

A negative regulation loop of long noncoding RNA HOTAIR and p53 in non-small-cell lung cancer

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Abstract: Non-small-cell lung cancer (NSCLC) is one of the leading causes of cancer-related death worldwide, and the 5-year survival rate is still low despite advances in diagnosis and therapeutics. A long noncoding RNA (lncRNA) HOX antisense intergenic RNA (HOTAIR) has been revealed to play important roles in NSCLC carcinogenesis but the detailed mechanisms are still unclear. In the current study, we aimed to investigate the regulation between the lncRNA HOTAIR and p53 in the NSCLC patient samples and cell lines. Our results showed that HOTAIR expression was significantly higher in the cancer tissues than that in the adjacent normal tissue, and was negatively correlated with p53 functionality rather than expression. When p53 was overexpressed in A549 cells, the lncRNA HOTAIR expression was downregulated, and the cell proliferation rate and cell invasion capacity decreased as a consequence. We identified two binding sites of p53 on the promoter region of HOTAIR, where the p53 protein would bind to and suppress the HOTAIR mRNA transcription. Inversely, overexpression of lncRNA HOTAIR inhibited the expression of p53 in A549 cells. Mechanistic studies revealed that HOTAIR modified the promoter of p53 and enhanced histone H3 lysine 27 trimethylation (H3K27me3). These studies identified a specific negative regulation loop of lncRNA HOTAIR and p53 in NSCLC cells, which revealed a new understanding of tumorigenesis in p53 dysfunction NSCLC cells.

Keywords: NSCLC, lncRNA HOTAIR, p53, negative loop

Introduction

Lung cancer is the second common malignant tumors and the leading cause of cancer death in the US and Asia.^{1,2} Lung cancer is characterized by high mortality and low cure rates, with an average of 5-year survival rate <18% based on SEER Cancer Statistics Review 1975–2012 conducted by the National Cancer Institute in the US.³ Among all the lung cancer incidents, non-small-cell lung cancer (NSCLC) accounts for ~85%.^{4,5} The treatment of NSCLC is mainly surgery, and radio- or chemotherapy; specific treatment varies from case to case depending on the stage of the tumor. Up to now, a number of site-specific targeting chemotherapeutic drugs have been used in treating NSCLC patients especially those in metastatic stages and have been received promising results in clinic.^{6–8} In all these therapies, the principle strategy is to induce DNA damage. As p53 mediates cellular response to DNA damage, the study of the roles of p53 in NSCLC becomes obviously essential.

p53 functions in response to a variety of cellular stress signals to induce cell cycle arrest, apoptosis, or senescence,^{9,10} which led to reduced cell proliferation rate and cell death.¹¹ One of p53 downstream effectors, p21, is a potent cyclin-dependent kinase inhibitor. When activated, p21 inhibits CDK1, CKD2, and CDK4/6 complex, resulting in cell cycle arrest at G1 and S phases.¹² New clinical research indicates that the

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activation of p53/p21 is associated with poor prognosis in NSCLC patients.¹³ The disruption of p53 pathway is a well-known mechanism to drive tumor formation, which was discovered in many types of cancers. The research by Mogi and Kuwano showed that *P53* gene is mutated in 50% of all human cancers and >70% of lung cancer.¹⁴ These cancer-driving mutations on *P53* gene cause the loss of the tumor suppressing function of p53. A recent study has shown that a longtime inactivation of *P53* gene is necessary for significant tumor growth, but the restoration of wild type p53 can stop this growth and trigger tumor elimination.¹⁵ However, the mechanisms revealing p53 dysfunction are still obscure so far.

Noncoding RNAs fulfill a diversity of biological functions, including gene regulation, RNA splicing, DNA synthesis, genome defense, and so on. They can be divided into small noncoding RNAs (<200 kb) and long noncoding RNAs (lncRNAs) (>200 kb) based on their length. Since the discovery, lncRNAs were found as the cause for a variety of diseases, such as cancers, neurological disorders, and fragile X syndrome. Recent evidence has shown that many lncRNAs are heavily involved in both tumor oncogenic and suppressive pathways;^{16–18} therefore, they have gained increasing attention in their gene regulatory functions. HOX antisense intergenic RNA (HOTAIR) is a lncRNA that locates on human chromosome 12 loci 13.¹⁹ Discovered by Howard Chang's group, HOTAIR was shown to recruit polycomb repressive complex 2 (PRC2), a transcriptional co-repressor, to decrease the transcription of the homeobox gene D cluster and some other genes.^{19,20} This is an example of gene silencing at long distance, which has been used frequently in development and disease initiation. HOTAIR's role in cancer has been shown by several studies. Its expression is significantly higher in hepatocellular carcinoma compared to adjacent noncancerous tissues.²¹ A similar role of HOTAIR was also found in pancreatic cancer.²² HOTAIR was also reported to facilitate breast cancer metastasis and loss of HOTAIR reduced cancer invasiveness.²³ A number of lncRNAs have been found under direct p53 regulation or shown correlation with p53 level;^{19,24} yet, the correlation between HOTAIR and p53 has not been studied.

In this study, we intended to determine whether there was a correlation between HOTAIR and P53 in NSCLC patients and cell lines. Furthermore, the effect of cell proliferation rate and invasion capability was measured in p53 overexpressed NSCLC cell lines in which HOTAIR was dysregulated. Finally, our bioinformatics analysis predicted that p53 regulates HOTAIR by direct interaction, which was confirmed by experiments.

Methods

Patient specimens and cell culture

A total of 48 NSCLC tissues were obtained from NSCLC patients receiving surgical treatment at the Department of Respiration Medicine, Affiliated Hospital of Binzhou Medical University between January 2014 and February 2015. All patients recruited in this study were not treated before they diagnosed with NSCLC based on histopathological evaluation. All collected tissue samples were immediately stored at liquid nitrogen until use. Informed consent was obtained from all patients with regard to surgical treatment and acquisition of tissue specimens. The study was approved by the Ethics Committee of the Binzhou Medical University. Human tissue acquisition and use in this study complied with the National Regulations on the Use of Clinical Samples in China. The NSCLC cell lines A549 (P53 wild-type), NCI-H157 (P53 mutant), and H1299 (P53 deletion) were obtained from Cellbio Company (Shanghai, People's Republic of China). These cell lines were cultured at 37°C in a CO₂ incubator in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with 10% fetal bovine serum and 1% penicillin–streptomycin.

The pcDNA3-flag-p53 (#10838) plasmid and the lncRNA HOTAIR expressing plasmid LZRS-HOTAIR (#26110) were purchased from Addgene (Cambridge, MA, USA); the pcDNA3-flag vector control and pGL3-basic vector were gifts from Dr Zhiqiang Liu, Department of Pathophysiology, Tianjin Medical University. The promoter of lncRNA HOTAIR was cloned into the pGL3-basic firefly luciferase reporter with the primers 5'-ACTGGTACCTAAGCGGAGAGAGTCCC-3' and 5'-ACTAAGCTTGAGTCAGAGTTCCCCAC-3'. Plasmid transfection was achieved by Lipofectamine 2000 reagent (Life Technology, Shanghai, People's Republic of China) with ratio of Lipo: DNA =2; after 6 hours, the media were changed to complete culture media for another 48 hours.

Real-time PCR analysis of mRNA expression levels

Total RNA was extracted by Trizol reagent (Invitrogen, Carlsbad, CA, USA). Quantitative polymerase chain reaction (PCR) was performed using specific primers (Sangon, Shanghai, People's Republic of China) for HOTAIR. The cycling conditions were 30 seconds polymerase activation at 95°C, followed by 40 cycles amplification at 95°C for 5 seconds, and extension at 60°C for 30 seconds. The first strand cDNA served as the template for the regular PCR performed using a DNA Engine

(Applied Biosystem, CA, USA). PCR primers: HOTAIR (200 bp), forward 5'-GGCAAATGTCAGAGGGTTCT-3' and reverse 5'-TTCTTAAATTGGGCTGGGTC-3'; glyceraldehyde 3-phosphate dehydrogenase (GAPDH) (173 bp), forward 5'-ACAGGGGAGGTGATAGCATT-3' and reverse 5'-GACCAAAAGCCTTCATACATCTC-3'. GAPDH as an internal control was used to normalize the data to determine the relative abundance of the target RNA. The amplification curves and melting curves were analyzed after completion of the reaction. Gene expression values were represented using the $2^{-\Delta\Delta C_t}$ method.

Western blot

Cells were lysed by radioimmunoprecipitation buffer and proteins were then extracted and quantified by Bradford reagent. An amount of 50 mg of total proteins were loaded on Nupage 4%–12% polyacrylamide gels (Thermo Fisher Scientific, Waltham, MA, USA) for each sample and transferred to polyvinylidene difluoride membranes (GE Healthcare Bio-Sciences Corp., Piscataway, NJ, USA), then they were probed with the following antibodies separately: mouse anti-p53 (1:1,000, Cat No: ab28, Abcam, Cambridge, UK) and rabbit anti- β -actin (1:2,000, Cat No: ab3280, Abcam). For detection, goat antirabbit or goat antimouse secondary antibodies conjugated to horseradish peroxidase (1:2,000, GE Healthcare Bio-Sciences Corp.) were used. Protein abundance was measured by chemiluminescence reaction (ECL, GE Healthcare Bio-Sciences Corp.). Western blot and quantification were performed using ImageJ software analysis (National Institutes of Health, Bethesda, MD, USA).

Cell proliferation assays

Cell Proliferation Reagent Kit I (MTT) (Roche Applied Science, IN, USA) was used to monitor cell proliferation rate. A total of 3,000 cells were seeded into each well of a 96-well plate. All experiments were performed in triplicate and cell proliferation rates were recorded every 24 hours according to the manufacturer's protocol.

In vitro invasion assay

A modified in vitro Boyden chamber invasion assay with Matrigel-coated Transwell chambers was performed. Cells were trypsinized and resuspended in serum-free DMEM medium, and then were added to rehydrated Matrigel-coated inserts (BioCoat Matrigel Invasion Chamber; NJ, USA); the inserts were then placed in 24-well companion trans plates with DMEM medium and 10% fetal bovine serum

(chemoattractant) for cell migration through the pores to the trans side. After 24 hours, the cells and matrigel in the upper chambers were removed, and the cells in the bottom trans chambers were fixed with 3% glutaraldehyde and stained with 4',6-diamidino-2-phenylindole (DAPI) (1:5,000; Sigma-Aldrich Co., St Louis, MO, USA) fluorescent for 15 minutes. DAPI-stained cells were counted in five randomly different fields under an inverted fluorescence microscope. The experiments were performed in triplicate wells and each experiment was performed in triplicate.

Chromatin immunoprecipitation assay

Cells were cross-linked in 1% (vol/vol) formaldehyde for 10 minutes at room temperature. Then, 0.125 M glycine was used to quench the cross-linking reaction. Cells were pelleted and washed with ice-cold phosphate-buffered saline. Next, cell nuclei were isolated and a Bioruptor sonicator (Diagenode) was used to shear chromatin DNA. An amount of 5 μ L of p53 antibody (1:2,000, Cat No: ab28, Abcam) diluted in phosphate-buffered saline buffer was added to Dynabeads Protein A (Invitrogen) beads and incubated for 3 hours at 4°C with rotation. An amount of 5 μ L of normal rabbit immunoglobulin G (Cell Signaling, CA, USA) was used as negative control. Then, the dynabeads-antibody complexes were incubated with sheared chromatin DNA overnight at 4°C. After immunoprecipitation, the precipitated complex was treated with RNase A and Proteinase K, and incubated at 65°C overnight to reverse cross-links. Primers were designed and quantitative RT-PCR (qRT-PCR) was performed. The sequence of primers for HOTAIR promoter (110 bp) was as follows: F 5'-GCCCTTCTCCTAGCCCACCG-3', R 5'-GTGGGGACCCGCTAGACCTG-3'; sequence of primers for p53 promoter (125 bp) was as follows: F 5'-CA AAGAAATGGAGCCGTGTA-3', R 5'-GCCTCCTAA AGTGCCAAGAT-3'. Chromatin immunoprecipitation (ChIP)-qPCR data were normalized either by the percent input method or relative to the immunoglobulin G control.

Statistical analysis

Statistical analysis was performed using the SPSS statistical software program Version 15.0 (SPSS Inc., Chicago, IL, USA). Data from all these experiments were reported as mean \pm standard deviation for each group. Intergroup differences were analyzed by one-way analysis of variance. Correlation between expressions of p53 and lncRNA HOTAIR was analyzed with regression analysis. Differences with P -value of <0.05 were considered statistically significant.

Results

The lncRNA HOTAIR expression was negatively correlated with p53 in NSCLC

First of all, we measured the expression of p53 via real-time PCR in all 48 NSCLC patients' samples, and found that the expression was differed from patient to patient (data not shown). The expression of p53 in 12 samples from all 48 patients was also measured by Western blot (Figure 1A), and mRNA level of HOTAIR was measured by qRT-PCR as well (Figure 1B). In addition, expressions of the lncRNA HOTAIR in the cancer tissues or the adjacent-normal tissues from all 48 samples were detected using real-time PCR, and the result showed that expression of HOTAIR in cancer tissue group was significantly higher than that in the adjacent-normal tissue control group ($P<0.001$) (Figure 1C). By comparing the expression of p53 and HOTAIR, we found that the higher level of HOTAIR was associated with a lower p53 level. The same negative correlation between lncRNA HOTAIR and P53 was confirmed in all of our 48 NSCLC patients' samples using regression analysis (Figure 1D).

In order to further investigate the correlation between the expressions of HOTAIR and p53, the expressions of

both molecules were measured in three NSCLC cell lines based on their p53 function and presence: A549 (wild-type p53), NCI-H157 (mutant p53), and H1299 (p53 deletion). We confirmed the expression characteristics of p53 in these three cell lines, among which H1299 cells had almost invisible p53, while the A549 and NCI-H157 cells had considerable p53 expression (Figure 2A). However, we did not detect the mutation of p53 in NCI-H157 cells in this study. Next, the expression level of HOTAIR was measured in each of the three cell lines by qRT-PCR. We found that HOTAIR level was significantly higher in H1299 cells and NCI-H157 cells compared to A549 cells (Figure 2B), which suggested that the expression levels of HOTAIR were negatively correlated with p53 functionality rather than expression.

Modulation of p53 expression alternated the lncRNA HOTAIR expression and cell invasion in NSCLC cells

As we observed the negative correlation between p53 and HOTAIR, we next investigated if the expression of p53 suppressed HOTAIR level. p53 was overexpressed in A549 cells using the pcDNA3-flag-p53 plasmid, and Western blot was

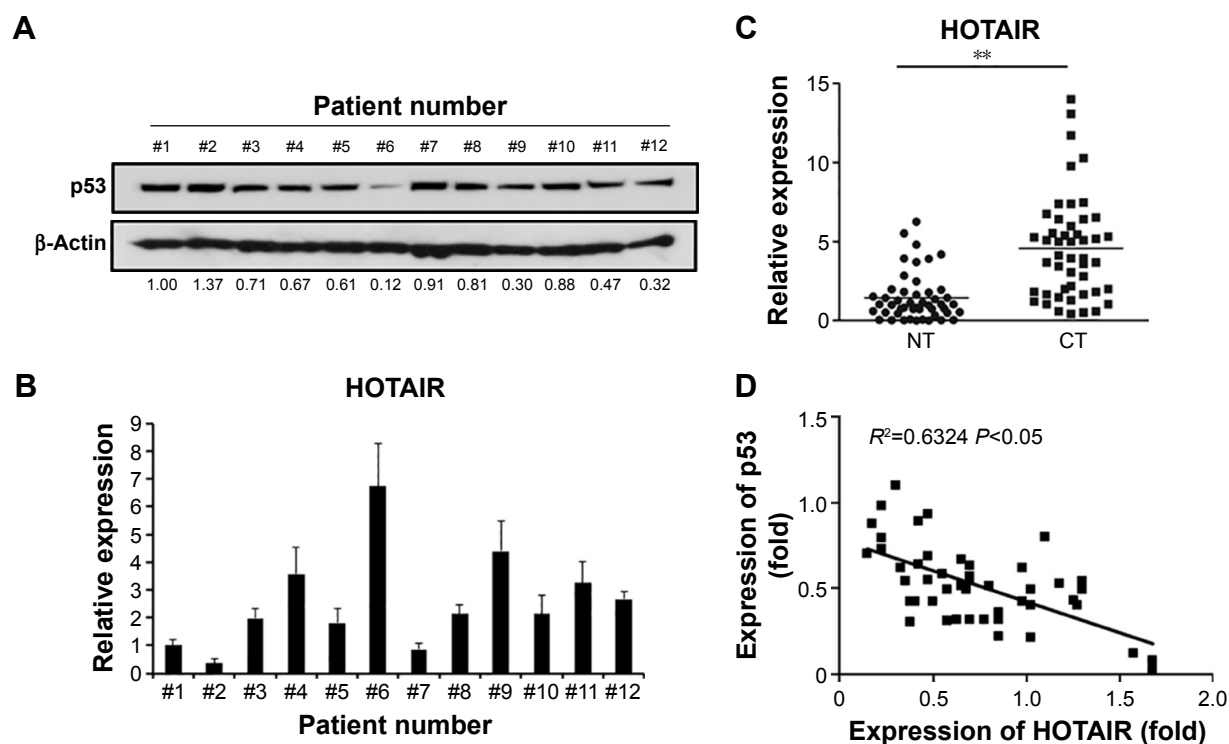


Figure 1 The negative correlation between lncRNA HOTAIR and p53 expressions in NSCLC.

Notes: (A) Western blot assay showed the p53 protein level in 12 NSCLC patients' samples, and β -actin was used as an endogenous control. The relative expressions of p53 in different samples were standardized to the No. 1 by gray scale scanning, and the number below each band showed its relative p53 expression. (B) The relative expressions of lncRNA HOTAIR in the same patients' samples with (A) detected by real-time PCR assay. (C) Real-time PCR assay showed the expression of the lncRNA HOTAIR in the cancer adjacent NT or CT in all 48 patients. (D) The correlation between the expressions of HOTAIR and p53 was analyzed with regression analysis. $**P<0.01$.

Abbreviations: CT, cancer tissues; HOTAIR, HOX antisense intergenic RNA; lncRNA, long noncoding RNA; NSCLC, non-small-cell lung cancer; NT, normal tissues; PCR, polymerase chain reaction.

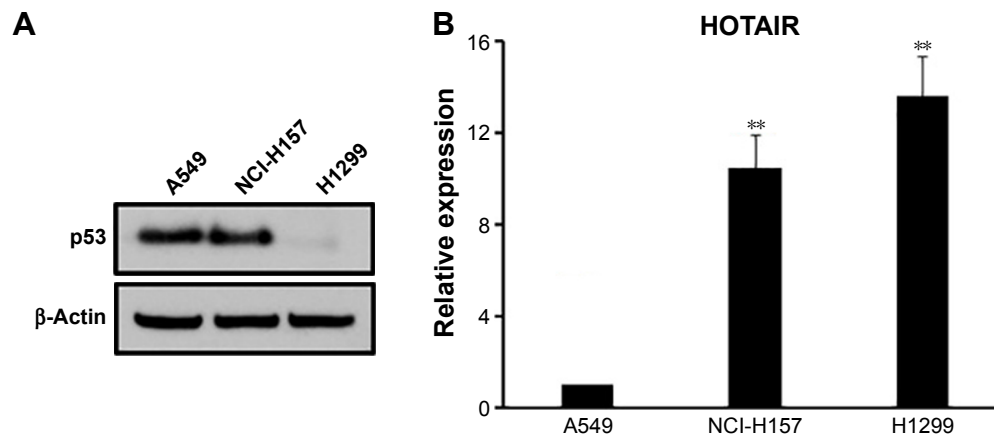


Figure 2 IncRNA HOTAIR negatively correlates with p53 functionality.

Notes: (A) Western blot showed the p53 level in the A549 (p53 wild-type), NCI-H157 (p53 mutant), and H1299 (p53 deletion) cell lines, and the IncRNA HOTAIR level in these three cell lines were detected using real-time PCR (B). For all real-time PCR results, the formula $2^{-\Delta\Delta C_t}$ was used, and data here were represented as mean \pm SD of three independent experiments. ** $P < 0.01$.

Abbreviations: HOTAIR, HOX antisense intergenic RNA; IncRNA, long noncoding RNA; PCR, polymerase chain reaction; SD, standard deviation.

used to confirm the high protein level of p53 48 hours after transfection (Figure 3A). Then, the expression of HOTAIR was measured by qRT-PCR, and our result showed that the expression of HOTAIR was significantly decreased in the p53 overexpressing cells compared to the cells transfected with vector control (Figure 3B). Next, phenotypes changed in cell

biology of the A549 cells were examined. We confirmed that p53 overexpression affected cell proliferation and migration in the A549 NSCLC cells, as overexpression of p53 significantly decreased the proliferation rate of A549 cells (Figure 3C) as well as the invasion capability in vitro (Figure 3D), compared to the cells transfected with vector control, respectively.

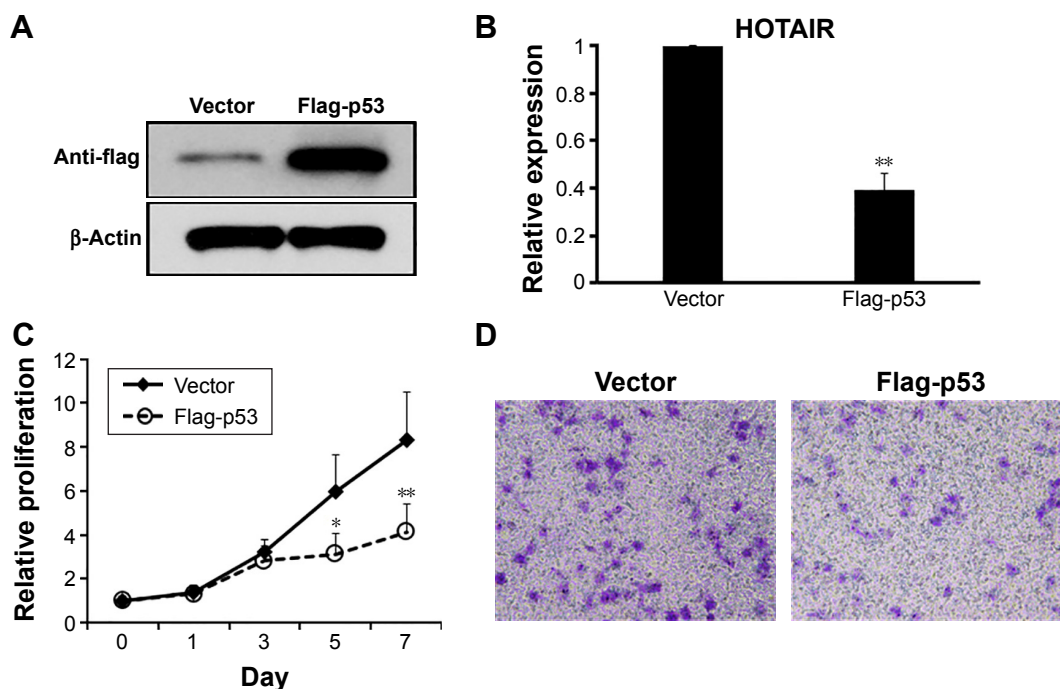


Figure 3 Modulation of p53 expression alternated the IncRNA HOTAIR expression and cell invasion in NSCLC cells.

Notes: (A) pcDNA-flag-P53 expressing plasmid and the pcDNA3-flag vector control plasmids were transfected into A549 cells for 48 hours, and the expression of flag-tagged p53 was measured by Western blot assay. (B) The relative expression of IncRNA HOTAIR in A549 cells transfected with vector control or flag-p53 expressing plasmid was detected using real-time PCR 48 hours after transfection. (C) Cell proliferation of A549 cells transfected with vector control or flag-p53 expressing plasmid was measured by an MTT assay at days 0, 1, 3, 5, and 7 after transfection. (D) The cell invasion capability was measured by a transwell assay at 48 hours after p53 overexpression in A549 cells. Representative images were shown from five independent experiments. Data represent mean \pm SD of three independent experiments. Magnification, 200 \times . * $P < 0.05$; ** $P < 0.01$.

Abbreviations: HOTAIR, HOX antisense intergenic RNA; IncRNA, long noncoding RNA; NSCLC, non-small-cell lung cancer; PCR, polymerase chain reaction; SD, standard deviation.

p53 Could bind and regulate the HOTAIR promoter

As the negative correlation between p53 and HOTAIR was known, we next studied possible interaction between these two molecules via bioinformatics analysis. Based on our computational prediction, two p53 binding sites were found on the promoter region of HOTAIR (Figure 4A, taller). In order to verify the prediction, the HOTAIR promoter was cloned into the pGL3-luciferase plasmid to construct HOTAIR-luc, which fused luciferase to HOTAIR promoter. Next, the HOTAIR-luc and p53 plasmids were cotransfected into A549 cells, in which the expression of p53 was then confirmed (data not shown). We found that the luciferase activity of the HOTAIR promoter was significantly decreased in cells overexpressing p53 compared with the vector control (Figure 4A, shorter). In order to further elaborate how P53 regulates HOTAIR expression at transcriptional level, ChIP assay was carried out in A549 cells. We found the HOTAIR promoter was pulled down significantly more in the p53 overexpressing cells than that in cells transfected with empty vector (Figure 4B), suggesting p53 binds and regulates the lncRNA HOTAIR promoter.

lncRNA HOTAIR reversely regulated the histone H3 lysine 27 trimethylation of p53 gene promoter

HOTAIR interacts with and recruits PRC2 to regulate the chromosome occupancy through the EZH2 molecule (a subunit of PRC2), which leads to histone H3 lysine 27 trimethylation (H3K27me3) of some genes.²⁵ Based on this known information, we explored whether there is a negative

loop between p53 and HOTAIR. When the HOTAIR overexpression plasmid was transiently transfected into A549 cells for 48 hours, the protein level of p53 was obviously down-regulated when detected by Western blot assay (Figure 5A), and we found that *P53* mRNA level was also significantly downregulated shown by real-time PCR assay (Figure 5B). In addition, the epigenetic modification of p53 promoter was checked via ChIP assay, which showed that overexpression of HOTAIR could increase the H3K27me3 level (Figure 5C), whereas the H3K4me3 was shown slightly decreased in both cells (data not shown). This result implies that HOTAIR might epigenetically regulate the *P53* gene promoter in a negative feedback loop model.

Discussion

Lung carcinoma is one of the most common types of cancer worldwide and exists as a major concern of health problem. As the most popular subtype of lung cancer, NSCLC occurred >50% in Asian female lung cancer patients. Although significant progress has been made in the past decades in chemotherapy, NSCLC is still resistant to current treatments. Therefore, better understanding of the mechanisms leading to NSCLC formation and development is essential. In this study, we found that p53 is involved in NSCLC cell proliferation and migration, and a negative correlation was identified between p53 functionality and an lncRNA molecule, HOTAIR. Further analysis discovered that p53 represses HOTAIR expression by direct binding to its promoter. Inversely, lncRNA HOTAIR inhibited the expression of p53 in NSCLC cells, and mechanistic studies revealed that HOTAIR modified the promoter of p53 and

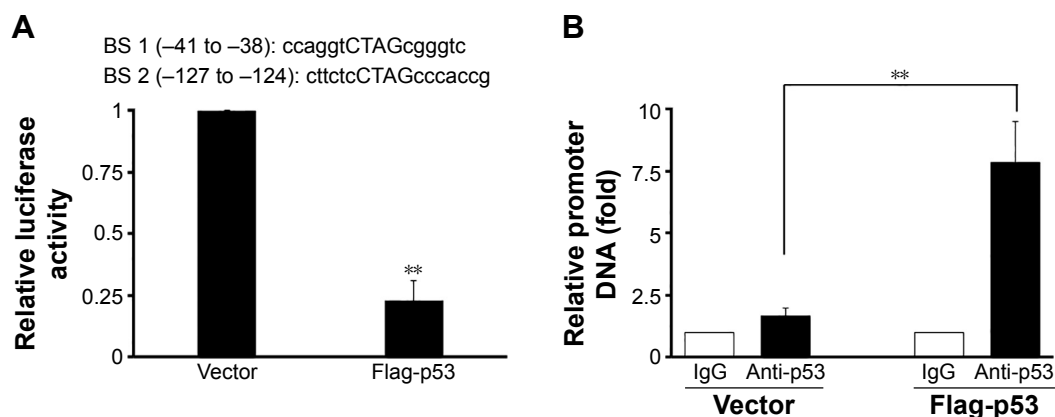


Figure 4 The regulation of p53 on the HOTAIR promoter.

Notes: (A) Two sequences shown at the top were two binding sites of p53 at the promoter of HOTAIR gene by bioinformatics prediction. Luciferase assay was carried out in A549 cells transfected with pGL3-basic or pGL-HOTAIR combined with pcDNA3-flag or flag-p53, respectively, for 48 hours. (B) The immunoprecipitation assay was performed to detect the activities of P53 binding to the promoter of lncRNA HOTAIR. The A549 cells were transfected with pcDNA-flag vector or flag-p53 expressing plasmids for 48 hours, and cell lysate was pulled down using IgG control or anti-p53 monoclonal antibody. Data represent mean \pm SD of three independent experiments. ** $P < 0.001$.

Abbreviations: HOTAIR, HOX antisense intergenic RNA; IgG, immunoglobulin G; lncRNA, long noncoding RNA; SD, standard deviation; BS, binding sites.

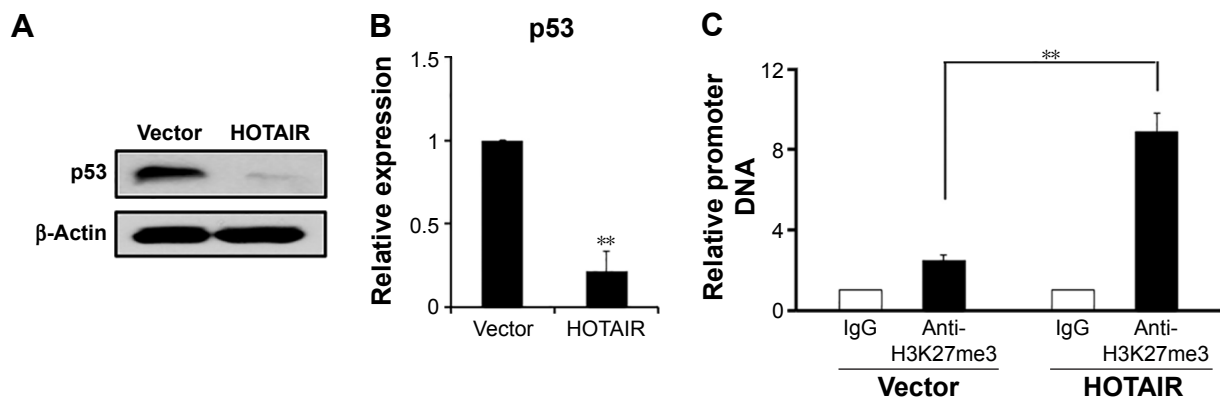


Figure 5 The epigenetic regulation of HOTAIR on the promoter of p53 gene.

Notes: (A) Western blot showed the p53 protein level in A549 cells transfected with vector control or LZRS-HOTAIR plasmid for 48 hours. β -Actin was used as the endogenous control. (B) p53 mRNA level in A549 cells transfected with vector or LZRS-HOTAIR plasmid for 48 hours. (C) The activities of H3K27me3 level were determined with ChIP assay in A549 cells transfected with LZRS-HOTAIR plasmid or vector control for 48 hours. Data represent mean \pm SD of three independent experiments. ** $P < 0.001$.

Abbreviations: ChIP, chromatin immunoprecipitation; HOTAIR, HOX antisense intergenic RNA; IgG, immunoglobulin G; SD, standard deviation.

enhanced histone H3 lysine 27 trimethylation (H3K27me3). Therefore, we for the first time revealed a negative regulation loop between p53 and lncRNA HOTAIR in NSCLC.

lncRNAs have attracted increasing attentions in recent years as they appeared to be involved in a number of cancer formations and metastases. Many lncRNAs have shown the capacity to regulate the initiation, progression, and response to therapy in a variety of solid tumors and hematological malignancies.^{26,27} Gene expression microarrays identified that hundreds of lncRNAs are associated with lung cancer and massively parallel RNA sequencing of tumor tissues and paired adjacent nontumor tissues in the lung.^{28–31} Thus, further study on the role of lncRNAs in NSCLC is worth conducting. Recent research identified HOTAIR as a promising target to diagnose and treat lung cancer. A study discovered that NSCLC tumor samples had higher HOTAIR expression levels compared to their corresponding noncancerous lung tissue.³² Besides lung cancer and breast cancer,²³ the expression levels of HOTAIR were found elevated in several other types of tumor tissues as well, including liver cancer³³ and certain colorectal cancer.³⁴ Colorectal cancer patients who showed a high HOTAIR expression level also had a worse prognosis than those patients with low HOTAIR levels. Besides primary tumor, HOTAIR is also found involved in tumor metastasis, for example, lymph node metastasis in hepatocellular carcinoma.²¹ Therefore, HOTAIR could be a promising diagnostic target, which needs further confirmation.

Here, we discovered that p53 was involved in the regulation of HOTAIR expression in NSCLC cell lines by direct binding to its promoter. Our study identified a new p53 downstream regulatory mechanism that led to cancer

when nonfunctional. *P53* is one of the most powerful tumor suppressor genes identified in human cells. Its mutation is most frequently found in cancer cells and particularly lung cancer, and >70% of lung cancer cases have a *P53* mutation. As HOTAIR is under the regulation of p53, deregulation of HOTAIR expression level in lung cancer could be common as well. Therefore, potential therapeutic drugs targeting this lncRNA might be able to provide promising results.

The function of HOTAIR in NSCLC is still not clear and needs further investigation. However, a previous study on breast cancer found that HOTAIR promotes primary cancer formation and metastasis by recruiting PRC2, which methylate H3K27 and silence metastasis suppressor genes.²³ Furthermore, HOTAIR can serve as a scaffold for multiple histone modification complexes: the 5' end of HOTAIR binds with PRC2, and its 3' end binds to lysine (K)-specific demethylase 1A complex.³⁵ Therefore, this bifunctional character of HOTAIR could be required for coordinating histone modifications of H3K27 methylation and H3K4 demethylation for epigenetic gene silencing in the metastatic processes.³⁶ In the current study, the relationship between HOTAIR and p53 has been verified. We found that the H3K27me3 modification on the promoter of p53 was changed, which might be relative with HOTAIR. Potential research might focus on the negative feedback loop between p53 and HOTAIR.

Conclusion

In conclusion, our study demonstrated that the p53 levels was negatively correlated with the amount of HOTAIR in NSCLC patients' samples and cell lines, suggesting a regulatory role of p53 in HOTAIR transcription. Moreover, the

HOTAIR might regulate the expression of p53 epigenetically via changing the modification of H3K27me3 with a negative feedback loop. This work added another piece of evidence in HOTAIR's role in cancer promotion and metastasis, and highlighted the importance of HOTAIR as a potential target in cancer diagnostic and treatment.

Disclosure

The authors report no conflicts of interest in this work.

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