Modulation of neurotrophic signaling pathways by polyphenols

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Abstract: Polyphenols are an important class of phytochemicals, and several lines of evidence have demonstrated their beneficial effects in the context of a number of pathologies including neurodegenerative disorders such as Alzheimer’s and Parkinson’s disease. In this report, we review the studies on the effects of polyphenols on neuronal survival, growth, proliferation and differentiation, and the signaling pathways involved in these neurotrophic actions. Several polyphenols including flavonoids such as baicalein, daidzein, luteolin, and nobiletin as well as nonflavonoid polyphenols such as auraptene, carnosic acid, curcuminoinds, and hydroxycinnamic acid derivatives including caffeic acid phenyl ester enhance neuronal survival and promote neurite outgrowth in vitro, a hallmark of neuronal differentiation. Assessment of underlying mechanisms, especially in PC12 neuronal-like cells, reveals that direct agonistic effect on tropomyosin receptor kinase (Trk) receptors, the main receptors of neurotrophic factors including nerve growth factor (NGF) and brain-derived neurotrophic factor (BDNF) explains the action of few polyphenols such as 7,8-dihydroxyflavone. However, several other polyphenolic compounds activate extracellular signal-regulated kinase (ERK) and phosphoinositide 3-kinase (PI3K)/Akt pathways. Increased expression of neurotrophic factors in vitro and in vivo is the mechanism of neurotrophic action of flavonoids such as scutellarin, daidzein, genistein, and fisetin, while compounds like apigenin and ferulic acid increase cyclic adenosine monophosphate response element-binding protein (CREB) phosphorylation. Finally, the antioxidant activity of polyphenols reflected in the activation of Nrf2 pathway and the consequent upregulation of detoxification enzymes such as heme oxygenase-1 as well as the contribution of these effects to the neurotrophic activity have also been discussed. In conclusion, a better understanding of the neurotrophic effects of polyphenols and the concomitant modulations of signaling pathways is useful for designing more effective agents for management of neurodegenerative diseases.

Keywords: flavonoids, hydroxycinnamic acids, neuroprotective, neurodegeneration, Trk

Introduction
Polyphenolic compounds

Polyphenols are an important group of phytochemicals that are abundantly present in food sources.1 Several lines of evidence have demonstrated beneficial effects of these compounds in the context of several pathologies,2–8 and it has been repeatedly shown that consumption of foods rich in phenolic compounds can lower the risk of several diseases.9,10

Polyphenols have been reported to be of therapeutic value in neurodegenerative diseases,11–13 hypertension14 and other cardiovascular diseases,15 cancer,16–18 inflammation,19,20 diabetes,21 dyslipidemia,22–24 allergy, and immune system diseases.25,26 They have also been reported to have a role in the prevention of different types of cancer ranging from liver,1 prostate,2 and colorectal28 cancer to lymphoblastic leukemia.29
Neurodegenerative disorders such as Alzheimer’s, Parkinson’s, and Huntington’s disease represent rapidly growing causes of disability and death, which have profound economic and social implications; nonetheless, only few effective disease-modifying therapies are available for these diseases.30–32 Recent research has shown that certain polyphenols may have considerable neuroprotective effects in different brain pathologies including neurodegenerative diseases.8,11,33,34 Polyphenols from grape juice administered to older adult subjects have been shown to ameliorate the mild cognitive impairment.35 Similarly, Witte et al36 have shown that administration of resveratrol, a polyphenol present in wine, in older adults significantly improves memory performance, indicating potential strategies to maintain brain health during aging. Assessment of cognitive performance in middle-aged individuals has indicated that consumption of different polyphenols such as catechins, flavonols, and hydroxybenzoic acids is strongly associated with language and verbal memory.37 Other studies have also shown similar effects of other polyphenols on cognitive function and memory in older individuals, who are at risk for neurodegenerative diseases.38,39 In another human study, epigallocatechin–gallate (EGCG) significantly improved cognitive deficits in individuals with Down syndrome.40 Several studies have demonstrated that different polyphenolics with neuroprotective activity, such as EGCG,41,42 epicatechin,43 curcumin,44 resveratrol,45 quercetin,46 and citrus flavonoids (naringenin and hesperetin),47 are able to cross the blood–brain barrier and therefore indicate that these compounds localize within the brain tissue and may well exert neuroprotective and neuromodulatory actions in the settings of different brain pathologies.

Polyphenolic compounds can be classified into different groups as described in Figure 1.1 Among them, flavonoids constitute a major subgroup, which are highly present in fruits and vegetables and possess several biological activities.1

Polyphenols have been extensively studied for their antioxidant action48–52 and some of their beneficial effects, including neuroprotective activity, have been attributed to this capacity.53–55 The neuroprotective effect of polyphenols against the diseases of nervous system has also been addressed in a number of studies.56–59 The aim of this review, however, is to extensively address the alterations of signaling pathways

![Figure 1](https://www.dovepress.com/)

**Figure 1** Main classes of polyphenols and their chemical structures.
involved in the neurotrophic action of polyphenols that lead to neuronal survival, growth, proliferation, and differentiation. The other aspects of neuroprotective activity of these compounds, which are mainly ascribed to the antioxidant action and mitigation of oxidative stress, have been reviewed elsewhere.\textsuperscript{5,7,8,60–62} and are only briefly mentioned here. It should also be mentioned that in in vitro studies, which are the main focus of this report, supraphysiological doses of polyphenols may have been used and, therefore, caution should be exerted in extrapolation of these data to in vivo conditions.\textsuperscript{56}

The structures of some of the most highly studied neurotrophic polyphenols are depicted in Figures 2 and 3.

**Neurotrophic effects of polyphenols: Enhancement of survival in neuronal cells**

Survival signaling is important to suppress the cell death machinery and counterbalance apoptotic signaling in the nervous system.\textsuperscript{63} Several studies have shown that polyphenolic compounds enhance neuronal survival in serum-deprived conditions. Lin et al.\textsuperscript{64} showed that in neuronal-like PC12 cells, luteolin (3′,4′,5,7-tetrahydroxyflavone, Figure 2) attenuates serum withdrawal-induced cytotoxicity. Other examples of neuroprotectants that exerted prosurvival action in PC12 cells include curcuminoids, the predominant polyphenolic compounds of *Curcuma longa* Linn.;\textsuperscript{65} EGCG (Figure 2), the major polyphenol of green tea extract;\textsuperscript{66} caffeic acid phenethyl ester (CAPE, Figure 3), an active component of propolis;\textsuperscript{67} 5-hydroxy-3,6,7,8,3′,4′-hexamethoxyflavone (HHMF) from the *Citrus* genus; and nobiletin (Figure 2), the most abundant polymethoxyflavone in orange peel extract.\textsuperscript{68} Furthermore, other investigators have shown that treatment with genistein and daidzein (Figure 2), isoflavones present in soybeans and soy products, enhances proliferation and survival of the hippocampal neuronal cells.\textsuperscript{69}

![Figure 2 Chemical structures of flavonoids with neurotrophic activity.](image-url)
In addition, application of 7,8-dihydroxyflavone (7,8-DHF, Figure 2), a flavone, promotes survival of cultured motoneurons, spiral ganglion neurons (SGNs), and hippocampal neuronal cells. In another study, the protective ability of quercetin (3,3′,4′,5,7-pentahydroxyflavone, Figure 2) on P19-derived neurons was determined. Ferulic acid (4-hydroxy-3-methoxycinnamic acid, Figure 3) was able to significantly increase the survival of neural stem/progenitor cells (NSC/NPCs) cultured from rat embryo, and also increased the number and size of secondary formed neurospheres.

Promotion of neurite outgrowth in neuronal cells

Neurite outgrowth is a crucial step in the differentiation of neurons, which begins at the cell body and extends outward to form functional synapses. In response to extracellular signals, the growth of neurite processes starts, involving the addition of new plasma membranes, generation of new cytoplasm, and the continued expansion and modification of the cytoskeleton. A number of neurotypic proteins have been associated with neurite outgrowth, including growth-associated protein 43 (GAP-43), microtubule-associated protein (MAP) and tau, as well as presynaptic membrane-associated proteins, such as synaptophysin and synapsin. Many polyphenolic compounds from natural and synthetic sources have been demonstrated to induce neurite outgrowth in various primary neuronal cultures and neuronal cell lines (Table 1).

Several studies have shown that different polyphenols including flavonoids such as genistein, quercetin, liquiritin from Glycyrrhiza radix plant (Figure 2), isorhamnetin (a flavonolglycone from Ginkgo biloba plant, Figure 2), and acetylated flavonoid glycosides from Scoparia dulcis as well as the stilbenoid compound resveratrol (a polyphenol present in grapes and red wine, Figure 3) cause a significant enhancement of neurotrophin (nerve growth factor [NGF] and brain-derived neurotrophic factor [BDNF])-mediated neurite outgrowth in PC12 cells.

Some of the phenolic compounds are capable of enhancing the expression levels of the differentiation markers

Figure 3 Chemical structures of polyphenols with neurotrophic activity that do not belong to the group of flavonoids.
### Table 1: Induction of neurite outgrowth by polyphenols and involvement of neurotrophic signaling pathways

<table>
<thead>
<tr>
<th>Compound</th>
<th>Source</th>
<th>Cell model</th>
<th>Involved signaling pathways</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Artepillin C</td>
<td>Chemical</td>
<td>PC12 cells</td>
<td>Y</td>
<td>145</td>
</tr>
<tr>
<td>CAPE</td>
<td>Chemical</td>
<td>PC12 cells</td>
<td>–</td>
<td>67</td>
</tr>
<tr>
<td>Daidzein</td>
<td>Chemical</td>
<td>DRG cultures</td>
<td>Y</td>
<td>69</td>
</tr>
<tr>
<td>7,8-DHF</td>
<td>Chemical</td>
<td>Cultured embryonic mouse motoneurons and SGNs</td>
<td>Y/104/N/70</td>
<td>70,104</td>
</tr>
<tr>
<td>EGCG</td>
<td>Chemical</td>
<td>PC12 cells</td>
<td>–</td>
<td>66</td>
</tr>
<tr>
<td>Fisetin</td>
<td>Chemical</td>
<td>PC12 cells</td>
<td>Y</td>
<td>146</td>
</tr>
<tr>
<td>Isoquercitrin</td>
<td>Chemical</td>
<td>NG108-15 neuroblastoma/glioma cells</td>
<td>–</td>
<td>193</td>
</tr>
<tr>
<td>Luteolin</td>
<td>Chemical</td>
<td>PC12 cells</td>
<td>Y</td>
<td>64,144</td>
</tr>
<tr>
<td>Methyl 3,4-dihydroxybenzoate</td>
<td>Chemical</td>
<td>Cortical neurons of neonatal rats</td>
<td>–</td>
<td>147</td>
</tr>
<tr>
<td>Resveratrol</td>
<td>Chemical</td>
<td>PC12 cells, neuro2a cells, and primary rat midbrain neuron-glia and neuron-astroglia cultures</td>
<td>–</td>
<td>79,194,195</td>
</tr>
<tr>
<td>7,8,3′-Trihydroxyflavone</td>
<td>Chemical</td>
<td>SGN</td>
<td>Y</td>
<td>104</td>
</tr>
<tr>
<td>Auraptene</td>
<td>Citrus plants</td>
<td>PC12 cells</td>
<td>Y</td>
<td>151</td>
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<tr>
<td>Baicalein</td>
<td>Scutellaria baicalensis</td>
<td>C17.2 NSCs</td>
<td>Y</td>
<td>196</td>
</tr>
<tr>
<td>Carnosic acid</td>
<td>Rosmarinus officinalis</td>
<td>PC12 cells</td>
<td>Y</td>
<td>184</td>
</tr>
<tr>
<td>Curcuminoids</td>
<td>Curcuma longa</td>
<td>PC12 cells</td>
<td>Y</td>
<td>65</td>
</tr>
<tr>
<td>3,7-Dihydroxy-2,4,6-trimethoxy-phenanthrene</td>
<td>Dioscorea nipponica</td>
<td>Mouse neuro2a (N2a) cells</td>
<td>–</td>
<td>197</td>
</tr>
<tr>
<td>Nobleitine</td>
<td>Citrus depressa</td>
<td>PC12D cells</td>
<td>Y</td>
<td>152</td>
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<tr>
<td>Oroxylin</td>
<td>Scutellaria baicalensis</td>
<td>Cultured rat primary neuron</td>
<td>Y</td>
<td>114</td>
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<tr>
<td>Protocatechuic acid</td>
<td>Alpinia oxyphylla</td>
<td>Cultured NSC/NPCs</td>
<td>–</td>
<td>198</td>
</tr>
<tr>
<td>Quercetin</td>
<td>Coesalpinia mimosoides</td>
<td>P19-derived neurons</td>
<td>–</td>
<td>71,199</td>
</tr>
</tbody>
</table>

Note: *Not determined.

Abbreviations: N, activation of the pathway is not involved; Y, activation of the pathway is involved; CAPE, caffeic acid phenethyl ester; 7,8-DHF, 7,8-dihydroxyflavone; DRG, dorsal root ganglia; EGCG, epigallocatechin–gallate; ERK, extracellular signal-regulated kinase; MEK, mitogen-activated protein kinase kinase; NSC/NPCs, neural stem/progenitor cells; PI3K, phosphoinositide 3-kinase; PKC, protein kinase C; PLCγ, phospholipase Cγ; SGNs, spiral ganglion neurons.
NGF has a higher binding rate with TrkA, while BDNF and NT-3 bind with TrkB, and finally, NT-3 is the main ligand for TrkC receptor, with a lower affinity for TrkA and TrkB.98

The neurotrophins-induced dimerization of the Trk receptors leads to activation through transphosphorylation of the cytoplasmic domain kinases and stimulates three major signaling pathways: phosphatidylinositol-3-kinase (PI3K)/Akt, mitogen-activated protein kinase (MAPK), and phospholipase C-γ1.92 Downstream signaling principally promotes survival, growth, and neuronal differentiation and mediates neurogenesis and plasticity in many neuronal populations.99–101

There are only few polyphenolic compounds that act as direct agonists of Trk receptors and mimic the binding of neurotrophins, while several others stimulate the more downstream pathways leading to the neurotrophic effect (discussed in the following sections). 7,8-DHF provokes TrkB dimerization and tyrosine phosphorylation and activates downstream Akt and extracellular signal-regulated kinase (ERK) as potently as BDNF. 7,8-DHF also inhibits neuronal death in T48, a stably transfected TrkB murine cell line, and hippocampal neurons, and its activity can be inhibited by K252a, a Trk receptor antagonist.102 In another study, 7,8-DHF strongly activated TrkB receptor and its downstream Akt and ERK1/2 pathways, prevented cell death, and promoted neuritogenesis in the retinal ganglion cells.103 It was also shown in a later report that 7,8-DHF as a BDNF agonist causes phosphorylation of TrkB receptors and stimulates survival and neurite growth of cultured motoneurons: PI3K/Akt but not MAPK pathway was responsible for the survival and growth promoting effects of 7,8-DHF,70 which is apparently different from MAPK activation mediated by 7,8-DHF in hippocampal neurons.102

Furthermore, 7,8,3′-trihydroxyflavone, a compound related to 7,8-DHF, has shown similar effects on the survival of SGNs, phosphorylation of TrkB receptor, and ERK.104 Diosmetin, another polyphenol compound belonging to flavonoids class of polyphenols, has also shown antipapoptotic effects, but it induces only weak phosphorylation of TrkB. Diosmetin also increased the phosphorylation of Akt and ERK.102 Another flavonoid, epicatechin, restores TrkA phosphorylation in diabetic animals and reduces diabetes-induced neuronal cell death. It also blocks diabetes-induced p75NTR expression and p75NTR apoptotic pathway in vivo and in Müller cells.105

**Modulation of expression of neurotrophic factors and Trk receptors**

Several polyphenols increase the expression levels of neurotrophins and Trk receptors (Table 2). Scutellarin, isolated
**Table 2** Increased expression of neurotrophic factors by polyphenols in cell and animal models

<table>
<thead>
<tr>
<th>Compound</th>
<th>Cell model</th>
<th>Animal model</th>
<th>Involved signaling pathways</th>
<th>Neurotrophic factors that are modulated</th>
<th>Function</th>
<th>References</th>
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<tr>
<td>Alpinetin</td>
<td>Primary rat astrocytes</td>
<td>–</td>
<td>Estrogen signaling</td>
<td>BDNF, NGF, GDNF</td>
<td>–</td>
<td>107</td>
</tr>
<tr>
<td>Astilbin</td>
<td>–</td>
<td>Mice</td>
<td>p-ERK/p-Akt</td>
<td>BDNF</td>
<td>Antidepressant-like effects</td>
<td>128</td>
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<tr>
<td>Baicalin</td>
<td>–</td>
<td>Mouse model of amnesia</td>
<td>p-ERK/p-CREB</td>
<td>BDNF</td>
<td>Antidepressant-like effects</td>
<td>109,129</td>
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<tr>
<td>Butein</td>
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<td>p-ERK/p-CREB</td>
<td>BDNF</td>
<td>Cognition enhancement</td>
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<tr>
<td>CAPE</td>
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<td>Nrf2/ARE</td>
<td>BDNF</td>
<td>Protection of nigral dopaminergic neurons</td>
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<td>Calycosin</td>
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<td>Estrogen signaling</td>
<td>BDNF, NGF, GDNF</td>
<td>–</td>
<td>107</td>
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<tr>
<td>Chrysin</td>
<td>–</td>
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<td>–</td>
<td>BDNF</td>
<td>Cognition enhancement</td>
<td>111</td>
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<tr>
<td>Curcumin</td>
<td>–</td>
<td>Rats</td>
<td>Akt/GSK-3β</td>
<td>BDNF</td>
<td>Suppression of β-amyloid-induced cognitive impairments</td>
<td>118</td>
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<td>Daidzein</td>
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<td>–</td>
<td>–</td>
<td>BDNF</td>
<td>–</td>
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<tr>
<td>Ferulic acid</td>
<td>NSC/NPCs</td>
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<td>p-CREB</td>
<td>BDNF</td>
<td>Antidepressant-like effects</td>
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<tr>
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<td>Mouse model of amnesia</td>
<td>p-ERK/p-CREB</td>
<td>BDNF</td>
<td>Cognition enhancement</td>
<td>110</td>
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<tr>
<td>Genistein</td>
<td>Primary rat astrocytes</td>
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<td>BDNF, NGF, GDNF</td>
<td>–</td>
<td>107</td>
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<tr>
<td>Genistein</td>
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<td>BDNF</td>
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<tr>
<td>HMF</td>
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<td>Luteolin</td>
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<td>Puerarin</td>
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<td>Nrf2/ARE</td>
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<td>Resveratrol</td>
<td>Dopaminergic neurons</td>
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<td>p-ERK/p-CREB</td>
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<td>p-CREB/p-Akt</td>
<td>BDNF, NGF, GDNF</td>
<td>–</td>
<td>106</td>
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<tr>
<td>Xiaochaihutang</td>
<td>–</td>
<td>Rats</td>
<td>–</td>
<td>BDNF, NGF</td>
<td>Antidepressant-like effects</td>
<td>127</td>
</tr>
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</table>

*Note:* Not determined in this study.

**Abbreviations:** CAPE, caffeic acid phenethyl ester; HMF, 3,5,6,7,8,3',4'-heptamethoxyflavone; BDNF, brain-derived neurotrophic factor; NGF, nerve growth factor; GDNF, glial cell line-derived neurotrophic factor; NSC/NPCs, neural stem/progenitor cells; ERK, extracellular signal-regulated kinase.
from the traditional Chinese herb, *Erigeron breviscapus* (Figure 2), alpinetin of the genus *Alpinia,* and also luteolin, calycosin, genistein, and isorhamnetin effectively upregulate the synthesis and release of NGF, glial cell line-derived neurotrophic factor (GDNF), and BDNF. The capacity of luteolin, genistein, calycosin, and isorhamnetin in increasing the expression of neurotrophins was mediated through estrogen signaling pathways. In the estrogen receptor (ER)-dependent pathway, the phosphorylated ER dimer was demonstrated to elevate the BDNF messenger RNA levels. Scutellarin induced the expression of neurotrophins’ messenger RNAs and proteins through cyclic adenosine monophosphate response element-binding protein (P-CREB) and p-Akt signaling in primary rat astrocytes.

Several polyphenolic compounds have been reported to increase BDNF levels; different flavonoids such as baicalein (5,6,7-trihydroxyflavone, Figure 2), rutin and fisetin of *Rhus verniciflua* (Figure 2),110 chrysin (5,7-dihydroxyflavone, Figure 2),111 daidzein and genistin,112 3,5,6,7,8,3′,4′-heptamethoxyflavone (HMF),113 oroxylin A (5,7-dihidridoxy-6-methoxyflavone, Figure 2),114 puerarin,115 and quercetin,116 as well as other polyphenols such as CAPE,117 curcumin,118 and resveratrol,19,119-122 have been reported to possess this property.

The aforementioned studies have examined BDNF levels under various experimental conditions. Some of the polyphenolic compounds, including fisetin and baicalin, one of the major flavonoids isolated from the roots of *Scutellaria baicalensis* Georgi, could activate the ERK-CREB pathway and exhibited memory-enhancing effects through the activation of CREB-BDNF pathway in the hippocampus of a mouse model of amnesia induced by scopolamine.109,110 In another study, chrysin was shown to rescue memory impairment and BDNF reduction caused by aging in mice.111

Other studies have demonstrated the increased BDNF expression and neuroprotective effects of polyphenolic compounds in the settings of in vitro and in vivo models of ischaemia-, 6-hydroxydopamine (6-OHDA)-, or amyloid-β (Aβ)-mediated injury.113,115,118,123,124 For example, HMF enhances BDNF production in astrocytes and induces neurogenesis in the hippocampus after brain ischemia, which is mediated by the activation of ERK1/2 and CREB pathways.119 Puerarin has also a protective effect in 6-OHDA-lesioned rats by modulating BDNF expression and activation of the nuclear factor E2-related factor 2/antioxidant response element (Nrf2/ARE) signaling pathway.115 Curcumin, on the other hand, was able to block BDNF reduction in the Aβ-infused rats through Akt/GSK-3β signaling pathway.118

Antiamnesic and protective effects of luteolin against Aβ toxicity in mice was associated with the increase of BDNF as well as TrkB expression in the cerebral cortex of mice.123

Many polyphenolic compounds have antidepressant effects that are associated with increased BDNF levels.125,126 *Xiaochaihutang,* a mixture of seven Chinese herbs that is traditionally used in People’s Republic of China for treatment of depressive-like symptoms, considerably increased BDNF, NGF, TrkB, and TrkA expressions in the hippocampus and also improved depression-like behaviors in chronic unpredictable mild stress (CUMS) model in rats.127 A flavonanol compound, astilbin, reverses depressive-like behaviors in a mouse model of depression and its effect is mediated by upregulation of BDNF and activation of ERK and Akt pathways.128 Antidepressant effects of baicalein, resveratrol, and rosmarinic acid (a caffeic acid ester) are also mediated by BDNF-ERK-mediated neurotrophic action.129-131 Another polyphenol, ferulic acid, not only increases the proliferation of NSC/NPCs in vitro and in vivo but also produces an additive antidepressant-like effect in corticosterone-treated mice, mediated by CREB-BDNF signaling pathway.72 Also, chrysin improves age-related memory decline in mice, an effect that is probably related to the modulation of BDNF production and free radical scavenging action of this compound.111

**ERK pathway activation**

Modulation of neuronal survival signaling pathways may represent a promising approach to the management of central nervous system diseases. Several pathways including ERK1/2 and PI3K promote cell survival in the nervous system as well as other tissues.132,133 Recently, the ERK pathway, a part of MAPKs, has been implicated in several physiological functions of neurons, including proliferation, differentiation, survival, and regulation of response to various growth factors.134-138 The activation of ERK1/2 requires phosphorylation of threonine and tyrosine residues that is carried out by the upstream activator kinase, mitogen-activated protein kinase kinase (MEK). Activated ERK1/2 then changes its localization and phosphorylates different target molecules, including transcription regulators and cytoskeletal proteins (Figure 4).139,140

ERK1/2 activation by some of the polyphenolic compounds promotes survival and antiapoptotic signaling in various cell lines (Table 3). It has been shown that ERK1/2 activation by luteolin protects PC12 cells against apoptosis. Furthermore, the cell viability of PC12 cells that were pretreated with U0126, a specific inhibitor of ERK1/2 kinase, was significantly reduced.64
Activation of the ERK1/2 pathway has mediated the neuroprotective activity against damaging insults.\textsuperscript{132} Aβ peptide can cause a significant decrease in cell viability and neurite outgrowth and can induce apoptosis in neuronal cells. Liquiritin’s protection against Aβ-induced neuronal apoptosis and its effect on the differentiation of rats’ primary cultured hippocampal neurons were inhibited with a MAPK inhibitor.\textsuperscript{141} Similarly, in another study, the decreased cell viability induced by Aβ\textsubscript{25–35} was reported to be blocked by icaritin (a prenyl flavonoid derivative from Chinese tonic herb Epimedium). A blocker of ERK/MAPK pathway weakened this protective effect, which implied that ERK1/2 pathway is involved in the neuroprotective action of icaritin.\textsuperscript{142} Likewise, rutin (3,3’4,5,7-pentahydroxyflavone-3-rhamnoglucoside) has also shown beneficial effects against Aβ-induced neurotoxicity in rats through the activation of MAPK and BDNF.\textsuperscript{143} As already mentioned, ERK1/2 signaling is crucial for neuronal differentiation and activation of associated cytoskeletal and synaptic proteins. It has been found that luteolin increases neurite outgrowth and expression of GAP-43 protein, a neuronal differentiation biomarker, and also heme oxygenase-1 (HO-1) expression in PC12 cells. All these effects could be blocked by pharmacological inhibition of ERK1/2.\textsuperscript{64} In another report, it was established that luteolin induced microRNA-132 expression in PC12 cells and induced neurite outgrowth, while these effects were suppressed by protein kinase A (PKA) and MEK1/2 inhibitors, but not by protein kinase C (PKC) inhibitors.\textsuperscript{144}
Lin et al\(^4\) have shown the involvement of both ERK and PKC signaling pathways in PC12 neurite outgrowth. Similarly, the involvement of the ERK and PKC signaling pathways in the process of neurogenesis in PC12 cells was demonstrated in response to curcuminoids.\(^5\) In another study, artepillin C-induced neurite outgrowth of PC12m3 cells has been shown to be inhibited by the ERK and p38 MAPK inhibitors. On the other hand, inhibition of ERK by U0126 entirely blocked artepillin C-mediated p38 MAPK phosphorylation of PC12m3 cells. It was suggested that the activation of p38 MAPK through the ERK signaling pathway is responsible for the artepillin C-induced neurite outgrowth of PC12m3 cells.\(^14\) Finally, fisetin has been very effective in induction of PC12 cell differentiation, while MEK inhibitors considerably decreased fisetin-induced ERK activation and neurite outgrowth.\(^15\) The different neurotrophic effects of polyphenols involving the ERK pathway are listed in Tables 1, 3–5.

### PI3K pathway activation

A number of studies have demonstrated that PI3K and its downstream effector Akt are involved in neuronal survival and increased neurite outgrowth as well as other neurotrophic effects of polyphenols.\(^82,106,118,128,147\) (Tables 1, 3–5). In this regard, Hoppe et al\(^118\) showed that PI3K/Akt pathway is involved in curcumin-mediated neuroprotection in Aβ-induced cognitive impairment in rats. Similarly, scutellarin is reported to protect neurons against hypoxia by increasing neurotrophins through pCREB and pAkt signaling, but not MAPKs.\(^106\) In cultured primary cortical neurons, methyl 3,4-dihydroxybenzoate, a phenolic acid derivative, promoted neuronal survival and neurite outgrowth via PI3K/Akt signaling pathway, and these effects could be inhibited by a PI3K-specific inhibitor.\(^147\) In another report, evidence was provided that oroxylin A increased BDNF production and neuronal differentiation in primary cortical neurons by activation of the Akt pathway. Puerarin also protected dopaminergic cells and potentiated the effect of NGF on neuritogenesis in PC12 cells via activation of the ERK1/2 and PI3K/Akt pathways.\(^82\) Finally, astilbin, a natural flavonoid, has been demonstrated to reduce depressive-like behaviors in mice models of depression by activation of the MAPK/ERK and PI3K/Akt pathways that are the downstream signaling pathways of BDNF.\(^128\)

### Table 3 Neuronal survival and differentiation induced by polyphenols and contribution of different neurotrophic pathways to these effects

<table>
<thead>
<tr>
<th>Compound</th>
<th>Cell model</th>
<th>Activation of pathway</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>MEK/ERK1/2</td>
<td>PI3K/Akt</td>
</tr>
<tr>
<td>Artepillin C</td>
<td>PC12m3 cells</td>
<td>Y</td>
<td></td>
</tr>
<tr>
<td>Auraptene</td>
<td>PC12 cells</td>
<td>Y</td>
<td></td>
</tr>
<tr>
<td>Baicalin</td>
<td>C17.2 NSCs</td>
<td>Y</td>
<td></td>
</tr>
<tr>
<td>Caffeic acid phenethyl ester</td>
<td>Rat organotypic midbrain slice cultures</td>
<td>Y</td>
<td></td>
</tr>
<tr>
<td>Carnosic acid</td>
<td>PC12 cells</td>
<td>Y</td>
<td></td>
</tr>
<tr>
<td>Carnosic acid</td>
<td>PC12 cells</td>
<td>Y</td>
<td></td>
</tr>
<tr>
<td>Curcuminoids</td>
<td>PC12 cells</td>
<td>Y</td>
<td></td>
</tr>
<tr>
<td>Daidzein</td>
<td>Primary rat DRG neuronal cultures</td>
<td>Y</td>
<td></td>
</tr>
<tr>
<td>7,8-Dihydroxyflavone</td>
<td>Mouse hippocampal cultures and mouse embryonic motoneuron cultures</td>
<td>N</td>
<td>Y</td>
</tr>
<tr>
<td>7,8-Dihydroxyflavone</td>
<td>Retinal ganglion cell cultures</td>
<td>Y</td>
<td>Y</td>
</tr>
<tr>
<td>4′-O-[β-D-Glucopyranosyl-3′, 4-dimethoxychalcone</td>
<td>PC12 Cells</td>
<td>Y</td>
<td>N</td>
</tr>
<tr>
<td>HMF</td>
<td>Mouse neuronal cells</td>
<td>Y</td>
<td></td>
</tr>
<tr>
<td>Hesperetin</td>
<td>PC12 cells</td>
<td>Y</td>
<td>Y</td>
</tr>
<tr>
<td>Luteolin</td>
<td>PC12 cells</td>
<td>Y</td>
<td></td>
</tr>
<tr>
<td>Luteolin</td>
<td>PC12 cells</td>
<td>Y</td>
<td></td>
</tr>
<tr>
<td>Methyl 3,4-dihydroxybenzoate</td>
<td>PC12D cells</td>
<td>–</td>
<td>Y</td>
</tr>
<tr>
<td>Nobiletin</td>
<td>PC12D cells</td>
<td>Y</td>
<td></td>
</tr>
<tr>
<td>Oxyroxylin A</td>
<td>Cortical neuronal cultures</td>
<td>–</td>
<td>Y</td>
</tr>
<tr>
<td>Puerarin</td>
<td>PC12 cells</td>
<td>Y</td>
<td>Y</td>
</tr>
<tr>
<td>Resveratrol</td>
<td>Astrocytes cultures</td>
<td>Y</td>
<td></td>
</tr>
<tr>
<td>Rosmarinic acid</td>
<td>PC12 cells</td>
<td>Y</td>
<td></td>
</tr>
</tbody>
</table>

**Note:** Not determined.

**Abbreviations:** N, activation of the pathway is not involved; Y, activation of the pathway is involved; DRG, dorsal root ganglion; ERK, extracellular signal-regulated kinase; HMF, 3,5,6,7,8,3′,4′-heptamethoxyflavone; MEK, mitogen-activated protein kinase kinase; NSCs, neural stem cells; PKC, protein kinase C; PLCγ, phospholipase Cγ.
Phosphorylation of CREB

Cyclic AMP response element (CRE) sequence is found in the regulatory region of several genes. CREB is a transcription regulator that recognizes CRE sequence and activates gene transcription, a process believed to play an important role in learning and memory in the brain. In neuronal cells, activation of the CREB pathway by neurotrophic factors such as NGF leads to the expression of genes that regulate survival, growth, synaptic plasticity, differentiation, dendritic spine formation, and long-term memory.148,149

Mantamadiotis et al150 provided evidence that mice lacking CREB function in the brain showed neurodegenerative process in the hippocampus and dorsolateral striatum. Considering the importance of this pathway in the nervous system, we hereby summarize the studies on some phenolic compounds that activate CREB pathway and discuss their possible therapeutic effects in neurodegenerative diseases (Figure 4).

Luteolin,144 auraptene (a coumarin derivative, Figure 3),151 curcumin and demethoxycurcumin,65 nobiletin,155 hesperetin (a flavonoid),153 and citrus HHMF158 have all been shown to increase CREB phosphorylation in PC12 cells. Chai et al150 have also suggested that one of the signaling pathways related to neuroprotective effect of scutellarin is pCREB, which stimulates the production and release of neurotrophic factors in primary rat astrocyte cultures.

Resveratrol has shown neurotrophic effect in dopaminergic neurons79 and antidepressant-like effects in rats via activation of CREB in the hippocampus and amygdala.138 In a study by Li et al,154 green tea catechins-treated mice showed significantly higher CREB phosphorylation than the aged control mice. Ferulic acid has also been reported to increase CREB phosphorylation in the hippocampus of corticosterone-treated mice.72 In an animal model of traumatic brain injury, 7,8-DHF restores the levels of significantly reduced CREB phosphorylation.51 Baicalein increased pCREB in the hippocampus of mice109 and also in NPC.155 Administration of baicalein to rats with cognitive impairment induced by corticosterone significantly improved memory-associated decrease in the expression levels of BDNF and CREB proteins in the hippocampus of these animals.156 Activation of CREB by apigenin (4′,5,7-trihydroxyflavone, Figure 2)157 and pinocembrin (5,7-dihydroxyflavanone)158 was seen in transgenic Alzheimer’s disease mouse model. Finally, rutin and fustin flavonoids have been demonstrated to increase the expression of CREB in the hippocampi of rats and to attenuate Aβ-induced learning impairment in mice, respectively.143,159

Antioxidant activity

The CNS is unique in its high susceptibility to oxidative stress, which is a result of its high oxygen consumption.
Table 5 Inhibition of neurotoxin-induced damage by polyphenols and neurotrophic signaling pathways that possibly contribute to this effect

<table>
<thead>
<tr>
<th>Compound</th>
<th>Animal or cell model</th>
<th>Action of neurotoxin</th>
<th>Involved signaling pathway</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Baicalein</td>
<td>C17.2 NSCs^{155,196}</td>
<td>Irradiation-induced cell death and neurogenesis deficit^{155}</td>
<td>Y^{196}</td>
<td>155,196</td>
</tr>
<tr>
<td>Baicalein</td>
<td>SH-SYSY cells^{174}</td>
<td>6-OHDA-induced damage in SH-SYSY cells^{174}</td>
<td>Y^{196}</td>
<td>174,196</td>
</tr>
<tr>
<td>Baicalein</td>
<td>C17.2 NSCs^{196}</td>
<td></td>
<td>Y^{196}</td>
<td>196</td>
</tr>
<tr>
<td>Caffeic acid</td>
<td>SH-SYSY cells^{60}</td>
<td>6-OHDA-induced damage in SH-SYSY cells^{60}</td>
<td>N^{106} Y^{106}</td>
<td>60,106</td>
</tr>
<tr>
<td>Caffeic acid</td>
<td>Primary rat astrocytes culture^{106}</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CAPE</td>
<td>PC12 cells^{67}</td>
<td>Dopaminergic neurotoxin MPP+ induced injury in PC12 cells^{67}</td>
<td>Y^{117}</td>
<td>67,117</td>
</tr>
<tr>
<td>CAPE</td>
<td>Rat midbrain slice cultures^{117}</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Carnosic acid</td>
<td>SH-SYSY cells^{173}</td>
<td>Amyloid-β-induced apoptosis in SH-SYSY cells^{173}</td>
<td>Y^{200} Y^{173}</td>
<td>173,184,200</td>
</tr>
<tr>
<td>Carnosic acid</td>
<td>PC12 cells^{94,200}</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Curcumin</td>
<td>PC12 cells^{60,204}</td>
<td>Cisplatin-induced neurite toxicity^{204}</td>
<td>Y^{65}</td>
<td>65,204</td>
</tr>
<tr>
<td>Curcumin</td>
<td>Rats</td>
<td>Amyloid-β-induced neuronal injury</td>
<td>Y</td>
<td>118</td>
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<tr>
<td>7,8-DHF</td>
<td>Mice</td>
<td>Kainic acid-induced neurotoxicity</td>
<td>Y Y</td>
<td>102</td>
</tr>
<tr>
<td>7,8-DHF</td>
<td>HT-22 cells^{55}</td>
<td>Glutamate-induced toxicity in HT-22 cells^{55}</td>
<td>Y^{55} Y^{55}</td>
<td>55,103</td>
</tr>
<tr>
<td>EGC</td>
<td>Mice^{205}</td>
<td>Striatal and substantia nigra dopaminergic neuron loss induced by MPT^{206}</td>
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<td>205,206</td>
</tr>
<tr>
<td>EGCG</td>
<td>Cortical neuronal cells</td>
<td>Amyloid-β-induced neurotoxicity</td>
<td>Y</td>
<td>142</td>
</tr>
<tr>
<td>EGCG</td>
<td>Primary cultured rat hippocampal neurons</td>
<td>Corticosterone-induced apoptosis</td>
<td>Y Y</td>
<td>177</td>
</tr>
<tr>
<td>EGCG</td>
<td>Primary cultured hippocampal neurons</td>
<td>Amyloid-β-induced neurotoxicity</td>
<td>Y Y</td>
<td>141</td>
</tr>
<tr>
<td>EGCG</td>
<td>PC12 cells</td>
<td>Dopaminergic neuronal damage in PC12 cells and behavioral impairments in MPTP-induced neurotoxicity in mice</td>
<td>Y Y</td>
<td>82</td>
</tr>
<tr>
<td>EGCG</td>
<td>Mice</td>
<td>Ethanol-induced injury^{21}</td>
<td>Y^{119} Y</td>
<td>119,121</td>
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<tr>
<td>Resveratrol</td>
<td>Schwann cells^{121}</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Resveratrol</td>
<td>Primary astroglia cultures^{119}</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Rutin</td>
<td>Rat</td>
<td>Protective effects against neurotoxicity of Aβ</td>
<td>Y</td>
<td>143</td>
</tr>
</tbody>
</table>

Note: N, activation of the pathway is not involved; Y, activation of the pathway is involved; Aβ, Amyloid-β; CAPE, caffeic acid phenethyl ester; 7,8-DHF, 7,8-dihydroxyflavone; EGC, epigallocatechin–gallate; ERK, extracellular signal-regulated kinase; MEK, mitogen-activated protein kinase kinase; MPP+, (1-methyl-4-phenylpyridinium); MPTP, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; NSC, neural stem cell; 6-OHDA, 6-hydroxodopamine; P3K, Phosphoinositide 3-kinase; PKC, protein kinase C; PLCγ, phospholipase Cγ.

Abbreviations: NSC, neural stem cell; MEK, mitogen-activated protein kinase kinase; PI3K/Akt, phosphoinositide 3-kinase/Akt; PLCγ, phospholipase Cγ.
and also the presence of large amounts of fatty acids and metals.\textsuperscript{160} Several lines of evidence suggest that oxidative stress is involved in the pathogenesis of neurodegenerative disorders such as Alzheimer’s and Parkinson’s disease.\textsuperscript{61,161} The antioxidant effect of polyphenols has long been studied as a mechanism of their neuroprotection against neurodegenerative disorders\textsuperscript{61,162} and other neurological diseases.\textsuperscript{163} Although the direct interaction of polyphenols with reactive oxygen species (ROS) does not seem to be very likely to happen in vivo as a main mechanism of action, their antioxidant effects are most probably exerted through other mechanisms such as activation of Nrf2 pathway, upregulation of antioxidant enzymes, induction of hypoxia signal transduction (HIF-1-α pathway), and interaction with metal ions as sources of ROS.\textsuperscript{12,161,164}

For example, EGCG\textsuperscript{165} and resveratrol\textsuperscript{166} have been reported to exert their neuroprotective action through activation of the HIF-1 pathway. Mangiferin and morin (3,5,7,2′,4′-pentahydroxyflavone) revealed considerable antioxidant and antiapoptotic properties through the activation of antioxidant enzymes.\textsuperscript{167}

**Protection against damage induced by neurotoxins**

Cellular, biochemical, and animal studies have shown that Aβ is a crucial factor in the pathogenesis of Alzheimer’s disease.\textsuperscript{168,169} There are several reports suggesting that some polyphenols protect neuronal cells against Aβ-induced neuronal cell death or other forms of neuronal injury (Table 5). For instance, icariin was shown to protect primary rat cortical neuronal cells against apoptosis induced by Aβ\textsubscript{25-35} insult.\textsuperscript{142} Similarly, Ushikubo et al.\textsuperscript{170} demonstrated that 3,3′,4′,5′,5′-pentahydroxyflavone prevents Aβ fibril formation and that lowering fibril formation decreases Aβ-induced cell death in rat hippocampal neuronal cells. In another study, ursolic acid, p-coumaric acid, and gallic acid extracted from Cornifrustus plant were shown to attenuate apoptotic features such as morphological nuclear changes, DNA fragmentation, and cell blebbing induced by Aβ peptide in PC12 cells.\textsuperscript{171} The major flavonoids of cocoa, epicatechin and catechin, protect PC12 cells from Aβ-induced neurotoxicity.\textsuperscript{172} Carnosic acid (Figure 3), a highly bioactive phenolic compound found in rosemary (Rosmarinus officinalis), and liquiritin have shown a protective effect against Aβ in SH-SY5Y human neuroblastoma cells and primary cultured hippocampal neurons, respectively.\textsuperscript{141,173}

6-OHDA is a compound used to induce a Parkinson-like pathology in vitro and in vivo. It has been reported that baicalein\textsuperscript{174} and also ferulic acid and caffeic acid derivatives, both belonging to hydroxycinnamic acid family (Figure 1),\textsuperscript{66} protect neuronal SH-SY5Y cells against 6-OHDA.

Hydrogen peroxide is a ROS that has been shown to induce neuronal cell damage in experimental models.\textsuperscript{175} It has been shown that several polyphenols such as quercetin in cultured neuronal precursor cells,\textsuperscript{176} 7,8-DHF in retinal ganglion and RGC-5 cells,\textsuperscript{103} and caffeic acid esters in PC12 cells\textsuperscript{53} are able to exert protective effects against ROS. In addition, other authors suggest that the neuroprotective effects of 7,8-DHF are mediated by its ability to scavenge ROS and increase cellular glutathione levels.\textsuperscript{55}

Other neurotoxins have also been applied to produce experimental models to assess the neuroprotective capacity of polyphenolic compounds. CAPE protects PC12 cells against dopaminergic neurotoxin MPP\textsuperscript{+} (1-methyl-4-phenylpyridinium).\textsuperscript{67} Administration of 7,8-DHF prevented neuronal death in mouse brain induced by kainic acid.\textsuperscript{102} Icariin is another polyphenol compound that protects primary cultured rat hippocampal neuronal cells against corticosterone-induced apoptosis.\textsuperscript{177} In addition, baicalein was also shown to inhibit necrotic cell death damage in NPCs and to attenuate impairment of hippocampal neurogenesis caused by irradiation.\textsuperscript{155}

Polyphenols have also shown therapeutic potential in animal models of neurodegenerative diseases induced by various neurotoxins. In an Aβ-induced amnesia model in mice, oral administration of luteolin mitigated learning and memory impairment.\textsuperscript{123} Curcumin has also been shown to be effective in preventing neuroinflammation, tau hyperphosphorylation, and behavioral impairments, triggered by Aβ in vivo.\textsuperscript{118}

**Activation of the Nrf2 signaling pathway**

One of the elegant mechanisms that neuronal cells have adapted to protect themselves against oxidative stress and other insults is Nrf2 pathway and the binding of this master transcriptional regulator with ARE in the regulatory region of many genes, which leads to the expression of several enzymes with antioxidant and detoxification capacities (Figure 5).\textsuperscript{178} The main enzymes that are transcribed under the control of ARE include γ-glutamylcysteine synthetase, glutathione peroxidase, glutathione S-transferase, HO-1, NADPH quinone oxidoreductase 1, peroxiredoxin, sulfiredoxin, thioredoxin, and thioredoxin reductase all of which have important roles in the protection of cells.\textsuperscript{179-181} The pivotal finding that Nrf2-knockout mice exhibit a severe deficiency in the coordinated regulation of gene expression and their susceptibility to oxidative damage
indicate the crucial role of Nrf2 in maintaining intracellular redox homeostasis and antioxidant defense mechanism. The role of this pathway and the detoxification enzymes that it regulates has been especially emphasized in CNS diseases. Some phenolic compounds such as luteolin and puerarin in PC12 cells and also CAPE in dopaminergic neurons enhance the binding of Nrf2 to ARE and increase the expression of HO-1. In cultured neurons, EGCG was able to protect cells against oxidative stress by increasing HO-1 expression via activation of the transcription factor Nrf2. Li et al evaluated the neuroprotective effect of puerarin on lesioned substantia nigra induced by 6-OHDA. Their findings showed that puerarin effectively protects neurons in substantia nigra by modulating brain-derived neurotrophic factor (BDNF) expression and also by activating the Nrf2/ARE pathway. Carnosic acid and common sage (Salvia officinalis) plants also showed neuroprotective effects by activating Nrf2 in PC12h cells.

**Polyphenols and activation of other neurotrophic pathways**

Polyphenols having multiple beneficial effects on the nervous system could provide an important resource for the development of new drugs for management of neurodegenerative diseases. In addition to the aforementioned signaling pathways involved in the neurotrophic action of polyphenols, other mechanisms may also be involved. Daidzein has caused significant axonal outgrowth via upregulation of GAP-43 expression in hippocampal neurons in culture. Interestingly, daidzein-promoted phosphorylation of PKC, and GAP-43 was abolished by pretreatment with ER and PKC antagonist. These results suggest that ER-mediated PKC phosphorylation of GAP-43 may play a role in daidzein-mediated axonal outgrowth. Hesperetin can also exhibit multiple neurotrophic effects via ER- and TrkA-mediated parallel pathways. The Na+/K+/2Cl cotransporter (NKCC) is a member of the cation–chloride cotransporter family and is involved in the transport of chloride ion(s) coupled with cation(s) across the plasma membrane. A previous study has shown that NGF treatment of PC12D cells increased the expression of NKCC1 protein. In another report, it has been demonstrated that knockdown of NKCC1 strongly inhibits NGF-induced neurite outgrowth in PC12 cells. Quercetin also promoted NGF-induced neurite outgrowth by increasing Cl−, and knockdown of NKCC1 inhibited this.
stimulatory effect. In these cells, intracellular Cl\textsuperscript{−} affects microtubule polymerization via modulation of intrinsic GTPase activity of tubulin.\textsuperscript{187}

A2A subtype of adenosine receptors has been reported to elevate the expression of BDNF as well as synaptic actions of BDNF.\textsuperscript{188,189} This receptor also activates TrkB receptor and Akt signaling pathway, which induces neuronal survival and modulates neurite outgrowth in several different cell types.\textsuperscript{190–192} Jeon et al\textsuperscript{114} have recently shown that oroxylin A could induce BDNF production in cortical neurons via activation of A2A receptor, which induced cellular survival, synapse formation, and neurite outgrowth. In another report, the adenosine A2A receptor inhibitor could block methyl 3,4-dihydroxybenzoate-induced neuronal survival and neurite outgrowth in cultured primary cortical neurons.\textsuperscript{147}

**Conclusion**

Polyphenols with multiple beneficial effects in the nervous system could provide an important resource for the discovery of new neurotrophic agents. Agonistic action on Trk receptors, activation of the ERK, PI3Kinase/Akt and CREB pathways, activation of the Nrf2 pathway and upregulation of antioxidant and detoxification enzymes, as well as several other mechanisms underlie the neurotrophic action of different polyphenols. A better understanding of the neurotrophic effects and the molecular mechanisms of action of these compounds could help design better agents for management of neurodegenerative diseases and other disorders of the nervous system.

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**Disclosure**

The authors report no conflict of interest in this work.

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