

Quantitative [Fe]MRI of PSMA-targeted SPIONs specifically discriminates among prostate tumor cell types based on their PSMA expression levels

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Abstract: We report the development, experimental verification, and application of a general theory called [Fe]MRI (pronounced fem-ree) for the non-invasive, quantitative molecular magnetic resonance imaging (MRI) of added magnetic nanoparticles or other magnetic contrast agents in biological tissues and other sites. [Fe]MRI can easily be implemented on any MRI instrument, requiring only measurements of the background nuclear magnetic relaxation times (T_1 , T_2) of the tissue of interest, injection of the magnetic particles, and the subsequent acquisition of a pair of T_1 -weighted and T_2 -weighted images. These images, converted into contrast images, are subtracted to yield a contrast difference image proportional to the absolute nanoparticle, iron concentration, ([Fe]) image. [Fe]MRI was validated with the samples of superparamagnetic iron oxide nanoparticles (SPIONs) both in agarose gels and bound to human prostate tumor cells. The [Fe]MRI measurement of the binding of anti-prostate specific membrane antigen (PSMA) conjugated SPIONs to PSMA-positive LNCaP and PSMA-negative DU145 cells in vitro allowed a facile discrimination among prostate tumor cell types based on their PSMA expression level. The low [Fe] detection limit of $\sim 2 \mu\text{M}$ for SPIONs allows sensitive MRI of added iron at concentrations considerably below the US Food and Drug Administration's human iron dosage guidelines ($< 90 \mu\text{M}$, 5 mg/kg).

Keywords: contrast difference, RT-PCR, flow cytometry

Introduction

Superparamagnetic iron oxide nanoparticles (SPIONs) are a very useful class of magnetic resonance imaging (MRI) contrast agents¹ that can be targeted to specific biological sites by recognition ligands attached to their surface. Such SPIONs loaded with therapeutic molecules, such as drugs,² microRNAs,³ or shRNA,⁴ are useful both as diagnostic and therapeutic agents, and an important, open question is how to accurately determine their concentrations in the target tissues in vivo, in a non-invasive manner. The direct methods that have been applied to SPION quantitation include both non-magnetic schemes, such as positron emission tomography-computed tomography^{5,6} and those utilizing the magnetic properties of the SPIONs, which include SQUID studies.⁷⁻⁹ An attractive alternative to direct detection of the particles is to indirectly measure their concentration by examining their influence on the magnetic resonance relaxation characteristics of tissue water¹⁰ or on the MRI-determined phase of the water signal, as in quantitative susceptibility mapping (QSM).¹¹ These indirect techniques offer the promise of high sensitivity due to the large number of water protons present in biological tissues ($\sim 10^{22}/\text{g}$), although it is still not clear whether QSM is sensitive enough to measure SPION iron concentrations less than millimolar. Relaxometry, which is based on the large increase in proton transverse relaxation rates, particularly

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for R_2^* , that occur as water molecules diffuse through the susceptibility gradients surrounding magnetic nanoparticles, has been shown to be able to detect iron concentrations, in vitro, in the parts per billion range.¹² Therefore, many authors have attempted to use simple R_2 , or R_2^* measurements to determine nanoparticle iron concentrations.^{13–19} However, R_2^* is determined by a number of tissue features independent of added SPIONs, predominant among these are susceptibility artifacts arising from tissue/air interfaces which increase R_2^* and lead to an overestimation of the SPION concentration.¹³ Furthermore, the MRI signal loss associated with large increases in R_2^* results in hypointense regions in tissue that are often difficult to distinguish from localized SPIONs. This has led to a number of complicated schemes to produce MR images with positive, rather than negative contrast^{20–24} many of which require extensive scanner reprogramming and would appear rather difficult to translate into the clinic.

We have, therefore, developed a simple, general, multi-parametric, quantitative MRI method, which we call [Fe]MRI (pronounced fem-ree) for the imaging of the concentration of added nanoparticle iron ([Fe]), and other magnetic objects, in cells and tissues that produces MR images with positive contrast and avoids many of these earlier issues and can be easily applied to cells, tissues, organs, animals, and patients. [Fe]MRI utilizes the enhancements produced by SPIONs on both the longitudinal and transverse relaxation rates, R_1 and R_2 , but not R_2^* , and is based on the contrast difference between T_1 -weighted (T_1w) and T_2 -weighted (T_2w) MR images rather than their intensities. Application of [Fe]MRI theory enabled us to correctly interpret the qualitative contrast in T_1w and T_2w MR images of SPIONs and SPION-labeled prostate tumor cells in agarose gels. We then quantitatively validated the method through the measurement of iron for SPIONs in the same samples.

The biological specificity of a SPION is determined by the recognition ligand attached to the surface of a nanoparticle. Our previous work^{25,26} had suggested that prostate specific membrane antigen (PSMA) would serve as a good target epitope for prostate cancer cells. PSMA is a cell-surface protein that is vastly overexpressed on prostate tumors.^{27–29} SPIONs were constructed with an attached antibody against PSMA. Several human prostate tumor cell lines, including LNCaP, C4-2,³⁰ and DU145 cells, were cultured and the expression of PSMA was measured by flow cytometry. We found that LNCaP and C4-2 cells expressed large numbers of PSMA molecules, while DU145 cells were virtually devoid of expression. We applied [Fe]MRI to an analysis of the specific binding of anti-PSMA-conjugated SPIONs to PSMA-positive

LNCaP and PSMA-negative DU145 cells in vitro in agarose gels and found that [Fe]MRI measurements quantitatively discriminated among prostate tumor cell types based on their PSMA expression level. These results supported the subsequent use of [Fe]MRI for the specific monitoring of [Fe] in tumors in vivo in mice, which we will report separately.

Theory of [Fe]MRI

Of fundamental significance for the quantitation of [Fe] by means of MRI, is the image contrast, C , defined as

$$C(S_A, S_B) = (S_A - S_B)/S_B \quad (1)$$

where S_A is the intensity of the voxel of interest and S_B is the intensity of a reference voxel. Note that if $S_A = S_B$ then $C = 0$, and if $S_A = 0$, then $C = -1$, and the contrast of a voxel formally varies from $C = -1$ to $C = +\infty$. MRI contrast agents alter S_A while, presumably leaving S_B unaffected.

The intensity of the nuclear magnetic resonance (NMR) signal from a voxel is a function of its intrinsic relaxation characteristics (T_1 , T_2) modified both by the presence of extrinsic agents and by the acquisition parameters (T_R , T_E), where T_1 and T_2 are the longitudinal and transverse relaxation times, and T_R and T_E are the recycle and echo times, respectively. MRI contrast agents alter the T_1 and T_2 relaxation times of the abundant water NMR signal in tissues. Therefore, the relaxivity, r , of a contrast agent is determined by its perturbation of the water proton relaxation times. This relaxation rate enhancement of a contrast agent is a linear function of its concentration, and has different values for the water T_1 or T_2 because each of these tissue relaxation characteristics arises from different physical correlation times. Therefore, the unperturbed water longitudinal and transverse relaxation rates, $R_1 (=1/T_1)$ and $R_2 (=1/T_2)$ are modified in the presence of a contrast agent by

$$R_1 = 1/T_1 + r_1 c \quad (2)$$

$$R_2 = 1/T_2 + r_2 c \quad (3)$$

where c is the concentration of the contrast agent and r_1 and r_2 are its longitudinal and transverse relaxivities, which are easily measured (Figure 1) for both free SPIONs and for SPIONs attached to or internalized by cells.

In the absence of contrast agents, $c = 0$, and the NMR signal, S , in a voxel is a function of the intrinsic relaxation times and the acquisition parameters:

$$S = M e^{-R_2 T_E} (1 - e^{-R_1 T_R}) \quad (4)$$

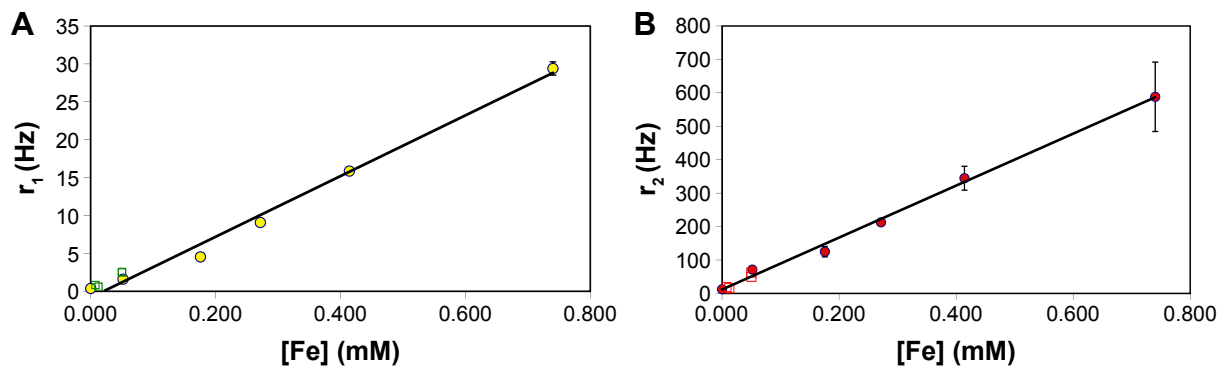


Figure 1 The relationship between the measured iron concentration and the longitudinal (r_1) (**A**) and transverse (r_2) (**B**) water relaxation rate enhancements at 1.0 T in 1% agarose gels (filled circles) containing μ MACS beads, and (open squares) anti-PSMA-conjugated μ MACS beads bound to LNCaP cells in 1% agarose.

Notes: The error bars reflect the standard error from the fits to the relaxation time measurements. The slopes give relaxivities of $r_1 = 38.3 \pm 1.1$ Hz/mM, and $r_2 = 800.4 \pm 15.9$ Hz/mM. Note that we observed no differences between the relaxivities of the SPIONs when free or bound to cells.

Abbreviations: PSMA, prostate specific membrane antigen; SPIONs, superparamagnetic iron oxide nanoparticles.

where M is an overall, machine-dependent amplitude factor proportional to the number of nuclei in the voxel. We refer to this as the tissue background signal. Note that based on Equation 4 the NMR signal in a voxel can be obtained in two rather different ways: one in which $T_E \sim T_2$ and $T_R \gg T_1$, which is called T_2w where contrast is primarily determined by differences in the tissue transverse relaxation times from voxel to voxel, and another in which $T_E \ll T_2$ and $T_R \sim T_1$, which is called T_1w where voxel to voxel contrast is primarily determined by differences in their longitudinal relaxation times.

In the presence of a contrast agent, Equation 4 needs to be modified by inserting Equations 2 and 3 into the exponentials so that the voxel signal additionally depends on the concentration, c , of the contrast agent.

$$S(c) = Me^{-(R_2 + r_2 c)T_E} (1 - e^{-(R_1 + r_1 c)T_R}) \quad (5)$$

Now, since Equation 4 shows that the NMR signal is a function of both the recycle and the echo times used to acquire the data, and that the relaxivities of the contrast agent enter separately in the exponentials for the echo and recovery amplitudes, one can imagine that contrast agents with differences in r_1 and r_2 will have separate effects on the signal amplitude depending on the set of acquisition parameters. Note here that SPIONs have $r_2 \gg r_1$, and hence are more potent in influencing tissue water T_2 than T_1 on a molar basis.

By using Equation 4 for the reference (or, from now on, we refer to this as a tissue background) voxel signal, and Equation 5 for the NMR signal from the voxel in the presence of the contrast agent, we can define the contrast, $C(c)$, in a voxel, with respect to the tissue background as

$$C(c) = \frac{e^{-(R_2 + r_2 c)T_E} (1 - e^{-(R_1 + r_1 c)T_R}) - e^{-R_2 T_E} (1 - e^{-R_1 T_R})}{e^{-R_2 T_E} (1 - e^{-R_1 T_R})} \quad (6)$$

The tissue contrast depends on Equation 6, in a complex manner, on the concentration of the contrast agent. At first glance, it would seem that Equation 6 is too complex to be useful, but, on closer inspection, it provides the key to quantitation of the contrast agent because we can write a separate version of Equation 6 for each of the two important types of nuclear resonance images; one for the T_1w case and another for the T_2w case. Furthermore, to simplify the analysis, the tissue background signal, while computed in the theory, is simply measured from the voxels of interest in an MR image in the absence of contrast agent, and is therefore just a number. Then, we have two equations for voxel contrast, one for the T_1w case and another for the T_2w case. Let us denote them by subscripts 1 and 2, respectively.

$$C_1(c) = \frac{e^{-(R_2 + r_2 c)T_{E1}} (1 - e^{-(R_1 + r_1 c)T_{R1}}) - e^{-R_2 T_{E1}} (1 - e^{-R_1 T_{R1}})}{e^{-R_2 T_{E1}} (1 - e^{-R_1 T_{R1}})} \quad (7)$$

$$C_2(c) = \frac{e^{-(R_2 + r_2 c)T_{E2}} (1 - e^{-(R_1 + r_1 c)T_{R2}}) - e^{-R_2 T_{E2}} (1 - e^{-R_1 T_{R2}})}{e^{-R_2 T_{E2}} (1 - e^{-R_1 T_{R2}})} \quad (8)$$

These can then be rewritten in an obvious notation as

$$C_1(c) = \frac{(S_1(c) - B_1)}{B_1} \quad (9)$$

$$C_2(c) = \frac{(S_2(c) - B_2)}{B_2} \quad (10)$$

where S_1 (S_2) is the (measured) T_1 w (T_2 w) voxel signal amplitude in the presence of the contrast agent, and B_1 (B_2) is the measured T_1 w (T_2 w) amplitude in the absence of the contrast agent. Equations 9 and 10 were modeled in Mathematica® as a function of the concentration of several popular SPIONs (μ MACS, Miltenyi Biotec, Bergisch Gladbach, Germany, and Dynabeads, Dynal Biotech, Oslo, Norway) with the subsequent prediction that the contrast in T_1 w images would differ by sign from that in T_2 w images depending on the SPION [Fe] and the image acquisition parameters. While the contrast in T_2 w images was usually negative, that for T_1 w images could be of either sign. What was most important, however, was the discovery that the difference in contrast,

$$\Delta C(c) = C_1(c) - C_2(c) \quad (11)$$

was always positive, and, for physiologically-relevant, US Food and Drug Administration-limited, [Fe], a monotonic,

single-valued function of [Fe]. However, Equation 11 is obviously of limited value for obtaining [Fe] from the measured ΔC , so we used Mathematica® to compute the empirical relationship between the contrast difference as a function, f , of the iron concentration, $\Delta C = f([Fe])$, and then we fitted this to high-accuracy with a polynomial, which enabled us to invert it to give

$$[Fe] = g(\Delta C) \quad (12)$$

and to determine the unknown function, g . We found that a typical set of MRI parameters gave (Figure 2) $[Fe] = 0.0005051 + 0.03961 \Delta C + 0.009258 (\Delta C)^2 + 0.008857 (\Delta C)^3$. Then, one needs to measure ΔC from a pair of images and use Equation 12 to convert the difference image into one directly giving the [Fe] in each voxel, in units of millimoles/liter. Since the background is subtracted during the calculation of the contrast images, C_1 and C_2 , the resultant [Fe] map is always bright against a zero background. Since the [Fe] MR

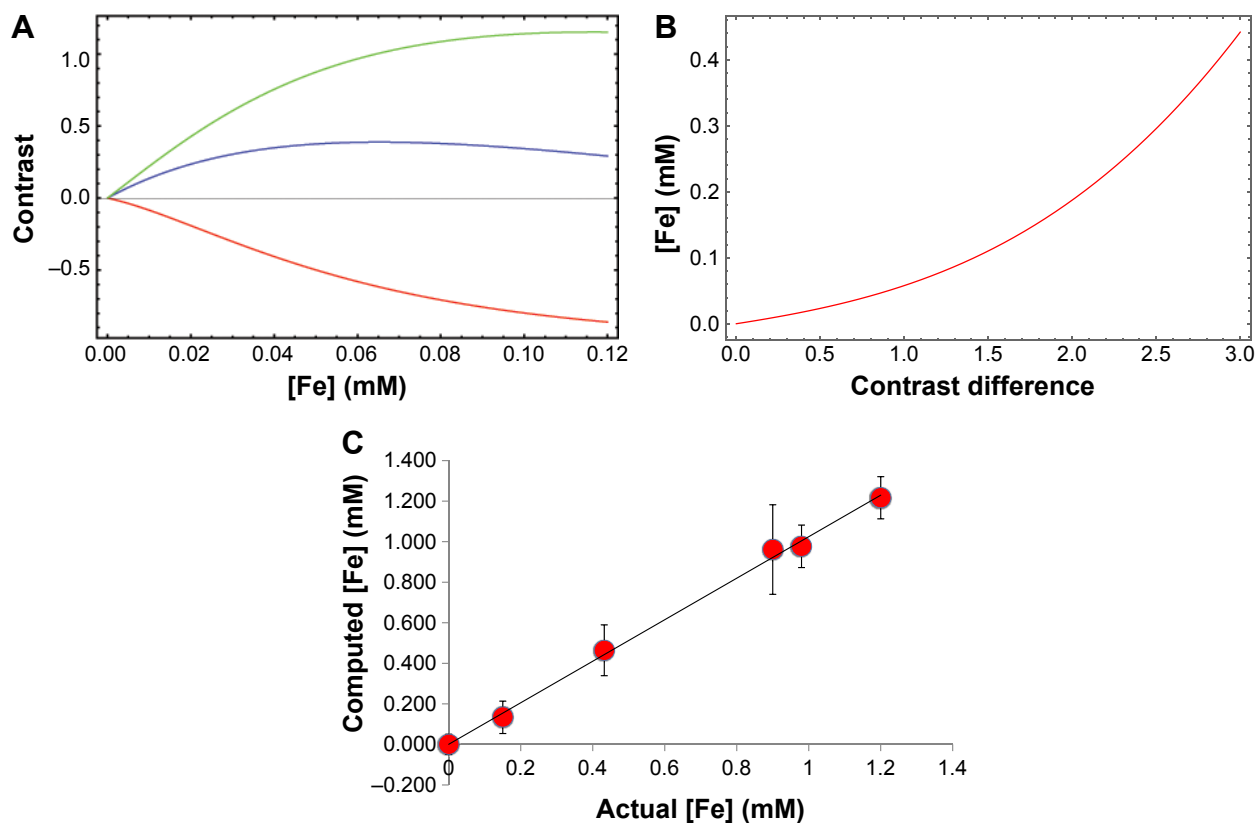


Figure 2 The theoretical and experimental relationship between MRI contrast and [Fe].

Notes: (A) Theoretical dependence of the tissue contrast on measured and set tumor MRI parameters as a function of the iron concentration. Measured parameters at 1.0 T: the r_1 relaxivity of the SPIONs was: 38.3 Hz/mM. The r_2 relaxivity of the SPIONs was: 800.4 Hz/mM. The tumor native T_1 was: 0.72 seconds. The tumor native T_2 was: 0.108 seconds. Set parameters: the echo time for the T_1 -weighted image was: $TE_1 = 0.004$ seconds. The echo time for the T_2 -weighted image was: $TE_2 = 0.0235$ seconds. The recycle time for the T_1 -weighted image was: $TR_1 = 0.5$ seconds. The recycle time for the T_2 -weighted image was: $TR_2 = 1.0$ seconds. The blue curve is for a T_1 -weighted image, the red curve is for a T_2 -weighted image, and the green curve is the contrast difference. (B) Inversion of the contrast difference relationship from (A) to give the empirical function (Equation 12) needed to convert contrast difference images into [Fe] images: $[Fe] = 0.0005051 + 0.03961 \Delta C + 0.009258 (\Delta C)^2 + 0.008857 (\Delta C)^3$, for the acquisition parameters in (A). Even though this function is non-linear, it is monotonic and well-defined. (C) Validation of the theoretical dependence of contrast on [Fe]. Suspensions ($n=3$) of varying concentrations of μ MACS SPIONs were prepared in 1% agarose gels and imaged with $TR = 0.5$ and 5 seconds, $TE = 7$ and 20 ms at 4.7 T. The relaxivities of the SPIONs at this field strength was $r_1 = 1.668$ Hz/mM, and $r_2 = 21.35$ Hz/mM. The least squares line fitted to the data has a slope of 1.024 and an intercept of +0.5 μ M. The background T_1 and T_2 of the agarose were 1.55 seconds and 0.108 seconds, respectively. The computed [Fe] was calculated from the images using the [Fe]MRI theory given in the text.

Abbreviations: MRI, magnetic resonance imaging; SPIONs, superparamagnetic iron oxide nanoparticles; TR, repetition times; TE, echo times.

image is derived from images obtained in the same gradient coordinates as the T_1w and T_2w images, registration of the [Fe]MRI data with the anatomical data is absolute.

Prior to embarking on an experimental examination of this theory, it was useful to estimate the expected additional iron concentration in cells and tissues from injected SPIONs. A simple model for an LNCaP prostate tumor cell is a sphere with a diameter $\sim 18 \mu\text{m}$. Assuming that SPIONs can completely cover the surface, we find that ~ 325 $1.05 \mu\text{m}$ Dynabeads, and 9×10^5 50 nm μMACS SPIONs can bind to each cell at saturation. With an iron content of $1.23 \times 10^{-13} \text{ g Fe per Dynabead}$ and $1.67 \times 10^{-17} \text{ g Fe per bead for } \mu\text{MACS SPIONs}$, we find that a sample of 2×10^6 LNCaP cells/mL saturated with $\mu\text{MACS SPIONs}$ would give an iron concentration of $\sim 53 \mu\text{M}$ (15 pg/cell); the value for Dynabeads was $\sim 105 \mu\text{M}$ (40 pg/cell). These order of magnitude estimates are well within the range for [Fe]MRI detection and are congruent with our [Fe]MRI measurements (vide infra). Note that a cell concentration of 2 million cells/mL is only 3% of the cell density of a LNCaP prostate tumor, which contains $\sim 3 \times 10^5 \text{ cells/mm}^3$. We estimate that, in our current, unoptimized [Fe]MRI acquisition scheme, the limit of detection (LOD) is $[\text{Fe}] \sim 2 \mu\text{M}$ (vide infra).

Materials and methods

Materials

Fluorescein-conjugated streptavidin was obtained from Thermo Fisher Scientific (Waltham, MA, USA). Dynabeads[®] MyOne[™] Streptavidin were obtained from Dynal Biotech, and MACS[®] Streptavidin MicroBeads (μMACS) were obtained from Miltenyi Biotec. Anti-PSMA antibody (clone 3C6) was purchased from Northwest Biotherapeutics (Bothell, WA, USA). Antibody was biotinylated using EZ-Link[™] Sulfo-NHS-LC-Biotin from Pierce (Rockford, IL, USA). Fully humanized anti-PSMA antibody J591 was purchased from N Bander (Weill Cornell Medical College, New York, NY, USA). The prostate cancer cell lines LNCaP, PC-3, and DU-145 were purchased from the American Tissue Type Collection (Manassas, VA, USA). The C4-2 prostate cancer cell line was a kind gift from Dr GN Thalmann (University of Bern, Bern, Switzerland).³⁰

Iron assays

SPIONs, gels containing SPIONs, and SPION-labeled cell suspensions were dissolved in 0.5 mL of 6 N HCl (Sigma-Aldrich, St Louis, MO, USA) at 95°C overnight in sealed tubes. Aliquots (0.1 mL) of the resulting solution were assayed for iron using the Prussian blue reaction by adding them to 0.9 mL of $10\% \text{ K}_4\text{Fe}(\text{CN})_6$ (Sigma-Aldrich), incubating at 60°C for 30 minutes, and reading the absorbance

at 690 nm . Standard curves ($n=7$) prepared using 1.00 mM FeCl_3 (Sigma-Aldrich) gave an extinction coefficient of $9,165 \pm 148 \text{ L/mol cm}$.

Cell culture

LNCaP cells were cultured in Roswell Park Memorial Institute 1640 medium (Sigma-Aldrich) supplemented with 10% fetal calf serum; C4-2, PC-3, and DU-145 were cultured in T-medium with 10% fetal calf serum (HyClone Laboratories, Inc., Logan, UT, USA).³⁰ All cells were cultured at 37°C in a humidified 5% CO_2 atmosphere. Upon reaching 90% confluency, cells were collected either by detachment in 0.5% trypsin containing 0.02% ethylenediaminetetraacetic acid or detached in cold phosphate-buffered saline (PBS) using cell scrapers. No ethical approval was required in the use of these cells for this study.

Real-time RT-PCR

In order to determine the extent of PSMA expression in human tissues and cultured prostate cell lines, we performed PCR assays to measure the amount of PSMA messenger RNA produced. Total RNA was isolated from cells using the RNeasy Kit (Qiagen, Valencia, CA, USA). Contaminating DNA was removed by DNase I treatment (Clontech, Palo Alto, CA, USA). cDNA was synthesized using avian myeloblastosis virus reverse transcriptase (Promega Corporation, Fitchburg, WI, USA) and random hexamer primers (Amersham Biosciences Corporation, Piscataway, NJ, USA) in the presence of RNase inhibitor (Promega Corporation). Real-time RT-PCR was performed in a 7000 Sequence Detection System thermocycler (Thermo Fisher Scientific) using gene-specific primer pairs and fluorescent reporter probes (Integrated DNA Technologies, Coralville, IA, USA). To avoid amplification from genomic DNA, intron-spanning primers were used (designed with Primer Express software; Thermo Fisher Scientific). The primers and probe for PSMA were: 5-TGAGAGACTCCAGGACTTTGACAA-3 (forward), 5-GGATCAATAAATGCTCTTTCCAGAA-3 (reverse), and 5-AGCAACCCAATAGTATTAAGAATGATGAATGATCAACTCA-3 (probe). For glyceraldehyde-3-phosphate dehydrogenase, commercially available reagents (Thermo Fisher Scientific) were used. Three hundred and fifty nanograms of cDNA were used in a total volume of $25 \mu\text{L}$ of PCR MasterMix containing 900 nM of the primers and 300 nM of the probe. The cycling conditions were 95°C 10 minutes, $45 \times (95^\circ\text{C} 15 \text{ seconds}, 60^\circ\text{C} 1 \text{ minute})$. No-template and non-reverse transcribed RNA controls were included. The data were analyzed using the Sequence Detection System software (Thermo Fisher Scientific). The expression

differences were calculated by the $2^{-\Delta\Delta C_t}$ method for assessing relative expression (Thermo Fisher Scientific, User Bulletin #2, 2001). The signals for PSMA were normalized to the signals for glyceraldehyde-3-phosphate dehydrogenase to control for RNA input. The size of the amplicons was verified in agarose gels.

Flow cytometry

Since there is often an imperfect correlation between messenger RNA amounts in cells and the amounts of the translated proteins, we also performed flow cytometry to further investigate the levels of cell surface expression of PSMA. All cells were harvested by trypsinization, centrifuged and resuspended in PBS containing 0.1% azide and 1% bovine serum albumin. Cells were analyzed for cell surface expression of PSMA using either biotinylated or unconjugated MAb 3C6. Fluorescent labeling was achieved with either a fluorescein conjugate of streptavidin or with a phycoerythrin conjugate of rat anti-mouse antibody (BD Biosciences, San Jose, CA, USA). The data were acquired using a Becton Dickinson FACscan flow cytometer (Brea, CA, USA) equipped with a 488 nm argon laser and CellQuest software (CellQuest, San Jose, CA, USA).

Antibody conjugation and cell labeling

Sulfo-NHS-LC-Biotin groups were attached to primary amines of MAb 3C6 using the EZ-Link™ Sulfo-NHS-LC-Biotin kit according to the manufacturer's protocol. Conjugated MAb 3C6 was separated from low molecular weight compounds using a Millipore Ultrafree®-MC Centrifugal Filter Unit (EMD Millipore, Billerica, MA, USA) at a force of $3,000\times g$ in a fixed angle microfuge rotor. Antibody concentration was determined using the Pierce Protein Assay Reagent Kit, and the ratio of biotin/antibody was determined with a Pierce HABA (4'-hydroxyazobenzene-2-carboxylic acid) colorimetric assay, both according to the manufacturer's protocol. Later experiments utilized J591, which was biotinylated using the same method as 3C6 and attached to streptavidin-conjugated SPIONs by incubation for 30 minutes. The free antibody was removed by washing the beads with the aid of a magnetic bead separation device (Miltenyi Biotec).

SPION-binding to prostate tumor cells

Dynabeads® MyOne™ Streptavidin superparamagnetic beads (1.05 \pm 0.10 μ m diameter, 26% iron oxide w/w, polystyrene coating, 1.23×10^{-13} g Fe/bead) and μ MACS® Streptavidin MicroBeads (~50 nm diameter, 57% iron oxide w/w, dextran coating, 1.67×10^{-17} g Fe/bead) were used as

contrast agents for MRI. The Dynabeads® solution contained detergent (0.01% Tween 20) and preservative (0.09% sodium azide), which were removed by multiple washing with PBS. The μ MACS® solution contained 0.05% sodium azide, but no detergent, and was not washed. LNCaP and DU-145 cell suspensions were incubated with biotinylated PSMA for 30 minutes at 4°C, followed by washing with PBS. Labeled cells were incubated with the streptavidin superparamagnetic beads (10 μ L bead solution/ 10^7 cells) for 30 minutes at 4°C with gentle agitation. Cells were separated from unbound beads by repeated centrifugation at $300\times g$. LNCaP and DU-145 cells were each resuspended in ~100 μ L low melting point 1% agarose in PBS and layered into plastic tubes as described in the MRI sample preparation section below.

MRI sample preparation

For the SPION-only samples, SPIONs at various concentrations were mixed with 1% agarose gel in 10 mm outside diameter plastic tubes and MR images and relaxation rates were measured at 0.27 T, 1.0 T, 1.9 T, and 4.7 T. For the SPION-labeled cells, the MRI samples were prepared with the layers of SPION-labeled LNCaP and DU-145 cells, in between layers of agarose gel, in 10 mm outside diameter plastic tubes, to measure the MRI signal intensities of the control DU-145 cells, the PSMA-positive LNCaP cells, and the agarose simultaneously in the same sample.

Magnetic resonance imaging

MRI was carried out using horizontal bore instruments at 1.0 T (MRT Systems, Tsukuba, Japan), at 1.9 T (Oxford Research, Oxfordshire, UK), and at 4.7 T (Bruker Biospec, Billerica, MA, USA). T_2 -weighted images were acquired using spin echo (SE) imaging sequences with echo times (TE) varying from 5.5 to 145.5 ms. T_1 w images were acquired by varying the repetition times (TR) from 0.3 seconds to 12 seconds (for TE ~4–7 ms). T_1 values were measured with a series of one-dimensional SE images with TE = 10.7 ms, and T_2 decay curves were generated using a series of SE images with different TE values. The relaxivities of the SPIONs were determined separately, and bound to cells, in 1% agarose gels by measuring the relaxation rates ($r_1 = 1/T_1$, $r_2 = 1/T_2$) for various [Fe] values (Figure 1).

Image processing

The complex, time-domain MRI datasets were baseline-corrected, Fourier-transformed, and co-added using Mathematica version 10.0.2 (Wolfram Research, Urbana, IL, USA) to produce the frequency domain images of each MRI slice. The theoretical dependence of the image contrast on [Fe] and

its inversion to produce [Fe] as a function of contrast and code to convert the MR images into [Fe] images was written in Mathematica. The average and standard deviation of the pixel intensities in the regions of interest were measured with the Mira AP software (Axiom Research, Inc., Tucson, AZ, USA). The errors in the computed contrast were calculated using a standard propagation-of-errors analysis³¹ based on the measured standard deviations of the pixel intensities for the cells and agarose.

Results

The relaxivity of μ MACS and Dynabeads

The NMR water relaxation rate enhancements (relaxivities) of SPIONs depend on the particle size. For this reason, we selected commercially available superparamagnetic particles of two quite different sizes in order to encompass a large range of particle sizes that might be used in practice. Dynabeads are iron oxide particles with a nominal diameter of 1 μ m, while μ MACS particles are much smaller, with diameters on the order of 50 nm. The magnetic relaxation of water protons is also a function of the rotational correlation time of the water molecule, so that the relaxivities of a SPION depend on the NMR Larmor frequency.³² For example, for μ MACS SPIONs, we found that r_1 (the longitudinal rate enhancement) decreased linearly from 175 Hz/mM at 0.27 T, to 1.67 Hz/mM at 4.7 T. Therefore, it was necessary to measure the relaxivity of a SPION at the same field strength at which it was to be used. At 1.0 T, we determined the r_1 and r_2 relaxivities of the μ MACS to be $r_1 = 38.3 \pm 1.1$ Hz/mM, and $r_2 = 800.4 \pm 15.9$ Hz/mM (mean \pm SD, Figure 1) and $r_1 = 0.080 \pm 0.0023$ Hz/mM and $r_2 = 32.02 \pm 0.91$ Hz/mM for the Dynabeads. We found that these relaxivities at 1.0 T were independent of the aggregation state of the SPIONs; they were the same for free SPIONs and SPIONs bound to prostate tumor cells (Figure 1). These measurements were also performed at 0.27 T, 1.9 T, and 4.7 T, and the results are given in the figure captions as needed.

Theoretical dependence of contrast on [Fe]

The MRI data were taken at 1.0 T with recycle times ranging from 500 to 5,000 ms, and with echo times from 4 to 145 ms. A typical theoretical calculation of the T_1 w and T_2 w image contrast, and contrast difference, using the measured relaxivities of the μ MACS SPIONs at 1.0 T and acquisition parameters within this range shows (Figure 2A) that the T_2 w image contrast is predominantly negative for all but the lowest [Fe] values, while the T_1 w image contrast is positive over the range of [Fe] anticipated in biological

tissues. Note that the contrast difference was always a positive, monotonically, increasing function of [Fe], for all iron concentration values expected. Inversion of the contrast difference function, $\Delta C = f([Fe])$, to produce the relationship between [Fe] and ΔC (Equation 12) is shown in Figure 2B; $[Fe] = 0.0005051 + 0.03961 \Delta C + 0.009258 (\Delta C)^2 + 0.008857 (\Delta C)^3$. While this inversion is straightforward to perform in Mathematica®, it needs to be computed for each set of tissue, SPION, and acquisition parameters. It also serves as an essential guide for planning experiments to achieve the optimal sensitivity of contrast to [Fe]. The units of [Fe] naturally arise as mM due to the fact that the longitudinal and transverse relaxivities of the SPIONs are given in Hz/mM; these units set a natural scale to the problem.

Comparison of the theoretical dependence of [Fe] with iron assay

To validate this method we compared the theoretical predictions with independent measurements of the [Fe] in MRI samples prepared from 1% agarose gels containing known concentrations of SPIONs. After the MRI measurements, the gels were removed from the tubes and the [Fe] was measured using the Prussian blue iron assay (see “Materials and methods”). Even though the relationship between pixel contrast difference and [Fe] was clearly non-linear (Figure 2B). It was still monotonic and single-valued and could therefore be used to convert ΔC into [Fe]. Thus, the data show (Figure 2C) that we can accurately measure [Fe] with this method; the slope of the fitted line is 1.024 ± 0.041 , which is not statistically, significantly different from 1.0, with an intercept of 0.5 μ M Fe.

Determination of the LOD of iron using [Fe]MRI

A Dynabead stock solution was diluted 1:6,250, 1:1,250, and 1:250 into 1% agarose and placed into 10 mm plastic tubes as bands surrounded by agarose. A T_1 w MR image was taken (Figure 3A) at TR=1 second, TE=5 ms, nex=6, and a T_2 -weighted image (Figure 3B) was taken at TR=1 second, TE=115 ms. The resulting [Fe]MR image (Figure 3C) showed detectable bands for [Fe]=9.7 and 48 μ M, with a barely detectable band at [Fe]=1.97 μ M. These bands were more easily seen and quantitated by means of a Mathematica-generated three-dimensional surface plot (Figure 3D) of the [Fe] as a function of the spatial coordinates in the tube, and therefore, we used this presentation in several of the following figures. The two bands with the highest [Fe] were easily detected, with signal to noise ratios of 4.7 (9.7 μ M Fe) and 6.6 (48 μ M Fe), but the band with only 1.97 μ M Fe had

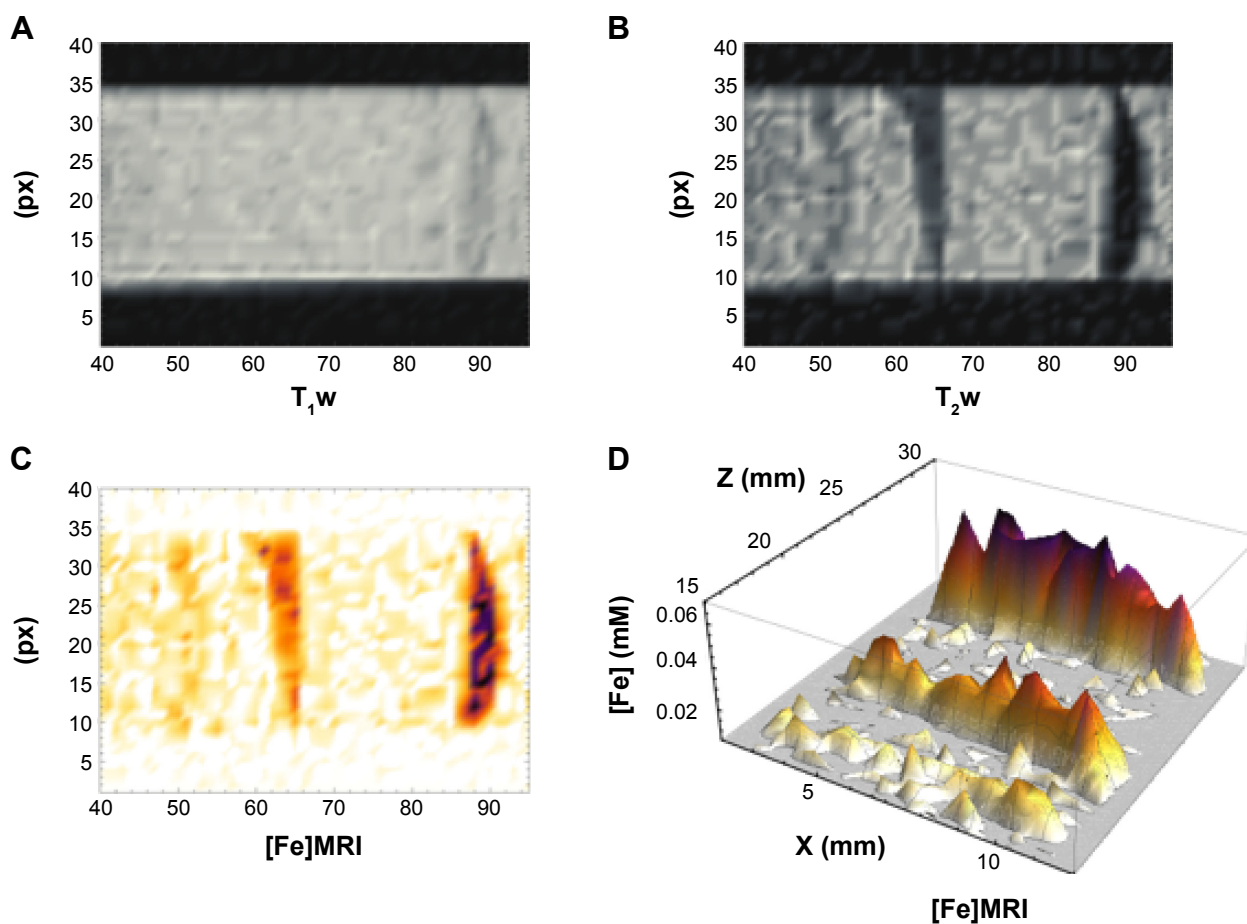


Figure 3 Determination of the limit of detection for iron using [Fe]MRI at 1.0 T. A Dynabead stock solution was diluted 1:6,250, 1:1,250, and 1:250 into 1% agarose and placed into 10 mm plastic tubes as bands surrounded by agarose.

Notes: (A) A T₁-weighted MR image at TR = 1 seconds, TE = 5 ms. (B) A T₂-weighted MR image at TR = 1 seconds, TE = 115 ms. (C and D) The resulting [Fe]MR images showing detectable bands for [Fe] = 9.7 and 48 μ M, with a barely detectable band at [Fe] = 1.97 μ M.

Abbreviations: MRI, magnetic resonance imaging; TR, repetition times; TE, echo times.

a signal to noise ratio of only 1.6, which was less than 2σ different from the noise, and therefore not a candidate detection. Thus, we found a detection limit of ~ 2 – 2.5 μ M Fe, for a signal to noise ratio of $\sim 45:1$ for the input T₁w and T₂w images. It appears to be an easy task to improve this LOD by acquiring higher signal to noise ratio images.

Specific binding of anti-PSMA-conjugated SPIONs to PSMA-positive prostate tumor cells

The [Fe]MRI measurement of [Fe] in SPION preparations in vitro was interesting in its own right, but the application of this method to biological samples was our primary goal. Our long history of applications of NMR to prostate cancer diagnosis^{2,25,26,33–37} suggested that we turn to this area for a candidate cell-surface epitope to discriminate between cancer cells and normal, non-malignant cells. Research on prostate cancer-specific cell markers^{25,27–29} led to the identification of

PSMA, the folate receptor, as a cell-surface protein that is highly-overexpressed on prostate tumors.

PSMA is a cell-surface protein expressed in the prostate at levels a 1,000-fold greater than that found in other tissues, such as kidney.³⁸ The expression of PSMA increases in prostate cancer and its expression is correlated with tumor grade, pathological stage, aneuploidy, and biochemical recurrence.²⁸ A humanized antibody to PSMA, J591, has been successfully used to image sites of metastasis.^{38–43} Ghosh and Heston³⁸ have demonstrated LNCaP tumor localization in nude mice with labeled [¹³¹I]-, and [¹¹¹In]-J591. In addition, Phase I studies using radiolabeled or cytotoxin (DM1)-linked J591 have also shown excellent tumor targeting.¹

Once a tumor-specific epitope had been identified, we sought cell lines that differed in its expression. The expression of PSMA at the level of mRNA was measured using reverse transcriptase polymerase chain reaction (RT-PCR), and the cell-surface protein expression confirmed using flow

cytometry²⁵ of several commonly cultured human prostate tumor cell lines, including LNCaP, C4-2, DU145, and PC3 cells. RT-PCR revealed significant expression of PSMA in both the LNCaP and C4-2 cell lines, while it was virtually undetectable in DU-145 and PC-3 cells under the PCR conditions used (see “Materials and methods”) in agreement with previous studies.^{28,39,44,45} Flow cytometry showed that LNCaP and C4-2 cells expressed $(1.05 \pm 0.10) \times 10^6$ and $(1.67 \pm 0.10) \times 10^6$ PSMA molecules per cell, while DU145 and PC3 cells expressed 100-fold less at only $(1.35 \pm 0.52) \times 10^4$ PSMA molecules per cell. Given their very different PSMA expression levels, we chose to use LNCaP and C4-2 cells as PSMA-positive samples, and DU145 cells as negative controls.

The anti-PSMA antibody, 3C6, was conjugated to the surface of the various SPIONs. Binding assays (Figure 4) in vitro showed that, on average, LNCaP cells ($n=100$) bound 80 ± 16 of

the $1 \mu\text{m}$ anti-PSMA-conjugated Dynabeads per cell, roughly consistent with our earlier order of magnitude estimates (vide supra). The control, PSMA-negative DU145 cells did not bind significant amounts of the anti-PSMA-conjugated SPIONs.

MRI contrast as a function of [Fe] for LNCaP cells with anti-PSMA-conjugated SPIONs

The theoretical relationship between MRI contrast and [Fe] (Figure 2A) predicted that for low [Fe] the T_1 w image contrast would be positive, while the T_2 w contrast would be negative for cells that bound SPIONs. While a test of this relationship (Figure 3) showed that these predictions were qualitatively correct, it was of further interest to numerically compare these predicted relationships with measurements. We found (Figure 5) that [Fe]MRI could quantitatively reproduce the dependence of contrast in T_1 w and T_2 w MR images on [Fe]

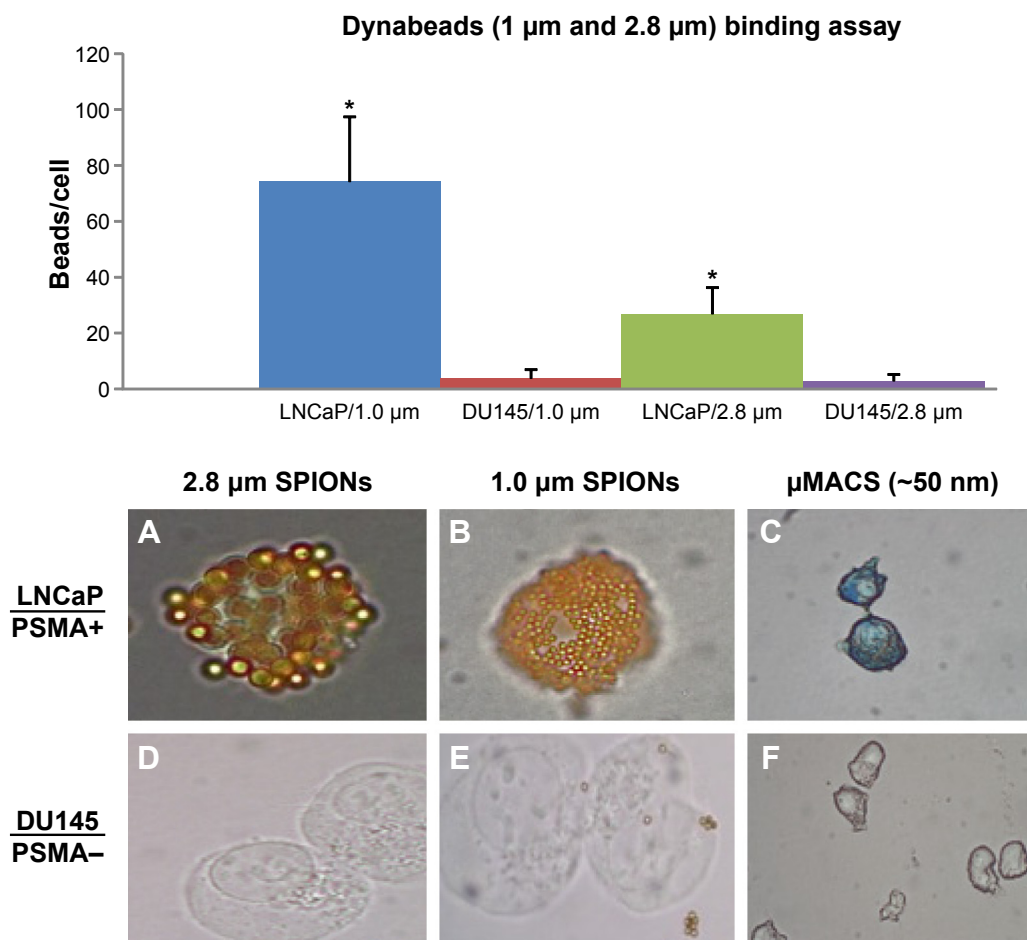


Figure 4 Specific binding of anti-PSMA antibody-conjugated SPIONs to prostate cancer cells, in vitro.

Notes: Fully humanized monoclonal antibody (J591) against PSMA was conjugated to Dynabeads (2.8 μm , [A, D]; 1.0 μm , [B, E]) and to μMACS (~50 nm, [C, F]) and then incubated with either PSMA-positive LNCaP (A, B, C) or PSMA-negative DU145 (D, E, F) cells at 4°C (to prevent uptake into the cytoplasm), washed and examined under the microscope. The SPIONs bound to each cell we either counted directly for $n=100$ cells, or in the case of the μMACS , Perl's stain for iron was applied to reveal the presence or absence of iron. Note that only the PSMA-positive LNCaP cells bound the SPIONs or stained blue indicating the presence of μMACS SPIONs. $*P<0.001$.

Abbreviations: PSMA, prostate specific membrane antigen; SPIONs, superparamagnetic iron oxide nanoparticles.

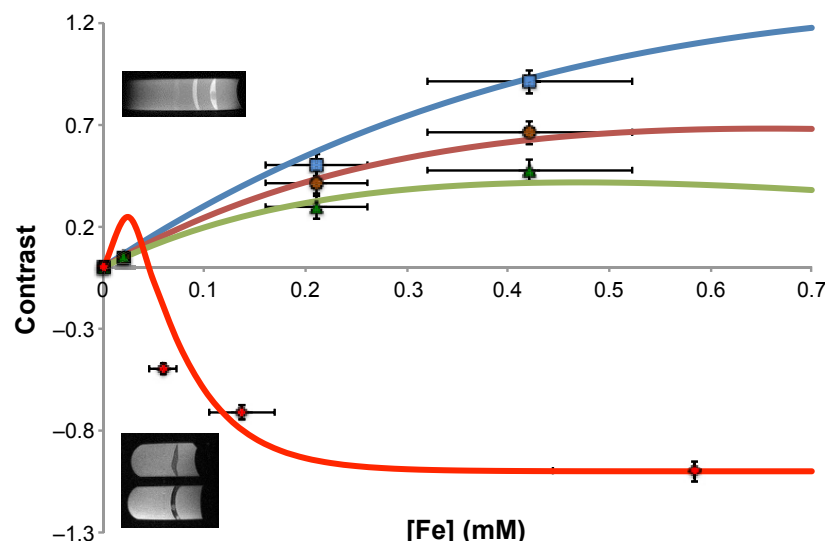


Figure 5 MRI contrast as a function of $[Fe]$ for T_1 -weighted (T_1w) (blue, brown, and green) and T_2 -weighted (T_2w) (red) MR images of suspensions of LNCaP cells with anti-PSMA-conjugated Dynabeads (top image and curves) and anti-PSMA-conjugated μ MACS SPIONs (bottom image and curve).

Notes: Note that the T_1w contrast was positive, while the T_2w contrast was negative, in agreement with Equations 7 and 8. The smooth lines are the theoretical predictions for the contrast given the measured relaxation and set acquisition parameters for each image: blue ($TR=0.5$ seconds, $TE=0.009$ seconds); brown ($TR=1.0$ seconds, $TE=0.009$ seconds); green ($TR=1.5$ seconds, $TE=0.009$ seconds); red ($TR=1.0$ seconds, $TE=0.023$ seconds). The insets show representative images from each set used to derive the contrast data. The LNCaP cells bound $(15.3 \pm 3.7) \times 10^4$ μ MACS SPIONs per cell and the cell densities ranged from 0 to 1.27×10^5 per μ L. These cells also bound ~ 80 Dynabeads per cell. The measured agarose relaxation times at 1.0 T gave $T_1=2.0$ seconds, $T_2=0.108$ seconds.

Abbreviations: MRI, magnetic resonance imaging; PSMA, prostate specific membrane antigen; SPIONs, superparamagnetic iron oxide nanoparticles; TR, repetition times; TE, echo times.

over essentially the entire physiological range of SPION iron concentrations. Note that the $[Fe]$ MRI theory contains no free parameters, only directly measured quantities.

$[Fe]$ MRI determination of $[Fe]$ for anti-PSMA-conjugated μ MACS SPIONs bound to PSMA-positive prostate tumor cells

There are two, equally-valid methods for the specific SPION labeling of PSMA-positive tumor cells. The first closely follows the logic for immunohistochemistry in which the PSMA-positive LNCaP cells were incubated with biotinylated anti-PSMA antibody, washed to remove free antibody, incubated with streptavidin-conjugated μ MACS SPIONs, washed to remove unbound SPIONs, and mixed with 0.1 mL of 1% agarose. The specificity for PSMA is preserved because the biotinylated antibody would bind only very weakly to PSMA-negative DU145 cells and produce only a small fraction of biotin-binding sites for the streptavidin-conjugated μ MACS SPIONs. The second method involves binding the streptavidin-conjugated μ MACS SPIONs to biotinylated PSMA, washing the SPIONs and incubating them with the cells and/or tissues. We initially used the first method in this work since one had direct access to the cell surface; we later switched to the second method because it was more appropriate for performing subsequent animal studies. The details for both techniques are given in the "Materials and methods" section. Since the results did not

depend on the order of the labeling, we refer to the SPIONs as PSMA-conjugated for simplicity.

Aliquots containing 2×10^6 , 1×10^6 , and 1×10^5 tumor cells were then layered onto an existing 1% agarose gel, and covered with additional agarose to form a tube containing bands of cells in agarose, and intervening bands of cell-free gel. $[Fe]$ MRI was performed on these cell phantoms to show that this method was capable of imaging the iron on SPION-bound cells, to measure the $[Fe]$, and to determine the limits of detection for SPION-bound cells. The results show (Figure 6) that LNCaP cells with anti-PSMA bound to their surface bound μ MACS SPIONs and gave rise to significant, positive T_1w MRI contrast. Computation of the $[Fe]$ MR image (Figure 6B and C) revealed the specific presence of iron only in the cell-containing bands. The bands containing 2 and 1 million cells were easily detectable, but the band containing only 100,000 cells did not reach the LOD. The measured $[Fe]$ ranged from 4 to 41 μ M, which combined with the band volume of 67 μ L and the number of cells per band yielded an iron value $\sim 9 \times 10^{-14}$ g Fe per cell, resulting from $\sim 3,200$ μ MACS per cell. This is approximately 3.5% of saturation, where 90,000 SPIONs could potentially tile the surface of a LNCaP cell (vide supra). We also found that the $[Fe]$ MRI signal scaled linearly with the number of cells present (Figure 6D). Again, we find that the limit of Fe detection was ~ 2 μ M. Other studies have found similar amounts of Fe in cells treated with SPIONs; for example,

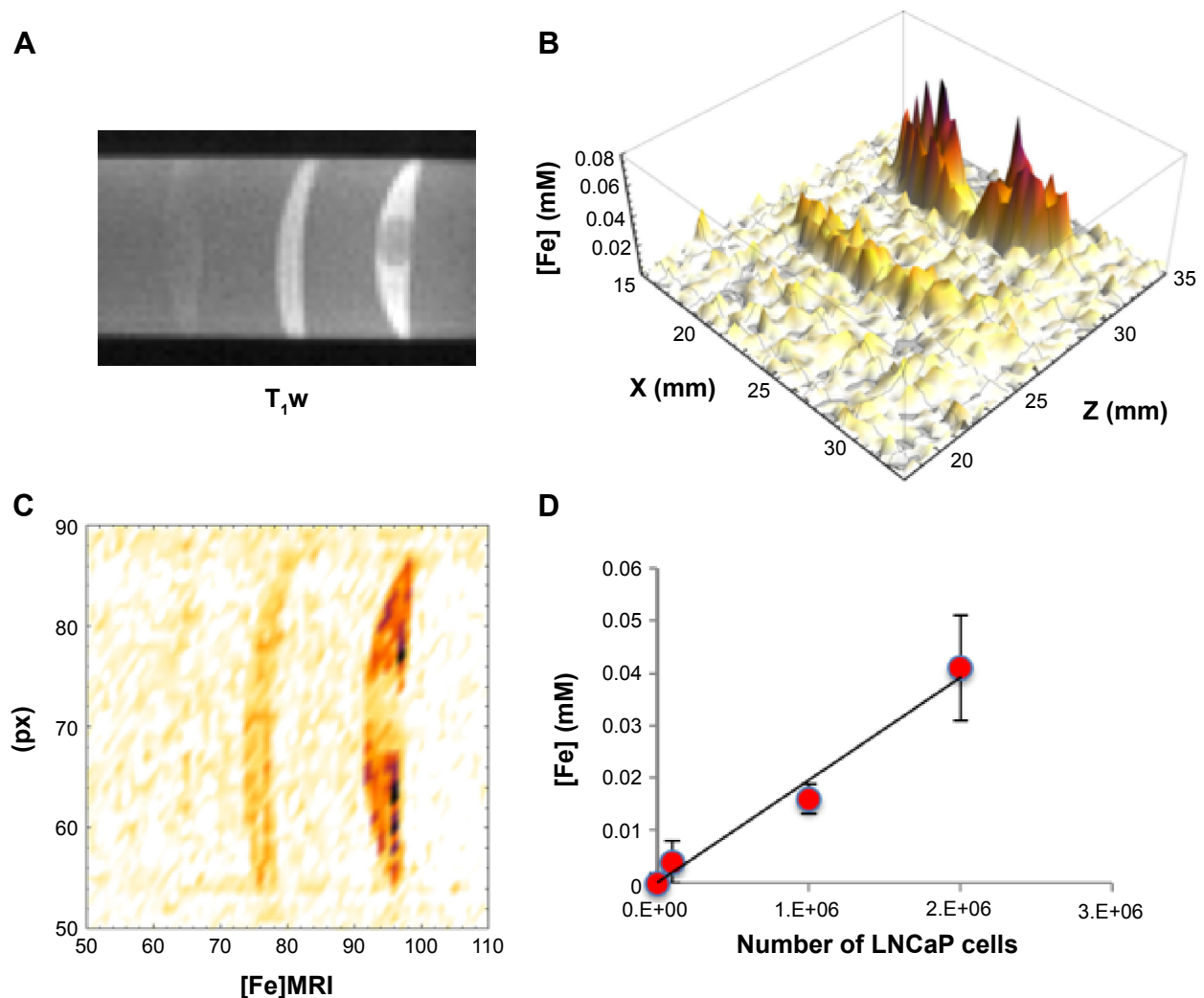


Figure 6 [Fe]MRI of anti-PSMA antibody-conjugated SPIONs to LNCaP prostate cancer cells, in vitro. Fully humanized anti-PSMA antibody J591 was conjugated to μ MACS SPIONs which were then bound to LNCaP cells and aliquots of cells containing 2×10^6 , 1×10^6 , and 1×10^5 cells were layered in 1% agarose gels.

Notes: (A–C) The bands containing 2 and 1 million cells were easily detectable, but the band containing only 100,000 cells barely reached the limit of detection. (D) The [Fe] in each band is plotted against the number of cells to yield the contribution per cell. From the band volume we calculate that there were $\sim 3,200$ μ MACS SPIONs per cell.

Abbreviations: MRI, magnetic resonance imaging; PSMA, prostate specific membrane antigen; SPIONs, superparamagnetic iron oxide nanoparticles.

Brisset et al,¹⁴ found that cells took up 8 pg Fe, while Wang and Cuschieri⁴⁶ measured 4 pg/cell.

[Fe]MRI determination of [Fe] for anti-PSMA-conjugated SPIONs internalized by LNCaP cells

We have shown that μ MACS bind to anti-PSMA-conjugated LNCaP Cells at 4°C, but are not taken up into the cytoplasm until the cells are warmed to 37°C, at which temperature the SPIONs are rapidly internalized with a time constant of 5–7 minutes.²⁶ It was of interest, therefore, to bind SPIONs to the cell surface at 4°C and then to allow internalization by raising the temperature. We prepared a cell phantom in agarose, as before, and used [Fe]MRI to measure the [Fe]. The results (Figure 7) showed that we could still observe a strong [Fe]MRI signal from the internalized SPIONs, and that

this signal scaled linearly with the number of cells, as shown in Figure 6. We found that these cells took up $(26.2 \pm 1.2) \times 10^4$ SPIONs per cell, in agreement with our previous order of magnitude estimate (vide supra) which corresponded to (1.74 ± 0.12) pg Fe/cell. The MR images (Figure 7) provided a strong visual confirmation of the theoretical prediction (Figure 2A) that for the acquisition parameters used here, T₁w images should show positive contrast and T₂w images should show negative contrast.

[Fe]MRI of anti-PSMA-conjugated SPIONs discriminates PSMA-positive from PSMA-negative prostate tumor cells

To use anti-PSMA-conjugated SPIONs for the specific detection of prostate tumors, one must first show that these SPIONs can distinguish cells based on their expression of PSMA.

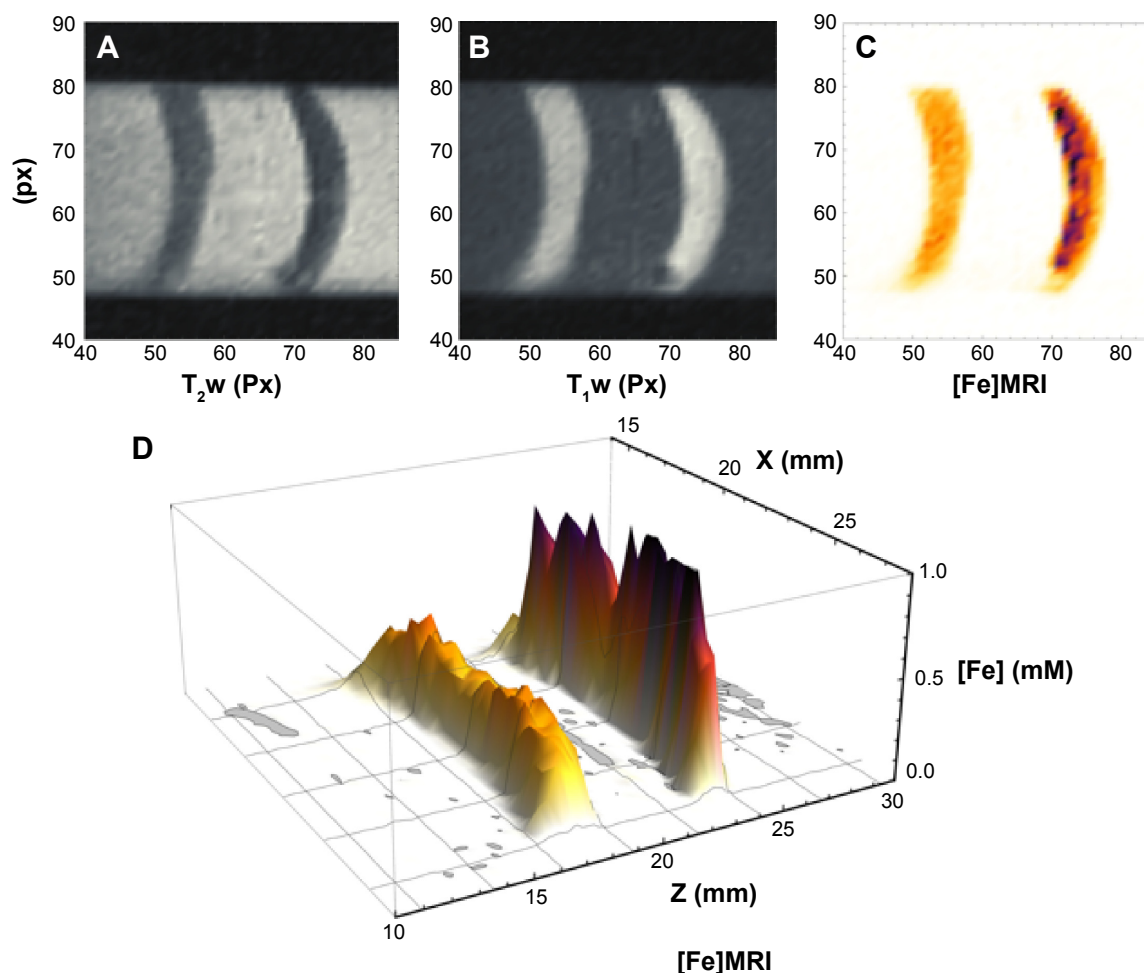


Figure 7 [Fe]MRI of LNCaP cells that have internalized anti-PSMA antibody-conjugated μ MACS SPIONs.

Notes: The cells were incubated with the SPIONs at 4°C, washed and then incubated for 60 minutes at 37°C so that the cells could take up the SPIONs. (A) T_2w (TR = 0.5 seconds; TE = 9 ms) and (B) T_1w (TR = 1.5 seconds; TE = 23.5 ms) images at 1.0 T. (C) [Fe]MRI map. (D) Three-dimensional plot of the [Fe]MRI map in (C). The bands in A–D contained 1 and 2 million cells, respectively, with [Fe] of 359 ± 34 and 658 ± 41 μ M.

Abbreviations: MRI, magnetic resonance imaging; PSMA, prostate specific membrane antigen; SPIONs, superparamagnetic iron oxide nanoparticles; TR, repetition times; TE, echo times; T_1w , T_1 -weighted; T_2w , T_2 -weighted.

Our RT-PCR and flow cytometry measurements (vide supra) showed that LNCaP cells were PSMA-Positive, while DU145 cells expressed 100-fold less PSMA and were therefore essentially PSMA-negative. We incubated biotinylated-anti-PSMA with LNCaP and DU145 cells, washed the cells to remove unbound 3C6, and then incubated equal numbers of each of these cells with streptavidin-conjugated SPIONs, washed away the unbound particles, and prepared agarose gels in plastic tubes. When we applied [Fe]MRI to these samples, we found (Figure 8) that the LNCaP cells bound 8.7-fold more of these SPIONs than the DU145 cells, indicating excellent specificity for the detection of cell-surface PSMA.

Discussion

We have developed, presented and applied a general, quantitative, multiparametric MRI theory for the measurement

of nanoscale magnetic objects introduced into biological systems. This new ability to directly, and simply, measure the amount of a nanoparticle, or other magnetic material using standard MRI, will enhance a large class of nanotechnology studies. The present LOD for [Fe]MRI was found to be ~ 2 μ M, but that could evidently be lowered through judicious choices of the relaxivity of the objects, which in this case were SPIONs, and by calculating and optimizing the contrast and signal to noise ratio based on varying the MRI acquisition parameters. We found that expected concentrations of iron could be detected from SPIONs and from SPIONs bound to and internalized by cells. This allowed us to show that [Fe]MRI could discriminate among prostate tumor cell types based on the expression, or lack thereof, of cell surface tumor markers; in this particular case of PSMA. One can envision a large range of other applications of [Fe]MRI.

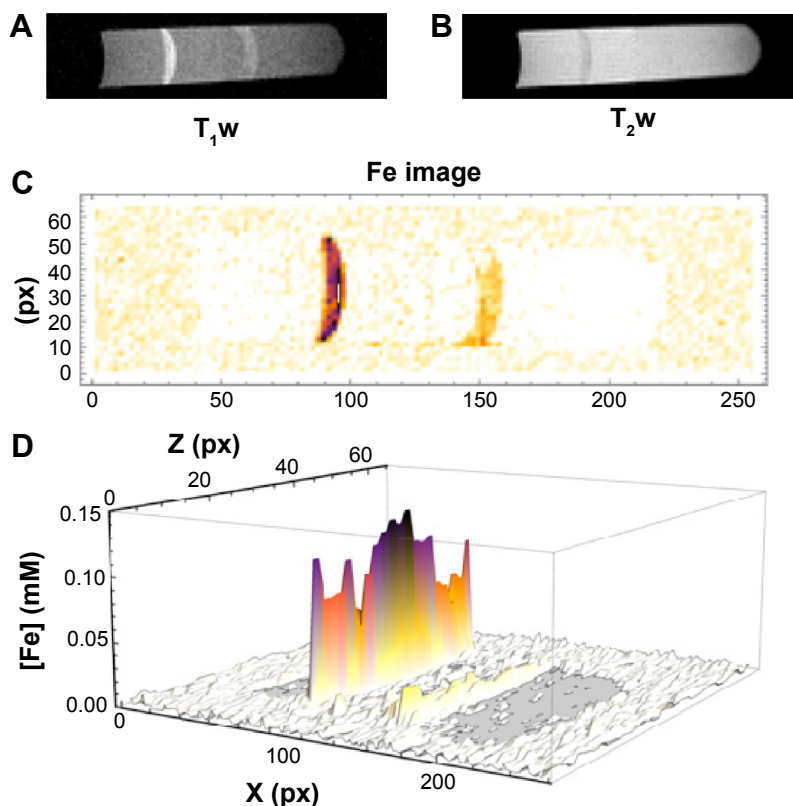


Figure 8 [Fe]MRI with anti-PSMA antibody-conjugated μ MACS SPIONs can discriminate PSMA-positive LNCaP from PSMA-negative DU145 cells.

Notes: The cells were incubated with the SPIONs for 30 minutes, washed and then mixed with 1% agarose and layered into a tube containing 1% agarose. (A) T₁-weighted MR image. The DU145 cells were inserted first and appear as a weak band near the bottom of the tube, while the LNCaP cells were inserted second and appear as a strong band near the top of the tube (B) T₂-weighted MR image. (C) [Fe] image. (D) Three-dimensional plot of the [Fe] image in (C). The [Fe] = (78 \pm 29) μ M for the LNCaP cells and only (9 \pm 4) μ M for the DU145 cells indicating a >8.7-fold difference in iron binding by the two cell lines.

Abbreviations: MRI, magnetic resonance imaging; PSMA, prostate specific membrane antigen; SPIONs, superparamagnetic iron oxide nanoparticles; T₁w, T₁-weighted; T₂w, T₂-weighted; px, pixels.

Previous attempts^{13–19} at iron-specific imaging have relied on the alteration in T₂ or T₂* by iron. These methods do not produce images, as we do, where the intensity is solely, and unambiguously, dependent on [Fe] as we propose here. They rely on the assumption that the [Fe] is the sole determinant of either T₂ or T₂*, and ignore the many other contributions to transverse relaxation. Other methods, such as QSM,^{47–49} require measurement of the absolute tissue susceptibility and suffer from the large background variations in the phase of the MR image due to susceptibility variations independent of [Fe] or from susceptibility jumps at the air-tissue boundaries of the body. Also, the expertise and software to accomplish QSM are not widely available. We have developed a method that is easy to apply, using only T₁w and T₂w images available on any MRI system. This ease of implementation should result in the widespread application of [Fe]MRI to a large class of biological, and physical–chemical problems. Note that [Fe]MRI is primarily a method for the quantitative imaging of added Fe arising, for example, from the injection of magnetic agents, and is not a method for the determination of total

tissue iron, because the background MRI signals are removed from the analysis to produce the basic contrast images.

The sensitivity of [Fe]MRI compares favorably with other MRI methods. Our current LOD was 2 μ M Fe. Dahnke and Schaeffter¹³ report an LOD for SPIONs, using a relatively complicated scheme for Δ Bo corrections to T₂* mapping, of 43 μ M in the brain and 144 μ M in the liver. At the extreme upper range of potential physiological concentrations that might be used for tumor hyperthermia, Garwood's laboratory used SWIFT to map [Fe] up to ~50 mM.⁵⁰ Girard et al¹⁹ using QSM report slopes of 1.760 ppm/mM Fe in vitro, so that our LOD corresponds to a susceptibility shift of 0.45 ppm at 3 T, which is easily measurable. He et al¹¹ measured [Fe] in the brain using QSM but the determined slope of the relationship between Δ X and [Fe] is so small in vivo (~16 ppm/mol Fe) that [Fe] smaller than ~1 mM results in Δ X values too small to measure. Our LOD is 500 times lower than this. Gunn et al,¹² used proton NMR spectroscopy in vitro to measure [Fe] with a LOD of ~0.2 nM, but this in vitro method is destructive and cannot be used in vivo. The current US Food

and Drug Administration upper limit for Fe administration to humans is 5 mg/kg, which corresponds to an [Fe] of 90 μ M, a value 45-fold higher than our determined LOD.

Given the ability to produce quantitative maps of the distribution of nanoscale magnetic objects (superparamagnetic nanoparticles, soluble paramagnetic agents like gadolinium, manganese, cobalt, etc) one can anticipate extensions of the work presented here. Clearly, it is possible to measure, monitor, and follow the time course of injected magnetic materials in the body. For example, in the work to be reported separately, we have used [Fe]MRI to image the [Fe] as a function of time for SPIONs injected into prostate tumor xenografts in mice, and to follow the time course of anti-PSMA-conjugated SPIONs as they infiltrate and bind to prostate tumor xenografts after intravenous injection. We have previously published MRI studies in which chemotherapeutic drugs were incorporated into superparamagnetic FePt nanomicelles for the simultaneous imaging³³ and the treatment of prostate tumor xenografts in mice.² In the present situation, by knowing the relationship between the amount of drug, and the amount of iron, in the nanoparticles, it becomes feasible to envision the [Fe]MRI signal as a surrogate for the [drug] in the tissue of interest to measure drug dosage to the tumor, non-invasively. One area in which this information would be of critical importance is for the chemotherapy of brain tumors where it is not accurately known if the drug actually reaches the tumor in therapeutic concentrations.⁵¹ Such information is currently only available with positron emission tomography imaging of expensive, short-lived, radioactively-labeled molecules. Our [Fe]MRI technology will open this field to a large variety of additional studies that were previously thought to be too expensive, or impractical.

Another of the many potential applications of [Fe]MRI would be to measure the infiltration of tissues by macrophages. For example, diabetic osteomyelitis is accompanied by a massive influx of these cells as the body attempts to clear the infection. Macrophages take up iron-containing nanoparticles, such as ferumoxytol, and serve as carriers of MRI contrast into the tissue. By means of [Fe]MRI, one could potentially infer the macrophage density, and hence the severity of infection, through [Fe]MRI measurements of the [Fe] in the affected tissues.^{52,53} We are currently pursuing a human clinical trial of these concepts.

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Disclosure

The author reports no conflicts of interest in this work.

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