Cycloid psychoses in the psychosis spectrum: evidence for biochemical differences with schizophrenia

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Abstract: Cycloid psychoses (CP) differ from schizophrenia regarding symptom profile, course, and prognosis and over many decades they were thought to be a separate entity within the psychosis spectrum. As to schizophrenia, research into the pathophysiology has focused on dopamine, brain-derived neurotrophic factor, and glutamate signaling in which, concerning the latter, the N-methyl-D-aspartate receptor plays a crucial role. The present study aims to determine whether CP can biochemically be delineated from schizophrenia. Eighty patients referred for psychotic disorders were assessed with the Comprehensive Assessment of Symptoms and History, and (both at inclusion and after 6 weeks of antipsychotic treatment) with the Positive and Negative Syndrome Scale and Clinical Global Impression. From 58 completers, 33 patients were diagnosed with schizophrenia and ten with CP according to the Diagnostic and Statistical Manual of Mental Disorders, Fourth Edition, and Leonhard criteria, respectively. Fifteen patients were diagnosed with other disorders within the psychosis spectrum. At both time points, blood levels of the dopamine metabolite homovanillic acid, brain-derived neurotrophic factor, and amino acids related to glutamate neurotransmission were measured and compared with a matched control sample. Patients with CP showed a significantly better response to antipsychotic treatment as compared to patients with schizophrenia. In CP, glycine levels were elevated and tryptophan levels were lowered as compared to schizophrenia. Glutamate levels were increased in both patient groups as compared to controls. These results, showing marked differences in both treatment outcome and glutamate-related variable parameters, may point to better neuroplasticity in CP, necessitating demarcation of this subgroup within the psychosis spectrum.

Keywords: cycloid psychoses, schizophrenia, glutamate, glycine, tryptophan, neuroplasticity

Introduction

Over many decades, dopamine (DA) neurotransmission has been thought to be the main mechanism in the pathophysiology of psychotic disorders, and DA receptor antagonists have been demonstrated to be effective in reducing – mainly positive – psychotic symptoms.1 Subsequent research has disclosed that other neurotransmitter systems, particularly that of glutamate, are associated with negative symptoms and cognitive dysfunctions via the N-methyl-D-aspartate (NMDA) receptor.2–5

Glutamate is the most abundant excitatory neurotransmitter in human brain, and glutamatergic receptors are involved in regulating neuronal migration, growth, and pruning (ie, neuroplasticity).6,7 Glutamate acts at different types of receptors, of which the NMDA receptor is the most investigated.8,9 Glycine acts as a co-agonist at the NMDA receptor and can potentiate glutamatergic neurotransmission.10 In contrast,
antagonizing the NMDA receptor by means of phencyclidine or ketamine induces schizophrenia-like symptoms.\textsuperscript{11,12}

These observations have led to the hypothesis that the pathophysiology of schizophrenia is at least partly related to impairment in NMDA neurotransmission.\textsuperscript{13,14} Hence, it seems likely that enhancing NMDA activity might benefit patients with schizophrenia, which could possibly be achieved by administering components targeting the glutamate system.\textsuperscript{8,10,15,16}

Cycloid psychoses (CP) as described by Leonhard in the 1950s, partly operationalized by Perris and Brockington and included in the International Classification of Diseases, Tenth Edition, as acute polymorphic psychotic disorders, differ from schizophrenia with respect to symptom profile, course, and prognosis.\textsuperscript{17–20} According to Leonhard, three subtypes can be delineated: anxiety–happiness psychosis, confusion psychosis, and motility psychosis, all showing a pleomorphic symptom profile with intraphasic bipolarity. In general, full recovery is reached without residual negative symptoms or cognitive decline.\textsuperscript{21–23} CP, as such, are not included in the Diagnostic and Statistical Manual of Mental Disorders, Fourth Edition (DSM-IV), although pananxiety, perplexity, and motility disorders have been demonstrated in a substantial percentage of patients.\textsuperscript{24–30} An overview of the characteristics of schizophrenia and CP is presented in Table 1.

Since CP differ significantly from schizophrenia, the hypothesis was formulated that they could also be distinct on their biochemical profile, especially regarding glutamatergic transmission. Therefore, in the present study, peripheral levels of amino acids related to glutamate signaling (ie, glutamate, glycine, and tryptophan) were measured in a group of patients with psychosis spectrum disorders. In addition, levels of brain-derived neurotrophic factor (BDNF) and the DA metabolite homovanillic acid (HVA) were determined, reflecting neuroplasticity and dopaminergic activity, respectively.

| Table 1 Characteristic differences between schizophrenia and cycloid psychoses |
|-----------------------------|-----------------------------------|
| **Schizophrenia**           | **Cycloid psychoses**             |
| Onset                       | Slow and insidious (weeks to months) |
| Age at onset                | Adolescence/young adulthood       |
| Symptomatology              | Delusions, hallucinations, disorganization, negative symptoms |
| Course                      | Relapsing and remitting, chronic, gradual functional and cognitive decline |
| Prognosis                   | Unfavorable                        |
| Treatment                   | Antipsychotic agents              |
| Sex distribution            | Male = female                      |

**Methods**

**Patients**

Patients were recruited over a 2.5-year period at the Vincent van Gogh Institute for Psychiatry. Included were adult male and female patients meeting DSM-IV criteria for psychosis spectrum disorders, who required antipsychotic treatment intervention. Intervention was defined as start or switch of antipsychotic agent or use of augmentation strategies. Exclusion criteria were known genetic syndromes or intellectual disability, and somatic or neurologic disorders interfering with treatment as usual. Research was performed in accordance with the Declaration of Helsinki and was approved by the regional ethics committee (METI\GG) and board of directors of Vincent van Gogh Institute. Written informed consent was obtained following Dutch ethical guidelines (CCMO registration number NL20469.097.07). After inclusion, all patients were prescribed first-generation antipsychotic (FGA) or second-generation antipsychotic (SGA) agents by their treating psychiatrist following standard professional guidelines. Concomitant treatment with two antipsychotic agents, mood stabilizer, and/or antidepressant was allowed and documented.

During the study period, 194 patients were identified as eligible for inclusion. However, from these 194 patients, 71 were considered unable to provide informed consent because of severity of illness. In another 23 patients, the intended pharmacological intervention was not performed, yielding 100 patients who were eligible for inclusion. Twenty patients refused to participate, mostly because of the need for venipuncture, resulting in a study group of 80 patients. Of them, 58 completed all clinical and biochemical assessments at baseline and after 6 weeks.

**Diagnostic procedures**

All patients were assessed by the first author (NvdK) before or within 1 week after start of treatment with antipsychotics. At baseline, the Comprehensive Assessment of Symptoms and History was applied to establish diagnoses according
to DSM-IV. PANSS and Clinical Global Impression of Severity and Improvement (CGI-S/I) were used to measure symptom profile and overall disease severity at baseline and after 6 weeks. PANSS cognitive score was calculated according to the procedure as described by Lindenmayer et al. To establish diagnosis of CP, a symptom checklist according to Leonhard was completed by NvdK and a clinician (MKF Schneider) trained in establishing diagnoses according to Leonhard. Final diagnoses were made in a so-called Longitudinal Evaluation of All Data (LEAD) conference, in which all possible and probable cases of CP were discussed with two experts in the field of CP (GS and WV).

Biochemical analyses

Patients

Blood samples were collected by means of venipuncture between 8 and 10 am at both assessment dates to determine BDNF, HVA, the amino acids glutamate, glycine, and tryptophan, and the ratio between tryptophan and the five large neutral amino acids (LNAAs) competing for the same transport system, being leucine, isoleucine, valine, phenylalanine, and tyrosine (Trp/LNAAs ratio).

Plasma was obtained after centrifugation of ethylenediaminetetraacetic acid blood for 20 minutes at 2,650 x g and 20°C, and stored at −80°C until analysis. All biochemical analyses were performed at the Neuropsychiatric Laboratory of Erasmus University Medical Center, Rotterdam, and the technician was blinded to the clinical situation and diagnosis of the patients.

Plasma was deproteinized, and amino acids were separated by high-performance liquid chromatography and detected by fluorescence after derivatization with orthophthalldialdehyde as described previously. HVA levels were measured after deproteinization of plasma by high-performance liquid chromatography using a Zorbax Eclipse XDB-C8 column (5 μm particle size, 250x3 mm; Agilent Technologies, Santa Clara, CA, USA) for separation. For the detection of HVA, an electrochemical detector (oxidation potential was set to 0.7 V) and the Intro controller (Antec Leyden, Zoeterwoude, the Netherlands) were used. Quantification was done by measuring peak heights relative to two internal standards (isoprenaline and 5-methylserotonin). The mean recovery (±standard deviation [SD]) of HVA added to the plasma samples was 95%±7%. BDNF was measured in serum by a double-antibody sandwich enzyme-linked immunosorbent assay (Promega Corporation, Madison, WI, USA).

Controls

Biochemical parameters from age-matched controls were taken from a large database of the aforementioned Neuropsychiatric Laboratory. Age-matched controls were taken from a large database, consisting of hospital staff and students and subjects from the general community, all without a personal or family history of psychiatric illness. Mean age of the control group (n=75) was 37.9±10.1 years.

Statistics

From the 58 completers, only patients with a diagnosis of schizophrenia (n=33) or CP (n=10) were included in the statistical analyses. To compare means, nonparametric Mann–Whitney U test was used for normally distributed data (biochemical parameters), whereas Student’s t-test was applied for normally distributed data. To investigate changes between time points, Wilcoxon signed rank test (biochemical parameters) or paired t-test (PANSS scores) was used. For binary variables, a chi-square test was done. Significance was set at P<0.05. All data are presented as mean ± SD, unless stated otherwise.

Results

Patients

By excluding patients diagnosed with other psychotic disorders (n=15), the study sample (n=43) consisted of 30 males and 13 females with a mean age of 34.8±11.3 years. Thirty-three patients received a diagnosis of schizophrenia, and in ten, a diagnosis of CP according to Leonhard was made (anxiety–happiness: n=5; confusion: n=2; motility: n=3). Mean age at first psychosis was 26.5±9.8 years, and mean duration of psychotic illness was 8.3±8.1 years. Fourteen patients (CP: n=5; schizophrenia: n=9) did not receive psychotropic medication prior to inclusion in the study (referred to as “at baseline”), of which eight were antipsychotic naïve (CP: n=3; schizophrenia: n=5). Eight out of the total of 43 patients were first-episode psychosis (FEP) patients. Five patients were classified as both medication naïve and having FEP (CP: n=3; schizophrenia: n=5). Details on symptom clusters, effect of antipsychotic treatment, and severity of illness are summarized in Table 2.

Since each patient received an individually targeted treatment, data were scrutinized for possible confounding effects of psychopharmacological heterogeneity. With respect to the two patient groups (CP: n=10; schizophrenia: n=33), no differences were found as to medication status (naïve, free >2 weeks, or medicated), use of co-medication (antidepressant or mood stabilizers), and antipsychotic class (FGA or SGA). Regarding the latter, in the CP group, none of the patients was treated with clozapine, whereas nine patients in the schizophrenia group did receive this agent. The CP group comprised significantly
more females, who had higher ages at first presentation of psychotic illness as well as at time of inclusion. Finally, five patients with CP were defined as FEP patients.

As can be inferred from Table 2, CP patients had significantly lower scores on the PANSS total, positive, and negative scale at both time points. Symptomatic improvement was most marked in the positive symptom cluster. While both groups showed symptomatic improvement after 6 weeks of treatment, a significant difference was found in PANSS cognitive score and in CGI-S for the CP group.

**Biochemical parameters**

As compared to controls, serum BDNF levels were lowered in patients with schizophrenia, and plasma glutamate levels appeared to be increased in both patient groups. Concerning plasma levels of glycine, significant higher values were found in CP patients at both time points as compared to patients with schizophrenia as well as control subjects. Also, at both time points, plasma tryptophan levels were significantly lower in patients with CP as compared to controls but differed only at baseline from the schizophrenia group. Trp/LNAAs ratio was lowered in both patient groups as compared to controls. Plasma levels of HVA did not differ between the groups. All data are presented in Table 3.

In both the subgroup of patients with schizophrenia who were treated with clozapine (n=9) and the subgroup of eight medication-naive patients (CP: n=3; schizophrenia: n=5), lowered serum levels of BDNF and elevated levels of glutamate were found at both time points. However, in the clozapine subgroup, glycine levels were significantly lowered as compared to controls at baseline (177.6±45.1 versus 224.2±47.8 μmol/L), whereas in the medication-naive subgroup, glycine levels did not differ from controls at both time points.

The 15 patients with diagnoses other than CP or schizophrenia (schizoaffective disorder: n=1; bipolar disorder: n=5; delusional disorder: n=1; brief psychotic disorder: n=2; schizotypal personality disorder: n=1, psychotic disorder not otherwise specified: n=5) were analyzed separately. Glutamate levels were significantly elevated as compared to controls at both time points. Data on biochemical parameters in this group are depicted in Table 4.

As to intercorrelations of biochemical and clinical parameters, both in the total patient sample and in the schizophrenia subgroup, baseline glutamate levels were positively correlated with changes in PANSS cognitive scores (total sample: T=0.280, P=0.012; schizophrenia: T=0.360, P=0.005). Glutamate levels after 6 weeks were positively correlated with PANSS cognitive scores at both time points in the total patient group (baseline: T=0.221, P=0.043; 6 weeks: T=0.279, P=0.011) and with PANSS cognitive scores after 6 weeks in schizophrenia patients (T=0.289, P=0.023).

Baseline glycine levels did not correlate with any of the clinical (sub)scales. Glycine levels after 6 weeks correlated significantly with changes in PANSS cognitive scores (T=0.523, P=0.038) in CP patients. No other correlations were found in CP between glutamate or glycine levels and other PANSS or CGI scores at both time points.

No significant correlations were found for (changes in) any of the biochemical parameters from Table 3 and (changes in) other clinical parameters (PANSS or CGI) in the two subgroups at both time points.

**Discussion**

In this study, patients with CP and schizophrenia were assessed on both clinical and biochemical profiles. Essentially, the CP group showed better clinical outcome than the schizophrenia group and differed significantly in terms of
biochemical parameters, particularly plasma levels of glycine and tryptophan. Serum levels of BDNF were significantly lower in the schizophrenia group as compared to controls. With respect to glycine, in CP, plasma levels were elevated as compared to both schizophrenia and healthy controls. Although in 2004 Sumiyoshi et al found lowered levels of glycine for schizophrenia in comparison with controls and tryptophan. Serum levels of BDNF were significantly elevated as compared to both schizophrenia and healthy controls. It could be postulated that in CP, glycine levels are elevated as a compensatory reaction to NMDA receptor hypofunction, which is corroborated by the present lowered serum BDNF levels in the schizophrenia group and by other studies showing that phencyclidine-induced psychotic symptoms can be ameliorated by administration of glycine. It is speculated that in CP, glycine levels are elevated as a compensatory reaction to NMDA receptor hypofunction, whereas patients with schizophrenia would have insufficient neuroplasticity to produce such a response. This hypothesis is corroborated by the present lowered serum BDNF levels in the schizophrenia group and by other studies showing that phencyclidine-induced psychotic symptoms can be ameliorated by administration of glycine. Further support is found in earlier studies that show beneficial effects of adjuvant high-dose glycine augmentation to other anti-psychotic agents.

In patients with other psychotic disorders, both BDNF and glycine levels did not differ from controls. It could be speculated that, even in the presence of sufficient neuroplasticity, the ability to adaptively increase glycine levels might be characteristic for CP.

Table 3 Blood levels of biochemical parameters in CP, schizophrenia, and controls

<table>
<thead>
<tr>
<th></th>
<th>CP (n=10)</th>
<th>Schizophrenia (n=33)</th>
<th>Controls (n=75)</th>
</tr>
</thead>
<tbody>
<tr>
<td>BDNF (μg/L)</td>
<td>20.4±7.1</td>
<td>19.6±5.8***</td>
<td>24.4±6.7</td>
</tr>
<tr>
<td>BDNF 6 weeks (μg/L)</td>
<td>19.9±8.1</td>
<td>19.1±6.0***</td>
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</tr>
<tr>
<td>HVA baseline (nmol/L)</td>
<td>60.4±21.3</td>
<td>58.5±19.7</td>
<td>53.4±13.7</td>
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<tr>
<td>HVA 6 weeks (nmol/L)</td>
<td>51.8±20.4</td>
<td>52.1±15.8</td>
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<tr>
<td>Glutamate baseline (μmol/L)</td>
<td>56.3±24.8**</td>
<td>69.5±30.5***</td>
<td>34.4±16.1</td>
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<tr>
<td>Glutamate 6 weeks (μmol/L)</td>
<td>64.9±44.6**</td>
<td>66.2±30.9***</td>
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<tr>
<td>Glycine baseline (μmol/L)</td>
<td>292.2±96.9†</td>
<td>215.1±62.3*</td>
<td>224.2±47.8</td>
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<tr>
<td>Glycine 6 weeks (μmol/L)</td>
<td>280.9±86.2*</td>
<td>223.7±62.3*</td>
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<tr>
<td>Tryptophan baseline (μmol/L)</td>
<td>37.4±14.0**</td>
<td>48.3±11.5**</td>
<td>47.5±7.7</td>
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<td>Tryptophan 6 weeks (μmol/L)</td>
<td>38.1±10.8**</td>
<td>43.5±11.7</td>
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<tr>
<td>Trp/LNAAs ratio baseline</td>
<td>7.1±1.4***</td>
<td>7.7±1.5*</td>
<td>8.6±1.5</td>
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<tr>
<td>Trp/LNAAs ratio 6 weeks</td>
<td>7.4±1.6*</td>
<td>7.8±1.7*</td>
<td></td>
</tr>
<tr>
<td>Phenylalanine baseline (μmol/L)</td>
<td>52.4±10.2</td>
<td>63.4±15.1*</td>
<td>56.3±8.9</td>
</tr>
<tr>
<td>Phenylalanine 6 weeks (μmol/L)</td>
<td>51.7±10.1</td>
<td>55.3±11.2</td>
<td></td>
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<tr>
<td>Isoleucine baseline (μmol/L)</td>
<td>69.1±22.1</td>
<td>82.7±32.8</td>
<td>71.0±22.2</td>
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<tr>
<td>Isoleucine 6 weeks (μmol/L)</td>
<td>63.6±19.6</td>
<td>71.8±22.1</td>
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<tr>
<td>Leucine baseline (μmol/L)</td>
<td>121.7±36.0</td>
<td>158.5±60.0*</td>
<td>130.3±32.4</td>
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<td>Leucine 6 weeks (μmol/L)</td>
<td>123.3±39.5</td>
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<td>Valine baseline (μmol/L)</td>
<td>218.7±65.2</td>
<td>267.6±76.1</td>
<td>245.3±56.6</td>
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<td>Valine 6 weeks (μmol/L)</td>
<td>218.7±49.8</td>
<td>239.6±58.0</td>
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<tr>
<td>Tyrosine baseline (μmol/L)</td>
<td>61.5±14.9</td>
<td>72.2±25.6</td>
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<td>Tyrosine 6 weeks (μmol/L)</td>
<td>61.5±18.7</td>
<td>62.1±15.4</td>
<td></td>
</tr>
</tbody>
</table>

Notes: *Difference versus controls (Mann–Whitney U), P<0.05. **Difference versus controls (Mann–Whitney U), P<0.01. ***Difference versus controls (Mann–Whitney U), P<0.001. 
#Difference between schizophrenia and CP (Mann–Whitney U), P<0.05. Data presented as mean ± SD.

Abbreviations: CP, cycloid psychoses; BDNF, brain-derived neurotrophic factor; HVA, homovanillic acid; Trp/LNAAs, tryptophan/large neutral amino acids; SD, standard deviation.

Table 4 Blood levels of biochemical parameters in patients with other psychotic disorders (n=15) and controls (n=75)

<table>
<thead>
<tr>
<th></th>
<th>Baseline</th>
<th>6 weeks</th>
<th>Controls</th>
</tr>
</thead>
<tbody>
<tr>
<td>BDNF (μg/L)</td>
<td>21.9±8.1</td>
<td>20.4±7.6</td>
<td>24.4±6.7</td>
</tr>
<tr>
<td>HVA (nmol/L)</td>
<td>75.5±36.6</td>
<td>63.5±23.3</td>
<td>53.4±13.7</td>
</tr>
<tr>
<td>Glutamate (μmol/L)</td>
<td>51.1±13.9*</td>
<td>50.9±16.6*</td>
<td>34.4±16.1</td>
</tr>
<tr>
<td>Glycine (μmol/L)</td>
<td>268.5±137.5</td>
<td>263.7±124.9</td>
<td>224.2±47.8</td>
</tr>
<tr>
<td>Tryptophan (μmol/L)</td>
<td>47.4±9.6</td>
<td>46.0±7.8</td>
<td>47.5±7.7</td>
</tr>
<tr>
<td>Trp/LNAAs ratio</td>
<td>7.9±2.2</td>
<td>8.0±1.6</td>
<td>8.6±1.5</td>
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<tr>
<td>Phenylalanine (μmol/L)</td>
<td>59.8±12.0</td>
<td>58.3±10.4</td>
<td>56.3±8.9</td>
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<tr>
<td>Isoleucine (μmol/L)</td>
<td>81.1±23.1</td>
<td>72.7±22.3</td>
<td>71.0±22.2</td>
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<tr>
<td>Leucine (μmol/L)</td>
<td>149.4±43.2</td>
<td>141.6±38.7</td>
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<tr>
<td>Valine (μmol/L)</td>
<td>264.4±52.4</td>
<td>252.1±65.1</td>
<td>245.3±56.6</td>
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<tr>
<td>Tyrosine (μmol/L)</td>
<td>73.7±21.1</td>
<td>70.3±20.7</td>
<td>63.9±17.0</td>
</tr>
</tbody>
</table>

Notes: *Difference versus controls (Mann–Whitney U), P<0.001. Data presented as mean ± SD.

Abbreviations: BDNF, brain-derived neurotrophic factor; HVA, homovanillic acid; Trp/LNAAs, tryptophan/large neutral amino acids; SD, standard deviation.
The observation that plasma levels of glutamate are increased in all patient groups as compared to controls is in accordance with reports that relate elevated glutamate levels with psychosis and psychotic relapse, albeit that decreased levels in schizophrenia and bipolar disorders have also been reported.\textsuperscript{29,45,46} Elevated glutamate levels are hypothesized to be the result of NMDA receptor hypofunction, leading to diminished glutamatergic neurotransmission and to the evolution of schizophrenic symptoms, including negative and cognitive symptoms.\textsuperscript{15,16}

The Trp/LNAAs ratio was significantly lower in CP and schizophrenia as compared to controls. This finding is suggestive of a decreased central serotonergic activity in both patient groups. The lower tryptophan levels in CP might point to an increased breakdown of tryptophan via the kynurenine pathway. One of the products in this pathway is kynurenine acid, which is not only an antagonist of the NMDA receptor but also a neuroprotectant.\textsuperscript{47,48} It could be speculated that in CP, other than in schizophrenia, this neuroprotective mechanism is still activated which would correspond with better neuroplastic properties and a more favorable course of disease in CP.

Apart from small sample size and the use of peripheral measurements only, a limitation can be identified in that a substantial number of patients were treated earlier with a wide range of FGA and SGA agents. The latter, however, most probably did not interfere with the results, since no influences were found regarding medication status in the two patient groups.

Conclusion

In both CP and schizophrenia, systemic glutamate metabolism and the Trp/LNAAs ratio were altered, which could be related to changes in glutamate signaling. Moreover, differences in especially glycine between CP and schizophrenia were found, which may point at a better neuroplasticity in CP than in schizophrenia. This may be in line with better clinical outcome in CP. Therefore, it is crucial to identify CP as a separate group of disorders within the psychosis spectrum.

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Disclosure

The authors declare that they have no conflicts of interest in this work.

References


