Detection of AR-V7 mRNA in whole blood may not predict the effectiveness of novel endocrine drugs for castration-resistant prostate cancer

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Abstract: A splice variant of androgen receptor (AR), AR-V7, lacks in androgen-binding portion and leads to aggressive cancer characteristics. Reverse transcription-polymerase chain reactions (PCRs) and subsequent nested PCRs for the amplification of AR-V7 and prostate-specific antigen (PSA) transcripts were done for whole blood of patients with prostate cancer and male controls. With primary reverse transcription PCRs, AR-V7 and PSA were detected in 4.5% and 4.7% of prostate cancer, respectively. With nested PCRs, AR-V7 messenger RNA (mRNA) was positive in 43.8% of castration-sensitive prostate cancer and 48.1% of castration-resistant prostate cancer (CRPC), while PSA mRNA was positive in 6.3% of castration-sensitive prostate cancer and 18.5% of CRPC. Whole-blood samples of controls showed AR-V7 mRNA expression by nested PCR. Based on multivariate analysis, expression of AR-V7 mRNA in whole blood was not significantly correlated with clinical parameters and PSA mRNA in blood, while univariate analysis showed a correlation between AR-V7 mRNA and metastasis at initial diagnosis. Detection of AR-V7 mRNA did not predict the reduction of serum PSA in patients with CRPC following abiraterone and enzalutamide administration. In conclusion, AR-V7 mRNA expression in normal hematopoietic cells may have annihilated the manifestation of aggressiveness of prostate cancer and the prediction of the effectiveness of abiraterone and enzalutamide by the assessment of AR-V7 mRNA in blood.

Keywords: AR-V7, prostate cancer, castration-resistant

Introduction

Androgen deprivation therapy for prostate cancer first reported by Huggins1 is effective at first, but eventually castration-resistant prostate cancer (CRPC) occurred and often caused cancer death. The annual incidence of and death from prostate cancer in Japan are 126.6 and 18.9 per 1,000,000 men, respectively, and both are still growing.2,3 Novel endocrine drugs for CRPC, abiraterone acetate and enzalutamide, have been introduced and shown to prolong overall survival of patients with CRPC regardless of preceding docetaxel therapy.4–7 Abiraterone acetate is a potent and irreversible inhibitor of CYP17 (17α-hydroxylase/C17, 20-lyase) that suppresses androgen synthesis, while enzalutamide is an androgen receptor (AR) signaling inhibitor in addition to being a highly potent AR antagonist.

Recently, splice variants of AR were identified. One of them, AR-V7, lacks the androgen-binding portion but automatically transduces intracellular signals resulting in enhanced nuclear transcription and more aggressive characteristics of cancer.8,9 Detection of AR-V7 messenger RNA (mRNA) in circulating tumor cells (CTCs) in patients with CRPC is reported to predict the noneffectiveness of
abiraterone acetate and enzalutamide. In the report, it is described that AR-V7 is specific to tumor cells.\textsuperscript{10} If AR-V7 is tumor specific, it is not absolutely necessary to isolate CTCs from whole blood, and thus mRNA extracted from whole blood may be sufficient to detect AR-V7 mRNA in CTCs.

Here, we assessed the AR-V7 mRNA expression in whole blood of patients with prostate cancer and analyzed its linkage with clinical parameters and effectiveness of abiraterone acetate and enzalutamide.

Materials and methods

Whole blood of 43 patients (age 62–94 years, median 78 years) with prostate cancer and six male controls (age 32–79 years, median 53 years) without prostate cancer was collected in tubes containing ethylenediaminetetraacetic acid. Thirty-five of them had been on androgen deprivation therapy, while others were scheduled to undergo the therapy. Abiraterone acetate or enzalutamide was administered to eleven patients with CRPC. For those patients, AR-V7 in blood was measured before abiraterone acetate or enzalutamide administration. Total RNA was extracted from 0.4 mL of whole blood using oligo dT primer and a DNA Extractor\textsuperscript{®} WB-Rapid Kit (Wako Pure Chemical Industries, Ltd, Osaka, Japan). Complementary DNA was synthesized from total RNA using a PrimeScript\textsuperscript{™} II first strand cDNA Synthesis Kit (Takara Bio, Inc., Kusatsu, Japan). Thirty-five cycles of polymerase chain reactions (PCRs) (primary PCR) in 50 µL were performed using a Premix Taq DNA Polymerase (Takara Bio, Inc.) for the amplification of AR-V7, prototype AR, prostate-specific antigen (PSA), and glyceraldehyde 3-phosphate dehydrogenase transcripts. In addition, 30 cycles of nested PCRs for AR-V7, prototype AR, and PSA were conducted using 1 µL of primary PCR products by substituting each corresponding antisense primer. The PCR primers are listed in Table 1. PCR amplicons were electrophoresed in a 1.5% agarose gel.

For univariate and multivariate analyses, odds ratios of positive expression of AR-V7 mRNA in whole blood were estimated, adjusting with metastasis at diagnosis, Gleason score, initial PSA, duration of hormone therapy, presence of CRPC, presence of clinical recurrence, and detection of PSA mRNA in whole blood by using logistic regression models. A waterfall plot was drawn to show the maximum reduction rate of PSA after abiraterone acetate and enzalutamide administration to metastatic patients with CRPC.

The experiment was conducted with the human subjects’ understanding and written informed consent was obtained. The Ethical Committee of Kanto Rosai Hospital approved this study and the experiments.

Results

Patients with prostate cancer

Clinical characteristics of patients with prostate cancer are listed in Table 2. With primary PCRs, glyceraldehyde 3-phosphate dehydrogenase mRNA was positive in all whole blood samples of patients with prostate cancer, while prototype AR, AR-V7, and PSA were detected in 53.3%, 4.5%, and 4.7% of whole-blood samples, respectively. With nested PCR, AR-V7 mRNA was positive in 43.8% of patients with castration-sensitive prostate cancer and 48.1% of patients with CRPC, while PSA mRNA was positive in 6.3% of castration-sensitive prostate cancer and 18.5% of CRPC. Prototype AR mRNA was detected in 93.8% of samples. Representative data are shown in Figure 1. With multivariate analysis, expression of AR-V7 mRNA in whole blood was not significantly correlated with metastasis at diagnosis, Gleason score, initial PSA, duration of hormone therapy, presence of CRPC, presence of clinical recurrence, or detection of PSA mRNA in whole blood as shown in Table 3. Univariate analysis showed that AR-V7

<table>
<thead>
<tr>
<th>Table 1</th>
<th>Primers for PCR amplification</th>
</tr>
</thead>
<tbody>
<tr>
<td>Product</td>
<td>Type of primer</td>
</tr>
<tr>
<td>GAPDH</td>
<td>Sense</td>
</tr>
<tr>
<td>GAPDH</td>
<td>Antisense</td>
</tr>
<tr>
<td>Proto AR</td>
<td>Sense</td>
</tr>
<tr>
<td>Proto AR</td>
<td>Antisense</td>
</tr>
<tr>
<td>Proto AR</td>
<td>Antisense for nested PCR</td>
</tr>
<tr>
<td>AR-V7</td>
<td>Sense</td>
</tr>
<tr>
<td>AR-V7</td>
<td>Antisense</td>
</tr>
<tr>
<td>AR-V7</td>
<td>Antisense for nested PCR</td>
</tr>
<tr>
<td>PSA</td>
<td>Sense</td>
</tr>
<tr>
<td>PSA</td>
<td>Antisense</td>
</tr>
<tr>
<td>PSA</td>
<td>Antisense for nested PCR</td>
</tr>
</tbody>
</table>

Abbreviations: GAPDH, glyceraldehyde 3-phosphate dehydrogenase; Proto, prototype; AR, androgen receptor; PSA, prostate-specific antigen; PCR, polymerase chain reaction; NA, not applicable.
AR-V7 mRNA was correlated with metastasis at initial diagnosis, but not with other parameters (Table 3).

A waterfall plot analysis did not confirm whether detection of AR-V7 mRNA in whole blood could predict the reduction of serum PSA in patients with CRPC following abiraterone acetate or enzalutamide administration (Figure 2). Nondetection of AR-V7 mRNA was not correlated with the reduction of serum PSA ($P=0.8865$ by the two-tailed chi-square test).

### Controls without prostate cancer

Whole-blood samples of controls without prostate cancer similarly processed showed AR-V7 mRNA expression by nested PCR as shown in Figure 3.

### Discussion

In the present study, the detection rate of AR-V7 mRNA in blood was much higher than that of PSA, although not all CTCs in patients with prostate cancer may express PSA mRNA. Additionally, expression of AR-V7 mRNA in blood did not correlate with that of PSA mRNA in blood. Therefore, there is a high possibility that AR-V7 mRNA is expressed not only in CTCs but also in nontumorous cells in blood, that is, hematopoietic cells. Prototype AR mRNA is known to be expressed in various types of cells including hematopoietic cells, so it is quite reasonable to suppose that splice variants of AR mRNA are also expressed in hematopoietic cells. Actually, AR-V7 mRNA was detected in whole-blood samples of controls without prostate cancer.

It is supposed that prostate cancer cells expressing AR-V7 have more aggressive characteristics, but AR-V7 mRNA

### Table 2 Characteristics of prostate cancer patients

<table>
<thead>
<tr>
<th>Characteristics</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Age at diagnosis (years), range (median)</td>
<td>59–88 (73)</td>
<td></td>
</tr>
<tr>
<td>Metastasis at diagnosis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Negative</td>
<td>17</td>
<td></td>
</tr>
<tr>
<td>Positive</td>
<td>26</td>
<td></td>
</tr>
<tr>
<td>Biopsy Gleason score</td>
<td></td>
<td></td>
</tr>
<tr>
<td>GS ≤6</td>
<td>4</td>
<td></td>
</tr>
<tr>
<td>GS 7</td>
<td>5</td>
<td></td>
</tr>
<tr>
<td>GS 8–10</td>
<td>31</td>
<td></td>
</tr>
<tr>
<td>iPSA (ng/mL), range (median)</td>
<td>5.3–9,529 (130)</td>
<td></td>
</tr>
<tr>
<td>Sensitivity to castration</td>
<td></td>
<td></td>
</tr>
<tr>
<td>CSPC</td>
<td>20</td>
<td></td>
</tr>
<tr>
<td>CRPC</td>
<td>23</td>
<td></td>
</tr>
<tr>
<td>Duration of hormonal T (years), range (median)</td>
<td>0.0–14.1 (2.5)</td>
<td></td>
</tr>
<tr>
<td>Clinical recurrence</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Negative</td>
<td>35</td>
<td></td>
</tr>
<tr>
<td>Positive</td>
<td>8</td>
<td></td>
</tr>
<tr>
<td>Previous docetaxel T</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Negative</td>
<td>37</td>
<td></td>
</tr>
<tr>
<td>Positive</td>
<td>6</td>
<td></td>
</tr>
</tbody>
</table>

**Notes:** Gleason score is missing for three patients, as pathologist did not report it.

**Abbreviations:** iPSA, initial prostate-specific antigen; CSPC, castration-sensitive prostate cancer; CRPC, castration-resistant prostate cancer; T, therapy.

### Table 3 Factors contributing to positive AR-V7 mRNA in blood

<table>
<thead>
<tr>
<th>Clinical parameters</th>
<th>Positive AR-V7 mRNA</th>
<th>Multivariate</th>
<th>Univariate</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>n</td>
<td>%</td>
<td>OR</td>
</tr>
<tr>
<td><strong>Metastasis at diagnosis</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Negative</td>
<td>17</td>
<td>23.5</td>
<td>4.90</td>
</tr>
<tr>
<td>Positive</td>
<td>26</td>
<td>61.5</td>
<td></td>
</tr>
<tr>
<td><strong>Gleason score</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>≤7</td>
<td>9</td>
<td>33.3</td>
<td></td>
</tr>
<tr>
<td>8–10</td>
<td>31</td>
<td>51.6</td>
<td>1.08</td>
</tr>
<tr>
<td><strong>iPSA (ng/mL)</strong></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>0–20</td>
<td>13</td>
<td>38.5</td>
<td></td>
</tr>
<tr>
<td>≥20</td>
<td>30</td>
<td>50.0</td>
<td>0.65</td>
</tr>
<tr>
<td><strong>Sensitivity to castration</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CSPC</td>
<td>16</td>
<td>43.8</td>
<td></td>
</tr>
<tr>
<td>CRPC</td>
<td>27</td>
<td>48.1</td>
<td>0.68</td>
</tr>
<tr>
<td><strong>Duration of hormonal T (years)</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0–3</td>
<td>23</td>
<td>52.2</td>
<td></td>
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<tr>
<td>≥3</td>
<td>19</td>
<td>42.1</td>
<td>1.28</td>
</tr>
<tr>
<td><strong>Clinical recurrence</strong></td>
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<tr>
<td>Negative</td>
<td>35</td>
<td>45.7</td>
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<tr>
<td>Positive</td>
<td>8</td>
<td>50.0</td>
<td>0.92</td>
</tr>
<tr>
<td><strong>PSA mRNA in WB</strong></td>
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<td></td>
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</tr>
<tr>
<td>Negative</td>
<td>37</td>
<td>43.2</td>
<td></td>
</tr>
<tr>
<td>Positive</td>
<td>6</td>
<td>66.7</td>
<td>4.16</td>
</tr>
</tbody>
</table>

**Abbreviations:** AR, androgen receptor; mRNA, messenger RNA; iPSA, initial prostate-specific antigen; CSPC, castration-sensitive prostate cancer; CRPC, castration-resistant prostate cancer; T, therapy; WB, whole blood; multivariate, multivariate analysis; univariate, univariate analysis; OR, odds ratio.
expression in blood did not necessarily indicate aggressive parameters of prostate cancer such as high Gleason scores, high initial serum PSA levels, acquisition of castration resistance, or clinical recurrence in the present study. Only positive metastasis at diagnosis was linked with positive AR-V7 mRNA in blood. The AR-V7 mRNA expression in normal hematopoietic cells may have obscured the manifestation of aggressiveness of prostate cancer by the assessment of AR-V7 mRNA in blood.

In contrast to the preceding report, the resistance to further androgen-related cancer therapy in cases with positive AR-V7 mRNA in blood was not observed in the present study. AR-V7 mRNA expression in normal hematopoietic cells may have annihilated the prediction of the effectiveness of abiraterone acetate and enzalutamide for CRPC.

AR-V7 expression was first described in metastatic tissue of hormone-resistant prostate cancer. If AR-V7 is expressed in normal hematopoietic cells, it may be mandatory to completely eliminate infiltrating hematopoietic cells from cancer-containing tissue before conducting AR-V7 assays in order to show that AR-V7 of tumor origin is actually expressed. This maneuver seems much more difficult in solid tissues than in liquid materials like blood. Moreover, the amounts of AR-V7 mRNA in cells in blood are very low, as a lot of cycles of amplification are often necessary to show up AR-V7 transcripts. The threshold for positive AR-V7 detection could be very hard to determine. Considering the factors mentioned earlier, it may not be very convenient to utilize AR-V7 mRNA expression as a biomarker for abiraterone acetate and enzalutamide administration in patients with CRPC.

Conclusion

AR-V7 mRNA expression in normal hematopoietic cells may annihilate the manifestation of aggressiveness of prostate cancer and the prediction of the effectiveness of abiraterone and enzalutamide for CRPC by assessment of AR-V7 mRNA in blood.

Disclosure

The authors report no conflicts of interest in this work, and no conflict of interest regarding the publication of this paper.

References