A network pharmacology approach to discover active compounds and action mechanisms of San-Cao Granule for treatment of liver fibrosis

Shizhang Wei,1,2 Ming Niu,1 Jian Wang,2 Jiabo Wang,3 Haibin Su,4 Shengjiang Luo,5 Xiaomei Zhang,2,6 Yanlei Guo,4 Liping Liu,1 Fengqun Liu,1 Qingguo Zhao,1 Hongge Chen,1 Xiaohua Xiao,1 Pan Zhao,4,7 Yanling Zhao1,2

1Department of Pharmacy, 302 Hospital of People's Liberation Army, Beijing, People's Republic of China; 2Pharmacy College, Chengdu University of Traditional Chinese Medicine, Chengdu, People's Republic of China; 3China Military Institute of Chinese Medicine, 302 Hospital of People's Liberation Army, Beijing, People's Republic of China; 4Liver Failure Therapy and Research Center, 302 Hospital of People's Liberation Army, Beijing, People's Republic of China; 5Department of Integrative Medical Center, 302 Hospital of People's Liberation Army, Beijing, People's Republic of China; 6Clinical Trial Center, 302 Hospital of People's Liberation Army, Beijing, People's Republic of China; 7Department of Traditional Chinese Medicine, Key Laboratory of Chongqing TCM Resources, Chongqing, People's Republic of China.

Correspondence: Yanling Zhao
Department of Pharmacy, 302 Hospital of People's Liberation Army, 100 Western 4th Ring Road, Beijing 100039, People's Republic of China
Email zhaoyl12855@126.com

Pan Zhao
Clinical Trial Center, 302 Hospital of People's Liberation Army, 100 Western 4th Ring Road, Beijing 100039, People's Republic of China
Email doczhaopan@126.com

Ethnopharmacological relevance: San-Cao Granule (SCG) has been used in patients with liver fibrosis for many years and has shown good effect. However, its mechanism of therapeutic action is not clear because of its complex chemical system. The purpose of our study is to establish a comprehensive and systemic method that can predict the mechanism of action of SCG in antifibrotic efficacy.

Materials and methods: In this study, a “compound–target–disease” network was constructed by combining the SCG-specific and liver fibrosis–specific target proteins with protein–protein interactions, and network pharmacology was used to screen out the underlying targets and mechanisms of SCG for treatment of liver fibrosis. Then, some key molecules of the enriched pathway were chosen to verify the effects of SCG on liver fibrosis induced by thioacetamide (TAA).

Results: This systematic approach had successfully revealed that 16 targets related to 11 SCG compounds were closely associated with liver fibrosis therapy. The pathway-enrichment analysis of them showed that the TGF-β1/Smad signaling pathway is relatively important. Animal experiments also proved that SCG could significantly ameliorate liver fibrosis by inhibiting the TGF-β1/Smad pathway.

Conclusion: SCG could alleviate liver fibrosis through the molecular mechanisms predicted by network pharmacology. Furthermore, network pharmacology could provide deep insight into the pharmacological mechanisms of Chinese herbal formulas.

Keywords: San-Cao Granule, network pharmacology, liver fibrosis, TAA

Introduction

Liver fibrosis, a common pathological change of chronic liver injury induced by various etiological factors, is the early stage of cirrhosis.1,2 An important feature of liver fibrosis is the massive deposition of extracellular matrix (ECM) proteins. Activated hepatic stellate cells (HSCs) are the major source of ECM, which regulate the massive production and secretion of ECM.3 Although liver fibrosis is a common and serious disease, there has been no satisfactory treatment.

In recent years, the therapy against hepatic fibrosis with traditional Chinese medicines (TCMs) in the People’s Republic of China has achieved great progress.4 Chinese herbal formulas including more than two kinds of TCM have synergistic therapeutic efficacy because of multiple active compounds; hence, an increasing number of Chinese herbal formulas have been researched for their effect on antifibrotic fibrosis.5–7

SCG consists of Rubia cordifolia L., Siegesbeckia orientalis L., Gentiana macrophylla Pall., Salvia miltiorrhiza Bunge, Forsythia suspensa (Thunb.) Vahl., and Glycyrrhiza uralensis Fisch. In the ratio of 3:3:1.5:1.5:1.5:1:1.5:1 and has been used to...
treat liver fibrosis. Its clinical efficacy has been confirmed in hepatic fibrosis patients, but the mechanisms of action, precise targets, and the relationships between these herbs and diseases are still unanswered. Therefore, further research of SCG remains a big challenge for us.

Recently, network pharmacology has attracted the attention of Chinese medicine researchers because it can be used to analyze the relationship between drug and disease using network from a proteome or at systematic level. Especially, it can react and clarify the interactive relationship between multiple components and multiple targets of TCM and has become a research focus of the theory and method in drug discovery and development. Therefore, we applied network pharmacology in our study to understand the efficacy of SCG in the prevention of liver fibrosis.

Figure 1 The whole framework of this study based on an integrative strategy of network pharmacology and experimental verification.

**Materials and methods**

**Database construction**

The chemical structures of the composite compounds in SCG were obtained from TCM Database@Taiwan (TDT). Known compound targets were collected from Herbal Ingredients’ Targets Database (HIT), and the putative targets from these were screened out from Therapeutic Targets Database (TTD). Gene and protein targets associated with liver fibrosis therapy were collected from the Online Mendelian Inheritance in Man (OMIM) database. Other interacting human proteins of the aforementioned targets were obtained from Database of Interacting Proteins (DIP), and different ID types of the proteins were converted to UniProt IDs.

**Network construction and analysis**

To provide the scientific and reasonable interpretation of the complex relationships between the compounds and targets associated with liver fibrosis, network analysis was performed. The chemical components, SCG putative targets, liver fibrosis–significant targets, and interacting proteins were all connected to construct a “compound–target–disease” network by using protein–protein interaction (PPI) information. Cytoscape 2.8.3 was applied to visualize and analyze the network, and the topological features of each node in the network were calculated using CenSiCaPe 1.2 of the Cytoscape software. Only the nodes with higher values of “Degree”, “Betweenness centrality”, and “Closeness centrality” (above twofold of the median value) were identified as the candidate SCG targets for liver fibrosis.
Drugs and reagents
SCG consisted of Rubia cordifolia L., Siegesbeckia orientalis L., Gentiana macrophylla Pall., Salvia miltiorrhiza Bunge, Forsythia suspensa (Thunb.) Vahl and Glycyrrhiza uralensis Fisch., which were produced by Beijing Sheng Shi Long Herbs Co., Ltd (Beijing, People’s Republic of China). The aforementioned TCMs were extracted by the following process: crude medical material was extracted with ten-fold the volume of water, twice, 1 hour for each time. The extract was concentrated and was then purified with alcohol. The supernatant through purification was condensed and dried, then the dry powder was mixed with moderate dextrin and pelletized with 95% alcohol.

Thioacetamide (TAA) was purchased from Sinopharm Chemical Reagent Co., Ltd (Shanghai, People’s Republic of China). Serum alanine transaminase (ALT), aspartate aminotransferase (AST), and albumin (ALB) testing kits were purchased from Nanjing Jiancheng Co. Ltd (Nanjing, People’s Republic of China). Serum total bilirubin (TBIL), hydroxyproline (Hyp), hyaluronic acid (HA), laminin (LN), and type IV collagen (CIV) testing kits were purchased from Clong-Cloud Corp (Houston, TX, USA); α-SMA, CTGF, TGF-β1, p-Smad3, Smad3, and Smad7 primary antibodies were purchased from Abcam (Cambridge, UK).

Time-of-flight liquid chromatography mass spectrometry mass spectrometer system
The conditions of LC (SHIMADZU, LC-30AD) were as follows: column – Kinetex 00D-4462-AN C18 (100×2.1, 2.6 μm); mobile phase – acetonitrile with 0.1% (v/v) formic acid (A) and water containing 0.1% (v/v) formic acid (B); gradient – 0–1 minutes, 5:95; 1–8 minutes, 80:20; 8–15 minutes, 95:5; 15–20 minutes, 100:0 (A:B, v/v); column temperature – 35°C; flow rate – 0.3 mL/min; and injection volume – 3 μL.

The TOF-MS (Triple TOF 4600) parameters were as follows: Curtain Gas: 25,000; Ion Source Gas1:55,000; Ion Source Gas2:55,000; Ionspray Voltage Floating: 4,500,000; Temperature: 600,000.

Animals
Male Sprague Dawley rats (180±10 g) were obtained from the animal center of Military Medical Sciences Academy of the PLA (Permission no SCXX-(A) 2012-0004). The rats were acclimatized for 1 week under environmentally controlled conditions with temperature (25±0.5°C), humidity (55±5%), and light (12:12 hour light:dark cycle) prior to the experiment. The animals received humane care in compliance with the Chinese Animal Protection Act, according to the National Research Council criteria. The study was approved by the ethics committee of Beijing 302 Hospital.

TAA-induced liver fibrosis and treatments
TAA can induce liver fibrosis in Sprague Dawley rats. Ursodeoxycholic acid (UDCA) was taken as a positive medicine, because it is an accepted drug for treating liver fibrosis in clinical application and laboratory studies. In our study, 60 Sprague Dawley rats were divided into normal group (saline-treated), TAA-treated, TAA-treated + UDCA (60 mg/kg), TAA-treated + SCG (3.6 g/kg), TAA-treated + SCG (1.8 g/kg), and TAA-treated + SCG (0.8 g/kg) groups, with ten rats in each group. TAA was dissolved in saline and then intraperitoneally administered (200 mg/kg) to all the rats except the normal group twice a week for 6 weeks, and then the rats were orally administered with the corresponding drugs daily for 4 weeks. During the drug administration, TAA was continually administered once a week until the end of the experiment. All the animals were fasted for 24 hours before they were sacrificed.

Evaluation of serum and tissue indexes
The effect of SCG on liver injury and liver fibrosis induced by TAA was evaluated using serum ALT, AST, ALB, HA, AN, CIV, and tissue Hyp testing kits through microplate test or enzyme-linked immunosorbent assay.

Histopathological and immunohistochemical evaluation
The left lobe of livers of all rats were excised and fixed in 4% paraformaldehyde buffer for 24 hours. Then, the fixed liver tissues were dehydrated, embedded in paraffin, and sectioned into 4 μm thick sections. The section was stained with hematoxylin and eosin (HE) and Masson fiber stains. These experimental operations were performed by a professional pathologist (Lu Yinglin, Department of Pathology, Academy of Military Medical Sciences, Beijing, People’s Republic of China). Hepatic fibrosis was quantitatively assessed by analyzing Masson-stained paraffin sections under an optical microscope. The degree of liver fibrosis was graded based on the following criteria: 0: no fibrosis; I: slight fibrosis, fibrosis located in the central liver lobule; II: moderate fibrosis, widened central fibrosis; III: severe fibrosis, fibrosis extended to the edge of liver lobule; and IV: liver cirrhosis.16 The percentage of collagen content (collagen area/total area) was measured using the Image software (Image pro-plus 6.0, Media Cybernetics, Inc., Rockville, MD, USA).
For the immunohistochemical analysis of α-SMA, the 5 μm liver sections were dipped in 3% hydrogen peroxide for 30 minutes to quench endogenous peroxidase activity after being dewaxed and hydrated, and then were blocked and incubated overnight at 4°C with anti-α-SMA antibody. A detectable color was generated after being incubated with 3, 30-diaminobenzidine tetrachloride for 5 to 10 minutes. The morphological characteristics were visualized by light microscopy. The integrated optical density (IOD) value of α-SMA was detected using the Image software (Image pro-plus 6.0).

Western blotting
To detect the levels of protein expression of α-SMA, TGF-β1, p-Smad3, Smad3, Smad7, and CTGF, we performed Western blotting. The liver tissues (50 mg) stored in liquid nitrogen tank were homogenized in 450 mL PBS with proteinase inhibitor, and were then centrifuged at 12,000×g at 4°C for 10 minutes to obtain a supernatant. The total protein obtained was quantified using the BCA protein assay kit (Thermo Fisher Scientific, Waltham, MA, USA). The protein sample was boiled for 10 minutes. Next, equal amounts of protein sample were run on 12% SDS-PAGE, transferred onto polyvinylidene fluoride (PVDF) membrane, and incubated in blocking buffer (TBST; 5× TBS: Tween-20 =1:20 (V/V)) nonfat milk for 1–2 hours. The PVDF membrane was then incubated with anti-α-SMA (1:2,000), CTGF (1:500), anti-TGF-β1 (1:50), anti-p-Smad3 (1:500), anti-Smad3 (1:3,000), and anti-Smad7 (1:2,000) primary antibodies for 3 hours, respectively, and then washed three times with TBST. The PVDF membrane was incubated with secondary HRP-conjugated donkey anti-mouse IgG antibody (1:2,000) for 45 minutes and was washed three times again. The bound antibody was reacted with the chemiluminescence kit, and the intensity of the protein bands was analyzed with the ImageJ software program.

Statistical analysis
The data of the experiment are presented as means ± SEM. One-way ANOVA was used for statistical analysis. P<0.01 was regarded as highly significant and P<0.05 was regarded as significantly different.

Results
Targets screening of SCG and liver fibrosis
Based on database construction method, a total of 68 compounds and 387 targets of SCG, 59 candidate protein targets associated with liver fibrosis therapy, and 440 interacting proteins of the targets were screened out. Among the compounds and targets of SCG, 16 compounds and 51 targets were collected for Rubia cordifolia L., 24 compounds and 75 targets for Siegesbeckia orientalis L., 2 compounds and 3 targets for Gentiana macrophylla Pall., 4 compounds and 82 targets for Salvia miltiorrhiza Bunge, 11 compounds and 108 targets for Forsythia suspensa (Thunb.) Vahl, and 11 compounds and 68 targets for Glycyrrhiza uralensis Fisch. The obtained compounds and targets were all used to construct the “drug–target–disease” network.

Network construction and pathway analysis
The construction of target protein network of the TCM formula for a specific disease and the summary and analysis of the noteworthy features of the network would provide some important information for us to understand the drug–target interaction mechanism of the certain drugs on the specific disease. This study focused on the effect of SCG on liver fibrosis. Figure 2 shows the network for the compounds and their potential targets with color-coded nodes. The red triangles represent active chemical constituents of SCG, the blue dots represent the indirect targets for drugs, the yellow dots represent the targets of the specific disease of liver fibrosis, and the yellow squares represent the common targets of herbs and liver fibrosis. That is, the common targets are the key targets by which SCG treated liver fibrosis and are also the relatively important targets screened for further research.

Figure 2: The main network of compound–target–disease of SCG.
Notes: The red triangles represent active chemical constituents of SCG, the blue dots represent the indirect targets for drugs, the yellow dots represent the targets of the specific disease of liver fibrosis, the yellow squares represent the common targets of herbs and liver fibrosis, and the purple dots represent the interactive proteins with the targets of liver fibrosis and drugs.
Abbreviation: SCG, San-Cao Granule.
The purple dots represent the interactional proteins with the targets of liver fibrosis and drugs.

Table 1 indicates that 16 common targets were predicted. To further screen the more important targets, three topological parameters, “Degree”, “Betweenness centrality”, and “Closeness centrality”, were chosen to identify candidate hepatic fibrosis targets. We identified a node as the candidate SCG target for liver fibrosis if its three topological parameters were more than twofold of the median value, respectively. After calculating the value of the three parameters for each hepatic fibrosis significant protein in the PPI network, the twofold of the median value of “Degree”, “Betweenness centrality”, and “Closeness centrality” was 3.02, 0.41, and 0.07, respectively. Therefore, we determined that hepatic fibrosis significant proteins with “Degree” > 3.02, “Betweenness centrality” > 0.41, and “Closeness centrality” > 0.07 were candidate targets for hepatic fibrosis therapy. As a result, amyloid β A4 protein; actin, cytoplasmic 1; transforming growth factor β-1; and fibroblast growth factor two were identified as candidate hepatic fibrosis targets.

As shown in Figure 3, 22 KEGG pathways were enriched in the ArrayTrack v.3.5.0 pathway-enrichment analysis. As we know that inflammation plays an important role in liver fibrosis, the analysis of KEGG pathways manifests that most of the pathways, such as TGF-β signaling pathway, P53 signaling pathway, MAPK signaling pathway, Toll-like receptor signaling pathway, NOD-like receptor signaling pathway, and chemokine signaling pathway are either directly involved in immunity and inflammation or play other important roles.

Considering the “Degree”, “Betweenness centrality” and “Closeness centrality” of TGF-β1 were estimated to be relatively large, and TGF-β1 participated in many signaling pathways that were relatively important in the obtained 22 KEGG pathways such as pathways in cancer, pancreatic cancer, chronic myeloid leukemia, cytokine–cytokine receptor interaction, cell cycle, adherens junction, and TGF-β signaling pathway. Therefore, we selected TGF-β1 and its downstream important proteins Smad3 and Smad7 chosen from the TGF-β signaling pathway map in the forms of molecular networks for further verification study to validate the effectiveness of the signaling pathway for the treatment of SCG for liver fibrosis.

The identification of chemical ingredients of SCG by TOF LC/MS/MS

In our previous studies,17 we identified 15 active chemical ingredients from SCG through TOF LC/MS/MS, which were rubimaiillin, purpurin, gentiopicroside, tanshinone IIA, salvianolic acid B, danshensu, glycyrrhizinic acid, glycyrrhetinic acid, cryptotanshinone, alizarin, ferulic acid, formononetin, orientin, sweroside, and swertamarin as shown in Table 2.

Improvement of SCG on the serum parameters of liver function and liver fibrosis in TAA-induced liver fibrotic rats

The serum ALT, AST, ALB, and TB levels can indicate the degree of liver damage. As shown in Figure 4A, the levels of ALT, AST, and TB were significantly elevated in

Table 1 The targets and its relevant topological parameters and corresponding active compounds of SCG screened by network pharmacology

<table>
<thead>
<tr>
<th>Direct target</th>
<th>Target protein</th>
<th>Degree</th>
<th>Closeness centrality</th>
<th>Betweenness centrality</th>
<th>Compound</th>
</tr>
</thead>
<tbody>
<tr>
<td>P05067</td>
<td>Amyloid β A4 protein</td>
<td>12</td>
<td>0.9167</td>
<td>0.9818</td>
<td>Cryptotanshinone</td>
</tr>
<tr>
<td>P60709</td>
<td>Actin, cytoplasmic 1</td>
<td>9</td>
<td>0.5313</td>
<td>0.7794</td>
<td>Danshensu</td>
</tr>
<tr>
<td>P01137</td>
<td>Transforming growth factor β-1</td>
<td>8</td>
<td>0.4473</td>
<td>0.4265</td>
<td>Danshensu</td>
</tr>
<tr>
<td>P09038</td>
<td>Fibroblast growth factor 2</td>
<td>6</td>
<td>0.65</td>
<td>0.641</td>
<td>Ursolic acid</td>
</tr>
<tr>
<td>P11802</td>
<td>Cyclin-dependent kinase 4</td>
<td>11</td>
<td>0.2577</td>
<td>0.0729</td>
<td>Glycyrrhetinic acid</td>
</tr>
<tr>
<td>P31749</td>
<td>RAC-α serine/threonine-protein kinase</td>
<td>13</td>
<td>0.2673</td>
<td>0.061</td>
<td>Salvianolic acid A</td>
</tr>
<tr>
<td>P03372</td>
<td>Estrogen receptor</td>
<td>21</td>
<td>0.2407</td>
<td>0.0906</td>
<td>Formononetin</td>
</tr>
<tr>
<td>O15392</td>
<td>Bacuoloviral IAP repeat-containing protein 5</td>
<td>8</td>
<td>0.1717</td>
<td>0.0363</td>
<td>Ursolic acid</td>
</tr>
<tr>
<td>Q07817</td>
<td>Bcl-2-like protein 1</td>
<td>4</td>
<td>0.205</td>
<td>0.0054</td>
<td>Ursolic acid</td>
</tr>
<tr>
<td>P07550</td>
<td>β-2 adrenergic receptor</td>
<td>3</td>
<td>1</td>
<td>1</td>
<td>11,12,13-trinorguai-6-ene-4β,10β-diol</td>
</tr>
<tr>
<td>P22303</td>
<td>Acetylcholinesterase</td>
<td>1</td>
<td>0.5</td>
<td>0</td>
<td>Ursolic acid</td>
</tr>
<tr>
<td>P04141</td>
<td>Granulocyte-macrophage colony-stimulating factor</td>
<td>3</td>
<td>1</td>
<td>1</td>
<td>Glycyrrhizinic acid</td>
</tr>
<tr>
<td>P01008</td>
<td>Antithrombin-III</td>
<td>3</td>
<td>1</td>
<td>0</td>
<td>Cryptotanshinone</td>
</tr>
<tr>
<td>P62736</td>
<td>Actin, aortic smooth muscle</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>Salvianolic acid A</td>
</tr>
<tr>
<td>Q9UKK3</td>
<td>Poly [ADP-ribose] polymerase 4</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>Tanshinonell A</td>
</tr>
<tr>
<td>P09917</td>
<td>Arachidonate 5-lipoxygenase</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>Alizarin</td>
</tr>
</tbody>
</table>

Abbreviation: SCG, San-Cao Granule.
TAA-treated group (P<0.01), meanwhile the level of serum ALB was significantly reduced (P<0.01). TAA-treated + SCG (3.6 and 1.8 g/kg) and TAA + UDCA (60 mg/kg) markedly reduced the high serum levels of ALT, AST, and TB (P<0.05), but markedly increased the level of ALB (P<0.01). The improvement of SCG (3.6 and 1.8 g/kg) on liver injury was better than UDCA (60 mg/kg) and SCG (0.9 g/kg). These results showed that the improvement in liver injury was better on SCG treatment (3.6 and 1.8 g/kg) than on UDCA treatment.

The serum levels of HA, LN, and CIV and the Hyp level in liver tissue can indicate the degree of liver fibrosis. As shown in Figures 4B and 5D, the levels of HA, LN, CIV, and Hyp were significantly increased in the TAA-treated group (P<0.01). However, treatment with SCG (3.6 and 1.8 g/kg) and UDCA (60 mg/kg) markedly reduced the increased levels of HA, LN, and CIV (P<0.05). The improvement with SCG treatment (3.6 and 1.8 g/kg) on liver fibrosis was better than that with UDCA (60 mg/kg) and SCG (0.9 g/kg).

**Histological evaluation**

The histopathological changes in the liver tissues taken from all of the experiment groups were examined by HE staining. As shown in Figure 6A, the liver tissue structure of normal group exhibited regular hepatocellular arrangement with branching form and clearly showed hepatic cords. The hepatocytes had a single circular vesicular nuclei and acidophilic cytoplasm, except for a few hepatocytes that had two vesicular nuclei. The liver sections of the model group showed that many cytoplasmic vacuoles appeared in most of the
hepatocytes and that portal areas were infiltrated by massive inflammatory cells, and the nuclei were stained deeply. SCG (3.6 g/kg) administration significantly improved the disordered pathological structure of liver; SCG (1.8 and 0.9 g/kg) and UDCA (60 mg/kg) had moderately reduced the disorder of hepatocytes.

As shown in Figure 6B, the results of Masson’s trichrome staining indicated that few collagenous fibers were observed in normal group. However, the sections of the model group had much more collagenous fibers around the central veins and portal area. SCG (3.6 g/kg) treatment significantly reduced the content of collagenous fibers, while the treatment with SCG (1.8 and 0.9 g/kg) and UDCA (60 mg/kg) only slightly reduced the content of collagenous fibers. The results of staging scores and percentage of collagen content of liver fibrosis (Table 3 and Figure 5E) also showed that SCG (3.6, 1.8, and 0.9 g/kg) and UDCA markedly alleviated the score of liver fibrosis (P<0.01).

Effect of SCG on the expression of TGF-β1

TGF-β1 is an important multifunctional cytokine because of its special role in the development of liver fibrosis. We detected the expression of TGF-β1 using Western blot. As shown in Figure 7A and B, we found that the expression of TGF-β1 in TAA-treated group was promoted markedly, while the administration of SCG (3.6 and 1.8 g/kg) treatments significantly inhibited the expression of TGF-β1.

Effect of SCG on the expression of p-Smad3, Smad3, and Smad7

As shown in Figure 7, the expression of p-Smad3 was significantly elevated, while the expression of Smad3 was inapparent, and the expression of Smad7 was significantly reduced in TAA-treated group. SCG (3.6 and 1.8 g/kg) treatments remarkably attenuated the expression of p-Smad3 and induced the expression of Smad7.

The results indicated that SCG interdicted the transmission of TGF-β1/Smad through reducing the expression of TGF-β1 and p-Smad3 but promoting the expression of Smad7.

Discussion

To uncover the therapeutic effect on diseases and the active substances involved in the drugs, we structured an integrated model of system pharmacology to investigate the pharmacological mechanisms of SCG for treatment of liver fibrosis. In this study, we first predicted the active ingredients and

---

**Figure 4** The serum levels of ALT, AST, ALB, TB, HA, LN, and CIV of experimental groups. Notes: (A): ALT, AST, ALB, TB; (B): HA, LN, CIV. Data are expressed as the mean ± SD. (SCG-H: TAA + SCG [3.6 g/kg]; SCG-M: TAA + SCG [1.8 g/kg]; SCG-L: TAA + SCG [0.9 g/kg]). *P<0.01 compared with normal group, **P<0.01 compared with model group, #P<0.05 compared with model group.

**Abbreviations:** SCG, San-Cao Granule; UDCA, ursodeoxycholic acid; TAA, thioacetamide; ALT, alanine transaminase; AST, aspartate aminotransferase; ALB, albumin; TB, total bilirubin; HA, hyaluronic acid; LN, laminin; CIV, IV collagen.
Figure 5 The effect of SCG on the expressions of α-SMA and CTGF.

Notes: (A) The expression of α-SMA detected by immunohistochemistry staining (a: Normal; b: Model; c: UDCA; d: SCG-H; e: SCG-M; f: SCG-L); (B, C) the expression of α-SMA and CTGF detected by Western blot; (D) the level of Hyp in liver tissue; (E) the percentage of collagen content detected by image software through liver tissue stained by Masson (100 ×); (F) the integrated optical density (IOD) value of α-SMA. (SCG-H: TAA + SCG [3.6 g/kg]; SCG-M: TAA + SCG [1.8 g/kg]; SCG-L: TAA + SCG [0.9 g/kg]). *P<0.01 compared with normal group, **P<0.01 compared with model group.

Abbreviations: SCG, San-Cao Granule; UDCA, ursodeoxycholic acid; TAA, thioacetamide; SMA, smooth muscle actin; CTGF, connective tissue growth factor; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; Hyp, hydroxyproline.
potential targets of SCG related to liver fibrosis through constructing the biological network of interaction between chemical compounds and protein targets from a molecular or systemic level.

The predicted results indicated that 11 ingredients in SCG interacted with 16 targets related to liver fibrosis in this formula. Some ingredients could modulate the same target simultaneously, which gives a good explanation of synergistic mechanisms of TCMs. It is worth mentioning that the computational models can only give hints, which still need to be verified by real experiment.

Based on the results from network pharmacology analysis, we further explored the effect of SCG on the important parameters related to liver injury and liver fibrosis by a TAA-induced liver fibrotic rat model and the key proteins analyzed by KEGG pathways and Cytoscape. The levels of ALT, AST, ALB, and TB were significantly improved by SCG, which indicated that SCG alleviated liver injury. The reduction of the levels of HA, LN, CIV, and Hyp showed that liver fibrosis induced by TAA was significantly alleviated. From the results of histological evaluation, we could find that the inflammation and damage conditions and the collagen content of hepatic tissue were markedly reduced by SCG.

It has been demonstrated that the overexpression of α-SMA can illustrate the activation of (HCSs), which are the primary cells responsible for the development of liver fibrosis. CTGF is an important fibrogenic factor synthesized by hepatocytes and HSCs and is a general mediator of the interactions between fiber–fiber, fiber–matrix, and matrix–matrix. Therefore, we further evaluated the effect of SCG on the liver fibrosis by testing the expressions of α-SMA and CTGF. Our results demonstrated that α-SMA and CTGF were significantly reduced by SCG, which also indicated that SCG inhibited the activation of HSCs and alleviated liver fibrosis. To validate the results predicted by network pharmacology, we evaluated the expression of TGF-β1 and its downstream signaling such as Smad3 and Smad7. The results showed that SCG significantly reduced the expression of TGF-β1 and p-Smad3 and increased the expression of Smad7, which indicated that SCG alleviated liver fibrosis.

Table 3 The degree of hepatic fibrosis in rats

<table>
<thead>
<tr>
<th>Groups</th>
<th>n</th>
<th>0</th>
<th>I</th>
<th>II</th>
<th>III</th>
<th>IV</th>
<th>Staging scores (mean ± SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal</td>
<td>10</td>
<td>10</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Model</td>
<td>10</td>
<td>0</td>
<td>0</td>
<td>7</td>
<td>3</td>
<td>3.3±0.48*</td>
<td></td>
</tr>
<tr>
<td>UDCA</td>
<td>10</td>
<td>0</td>
<td>1</td>
<td>4</td>
<td>5</td>
<td>2.4±0.69**</td>
<td></td>
</tr>
<tr>
<td>TAA + SCG (3.6 g/kg)</td>
<td>10</td>
<td>0</td>
<td>8</td>
<td>2</td>
<td>0</td>
<td>1.2±0.42**</td>
<td></td>
</tr>
<tr>
<td>TAA + SCG (1.8 g/kg)</td>
<td>10</td>
<td>0</td>
<td>5</td>
<td>4</td>
<td>1</td>
<td>1.6±0.68**</td>
<td></td>
</tr>
<tr>
<td>TAA + SCG (0.9 g/kg)</td>
<td>10</td>
<td>0</td>
<td>2</td>
<td>5</td>
<td>3</td>
<td>2.1±0.73**</td>
<td></td>
</tr>
</tbody>
</table>

Notes: *P<0.01 compared with normal group, **P<0.01 compared with model group.

Abbreviations: SCG, San-Cao Granule; UDCA, ursodeoxycholic acid; TAA, thioacetamide; HE, hematoxylin-eosin.
The effect of SCG on the expressions and semiquantitative analysis of TGF-β1, p-Smad3, Smad3, and Smad7 detected by Western blot.

Notes: (SCG-H: TAA + SCG [3.6 g/kg]; SCG-M: TAA + SCG [1.8 g/kg]; SCG-L: TAA + SCG [0.9 g/kg]). *P < 0.01 compared with normal group, **P < 0.01 compared with model group. (A) Western blot analysis for TGF-β1, p-Smad3, Smad3, and Smad7; Densitometry analysis of TGF-β1 (B), Smad7 (C), and p-Smad3/Smad3 (D) on Western blot.

Abbreviations: SCG, San-Cao Granule; TAA, thioacetamide; TGF-β1, transforming growth factor β1; GAPDH, glyceraldehyde 3-phosphate dehydrogenase.

Figure 7

fibrosis through the action mechanisms predicted by network pharmacology analysis. Meanwhile, tanshinone IIA,[21] danshensu,[22] salvianolic acid B,[23] glycyrrhetinic acid,[24] and glycyrrhizic acid[20] have antifibrosis effects, as reported by some previous studies. The five aforementioned components were also predicted as the active components of SCG, treating liver fibrosis by our method of network pharmacology. Therefore, network pharmacology can be considered to be an effective way to predict and discover new medicines treating specific diseases.

The understanding of the mechanism of Chinese herbal formulas is necessary for its modernization, and the integrative and holistic ideology of network pharmacology is consistent with the essence of Chinese herbal formulas in treatments. Therefore, network pharmacology can be applied to the action mechanism research of Chinese herbal formulas and promote its modernization. Moreover, the ability of revealing correlations between drugs and diseases can help network pharmacology develop into a new strategy for new drug research and development.
Acknowledgments
This work was supported by the Major Projects of the National Science and Technology (no 2012ZX10005010-002-002) and the National Natural Science Foundation Project of China (no 81303120) and (no 81173571).

Disclosure
The authors report no conflicts of interest in this work.

References