with nitroglycerin and the idiopathic cataract patients (27% and 32%, respectively) and the control group (40%) (Table 3).

Table 1 Basic demographic characteristics of control, cardiac patients (not treated with nitroglycerin), and cataract patients

<table>
<thead>
<tr>
<th>Item</th>
<th>Control</th>
<th>Cardiac patients (not treated with nitroglycerin)</th>
<th>Idiopathic cataract</th>
<th>Nitroglycerin users with cataract</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>55.5±4.3</td>
<td>54.7±4.9</td>
<td>55.1±4.8</td>
<td>55.1±4.6</td>
</tr>
<tr>
<td>Sex</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male (%)</td>
<td>34 (57)</td>
<td>34 (57)</td>
<td>34 (57)</td>
<td>34 (57)</td>
</tr>
<tr>
<td>Female (%)</td>
<td>26 (43)</td>
<td>26 (43)</td>
<td>26 (43)</td>
<td>26 (43)</td>
</tr>
<tr>
<td>Education</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Low (%)</td>
<td>48 (80)</td>
<td>45 (75)</td>
<td>50 (83)</td>
<td>47 (78)</td>
</tr>
<tr>
<td>High (%)</td>
<td>12 (20)</td>
<td>15 (25)</td>
<td>10 (17)</td>
<td>13 (22)</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>24.1±2.9</td>
<td>24.5±2.8</td>
<td>24.3±3.7</td>
<td>24.8±3.0</td>
</tr>
</tbody>
</table>

Notes: Values are presented as mean ± SD for age and BMI, n=60 for each group. Sex and education are presented as number and percent.

Abbreviations: BMI, body mass index; SD, standard deviation.

Table 2 Biochemical analysis of control, cardiac patients (not treated with nitroglycerin), and cataract patients

<table>
<thead>
<tr>
<th>Item</th>
<th>Control</th>
<th>Cardiac patients (not treated with nitroglycerin)</th>
<th>Idiopathic cataract</th>
<th>Nitroglycerin users with cataract</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fasting blood glucose (mmol/L)</td>
<td>5.4±0.3</td>
<td>5.5±0.1</td>
<td>5.4±0.1</td>
<td>5.3±0.2</td>
</tr>
<tr>
<td>HbA1c (% of hemoglobin)</td>
<td>5.2±0.1</td>
<td>5.1±0.2</td>
<td>5.3±0.1</td>
<td>5.4±0.2</td>
</tr>
<tr>
<td>Total cholesterol (mmol/L)</td>
<td>4.8±0.4</td>
<td>4.7±0.3</td>
<td>4.8±0.8</td>
<td>4.7±0.5</td>
</tr>
<tr>
<td>HDL-cholesterol (mmol/L)</td>
<td>1.2±0.1</td>
<td>1.1±0.2</td>
<td>1.3±0.2</td>
<td>1.2±0.2</td>
</tr>
<tr>
<td>LDL-cholesterol (mmol/L)</td>
<td>3.3±0.1</td>
<td>3.2±0.2</td>
<td>3.1±0.1</td>
<td>3.1±0.2</td>
</tr>
<tr>
<td>Triglyceride (mmol/L)</td>
<td>1.5±0.2</td>
<td>1.5±0.1</td>
<td>1.6±0.2</td>
<td>1.5±0.3</td>
</tr>
</tbody>
</table>

Notes: Values are presented as mean ± SD for each group. *p<0.05 calculated by one-way ANOVA.

Abbreviations: ANOVA, analysis of variance; HbA1c, glycated hemoglobin A; HDL-cholesterol, high-density lipoprotein cholesterol; LDL-cholesterol, low-density lipoprotein cholesterol; SD, standard deviation.
 Serum nitrite levels were found to be significantly higher in the cardiac patients not treated with nitroglycerin and in the idiopathic cataract patients compared with the control group (22% and 18%, respectively). Cardiac patients treated with nitroglycerin showed a significant increase in serum nitrite levels (1.6-fold), compared with the cardiac patients not treated with nitroglycerin and the idiopathic cataract patients (39% and 44%, respectively) and the control group (70%) (Table 3).

Table 4 demonstrates a significant positive correlation between serum CD, serum TBARS, and serum nitrite in the cardiac patients who were not treated with nitroglycerin. On the other hand, a significant negative correlation was found between plasma SOD activity and serum nitrite in the cardiac patients who were not treated with nitroglycerin. A significant negative correlation between plasma SOD activity, serum TBARS, and serum GSH was revealed in the idiopathic cataract patients. In addition, a significant negative correlation was found between serum GSH and serum nitrite in the idiopathic cataract patients. However, a significant positive correlation between serum TBARS, serum nitrite, and serum CD was observed in the idiopathic cataract patients. Cardiac patients treated with nitroglycerin showed a significant positive correlation between plasma SOD activity and serum GSH, between serum TBARS and serum nitrite, and between serum CD, serum TBARS, and serum nitrite. In contrast, a significant negative correlation was found between plasma SOD activity and serum TBARS, serum nitrite, and serum CD in the cardiac patients treated with nitroglycerin. In addition, a significant negative correlation was found between serum GSH and serum nitrite in the cardiac patients treated with nitroglycerin.

The cardiac patients treated with nitroglycerin showed a significant increase in the concentration of lens CD and TBARS, compared with the idiopathic cataract patients.

Table 3 Blood biomarkers of oxidative stress and nitrite in control, cardiac patients (not treated with nitroglycerin), and cataract patients

<table>
<thead>
<tr>
<th>Item</th>
<th>Control</th>
<th>Cardiac patients (not treated with nitroglycerin)</th>
<th>Idiopathic cataract</th>
<th>Nitroglycerin users with cataract</th>
<th>F value*</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>SOD (U/mL)</td>
<td>4.0±0.9</td>
<td>3.2±0.8‡</td>
<td>3.1±0.2†</td>
<td>2.0±0.1¹‡</td>
<td>110.3</td>
<td>0.000</td>
</tr>
<tr>
<td>GSH (µmol/mL)</td>
<td>2.1±0.1</td>
<td>1.9±0.1*</td>
<td>1.8±0.1*</td>
<td>1.4±0.3¹*</td>
<td>189.9</td>
<td>0.000</td>
</tr>
<tr>
<td>CD (nmol/mL)</td>
<td>1.7±0.1</td>
<td>2.6±0.4†</td>
<td>2.5±0.1*</td>
<td>2.9±0.1¹‡</td>
<td>292.2</td>
<td>0.000</td>
</tr>
<tr>
<td>TBARS (nmol/mL)</td>
<td>7.1±0.1</td>
<td>8.7±1.2‡</td>
<td>8.1±0.1*</td>
<td>14.1±0.3¹‡</td>
<td>1,523.8</td>
<td>0.000</td>
</tr>
<tr>
<td>Nitrite (µmol/L)</td>
<td>31.4±0.4</td>
<td>38.4±8.8†</td>
<td>37.1±1.0*</td>
<td>53.4±2.2¹‡</td>
<td>525.3</td>
<td>0.000</td>
</tr>
</tbody>
</table>

Notes: Values are presented as mean ± SD, n=40 for each group. ‡P<0.05 calculated by one-way ANOVA followed by Tukey post hoc test. *Significance compared with control group. †Significance compared with idiopathic cataract.

Abbreviations: ANOVA, analysis of variance; CD, conjugated dienes; GSH, reduced glutathione; SD, standard deviation; SOD, superoxide dismutase; TBARS, thiobarbituric acid reacting substances.

Table 4 Correlation between different biochemical parameters in control, cardiac patients (not treated with nitroglycerin), and cataract patients

<table>
<thead>
<tr>
<th>Item</th>
<th>Control</th>
<th>Cardiac patients (not treated with nitroglycerin)</th>
<th>Idiopathic cataract</th>
<th>Nitroglycerin users with cataracts</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bivariate correlation study</td>
<td>r₁₂ −0.05</td>
<td>r₁₂ 0.06</td>
<td>r₁₂ 0.76***</td>
<td>r₁₂ 0.92***</td>
</tr>
<tr>
<td></td>
<td>r₁₁ −0.07</td>
<td>r₁₁ 0.21</td>
<td>r₁₁ −0.79***</td>
<td>r₁₁ −0.92***</td>
</tr>
<tr>
<td></td>
<td>r₁₀ 0.06</td>
<td>r₁₀ −0.55**</td>
<td>r₁₀ 0.73**</td>
<td>r₁₀ −0.89**</td>
</tr>
<tr>
<td></td>
<td>r₁₀ 0.01</td>
<td>r₁₀ 0.08</td>
<td>r₁₀ 0.90**</td>
<td>r₁₀ −0.94**</td>
</tr>
<tr>
<td></td>
<td>r₁₀ 0.05</td>
<td>r₁₀ 0.08</td>
<td>r₁₀ −0.79***</td>
<td>r₁₀ −0.88**</td>
</tr>
<tr>
<td></td>
<td>r₁₀ 0.05</td>
<td>r₁₀ 0.09</td>
<td>r₁₀ −0.59***</td>
<td>r₁₀ −0.96**</td>
</tr>
<tr>
<td></td>
<td>r₁₀ 0.07</td>
<td>r₁₀ 0.01</td>
<td>r₁₀ 0.73**</td>
<td>r₁₀ −0.93**</td>
</tr>
<tr>
<td></td>
<td>r₁₀ 0.01</td>
<td>r₁₀ 0.10</td>
<td>r₁₀ 0.58**</td>
<td>r₁₀ −0.87**</td>
</tr>
<tr>
<td></td>
<td>r₁₀ 0.07</td>
<td>r₁₀ 0.39***</td>
<td>r₁₀ 0.66**</td>
<td>r₁₀ 0.87**</td>
</tr>
<tr>
<td></td>
<td>r₁₀ 0.02</td>
<td>r₁₀ 0.32*</td>
<td>r₁₀ 0.75**</td>
<td>r₁₀ 0.93**</td>
</tr>
</tbody>
</table>

Notes: r₁₂ Correlation between X₁ and X₂; r₁₁ Correlation between X₁ and X₁; r₁₀ Correlation between X₁ and X₀; r₁ Correlation between X₁ and X; r₂ Correlation between X₂ and X₂; r₂ Correlation between X₂ and X; r Correlation between X and X. *Correlation is significant at the 0.01 level (two-tailed). †Correlation is significant at the 0.05 level (two-tailed).
Table 5 Biomarkers of oxidative stress and nitrates in the lens of cataract patients

<table>
<thead>
<tr>
<th>Item</th>
<th>Idiopathic cataract</th>
<th>Nitroglycerin users with cataract</th>
</tr>
</thead>
<tbody>
<tr>
<td>SOD (U/mL)</td>
<td>5.3±0.2</td>
<td>4.7±0.2*</td>
</tr>
<tr>
<td>GSH (µmol/mL)</td>
<td>1.9±0.1</td>
<td>1.3±0.1*</td>
</tr>
<tr>
<td>CD (nmol/mL)</td>
<td>15.4±0.9</td>
<td>18.1±1.4*</td>
</tr>
<tr>
<td>TBARS (nmol/mL)</td>
<td>2.2±0.1</td>
<td>3.3±0.1*</td>
</tr>
<tr>
<td>Nitrite (µmol/L)</td>
<td>3.4±0.3</td>
<td>5.7±0.3*</td>
</tr>
</tbody>
</table>

Notes: Values are presented as mean ± SD, P<0.05, n=60 for each group.

Abbreviations: CD, conjugated dienes; GSH, reduced glutathione; SD, standard deviation; SOD, superoxide dismutase; TBARS, thiobarbituric acid reacting substances.

Table 6 Correlation between the levels of different biochemical parameters in the blood and lens in cataract patients

<table>
<thead>
<tr>
<th>Item</th>
<th>Idiopathic cataract</th>
<th>Nitroglycerin users with cataract</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD</td>
<td>0.88**</td>
<td>0.58**</td>
</tr>
<tr>
<td>Nitrite</td>
<td>0.59**</td>
<td>0.76**</td>
</tr>
<tr>
<td>GSH</td>
<td>0.96**</td>
<td>0.91**</td>
</tr>
<tr>
<td>TBARS</td>
<td>0.43**</td>
<td>0.85**</td>
</tr>
</tbody>
</table>

Note: **Correlation is significant at the 0.01 level (two-tailed).

Abbreviations: CD, conjugated dienes; GSH, reduced glutathione; SOD, superoxide dismutase; TBARS, thiobarbituric acid reacting substances.

Discussion

The current study was designed to evaluate the role of NO (as a metabolite of nitroglycerin) in the pathogenesis of cataract and to determine the levels of an NO metabolite (nitrite), lipid peroxidation, SOD, and reduced glutathione (GSH) as an indicator of oxidative stress.

NO and NO-derived RNS can damage and kill cells through many mechanisms such as inactivation of respiratory chain complexes, inhibition of DNA and protein synthesis, decreasing the activity of antioxidative enzymes, intracellular GSH, and adenosine triphosphate, and inactivation of genes by inducing DNA methylation, and irreversible oxidative modification and degradation of DNA, proteins, and membrane lipids. There are several reports concerning the effects of NO in the eye. NO has been implicated in the development of certain ocular diseases, such as glaucoma and retinal diseases. However, the action of NO in the lens has not yet been completely established. Yamamoto et al have studied the localization of nitric oxide synthase in the lens epithelium. However, few published studies have investigated the relationship between NO and cataract.

In the current study, serum nitrite levels were found to be significantly increased in the idiopathic cataract patients compared with the control group (18%). Cardiac patients who received nitroglycerin showed a significant increase in serum nitrite (1.6-fold), compared with the idiopathic cataract patients (44%) and the control group (70%). This finding is in agreement with Ornek et al, who found that NO has a role in the development of cataract.

Inomata et al found that aminoguanidine, a nitric oxide synthase inhibitor, inhibits the formation of cataract in a hereditary cataract model. Previous animal studies have demonstrated that there is an association between nuclear cataract and the NO molecule, but observed lower NO levels in such cataracts compared with those in other tissues. Inomata et al have demonstrated that increased levels of NO may freely affect the lens and induce the formation of cataract.

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systemic hypertension. These findings suggest a possible role of systemic diseases, especially those involving elevated NO levels, in the pathology of lens opacification in human eyes. Elevated NO levels in patients with cataractous lenses, especially in patients with posterior subcapsular cataract and hypertension, suggest a possible role for NO in the development of human cataracts. It can be concluded that NO is likely involved in the pathogenesis of human cataracts. Increased generation of free radicals and decreased antioxidant activity have been proposed to play a major role in the development of age-related cataract. In addition, Micelli-Ferrari et al found that the degree of lens opacity is related to the local level of preoxidative products. Serum TBARS and CD are a measure of systemic oxidative damage. In the present study, the significantly elevated level of serum TBARS in the idiopathic cataract patients provides evidence for the systemic imbalance of lipid redox status. On the other hand, plasma SOD activity was significantly decreased in these patients, which correlated well with the fact that a reduction in antioxidant defenses might be responsible for deviating the systemic redox balance toward oxidative stress that is a possible cause of cataract formation. In agreement with these results, Chakraborty et al demonstrated a significant reduction in SOD activity in cataract patients. In the current study, serum CD and TBARS, which are markers for oxidative stress, were found to be significantly increased in the idiopathic cataract patients compared with the control group (53% and 15%, respectively). Cardiac patients who had received nitroglycerin showed a significant increase in serum CD and TBARS, compared with the idiopathic cataract patients (11% and 73%, respectively) and the control group (70% and 98%, respectively).

It can be proposed that a successful alteration in an adaptive phenomenon against oxidative stress caused an elevation in the levels of plasma SOD in the aged persons. However, the cataract patients could not have had such an adaptive response to oxidative stress and demonstrated a significant decrease in SOD activity. This could have resulted from either genetic or environmental factors, or both. In the present study, the levels of plasma SOD, a marker of enzymatic antioxidant activity, were found to be significantly decreased in the idiopathic cataract patients compared with the control group (22%).

GSH is a potent antioxidant present in a clear lens and is severely reduced in cataract. In the present study, the levels of serum GSH, a marker of nonenzymatic antioxidant activity, were found to be significantly decreased in the idiopathic cataract patients compared with the control group (12%). Cardiac patients who received nitroglycerin showed a significant decrease in serum GSH, compared with the idiopathic cataract patients (32%) and the control group (40%).

Conclusion
We conclude that plasma SOD, serum reduced GSH, nitrite, and lipid peroxidation activity could be good predictors for the development of cataracts due to systemic redox imbalance caused by chronic administration of nitroglycerin in cardiac patients. NO, as a metabolite of nitroglycerin, played an important role in the pathogenesis of cataracts in cardiac patients who were administered nitroglycerin. Further studies are needed to establish the role of nitroglycerin in the genesis of cataracts.

Limitations of the study
There are limitations to this study. It is essential to increase the sample size in future studies by collaborating with multiple hospitals all over Saudi Arabia. There are likely more cataract patients, especially elderly patients, who do not attend hospital and they might not be represented by the population studied here.

Despite its small size, this study suggests that those who are administered nitroglycerin should receive antioxidants during their treatment and undergo periodic ophthalmological examinations.

Acknowledgment
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Disclosure
The authors report no conflicts of interest in this work.

References


