Mixed cryoglobulinemia-associated Sjögren's syndrome leading to spontaneous rupture of the kidney: a case report

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Background: Spontaneous rupture of the kidney is uncommon and is mainly caused by renal tumors. Only a few cases are caused by vasculitis. We report here the first case of spontaneous rupture of kidney resulting from mixed cryoglobulinemia.

Case presentation: A 44-year-old man presented with sudden onset of fever, acute pulmonary edema, left flank abdominal pain unassociated with trauma, and rapidly progressive renal failure requiring dialysis. Computed tomography of the abdomen revealed a large perirenal hematoma of the left kidney. During conservative surgery, the patient underwent renal biopsy that showed renal vasculitis and membranoproliferative glomerulonephritis with intracapillary microthrombi. Tests were positive for mixed cryoglobulinemia caused by Sjögren's syndrome. The patient was better after immunosuppressive therapy, with the disappearance of clinical symptoms and the recovery of baseline renal function.

Conclusion: We report on this case and discuss a possible link between spontaneous rupture of kidney and mixed cryoglobulinemia-associated Sjögren's syndrome.

Keywords: mixed cryoglobulinemia, Sjögren's syndrome, spontaneous rupture of kidney, renal hematoma, Wunderlich syndrome

Introduction

Spontaneous rupture of kidney (SRK) is uncommon. In most cases, it was a complication of benign or malignant kidney tumors. Only a few cases are a result of systemic vasculitis, particularly polyarteritis nodosa (PAN), microscopic polyangiitis, and in sporadic reports, Wegener's granulomatosis. To our knowledge, SRK has never been associated with mixed cryoglobulinemia.

Case presentation

A 44-year-old white man was admitted to our hospital because of acute pulmonary edema. His past medical history was significant for chronic tobacco use and recurrent episodes of arthralgias over the course of 2 years, which were treated symptomatically by the patient by self-medication with analgesics, without any specific medical management.

The patient had been well until 2 months earlier, when he began to have worsening of his physical state, anorexia, weakness, weight loss, and diffuse myalgias. Three days before admission, he complained of acute left flank pain unassociated with trauma. It was sudden in onset, intense, and associated with nausea. In addition, the patient reported a significant decrease of urine volume (<0.5 L urine in 24 hours) and dyspnea. Physical examination showed high blood pressure (170/100 mmHg), his temperature

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was 38.7°C, and his respiratory rate was 34 breaths per minute. His weight was 71 kg (he weighed 81 kg 2 months earlier). Chest auscultation revealed bilateral rales. Cardiac examination findings were unremarkable. The abdomen was tender, mainly over the left upper quadrant, and had no palpable hepatosplenomegaly. He had 1+ lower-extremity edema. Exam of the lower limb found petechial purpura on his left leg, with livedo reticularis. Otherwise, the examination was normal.

Laboratory findings revealed a rapidly progressive renal failure with a serum creatinine level of 1.71 mg/dL (normal value, 0.6–1.2 mg/dL), which increased to 4.02 mg/dL over the next 2 days; a blood urea nitrogen level of 57 mg/dL (normal value, 7–20 mg/dL), which also increased to 123 mg/dL; a hypervolemic hyponatremia of 124 mEq/L (normal value, 135–145 mEq/L); and normal serum electrolytes otherwise. Twenty-four hour urinary protein levels were 0.6 g/24 hours (normal value, <0.03 g/24 hours), and urinary sediment was positive for microscopic hematuria. Hemoglobin was 9.7 g/dL (normal value, 13–16 g/dL), white blood cell count was 18,300/mm³ (normal value, 4,000–1,000/mm³), and platelets were 626,000/mm³ (normal value, 150,000–300,000/mm³). Erythrocyte sedimentation rate was 80 mm/hour (normal value, <15 mm/hour), with a C-reactive protein level of 11.3 mg/L (normal value, 6–12 mg/L). Aminotransferase and creatinine kinase levels were normal. Lactate dehydrogenase levels were elevated (1,243 U/L; normal value, <250 U/L). CH50 was not checked, but complement component C3 was normal, at 80 mg/dL (normal, 80–170 mg/dL), and C4 was strongly reduced, at 6 mg/dL (normal, 16–40 mg/dL). Rheumatoid factor and antinuclear antibodies were present. There was no monoclonal paraprotein detected both on serum and urine protein immuno-electrophoresis. In addition, five blood samples for cultures (both aerobic and anaerobic)

were sent to the laboratory in view of the patient's fever, leukocytosis, and high erythrocyte sedimentation rate, all of which were negative.

Chest radiography showed cardiomegaly with bilateral pulmonary infiltrates, suggesting acute pulmonary edema. Renal ultrasound showed normal-sized kidneys without evidence of hydronephrosis, but it allowed us to identify a subcapsular renal hematoma that measured 63×52 mm in the upper pole of the left kidney, which was confirmed by abdominal computed tomography (CT) scan (Figure 1A). Multislice computed angiotomography scanning identified the hematoma and ruled out microaneurysms of the main renal arteries, other vascular abnormalities, and renal tumors (Figure 1B). Hemodialysis therapy was started because of oliguria, continuous decrease of renal function, and persisting pulmonary edema despite high doses of furosemide (500 mg/day), and then the patient was operated on with conservative surgery for renal hematoma. A drain was placed and then withdrawn after 3 days.

Renal biopsy was performed, during surgery, the third day after admission and was submitted for light microscopic and immunofluorescence examination (Figure 2). The specimen consisted of renal medulla and cortex with 14 glomeruli. One glomerulus was globally sclerosed. Some glomeruli were ischemic. The preserved glomeruli were enlarged and showed diffuse mesangial expansion. There was significant hypercellularity of glomeruli with numerous infiltrating neutrophils, minimal endocapillary proliferation, and one cellular crescent. There were thickening and reduplication of the glomerular basal membrane, suggesting membranoproliferative glomerulonephritis. In two glomeruli, we noted intracapillary amorphous deposits compatible with microthrombi. Small vessels showed extensive necrotizing angitis. Some vessels had also leukocytoclastic angitis.

Figure 1 Renal computed tomography.

Notes: (A) Enhanced abdominal computed tomography scan shows rupture of the left kidney with perirenal hematoma (green arrow). The point of rupture is clearly seen (red arrow). (B) Sagittal reconstruction of the computed angiotomography scan showing normal left renal artery with avascular upper pole of left kidney corresponding to the area of rupture (blue arrow).

Figure 2 Renal biopsy specimen.

Notes: (A, B and D) Membrano proliferative glomerulonephritis with intracapillary microthrombi. (A) Trichrome stain; original magnification, $\times 400$. (B) Eosin and hematoxylin stain; original magnification, $\times 400$. (D) Jones silver methenamine stain; original magnification, $\times 600$. (C) Extensive necrotizing arteritis affecting an interlobular artery. Eosin and hematoxylin stain, magnification $\times 400$.

There was considerable tubular necrosis and interstitial edema with minimal leukocyte infiltrates. Unfortunately, the immunofluorescence sample did not contain glomeruli.

The patient underwent an extensive workup to determine the cause of his renal vasculitis. The laboratory tests were sent immediately after we received the kidney biopsy results, 72 hours after the biopsy had been performed. Serological studies for antineutrophil cytoplasmic antibodies, antiglomerular basement membrane antibodies, anti-double-stranded DNA antibodies, and anti-Smith antibodies were negative, excluding respectively pauci-immune vasculitis, Goodpasture's syndrome, and lupus vasculitis. There was neither apparent sepsis nor visceral abscess on computed tomography scan. Transthoracic echocardiography showed no vegetations, thus excluding infectious endocarditis. Skin-biopsy specimen of a purpuric lesion showed leukocytoclastic vasculitis without immunoglobulin A deposition, and therefore did not support the diagnosis of Henoch-Schönlein purpura. Tests for cryoglobulinemia were positive, including the presence of cryocrit (12%) with type II mixed cryoglobulin (immunoglobulin G/immunoglobulin M *kappa*). We made additional investigations to determine the cause of mixed cryoglobulinemia, and assessment for hepatitis C virus (HCV) RNA and hepatitis B virus DNA by

polymerase chain reaction was negative. Tests for Sjögren's syndrome were positive with the presence of anti-Ro (SS-A), anti-La (SS-B), lymphocytic infiltrate in minor salivary gland biopsy (grade 3 according to Chisholm and Mason grading system), and positive Schirmer's test. Screening for lymphoma remained negative despite initial elevated lactate dehydrogenase levels. There were neither peripheral or visceral adenopathies nor lymphomatic infiltration on osteomedullary biopsy. All these data confirmed that the patient had SRK caused by type II mixed cryoglobulinemia-associated Sjögren's syndrome.

The patient received 3 days of pulses of methylprednisolone (1 g/day), followed by oral prednisone (1 mg/kg/day) with a decreasing regimen and 6 monthly pulses of intravenous cyclophosphamide (1 g) as induction of remission. The patient remained febrile during hospitalization, with temperatures ranging between 38.5°C and 38.8°C, until he received the third pulse of methyl prednisolone, which was administered 12 days after admission. Azathioprine (100 mg/day) was started as maintenance treatment. The patient had spectacular improvement of his physical state. Arthralgias and the other symptoms disappeared, proteinuria fell to only trace amounts, and renal function returned to its baseline value (0.9 mg/dL) over the next 2 weeks, and C4 levels

were again normal. Resorption of the renal hematoma was complete over the next 5 weeks. Cryoglobulins remained undetectable. Renal function and lactate dehydrogenase were still normal. Ethical approval was not needed in the present case report, because it didn't include research or experiments. There was no risk of causing harm or violating the patient's right, since it was presented in a way that respected the patient's privacy and confidentiality. The patient gave oral consent when informed that the data will be published.

Discussion

SRK, or Wunderlich syndrome, is a rare entity. It was first reported by Bonet in 1700 and later described in more detail by Wunderlich in 1856.^{1,2} To date, more than 200 cases have been reported in world literature. To the best of our knowledge, no similar case was described previously.

Several causes may lead to SRK. Tumors, particularly renal cell carcinoma and angiomyolipoma, are the most common cause of SRK, occurring in 57%–73% of cases.^{3,4} Vascular disease was reported in a meta-analysis of spontaneous renal hemorrhage as the next most common offender, with PAN occurring most frequently.⁵ Other vasculitis was also reported to be associated with SRK, especially microscopic polyangiitis and Wegener's granulomatosis.^{6–8} Our patient had SRK, probably as a result of vasculitis; however, he had neither PAN nor pauci-immune vasculitis. Microaneurysms and renal vessel thrombosis were not noted in computed angiotomography scan. Furthermore, the patient had also neither renal tumor, both in radiologic and histologic findings, nor renal infection. On the basis of clinical context and laboratory findings, we suggest that SRK was a result of cryoglobulinemic vasculitis associated with Sjögren's syndrome.

Mixed cryoglobulinemia is strongly associated with HCV. Prevalence of HCV infection in patients with mixed cryoglobulinemia ranges from 30% to 96%.^{9,10} The etiologies of negative-HCV mixed cryoglobulinemia are "essential" mixed cryoglobulinemia (27%); lymphoproliferative disorders, especially B-non-Hodgkin's lymphoma and myeloma (26%); autoimmune diseases, particularly Sjögren's disease and lupus (34%); and infectious disease (13%), including hepatitis B virus and HIV.¹¹ In the study of Trejo et al,¹² of 443 patients with cryoglobulinemia, 10% had Sjögren's syndrome. Inversely, in large series of patients with Sjögren's syndrome, mixed cryoglobulinemia, especially type II, was present in 20% of cases and was characterized by a low CH50, cutaneous vasculitis, and poor prognosis resulting from more severe internal organ involvement and frequent evolution to malignant lymphomas.^{13–15}

Vasculitis in Sjögren's syndrome can occur either as a cutaneous localized form (leukocytoclastic vasculitis) or as systemic necrotizing vasculitis involving small-sized and medium-sized arteries of various organs related to the presence of cryoglobulinemia.¹⁶ It is well known that in Sjögren's syndrome type B, there is aneurysmal dilatation of arteries. However, spontaneous rupture was rarely described. To the best of our knowledge, there are only three reports: Achkar et al described first a ruptured aneurysm of the inferior pancreaticoduodenal artery in the setting of rheumatoid vasculitis-associated Sjögren's syndrome,¹⁷ Curi et al reported a contained rupture of the visceral aorta in a patient with Sjögren's syndrome associated with osteoarthritis,¹⁸ and Klingler et al reported a rupture of a spinal artery aneurysm attributable to exacerbated Sjögren syndrome-associated cryoglobulinemic vasculitis.¹⁹ In contrast, it is also known that cryoglobulinemic vasculitis may induce injuries in the vascular wall. Herold et al described cryoglobulinemic vasculitis associated with aortic aneurysms in pediatric cases of Kawasaki disease,²⁰ and Au et al reported structural aortic abnormalities in four patients with negative-HCV cryoglobulinemia, including multiple dissections, aneurysms, and congenital aortic arch abnormalities.²¹ Therefore, we presume that an exacerbation of the Sjögren's syndrome-induced cryoglobulinemic vasculitis with necrotizing arteritis and fibrinoid necrosis of the vessel's media led to weakening of the arterial wall, formation of kidney aneurysm, and spontaneous rupture. This mechanism may also explain the occurrence of microaneurysms and SRK in PAN and in rare cases of microscopic polyangiitis and Wegener's granulomatosis.^{22–26}

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Author contributions

All authors made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; took part in either drafting the article or revising it critically for important intellectual content; gave final approval of the version to be published; and agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Disclosure

The authors report no conflicts of interest in this work.

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