

miR-34b inhibits nasopharyngeal carcinoma cell proliferation by targeting ubiquitin-specific peptidase 22

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Objectives: This study aimed to investigate the precise role of miR-34b in nasopharyngeal carcinoma (NPC).

Materials and methods: The expression of miR-34b and transcription of ubiquitin-specific peptidase 22 (USP22) were examined using quantitative reverse transcription-polymerase chain reaction. Western blot analysis was used to measure the protein expression of USP22. A dual-luciferase assay was used to investigate the interaction between miR-34b and USP22. Cell viability was determined by 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide assay. The cell cycle was analyzed by propidium iodide staining followed by flow cytometry analysis.

Results: miR-34b was significantly downregulated in NPC tissues and NPC cell lines. Overexpression of miR-34b in NPC SUNE-6-10B cells inhibited cell viability and proliferation. USP22 was highly expressed in NPC cells and promoted cell viability and proliferation. Restoration of USP22 expression could reverse the effect of miR-34b on NPC cell viability and proliferation.

Conclusion: miR-34b acts as a tumor suppressor in NPC, which is mediated via repression of the oncogene USP22.

Keywords: cell proliferation, miR-34b, nasopharyngeal carcinoma, overexpression, USP22

Introduction

Nasopharyngeal carcinoma (NPC) derived from the nasopharyngeal epithelium has a high incidence rate in Southeast Asia and Northern Africa.¹ Genetic susceptibility, endemic environmental factors, and Epstein-Barr virus infection are believed to be the major etiological factors of NPC.² Currently, radiotherapy is the most sensitive and effective treatment for NPC, but the prognosis is often not satisfactory due to the rates of recurrence and metastasis.³ Elucidation of the molecular mechanisms underlying its pathogenesis and progression is essential for the development of novel strategies for the diagnosis and treatment of NPC.

MicroRNAs (miRNAs) are noncoding small RNAs that can regulate their target gene expression to modulate cell proliferation, migration, differentiation, and apoptosis.⁴ Abnormal miRNAs can function as either oncogenes or tumor suppressors.⁵ Multiple miRNAs have been shown to be aberrantly expressed and to contribute to the development and progression of NPC.⁶ Although miR-34b has been reported to be a tumor suppressor gene in many types of cancers, its function in NPC remains not well understood. The miRNA expression profile showing that miR-34b is underexpressed in NPC compared to normal tissue samples caught our attention.^{7,8} In a recent study, we compared the expression of miR-34b in NPC cell lines and an immortalized nasopharyngeal epithelial cell line and found that miR-34b was also downregulated

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in NPC cells. Furthermore, we performed a bioinformatics analysis of the target genes of miR-34b and selected ubiquitin-specific peptidase 22 (USP22) as a gene of interest.

USP22, a newly discovered member of the ubiquitin hydrolase family, possesses deubiquitinating activity.⁹ USP22 expression is significantly elevated in tumor tissues, compared with normal tissues,¹⁰ and it has been suggested as a valuable biomarker for predicting the recurrence and metastasis of malignancies.^{11,12} USP22 plays a key role in cell cycle regulation, and interference with USP22 expression in tumor cells can arrest cell cycle progression in the G1 phase and inhibit tumor cell proliferation.^{13,14} USP22 was also identified as a subunit of the Spt-Ada-Gcn5 acetyltransferase coactivator complex, which regulates the expression of genes related to oncogenicity and proliferation.¹⁵ However, little is known about the oncogenic role played by USP22 in the growth of NPC.

In this study, we evaluated the impact of miR-34b overexpression on the proliferation of NPC. Furthermore, we identified the oncogene USP22 as a target gene of miR-34b. Overexpression of USP22 attenuates the inhibitory effect of miR-34b on NPC cell viability and proliferation, thus confirming our hypothesis that miR-34b inhibits NPC proliferation by downregulating USP22.

Materials and methods

Cell lines

All human/animal studies have been approved by the Institute Research Medical Ethics Committee of Guangzhou University of Chinese Medicine. All human studies have been performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki and its later amendments. All persons gave their informed verbal consent prior to their inclusion in the study. Human NPC cell lines CNE-1, CNE-2, SUNE-5-8F, and SUNE-6-10B were preserved in our laboratory and maintained in RPMI-1640 medium (Thermo Fisher Scientific, Waltham, MA, USA) supplemented with 10% fetal bovine serum (Thermo Fisher Scientific) at 37°C in a humidified atmosphere containing 5% CO₂. The control cell line NP69 was purchased from the Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences and maintained in Keratinocyte Serum-Free Medium (Thermo Fisher Scientific) in a humidified atmosphere (37°C, 5% CO₂). When the cells reached the logarithmic growth phase, the subsequent experimental analyses were performed.

RNA isolation and quantitative reverse transcription-polymerase chain reaction

Total RNA was isolated from the cell lines and tissue samples using TRIzol (Thermo Fisher Scientific), according to the

manufacturer's instructions. mRNA was reverse transcribed using the ReverTra Ace qPCR RT Kit (Toyobo Biochemicals, Tokyo, Japan), according to the manufacturer's protocol. Real-time polymerase chain reaction (PCR) was performed with GoTaq® qPCR Master Mix (Promega Corporation, Fitchburg, WI, USA) using a MiniOpticon™ Real-Time PCR detection instrument (Bio-Rad, Hercules, CA, USA) with the SyBr Green detection protocol provided by the manufacturer. Reverse transcription and quantitative reverse transcription-polymerase chain reaction (qRT-PCR) for miRNA were performed using the Stem-Loop miRNA qRT-PCR Primer Set (Forevergen, Guangzhou, People's Republic of China). Data analysis was performed using the 2^{-ΔΔCt} method. The level of USP22 was normalized to the expression of glyceraldehyde 3-phosphate dehydrogenase (GAPDH). The level of miR-34b was normalized to the expression of U6. Sequence-specific primers for USP22 and GAPDH were as follows: USP22, 5'-CCATTGATCTGATGTACGGAGG-3' (forward) and 5'-TCCTTGGCGATTATTTCCATGTC-3' (reverse) and GAPDH, 5'-GAGTCAACGGATTGGTCGT-3' (forward) and 5'-GACAAGCTTCCCGTTCTCAG-3' (reverse). The primers used for miR-34b and U6 were purchased from Forevergen.

Western blot analysis

Cells from all the experimental groups were collected using a cell scraper, and proteins were extracted using cell lysis buffer. The protein concentration was assessed by Bradford assay, and equal amounts of total protein were separated by sodium dodecyl sulfate polyacrylamide gel electrophoresis, transferred to polyvinylidene fluoride membranes, and blocked with 5% skim milk for 1 hour at room temperature. The polyvinylidene fluoride membranes were then washed with TBST (containing NaCl, Tris-HCl, and Tween-20) and incubated overnight with primary antibodies against target proteins at 4°C followed by two washes in TBST. The USP22 antibody was purchased from Abcam (Abcam, Cambridge, MA, USA). The membranes were incubated with the appropriate secondary antibodies for 1 hour at room temperature and then washed three times with TBST. Proteins were visualized by chemiluminescence (Forevergen).

Plasmid construction and transfection

The 3'-untranslated region (3'-UTR) of USP22 genes was cloned into the pmirGlo plasmid (Promega Corporation) between the *SacI* and *XbaI* sites using the following primers: USP22 wild-type 3'-UTR forward primer, 5'-ATGCGAGCTC TAGACAGCCAGGGAGTAAACAC-3' and reverse primer, 5'-ATGCTCTAGATGACATAGGAGAGACTACAAAGC A-3'. The pmirGlo plasmids containing the mutated USP22

3'-UTR were constructed using a KOD-plus Mutagenesis Kit (Toyobo Biochemicals) according to the manufacturer's instructions. Primers for mutated USP22 3'-UTR were as follow: 5'-CAAACTTACTGATAAATTGTCAAAGAACA-3' (forward) and 5'-AATTTATCAGTAAGTTTGT TACTGTAAGTTTG-3' (reverse). The open reading frame expression vectors pCMV-USP22 and the control vector pCMV were purchased from the GeneChem Corporation (GeneChem Corporation, Shanghai, People's Republic of China). All the inserted or mutated sequences were confirmed by sequencing. All the transfection reactions were performed using Lipofectamine 2000 (Thermo Fisher Scientific) in accordance with the manufacturer's instructions.

Transfection of miRNAs and small interfering siRNAs

All the small interfering RNAs (siRNAs) and miRNA mimics were purchased from Guangzhou RiboBio (RiboBio, Guangzhou, People's Republic of China) and transfected into cells using Lipofectamine 2000 (Thermo Fisher Scientific), as recommended by the manufacturer. The cells were transfected with miRNA mimics or siRNA at a concentration of 100 nM. The sequences of the siRNAs were as follows: siRNA-USP22: sense, 5'-CACGGACAGTCTCAACAAT-3' and antisense, 5'-ATTGTTGAGACTGTCCGTG-3'.

3-(4,5-Dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide assay

Harvested cells were seeded in 96-well plates at 5×10^3 cells/well and grown in normal medium for 1 day, 2 days, 3 days, and 4 days. Subsequently, the cells were incubated in 0.1 mg/mL 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) at 37°C for 3 hours and lysed in dimethyl sulfoxide at room temperature for 30 minutes. The absorbance in each well was measured at 490 nm using a microplate reader. Each experiment was performed in triplicate.

Colony forming assay

Colony forming assays were performed as described previously.¹⁶ Cells were seeded into triplicate six-well plates at a density of 1,000 cells/well, and the medium was replaced every 3 days. After 10 days, the cells were washed twice with phosphate-buffered saline (PBS), fixed, and stained with 0.5% crystal violet. The colonies were then counted under a microscope.

Cell cycle analysis by flow cytometry

Cells were trypsinized, collected, and washed three times with PBS. The cells were then fixed in 1 mL of 70% ice-cold ethanol

overnight at 4°C. After two washes with PBS and centrifugation for 10 minutes at 1,000 rpm, the supernatant was discarded and the cell pellets were stained with 50 µg/mL of propidium iodide and 100 U/mL of RNase A and incubated in PBS for 30 minutes at room temperature in dark conditions. The percentages of cells in different phases of the cell cycle were determined by evaluating the DNA content. A total of 10,000 cells/sample were counted. A cytometric analysis was performed with a flow cytometer (BD Biosciences, San Jose, CA, USA) using CellQuest software. Each assay was performed in triplicate.

Dual-luciferase assay

Cells were seeded in 48-well plates 1 day prior to transfection. The miRNA mimics (100 nM) and plasmid (5 ng/mL) were cotransfected into SUNE-6-10B cells. At 48 hours after transfection, the luciferase activity was measured using the Dual-Glo luciferase assay kit (Promega Corporation), according to the manufacturer's instructions.

Statistics analysis

SPSS 17.0 (SPSS Inc., Chicago, IL, USA) was used to analyze the experimental data. All the experiments were performed in triplicate, and the results were analyzed by analysis of variance or Student's *t*-test and are expressed as mean \pm SD. *P*-value <0.05 or <0.01 was considered to be statistically significant.

Results

miR-34b is downregulated in NPC tissue samples and cell lines

To investigate the role of miR-34b in NPC, we first compared its expression levels in NPC and adjacent noncancerous tissue samples (sample size: tumor group: $n=18$ and control group: $n=15$). NPC samples showed significant downregulation of miR-34b compared with adjacent noncancerous tissue samples (Figure 1A). We further compared the expression levels of miR-34b in NPC cell lines (SUNE-5-8F, SUNE-6-10B, CNE-1, and CNE-2) against expression in the immortalized nasopharyngeal epithelial cell line NP69. The qRT-PCR analysis showed that the basal expression levels of miR-34b were generally downregulated in the four NPC cell lines compared with NP69 (Figure 1B). The downregulation of miR-34b in NPC cell lines suggests that miR-34b might be involved in the progression of NPC.

Overexpression of miR-34b inhibits cellular proliferation of NPC

To evaluate whether miR-34b has an effect on NPC cell proliferation, we transfected the NPC cell line SUNE-6-10B

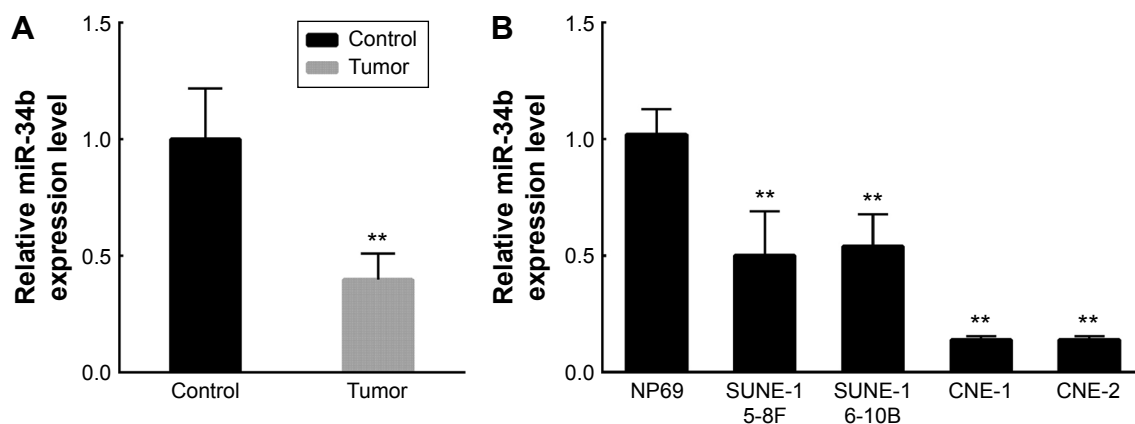


Figure 1 miR-34b expression in NPC tissue samples and cell lines.

Notes: (A) Relative miR-34b expression levels in NPC and adjacent peritumoral tissues. miR-34b displayed a decreased expression level in NPC tissue compared with the adjacent peritumoral tissue (** $P < 0.01$). (B) Relative miR-34b expression levels in various cell lines as determined by qRT-PCR analysis (** $P < 0.01$).

Abbreviations: NPC, nasopharyngeal carcinoma; qRT-PCR, quantitative reverse transcription-polymerase chain reaction.

with miR-34b mimics and negative control (NC) mimics. The qRT-PCR analysis revealed a significant upregulation of miR-34b after transfection (Figure 2A). The MTT assay showed that cell viability was significantly suppressed in miR-34b-transfected cells in comparison with NC-transfected cells (Figure 2B). Cell cycle analysis demonstrated that miR-34b overexpression led to an increased percentage of SUNE-6-10B cells in the G1 phase and a decreased percentage of cells in the S phase, which suggests that miR-34b can induce G1 arrest (Figure 2C and D). To further confirm that miR-34b has an effect on NPC cell proliferation, expression levels of cell cycle-related proteins P21, CyclinD and CyclinE, and apoptosis-related proteins Bcl-2, Bax, and Caspase-3 were analyzed (Figure 2E). P21 was increased, while CyclinD and CyclinE were suppressed in the miR-34b mimics group. In addition, Bax and Caspase were increased and Bcl-2 was decreased in the miR-34b mimics group. These results indicate that miR-34b suppresses the proliferation of NPC cells.

miR-34b targets USP22 to cause its downregulation

In order to gain insight into the mechanism by which miR-34b inhibits NPC, we searched for candidate target genes of miR-34b using bioinformatics and ultimately focused on USP22. TargetScan analysis revealed that the 3'-UTR sequence of USP22 contains one conserved miR-34b binding site (Figure 3A). To confirm that USP22 is a direct target of miR-34b, we cloned the 3'-UTR of USP22 into a reporter plasmid downstream of the luciferase gene. The reporter assays showed that the upregulation of miR-34b significantly

decreased the relative luciferase activity of the wild-type 3'-UTR of USP22, compared with the NC, but had no effect on the mutant USP22 3'-UTR. These results suggest that miR-34b directly binds to the 3'-UTR of USP22 (Figure 3B). Next, we detected the expression of USP22 in SUNE-6-10B cells following transfection with miR-34b mimics and NC mimics. qRT-PCR and Western blot data show that miR-34b transfection significantly suppressed the level of USP22 (Figure 3C and D). These results suggest that USP22 is a direct downstream target gene of miR-34b.

USP22 promotes the proliferation of NPC

qRT-PCR analysis showed that the expression levels of USP22 mRNA were higher in NPC tissues than that in normal nasopharyngeal mucosa (Figure 4A) (sample size: tumor group: $n=18$ and control group: $n=15$). We also examined the expression of USP22 in four NPC cell lines and NP69 cells. Western blot analysis showed that USP22 was highly expressed in four NPC cell lines compared to NP69 cells and exhibited an inverse expression pattern compared with miR-34b (Figure 4B). To explore the function of USP22 in NPC cells, we constructed the USP22 overexpression vector pCMV-USP22 (Figure 4C). The MTT assay showed that overexpression of USP22 increased the viability of SUNE-6-10B cells compared with pCMV (Figure 4D). The cell cycle distribution revealed that a high level of USP22 resulted in a decrease of SUNE-6-10B cells in the G1 phase and an increase of SUNE-6-10B cells in the S phase (Figure 4E and F).

Additionally, we transfected SUNE-6-10B cells with an siRNA against USP22 and observed that USP22 knockdown

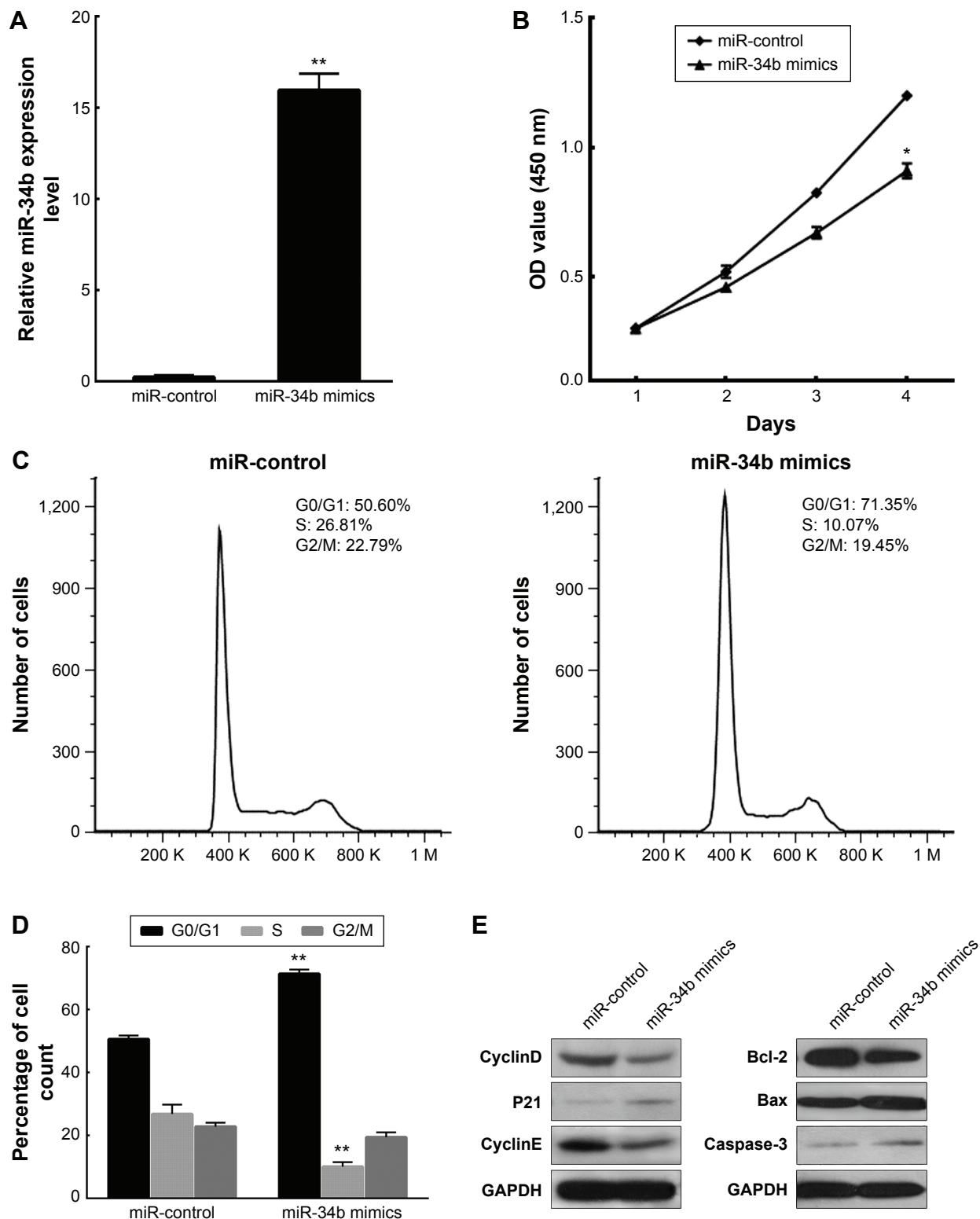


Figure 2 Ectopic overexpression of miR-34b suppressed the proliferation of SUNE-6-10B cells and induced G0/G1 cell cycle arrest in vitro.

Notes: (A) The expression levels of miR-34b in SUNE-6-10B cell lines were verified after transfection with 100 nM miR-34b mimics and miR-control. (B) The effect of miR-34b on cell proliferation was measured using the MTT assay in SUNE-6-10B cell lines (* $P < 0.05$). (C and D) Flow cytometric analysis of the cell cycle distribution of SUNE-6-10B cells transiently transfected with 100 nM miR-34b mimics and the control. (E) The expression levels of P21, CyclinD, CyclinE, Bcl-2, Bax, and Caspase-3 were measured by Western blot analysis. GAPDH was used as an internal control. Each independent experiment was repeated three times. All the experiments were performed in triplicate, and the results were analyzed by ANOVA and are expressed as mean \pm SD (** $P < 0.01$).

Abbreviations: MTT, 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; ANOVA, analysis of variance; OD, optical density.

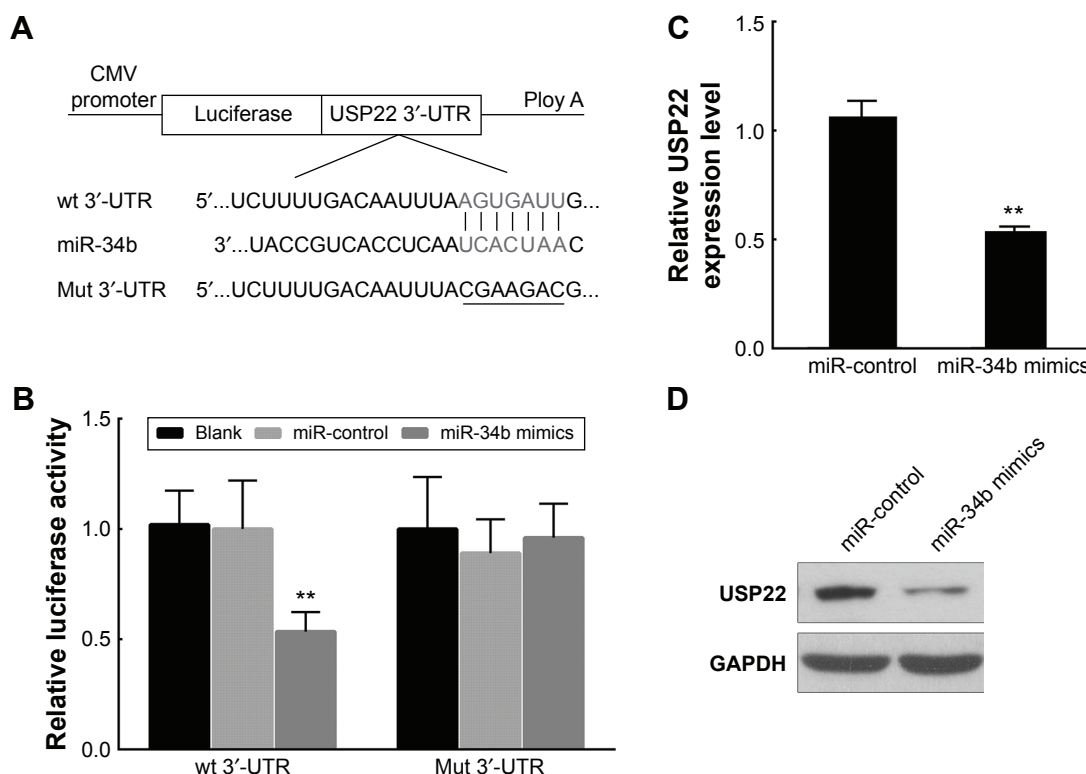


Figure 3 miR-34b targets the 3'-UTR of the USP22 gene.

Notes: (A) Schematic graph of the putative binding sites of miR-34b in the USP22 3'-UTR. Mut 3'-UTR indicates the USP22 3'-UTR with a mutation in the miR-34b binding sites. (B) miR-34b mimics downregulated the activity of a luciferase reporter containing wild-type USP22 3'-UTR (** $P < 0.01$) but not of a reporter containing mutant USP22 3'-UTR. (C) Forty-eight hours after miR-34b mimic and control transfection of SUNE-6-10B cells, the mRNA level of USP22 decreased significantly compared with the miR-control, as determined by qRT-PCR (** $P < 0.01$). (D) Forty-eight hours after miR-34b mimic and control transfection of SUNE-6-10B cells, the protein level of USP22 decreased significantly compared with the miR-control, as determined by Western blot analysis. GAPDH was used as an internal control. Each independent experiment was repeated three times. All the experiments were performed in triplicate. The results were analyzed by ANOVA and are expressed as mean \pm SD.

Abbreviations: 3'-UTR, 3'-untranslated region; USP22, ubiquitin-specific peptidase 22; qRT-PCR, quantitative reverse transcription-polymerase chain reaction; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; ANOVA, analysis of variance; wt, wild type; Ploy A, polyadenylic acid; CMV, Cytomegalovirus.

resulted in an impaired viability of SUNE-6-10B cells compared with the NC (Figure 4G). The MTT assay demonstrated that USP22 knockdown decreased the viability of SUNE-6-10B cells (Figure 4H). Flow cytometry revealed that USP22 siRNA transfection led to an increased percentage of SUNE-6-10B cells in the G1 phase and a decreased proportion of SUNE-6-10B cells in the S phase compared with the siRNA NC (Figure 4I and J).

These results demonstrate that USP22 promotes cell proliferation and plays an oncogenic role in NPC cells.

Overexpression of USP22 attenuates the inhibitory effect of miR-34b on NPC cell proliferation

To investigate whether miR-34b inhibits NPC cell proliferation by targeting USP22, we transfected SUNE-6-10B cells and a cell line that was stably expressing USP22 (Figure 5A) with miR-34b mimics. MTT (Figure 5B and C), cell cycle (Figure 5D and E), and colony formation analyses (Figure 5F) show that overexpression of USP22 can significantly

attenuate the suppressive effect of miR-34b. These results provide further support that USP22 is a downstream functional target of miR-34b.

Discussion

NPC has a high incidence in southern People's Republic of China and is difficult to detect during its early stages due to the lack of symptoms.¹⁷ At later stages, the reduced radiosensitivity and high invasiveness of NPC typically cause poor clinical outcomes.¹⁸ The development of more sensitive diagnostic markers and therapeutic targets are urgently needed. miRNAs, which are abundant in NPC,¹⁹ are ideal candidates because they are differentially expressed in NPC and normal nasopharyngeal tissues.²⁰ Aberrant expression of miRNAs may represent a novel marker in the clinical diagnosis and prognosis of NPC.²¹ Importantly, miRNAs can also play tumor suppressor or oncogenic roles in NPC cells through the downregulation of their downstream target genes.^{16,22–25} In the present study, we evaluated the oncogene USP22 as a potential target gene of miR-34b. Furthermore, we demonstrated

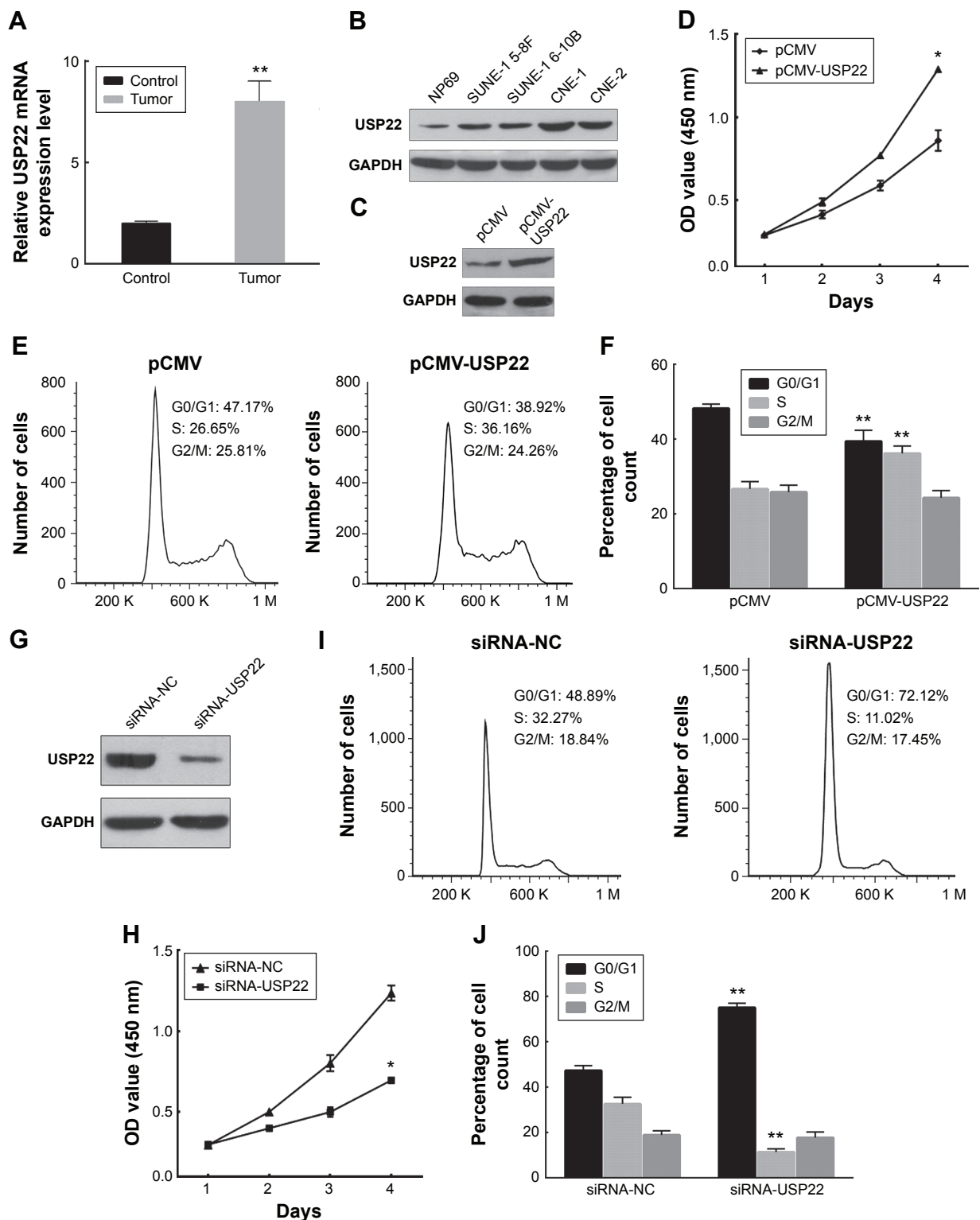


Figure 4 USP22 regulates the proliferation and cell cycle of SUNE-6-10B cells in vitro.

Notes: (A) Quantitative RT-PCR analysis of USP22 mRNA expression in nasopharyngeal carcinoma tissue and noncancerous nasopharyngeal mucosa. (B) Western blot analysis of USP22 in various NPC cell lines. (C) The abundance of USP22 proteins was assessed in SUNE-6-10B cells by Western blot analysis following ectopic overexpression of USP22. (D) The effect of USP22 overexpression on cell proliferation was measured using the MTT assay in SUNE-6-10B cell lines. (E and F) Flow cytometric analysis of the cell cycle distribution of SUNE-6-10B cells transiently transfected with USP22-overexpressing plasmid or control plasmid. (G) The abundance of USP22 proteins in SUNE-6-10B cells after siRNA transfection was analyzed by Western blot analysis. (H) The effect of siRNA-USP22 on cell proliferation in SUNE-6-10B cell lines was measured using the MTT assay. (I and J) Flow cytometric analysis of the cell cycle distribution of SUNE-6-10B cells that were transiently transfected with 100 nM siRNA-USP22 or the control. Each independent experiment was repeated three times. All the experiments were performed in triplicate, and the results were analyzed by ANOVA and are expressed as mean \pm SD (** P <0.01; * P <0.05).

Abbreviations: USP22, ubiquitin-specific peptidase 22; RT-PCR, reverse transcription-polymerase chain reaction; NPC, nasopharyngeal carcinoma; MTT, 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide; siRNA, small interfering RNA; ANOVA, analysis of variance; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; OD, optical density.

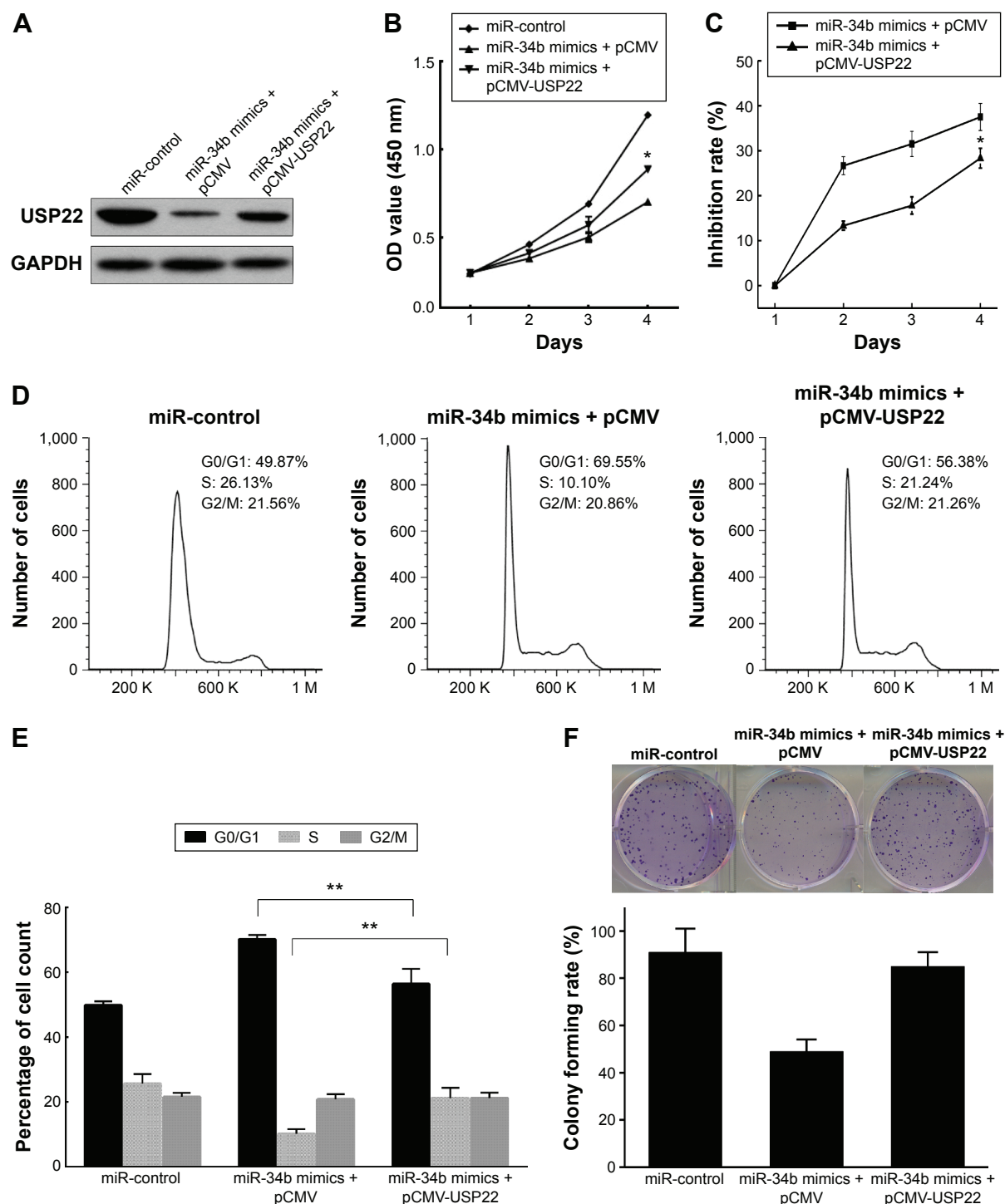


Figure 5 miR-34b represses USP22 to suppress SUNE-6-10B growth.

Notes: (A) Western blot analyses were performed to evaluate the efficiency of miR-34b overexpression and USP22 reintroduction in SUNE-6-10B cells. (B and C) The cell proliferation activity was analyzed using the MTT assay. (D and E) Flow cytometric analysis of the cell cycle distribution of the indicated SUNE-6-10B cells. (F) Colony formation ability was analyzed by colony formation assay. ** $P < 0.01$; * $P < 0.05$.

Abbreviations: USP22, ubiquitin-specific peptidase 22; MTT, 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; OD, optical density.

that miR-34b bound directly to the 3'-UTR of USP22 and downregulated its protein expression. The results of this study provide a novel insight of the mechanism by which miR-34b suppresses the proliferation of NPC cells.

Although miR-34b has been identified as a tumor suppressor in many types of cancers, it has also been reported to play an oncogenic role in breast cancer and esophageal squamous cell carcinoma.^{26,27} Therefore, we

sought to evaluate the precise effect of miR-34b in NPC. To achieve this goal, we overexpressed miR-34b by transfection into NPC SUNE-6-10B cells and observed an inhibition of cell viability. Furthermore, flow cytometry analysis revealed an arrest of SUNE-6-10B cells in the G1 phase, which suggests that miR-34b functions as a negative regulator of cellular proliferation in NPC. Moreover, a comparison of the expression levels of miR-34b in NPC cell lines and immortalized nasopharyngeal epithelial cells revealed that the basal expression level of miR-34b was generally downregulated in four NPC cell lines compared with NP69 (Figure 1A). These results are consistent with miRNA expression profiling, which shows a downregulation of miR-34b in eight NPC tissues and four normal nasopharyngeal tissues.⁷ The results suggest that the carcinogenesis of NPC could be ascribed to a loss of miR-34b, which raises questions regarding the cause of the downregulation of miR-34b. An increasing number of studies have proposed that the expression of miR-34b is inversely correlated with CpG island methylation.²⁸ In addition to epigenetic regulation, miR-34b is also a target gene of p53.^{29–31} Because p53 mutations are rare in NPC, whether miR-34b is regulated by p53 warrants further investigation.

USP22 belongs to the ubiquitin hydrolase family and has recently generated a large amount of attention due to its association with cancer. A microarray analysis revealed that a high level of USP22 is commonly seen in many types of cancers.¹⁰ In oral squamous cell carcinoma and cervical cancer cells, USP22 was identified as a novel molecular biomarker that is predictive of cancer progression and patient prognosis.^{11,12} However, the expression of USP22 in NPC has remained unclear. Here, we compared the expression of USP22 in NPC cell lines and immortalized nasopharyngeal epithelial cells. The data showed that USP22 was overexpressed in NPC cell lines when compared with NP69. Accentuated USP22 expression in many types of cancer cells usually indicates a poor clinical outcome. The influence of USP22 expression on the clinical prognosis of NPC requires further analysis. Recently, USP22 was identified as a subunit of the Spt–Ada–Gcn5 acetyltransferase coactivator complex, which regulates the expression of genes that are related to oncogenicity and proliferation.³² We assessed the function of USP22 in NPC proliferation. The overexpression or knockdown of USP22 promoted the proliferation of NPC. Notably, the overexpression of USP22 rescued the inhibitory effect of miR-34b on NPC, demonstrating that miR-34b suppresses the proliferation of NPC by targeting USP22.

Conclusion

In summary, our recent study showed that miR-34b was expressed at a low level in NPC and acted as positive regulator of NPC proliferation. By contrast, USP22 was overexpressed in NPC cells and promoted the proliferation of NPC. Furthermore, we demonstrated that miR-34b inhibited NPC proliferation by downregulating the expression of USP22. The results of this study provide a theoretical basis for miR-34b- or USP22-targeted diagnoses and therapies for NPC.

Acknowledgments

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Disclosure

The authors report no conflicts of interest in this work.

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