Enhanced effect of fibroblast growth factor-2-containing dalteparin/protamine nanoparticles on hair growth

Yuki Takabayashi¹
Masaki Nambu¹
Masayuki Ishihara²
Masahiro Kuwabara¹
Koichi Fukuda³
Shingo Nakamura²
Hidemi Hattori³
Tomoharu Kiyosawa¹

¹Department of Plastic and Reconstructive Surgery, ²Division of Biomedical Engineering, Research Institute, National Defense Medical College, Tokorozawa, Saitama, Japan

Purpose: Although treatments for alopecia are in high demand, not all treatments are safe and reliable. Dalteparin/protamine nanoparticles (D/P NPs) can effectively carry growth factors (GFs) such as fibroblast GF (FGF)-2. The purpose of this study was to identify the effects of FGF-2-containing D/P NPs (FGF-2&D/P NPs) on hair growth.

Patients and methods: In this study, the participants were 12 volunteers with thin hair. One milliliter of FGF-2 (100 ng/mL) and D/P NPs (56 μg/mL) was applied and massaged on the skin of the scalp by the participants twice a day. They were evaluated for 6 months. Participants were photographed using a digital camera for general observation and a hair diagnosis system for measuring hair diameter.

Results: The mean diameter of the hairs was significantly higher following the application of FGF-2&D/P NPs for 6 months. Objective improvements in thin hair were observed in two cases. Nine participants experienced greater bounce and hair resilience.

Conclusion: The transdermal application of FGF-2&D/P NPs to the scalp can be used as a new treatment for alopecia.

Keywords: hair growth, dalteparin/protamine nanoparticles, fibroblast growth factor, transdermal application

Introduction

Hair loss is not a life-threatening event, and the number of patients suffering from hair loss has dramatically increased. Hair loss affects the social interactions and the psychological well-being of the patient. Currently, two anti-hair loss drugs, ie, finasteride and minoxidil, are available. These drugs have been used clinically, but their effect is reported to be limited, transient, and somewhat unpredictable.¹ Therefore, it is important to develop an effective pharmacological treatment for hair loss.

Hair growth is a complex and cyclically controlled process categorized by the following three phases: anagen, a finite period of hair fiber production; catagen, a brief regression phase; and telogen, a resting phase.²,³ The precise mechanism regulating the hair growth cycle has not been fully characterized. However, it is well known that both the continuation and transition of each phase result from the elaborate interactions between mesenchymal dermal papillae (DP) and adjacent follicular epithelium, the matrix, and the outer root sheath.³,⁴ Particularly, the DP cells located at the base of the hair follicle and surrounded by the matrix indicated the potential to induce hair follicle formation.⁵ The hair follicles, including DP cells, are organs, which are responsible for the production of hair fibers by inducing and maintaining the anagen...
phase. Various cytokines and growth factors (GFs) are involved in the regulation of hair follicles and hair growth cycle. Specifically, fibroblast GF (FGF)-2 has been shown to promote DP cell proliferation and increase the hair follicle size in mice.6,7 FGF-2 along with other GFs has been reported to have a protective effect on neomycin-induced hair loss.8 A study on the controlled release of FGF-2 has also demonstrated that hydrogel enabled FGF-2 to positively act on the hair growth cycle of mice.9 Polyelectrolyte complexes (PECs) are generated by electrostatic interactions between oppositely charged polyelectrolytes, that is, dalteparin (low-molecular-weight heparin) and protamine. Nonstoichiometric PECs are produced when this interaction occurs at nonequivalent ratios. This causes each PEC particle to carry an excess charge.10,11 Proteins interact with both synthetic and natural PECs.12 Heparin and dalteparin specifically interact with functional proteins, including GFs, cytokines, extracellular matrix components, and adhesion molecules, with high affinity.13,14 Thus, heparin may be useful as a therapeutic agent in various pathological conditions involving functional proteins. However, high-dose heparin cannot be used on account of high risk of bleeding.15 In contrast, dalteparin (~5,000 Da) has pharmacological and practical advantages compared to heparin. The lower protein-binding activity of dalteparin produces a low, stable, and predictable anticoagulant response. This bypasses the need for laboratory monitoring of drug levels to adjust the dosage.16 In addition, given the longer plasma half-life, one or two subcutaneous injections per day were sufficient to maintain therapeutic concentrations.16 Conversely, protamine, a purified mixture of proteins, was obtained from fish sperm, neutralized heparin, and dalteparin by forming a stable complex that lacked anticoagulant activity.16 Protamine was also employed in clinical use to reverse the anticoagulant activity of heparin following cardiopulmonary bypass. It was also employed in clinical use to reverse the anticoagulant activity of heparin following cardiopulmonary bypass. The D/P NPs were prepared by modifying the method as described earlier.20,25 To summarize, 2.5 mL of dalteparin solution (Fragmin: 6.4 mg/mL; Kissei Pharmaceutical Co., Tokyo, Japan), followed by 1.2 mL of protamine solution (10 mg/mL; Mochida Pharmaceutical Co., Tokyo, Japan) was added into 500 mL of saline (Otsuka Normal Saline; Otsuka Pharmaceutical Co., Ltd. Tokyo, Japan). The white solution was mixed vigorously. One milliliter of recombinant human FGF-2 (50 μg/mL; Fiblast; Kaken Pharmaceutical Co., Tokyo, Japan) was added to 500 mL of saline to produce diluted FGF-2&D/P NPs. The final concentrations of FGF-2 and D/P NPs were estimated at 100 ng/mL and 56 μg/mL, respectively. Under these conditions, almost all FGF-2 molecules were immobilized on the D/P NPs because ~5 μg of FGF-2 was immobilized on 1 mg of D/P NPs.11,17 The FGF-2&D/P NPs were stored at 4°C.

Measurement of particle size and zeta charge
Particle size and zeta charge of FGF-2&D/P NPs were measured by ELSZ-1000 (Otsuka Electronics Co. Ltd., Osaka, Japan). Weekly measurements were performed for 10 weeks. FGF-2&D/P NPs were stored at 25°C (room temperature) for 10 weeks.

Mitogenic activity of standalone FGF-2 and FGF-2&D/P NPs on human microvascular endothelial cells
In this study, FGF-2 (100 ng/mL), D/P NPs (56 μg/mL), and FGF-2 (100 ng/mL) and D/P NPs (56 μg/mL) were incubated either at 4°C or 37°C for 2 weeks and diluted with Dulbecco’s Modified Eagle’s Medium (DMEM) containing 5% fetal bovine serum (FBS). Human microvascular endothelial cells (HMVECs) from Takara Bio Inc., Shiga, Japan manufactured...
The penetration of FGF‑2&D/P NPs in rat skin
Inbred male Fischer 344 rats (Japan SLC, Inc., Shizuoka, Japan) weighing 170–190 g were used in all the experiments. All rats were maintained on a standard laboratory diet with water ad libitum. Rats were anesthetized by intraperitoneal injections of 0.15 mg/kg medetomidine, 2 mg/kg midazolam, and 5 mg/kg butorphanol tartrate. The dorsal skin hair was shaved and depilated using a depilatory cream. Intraperitoneal injections of 0.15 mg/kg atipamezole roused the rats from the anesthesia. FGF‑2&D/P NPs, standalone FGF‑2, or saline (control) was applied to the dorsal skin twice daily for 3 days. In this experiment, each solution (0.2 mL) was rubbed into the skin, and then 0.1 mL of solution was in contact with the skin for 20 minutes or more to wake up from the anesthesia. On the third day, the rats were killed and their skin samples were taken.

Skin specimens were fixed in 10% formalin solution (Wako Pure Chemical Industries, Ltd., Osaka, Japan), which were embedded in paraffin and sectioned at 4 μm. The sections were mounted onto glass slides and stained with FGF-2 antibody, which was performed at Kyodo Byori Inc. (Hyogo, Japan). All animal experiments were approved and carried out following the approval and guidelines for animal experimentation set by the National Defense Medical College, Saitama, Japan.

Participants of clinical research
The participants were 12 individuals (eleven men, one woman) with ages ranging from 41 years to 77 years (mean age, 58.3 years) with thin hair and without any treatment of thin hair for at least 6 months. The participants visited the hospital monthly to receive inspections and bottles of FGF‑2&D/P NPs. Participants were photographed using a digital camera for general observation and a hair diagnosis system (Folliscope; LeadM Corp, Seoul, South Korea) for measuring hair diameter. All research protocols were submitted and approved by the ethics committee of the National Defense Medical College (clinical trial No. 1148 and No. 1241) and registered in the University Hospital Medical Information Network Center (UMIN000011748). Each participant was informed of the protocols and signed a written consent form before participating in the study.

The application and evaluation of FGF‑2&D/P NPs on the skin of the scalp
One milliliter of FGF‑2&D/P NPs was applied and massaged on the skin of the scalp by participants twice a day. Monthly observations were conducted on the participants for 6 months. The Folliscope captured ten dermoscopic images at 1 cm intervals toward the front left from the parietal region. The diameters of all hair shafts in each dermoscopic image were measured. The mean diameter of all hairs and the percentage of terminal hair (with diameter >0.040 mm) were calculated using the measured data.

All values are expressed as mean ± SD. Statistical analysis was performed with Wilcoxon signed-rank test using JMP Pro 11.2.0 software (SAS Institute Inc., Cary, NC, USA). *P* < 0.05 was considered statistically significant.

Results
Measurement of particle size and zeta charge
Measurements for particle size and zeta charge of FGF‑2&D/P NPs were performed every week for 10 weeks at 25°C (room temperature). The diameter of the FGF‑2&D/P NPs increased from ~170 nm to 210 nm during the first 1–2 weeks, and thereafter it was stable (~210–220 nm). The zeta charge was also stable (~−25 mV to −28 mV; Figure 1). In addition, the morphology of the FGF‑2&D/P NPs was unchanged for 10 weeks at 25°C (data not shown). Thus, the FGF‑2&D/P NPs were stable in terms of particle size, zeta charge, and morphology for at least 10 weeks at 25°C.
Mitogenic activity of standalone FGF-2 and FGF-2&D/P NPs on HMVECs

In this study, FGF-2, D/P NPs, and FGF-2&D/P NPs were incubated at 4°C or 37°C for 2 weeks, and diluted with DMEM containing 5% FBS. HMVECs were cultured and growth stimulated in DMEM containing 5% FBS, FGF-2 (final concentration: 5 ng/mL), and/or D/P NPs (final concentration: 2.8 μg/mL). Standalone D/P NPs incubated at both 4°C and 37°C for 2 weeks showed no cell growth-stimulatory effect. Although the FGF-2 without D/P NPs incubated at 37°C lost the cell growth-stimulatory effect, the FGF-2 without D/P NPs at 4°C retained a partial cell growth-stimulatory effect. The FGF-2&D/P NPs incubated at both 4°C and 37°C maintained the cell growth-stimulatory effect. Thus, D/P NPs have a stabilizing effect on FGF-2 bioactivity (Figure 2).

Penetration of FGF-2&D/P NPs in rat skin

FGF-2&D/P NPs, standalone FGF-2, or saline was applied to the dorsal skin without any complications. The dorsal skin showed no signs of acute inflammation, skin redness, or swelling during the experimental period (3 days). There were no differences in skin appearances between FGF-2&D/P NPs, standalone FGF-2, and saline (control) conditions.

**Figure 1** Diameter and zeta charge of FGF-2&D/P NPs stored at room temperature.

**Notes:** Measurements were performed weekly for 10 weeks. The diameter of FGF-2&D/P NPs was stable (~210–220 nm), and the zeta charge was also stable (~−25 mV to ~−28 mV).

**Abbreviations:** D/P, dalteparin/protamine; FGF-2, fibroblast growth factor-2; NPs, nanoparticles; W, week.

**Figure 2** The protective effect of D/P NPs on FGF-2 bioactivity.

**Notes:** FGF-2, D/P NPs, and FGF-2&D/P NPs were incubated at 4°C or 37°C for 2 weeks. The growth-stimulatory activity for HMVECs in vitro was stable for at least 2 weeks at 37°C compared to the growth-stimulatory activity for those without D/P NPs. Positive control implies fresh FGF-2. Negative control indicates the absence of FGF-2 and D/P NPs.

**Abbreviations:** D/P, dalteparin/protamine; FGF-2, fibroblast growth factor-2; HMVECs, human microvascular endothelial cells; NPs, nanoparticles; OD, optical density.
Immunohistological evaluations were performed to confirm the skin penetration of FGF-2&D/P NPs. Figure 3 shows the representative microphotographs of the FGF-2 antibody staining for saline (control), standalone FGF-2, and FGF-2&D/P NPs applied to the dorsal skin on day 3. The group that received FGF-2&D/P NPs applications was stained more strongly than the group that received standalone FGF-2 and saline application (control). Hence, we concluded that FGF-2&D/P NPs had deeper penetration in rat skin.

Discussion

Various cytokines and GFs are involved in the regulation of hair follicles and hair growth cycle. In particular, FGF-2 was found to act on hair follicles as a regulator of hair growth cycle. However, when GFs are directly applied to the scalp, the local biological activity of FGF-2 may be limited in vivo by the rapid clearance. Furthermore, it is unlikely that direct application of GFs to the scalp penetrates the pores and hair follicles, since FGF-2 molecule is aqueously dissolved through hydration reaction.

In this article, we reported about D/P NPs, which were originally prepared as PECs. The D/P NPs were specifically bound to FGF-2, hepatocyte growth factor, and other GFs in PRP. The local injection of D/P NPs provided an excellent carrier that immobilized, retained, and gradually released various GFs from the PRP for the induction of hair follicles. The local injection of GFs from PRP&D/P NPs also had a substantial ability to enhance vascularization and fibrous tissue formation, flap survival rate, and healing in split-thickness skin graft donor sites. Results of the clinical study on the injection of GFs from PRP&D/P NPs indicated significant improvements in hair thickness following localized injections (at least five times). However, the treatment was extremely painful. Thus, the development of a reliable, efficient, and safe transdermal FGF-2 delivery system using D/P NPs may present potential applications for hair growth.

The D/P NPs actually protect FGF-2 from heat and proteolytic inactivation and enhance their biological activities. In addition, FGF-2 immobilized onto D/P NPs was released with a half release time of ~7 days in vitro. Typically, NPs tend to be assemble with each other and grow to form larger micro-assemblies. This study showed that the diameter of FGF-2&D/P NPs was stable (~200–250 nm), and that the zeta charge was also stable (~25 mV to ~28 mV; Figure 1). Furthermore, the morphology of FGF-2&D/P NPs (data not shown) and the growth-stimulatory activity for HMVECs in vitro (Figure 2) were stable for at least 2 weeks at 37°C
compared to those without D/P NPs. Thus, FGF-2&D/P NPs may be a stable colloidal dispersion that is suitable as an effective hair grower.

Several techniques are employed to improve the transdermal absorption of drugs. These include liposomes, hydrogels, ointments, creams, and patches incorporating various permeation enhancers. Recently, strategies using NPs were developed and investigated as nanomedicines to improve skin penetration. The particle size influences the functionality of the nanomedicines in terms of skin permeability, residence time, adherence, and degradation. The immunohistological results in this study indicate that a small part of FGF-2&D/P NPs may penetrate into skin that has pores which are apertures of hair follicles. This is because FGF-2 immunostainings in FGF-2&D/P NPs-applied group were higher than those of the group that received FGF-2 application without D/P NPs as well as the control groups (Figure 3).

The clinical study showed that hair growth in terms of hair diameter and percentage of terminal hair were signific-
cantly enhanced after the treatment with FGF-2&D/P NPs for 6 months (Figure 4). Thus, the transdermal application of FGF-2&D/P NPs to the scalp may be used as a new treatment for alopecia. Furthermore, because all the components of FGF-2&D/P NPs, ie, dalteparin, protamine, and FGF-2, are in clinical use, it is likely that the FGF-2&D/P NPs are safe for clinical use.

Conclusion
Basic and clinical research was conducted using FGF-2&D/P NPs by itself in 12 participants (including one woman) with thin hair. The diameter, the zeta charge, and the growth-stimulatory activity were stable for HMVECs in vitro of FGF-2&D/P NPs. The hair diameter and percentage of terminal hair significantly increased after the administration of FGF-2&D/P NPs for 6 months. The method with FGF-2&D/P NPs is simpler and cheaper than conventional methods.

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Disclosure
The authors report no conflicts of interest in this work.

References

