The role of hypoxia in cancer progression, angiogenesis, metastasis, and resistance to therapy

Barbara Muz
Pilar de la Puente
Feda Azab
Abdel Kareem Azab
Department of Radiation Oncology, Cancer Biology Division, Washington University School of Medicine in St Louis, MO, USA

Abstract: Hypoxia is a non-physiological level of oxygen tension, a phenomenon common in a majority of malignant tumors. Tumor-hypoxia leads to advanced but dysfunctional vascularization and acquisition of epithelial-to-mesenchymal transition phenotype resulting in cell mobility and metastasis. Hypoxia alters cancer cell metabolism and contributes to therapy resistance by inducing cell quiescence. Hypoxia stimulates a complex cell signaling network in cancer cells, including the HIF, PI3K, MAPK, and NFκB pathways, which interact with each other causing positive and negative feedback loops and enhancing or diminishing hypoxic effects. This review provides background knowledge on the role of tumor hypoxia and the role of the HIF cell signaling involved in tumor blood vessel formation, metastasis, and development of the resistance to therapy. Better understanding of the role of hypoxia in cancer progression will open new windows for the discovery of new therapeutics targeting hypoxic tumor cells and hypoxic microenvironment.

Keywords: hypoxia, cancer, metastasis, angiogenesis, treatment resistance

Introduction

Twenty-five percent of deaths in the United States are caused by cancer, and the number of incidences increases due to population growth, prolonged life expectancy, and an abundance of risk factors including smoking, a lack of activity, and obesity.1 A common feature of most tumors is a low level of oxygen, called hypoxia, the severity of which varies between tumor types (Table 1). In intensively proliferating and expanding tumor tissue, oxygen demand is surpassed by oxygen supply, and the distance between cells and the existing vasculature increases, hampering oxygen diffusion and creating even more hypoxic milieu.2,3 It is generally accepted that the oxygen level in hypoxic tumor tissues is poorer than the oxygenation of the respective normal tissues and on average it is between 1%–2% O2 and below (Table 1). However, tumor oxygen level depends on the initial oxygenation of the tissue, the size and stage of the tumor, the method of oxygen measurement, and in which part of the heterogenic tumor tissue the measurement was performed (Table 1).4–7 Tissue normoxia, also known as physoxia, is the oxygenation in healthy tissues, which varies widely between the organs due to diversified blood vessel network and metabolic activity. Oxygen concentration in humans ranges between approximately 9.5% O2 in the renal cortex7 to 4.6% O2 in the brain with neurons extremely sensitive to hypoxia.8–10 These oxygen values are far from the experimental in vitro conditions. The oxygen concentration commonly used in the laboratory setting is 20.9% O2, which means that cell culture is performed in hyperoxic rather than physoxic conditions of respective organs. In order to better understand principles of oxygenation...
in vitro and in vivo, basic knowledge of the physics of gases is required for newcomers in the hypoxia research field which has been neatly described in a recently published review.\textsuperscript{11}

Cancer cells respond differently to decreased oxygenation leading to cell death or cell survival which partially depends on the time of exposure to hypoxia. The discrepancy and lack of consistency in experimental oncology regarding the definition of acute versus chronic hypoxia often with different biological consequences was thoroughly reviewed.\textsuperscript{12,13} In general, it is accepted that acute hypoxia is an abrupt and brief exposure to short-term hypoxia which occurs when blood vessel occlusion lasts for at least several minutes.\textsuperscript{14} It is reversible and often leads to oxygen fluctuations called cycling hypoxia. In acute hypoxia in vitro, cells are usually exposed to continuous hypoxia between a few minutes and up to 72 hours.\textsuperscript{12} Short-term hypoxia allows cells to survive in these adverse conditions by activating autophagy, an apoptotic and metabolic adaptation of cells. Autophagy is achieved by decreasing oxidative metabolism.\textsuperscript{15,16} On the contrary, others have shown that cycling hypoxia led to increased reactive oxygen species (ROS) production, what contributed to tumor cell survival and progression.\textsuperscript{17,18} Moreover, both short- and long-term hypoxia was shown to increase radio-resistance of cancer cells both in vitro\textsuperscript{17,19} and in vivo.\textsuperscript{17,20} In addition, acute hypoxia was associated with more aggressive tumor phenotype through induction of spontaneous metastasis.\textsuperscript{12,21,22}

Enduring changes in blood flow and low oxygen availability resulting in chronic hypoxia are especially pronounced in larger tumors and contribute to long-term cellular changes. In experimental settings, chronic conditions are considered when the cells are incubated in hypoxia between a few hours and as long as several weeks.\textsuperscript{12} Longer exposure to hypoxia is associated with high frequency of DNA breaks, accumulation of DNA replication errors since hypoxia hampers DNA repair systems including homologous recombination and mismatch repair, potentially leading to genetic instability and mutagenesis.\textsuperscript{23-25} Of note, acute hypoxia also leads to genomic instability due to delayed DNA damage response and rapid p53-dependent apoptosis.\textsuperscript{26} It was suggested that cells lacking functional p53 are more susceptible to genomic instability and potentially tumorigenesis if they experience reoxygenation after acute exposure to hypoxia.\textsuperscript{26}

Nonetheless, cycling hypoxia represents the situation of oxygenation in tumor tissues. Oxygen fluctuation occurs at irregular intervals in cancer with sporadic reoxygenation periods due to dysfunctional tumor vascularity and heterogeneous blood supply.\textsuperscript{27,28} Undoubtedly, both chronic and acute hypoxic regions in tumors directly affect clinical responses to therapy by influencing tumor growth, ability to metastasize, and resistance to cell death.

### Signaling pathways related to tumor hypoxia

Hypoxia induces a number of complex intracellular signaling pathways such as the major hypoxia-inducible factor (HIF) pathway. Other hypoxia-associated pathways include PI3K/AKT/mTOR,\textsuperscript{29,30} MAPK also known as ERK pathways,\textsuperscript{31-33} and the NFκB.\textsuperscript{34} These pathways are involved in cell proliferation, survival, apoptosis, metabolism, migration, and inflammation.

PI3K/AKT/mTOR, MAPK, and NFκB signaling pathways are also stimulated in a hypoxia-independent manner by a number of factors such as cytokines, chemokines, and growth factors which bind to receptor tyrosine kinases, G protein-coupled receptors, toll-like receptors (TLR), and alarmins receptors on the cell surface, which eventually may also lead to HIF-1α activation (Figure 1). In addition, in cancer cells epigenetic changes and acquired mutations of the pathways’ members and overactivation/overstimulation of receptors cause uncontrollable cancer cell growth.\textsuperscript{35}

Targeting non-HIF pathways provides a promising target for anti-neoplastic therapy and each pathway is a vast topic on its own. More information regarding the role of the non-HIF pathways in cancer can be found elsewhere. This review will
mainly concentrate on the HIF pathway and its involvement in tumor progression.

**HIF pathway**

Cellular adaptation to hypoxia is primarily mediated by a family of transcriptional regulators, HIF, which was identified 2 decades ago. The hypoxic induction and protein stabilization of HIF-α subunits (HIF-1α, HIF-2α, and HIF-3α) is regulated by oxygen sensors, including PHD and FIH-1 enzymes. PHDs and FIH-1 are upstream of HIF-α and their activity is oxygen-dependent. In oxygenated cells, HIF-α subunits are hydroxylated by oxygen sensors, including PHD and FIH-1 enzymes, causing polyubiquitination and proteasomal degradation of hydroxylated HIF-α subunits (red arrows). PHD and FIH-1’s activity is oxygen-dependent (red arrows); in hypoxia (blue arrows) these enzymes lose their activity due to decreased oxygenation, resulting in HIF-α protein stabilization, accumulation, and translocation into the nucleus resulting in gene transcription and biological consequences (black arrows). HIF is also modulated in a hypoxia-independent manner in response to nitric oxide (NO), reactive oxygen species (ROS), cytokines, lipopolysaccharides, and growth factors through receptor tyrosine kinases (RTK), G protein-coupled receptors (GPCRs), toll-like receptors (TLR) and alarmins receptors. The non-hypoxic HIF regulation is mediated by a number of different signaling pathways including NFκB, PI3K/AKT/mTOR, and MAPK/ERK (green arrows). These pathways, as well as ROS production, are additionally regulated by hypoxia, which results in multiple levels of HIF-α stimulation, both hypoxic and normoxic. As a result, HIF accumulation and activation alters blood vessel formation, apoptosis, metastasis, and metabolism via a number of genes including VEGF, SDF-1, Ang-2, MMPs, BNIP-3, p53, epithelial-to-mesenchymal transition (EMT), E-cad, CXCR4, LOX, CAIX, GLUT-1, and GSK (black arrows).

![HIF pathway diagram](https://www.dovepress.com/)

**Figure 1 Regulation of HIF in normoxic and hypoxic conditions.**

**Notes:** HIF-α, a transcription factor, can be regulated by both hypoxic and non-hypoxic factors. In normoxia, HIF-α subunits are hydroxylated by oxygen sensors, including PHD and FIH-1 enzymes, causing polyubiquitination and proteasomal degradation of hydroxylated HIF-α subunits (red arrows). PHD and FIH-1’s activity is oxygen-dependent (red arrows); in hypoxia (blue arrows) these enzymes lose their activity due to decreased oxygenation, resulting in HIF-α protein stabilization, accumulation, and translocation into the nucleus resulting in gene transcription and biological consequences (black arrows). HIF is also modulated in a hypoxia-independent manner in response to nitric oxide (NO), reactive oxygen species (ROS), cytokines, lipopolysaccharides, and growth factors through receptor tyrosine kinases (RTK), G protein-coupled receptors (GPCRs), toll-like receptors (TLR) and alarmins receptors. The non-hypoxic HIF regulation is mediated by a number of different signaling pathways including NFκB, PI3K/AKT/mTOR, and MAPK/ERK (green arrows). These pathways, as well as ROS production, are additionally regulated by hypoxia, which results in multiple levels of HIF-α stimulation, both hypoxic and normoxic. As a result, HIF accumulation and activation alters blood vessel formation, apoptosis, metastasis, and metabolism via a number of genes including VEGF, SDF-1, Ang-2, MMPs, BNIP-3, p53, epithelial-to-mesenchymal transition (EMT), E-cad, CXCR4, LOX, CAIX, GLUT-1, and GSK (black arrows).
survival via a number of growth factors and inhibition of pro-apoptotic pathways, ii) contribute to tumor neo-vascularization via VEGF, VEGF receptors, COX-2, iNOS, iii) regulate cell detachment via downregulation of adhesion molecules such as cadherins, and iv) induce cell migration and invasion through matrix degrading enzymes. The HIF and NFKB pathways are controlled by a negative feedback loop mechanism and also intersect via alarmins. Tissue damage and necrosis, which can be also induced by hypoxia, increases the presence of alarmins, the endogenous markers for damage, which are recognized by receptor for advanced glycation endproducts (RAGE) and some of TLRs. In addition, the expression of RAGE receptor is also upregulated by HIF-1α. In turn, alarmin receptors strongly activate NFκB and proinflammatory gene expression. Moreover, the basal HIF-1α mRNA expression is regulated by NFκB in non-hypoxic conditions since HIF-1α promoter was shown to be responsive to certain NFκB subunits.

The HIF pathway is required during physiological processes and is implicated in cancer biology by regulating hundreds of genes. This master regulator facilitates tumor growth by promoting angiogenesis via VEGF and SDF-1, metabolism via regulation of GLUT-1, GLUT-3, and glycolytic enzymes, and regulating cell apoptosis and cell survival via BNIP-3, p53, TGF-β, and bFGF. Moreover, HIF-α contributes to cancer metastasis by altering cancer cell adhesion and motility through regulation of epithelial-to-mesenchymal transition (EMT) and E-cad, ZEB1, -2 and TCF3 expression, as well as migration and invasion abilities through CXCR4, CAIX, LOX, MMP-2, and MMP-9.

The role of hypoxia in progression and metastasis in cancer

Pathological hypoxia is a common microenvironment factor in tumors that facilitates cell survival and propagation of the tumor. Key cellular responses to hypoxia triggered by overexpression of HIF-1α and HIF-2α subunits and their downstream targets increase blood vessel formation, aggressiveness, metastasis, and resistance to treatment.

Blood vessel formation

Blood vessels create a network of tubes and capillaries which nourish the entire body with oxygen and nutrients. Thus, the way they are formed and function is crucial in embryogenesis and physiology. Blood vessels consist of endothelial cells (ECs) which create a tight barrier between the blood and tissue, and interact with ECM. In embryogenesis, blood vessels are formed de novo by vasculogenesis involving bone marrow-derived endothelial progenitor cells (EPCs) followed by angiogenesis, a process where new blood vessels are created from pre-existing vasculature. Lastly, the vessels undergo maturation which includes physical interaction with smooth muscle cells and pericytes. Abnormal angiogenesis is a feature of pathological conditions including tumor progression, where hyperproliferating cancer cells surpass their blood supply and become hypoxic. Hypoxia induces the imbalance between pro- and anti-angiogenic factors' production, which leads to enhanced, rapid and chaotic blood vessel formation. Hypoxia and potent transcription factors HIF-1α and HIF-2α have been shown to be involved in all steps of blood vessel formation. Hypoxia and HIF-α subunits contribute to the EPCs' recruitment from the bone marrow and induction of their differentiation into ECs by regulation of VEGF, a primary regulator of vasculogenesis. This is also mediated through stimulation of pro-angiogenic molecule production such as VEGF-R2 (Flk-1), members of the FGF family and PDGF, important in the primitive vascular network formation. Hypoxia and HIF-α are also involved in the angiogenesis process by inducing enzymes' expression (ie, MMPs) in order to sprout and split the pre-existing vessels. In turn, neovessels allow ECs to migrate in response to chemoattractants across ECM. Additionally hypoxia induces ECs' proliferation by regulation of VEGF-R1 (Flt-1), Ang-1 and Ang-2 expression. Finally, hypoxia and HIF-α support vessel maturation via induction of Ang-1, PDGF, and TGF-β to recruit supporting cells such as smooth muscle cells and pericytes creating mature and stable blood vessels.

However, in tumors, neovessels are often abnormal, immature, and leaky. They are either insufficient or excessive depending on the tumor type. Neovasculogenesis maintains blood flow to the growing tumor tissue that expands rapidly, providing nutrients and oxygen for thriving cancer cells; however, more cells means more demand causing even more hypoxia. Again, hypoxia in turn stimulates angiogenesis to ameliorate hypoxic condition, closing the vicious circle. As a consequence the tumor tissue ends up being highly hypoxic with excessive but dysfunctional vasculature.

Folkman was the first to propose anti-angiogenic therapy to treat cancer in 1971. A successful use of monoclonal antibody against VEGF (bevacizumab) approved for treatment of metastatic colorectal cancer followed by multiple solid tumors, has stimulated development of other anti-angiogenic therapies. However, long-term exposure to these agents revealed not only reducing tumor growth, but also more malignant and invasive cancer phenotype increasing metastasis. Long-term exposure to anti-angiogenic agents reduce...
tumor; however, at the same time induce more aggressive and metastatic tumor phenotype.71,72

Metastasis
Enhanced angiogenesis is associated with metastasis since permeable and heterogeneous vasculature facilitates the extravasation, circulation, and relocation of tumor cells of tumor cells to new and unaffected tissues escaping the hostile hypoxic environment.68 Tumor oxygenation is a critical factor of cancer progression and the overexpression of HIF-α subunits in tumors and their metastases is associated with the aggressiveness of a majority of human cancers and correlates with poor overall survival.49,73,74

It was demonstrated previously that hypoxic cells are more aggressive and invasive with better ability to metastasize. For instance, multiple myeloma cancer cells cultured in hypoxic conditions in vitro and injected into mice were able to spread to the new bone marrow faster than the cells cultured in normoxic conditions.63,75 Also, exposing an orthotopic mouse model of cervical carcinoma to a dozen cycles of 10 minutes 7% O2, which was followed by 10 minutes of air exposure daily, increased the number of lymph node metastases.76 Similar observations were recorded in mice bearing sarcoma tumors, where exposure to acute hypoxia augmented the lung metastases.77

Mechanistically, hypoxia was shown to influence invasive and migratory behavior of cancer cells via EMT, a trans-differentiation of cells in order to acquire plastic and mobile abilities, a process which alters their gene expression prior to migration.78 EMT is physiologically active during embryogenesis and tissue regeneration, as well as in cancerogenesis in many types of solid tumors79 and hematologic malignancies.63 Hypoxia-induced EMT is characterized by a decrease in epithelial-associated gene expression, such as E-cad, β-catenin80 and an increase in mesenchymal-like gene expression, such as N-cad,81 vimentin, SMA,82 and CXCR4.63,75 EMT is promoted by a master regulator TGF-β, also increased by hypoxia, which activates downstream transcription factors such as Smads, Snail, Slug, and Twist, and inhibits expression of E-cad.63,81 Interestingly, radio- and chemoresistance was also shown to be associated with EMT phenotype; expression of Snail and Slug antagonizes p53-mediated apoptosis and promotes resistance to radiation and chemotherapeutic agents such as paclitaxel and cisplatin in ovarian cancer cells.84

Moreover, HIF-1α was shown to be expressed in 90% of human gastric cancer biopsies at the front edge of the invading tumor compared to HIF-1α negative normal tissues.74 HIF inhibition significantly reduced the metastasis of gastric cancer cells in vivo, and HIF deficient cells were less motile, invasive, and adhesive in vitro.74 High involvement of the main hypoxic regulator, HIF-α, in all steps of metastasis led to many trials of inhibiting this molecule to diminish cancer cell trafficking thus reducing metastasis. Inhibition of HIF-1 activity using antisense oligonucleotide (EZN-2968) gave effective results and a safe toxicity in a Phase I clinical trial in metastatic, advanced solid malignancy.83 Targeting hypoxic cells with a pro-drug, activated only in a hypoxic environment, is one of the newest and highly promising strategies to reduce metastasis, currently undergoing phase I/II clinical trials in multiple myeloma, a model of a process of metastasis.86

Apart from targeting HIF-α molecules, another strategy to inhibit metastasis is to target genes downstream of HIF-α. For instance, CAIX is a hypoxia-inducible enzyme widely present in tumors; it is crucial in regulating intra- and extracellular pH, thus CAIX promotes survival and invasion of

Figure 2 Hypoxia as a driving force of tumor progression and metastasis. Notes: Hypoxia stimulates tumor i) vasculogenesis through endothelial progenitor cells’ mobilization from the bone marrow to the tumor site by VEGF, VEGF-R2, fibroblast growth factor (FGF), platelet-derived growth factor (PDGF) and stromal-derived growth factor-I (SDF-I) and ii) angiogenesis by sprouting of the pre-existing vessels caused by increased production of VEGF, VEGF-R1, VEGF-R2, Ang-I, Ang-2, and MMPs. New blood vessels facilitate cancer cells leaving the primary tumor site, which is enhanced by increased expression of lysyl oxidase (LOX), carbonic anhydrase IX (CA IX), MMPs, integrins, and CXCR4. Hypoxic cancer cells also undergo epithelial-to-mesenchymal transition (EMT) acquiring plastic and mobile phenotype by increasing transcription factors such as Slug, Snail, and Twist and decreasing expression of adhesion molecules such as β-catenin and E-cadherin (E-cad). Chemo- and radio-resistance of patients is caused by EMT-related stemness of cancer cells and hypoxia-induced cell cycle arrest in G1 phase. Hindered drug diffusion due to anomalous vascularity is another mechanism of chemoresistance.
cancer cells.\textsuperscript{87} It was demonstrated that, inhibition of CAIX decreased tumor growth and metastasis in pre-clinical breast tumor models.\textsuperscript{87} LOX is another protein elevated in hypoxic human tumor cells (such as breast, and head and neck cancer) and is HIF-α-dependent.\textsuperscript{61} LOX secreted by tumor cells accumulates at new sites and mediates bone marrow-derived cell recruitment which forms a “pre-metastatic niche” for future metastasis.\textsuperscript{88} Inhibition of LOX reduced hypoxia-induced recruitment and metastasis in the breast cancer mouse model.\textsuperscript{89} Another target candidate induced by hypoxia in cancer cells is CXCR4 involved in cell trafficking.\textsuperscript{89,90} It was demonstrated that metastatic tumor cells expressing high levels of CXCR4 home to tissues rich in its ligand, SDF-1,\textsuperscript{63,91} and inhibitors of CXCR4/SDF-1 axis resulted in disruption of the metastatic process (Figure 2).\textsuperscript{90}

Radiation and drug resistance
Resistance of cancer cells to treatment-induced apoptosis is one of the biggest obstacles in cancer therapy.\textsuperscript{92} A vast number of cancer patients relapse and suffer from recurring tumors as a result of micro-residual disease, the resistant subpopulation of cancer cells, leading to local recurrence and/or metastasis.\textsuperscript{93} Tumor hypoxia develops due to uncontrollable cell proliferation, altered metabolism, and abnormal tumor blood vessels resulting in reduced transport of oxygen and nutrients.\textsuperscript{13} Hypoxia is one of the main features of solid tumors and was shown to correlate with poor prognosis of cancer patients.\textsuperscript{73} While hypoxia is lethal for many cells, a subpopulation of tumor cells is able to not only adapt to hypoxic conditions but also become resistant to chemotherapeutics and radiotherapy. The role of hypoxia in the phenomenon of therapy resistance has been acknowledged for at least 60 years.\textsuperscript{94,95}

Hypoxia confers treatment resistance of cancer cells by regulating processes such as i) inducing cell cycle arrest (quiescence), a state of reduced cell proliferation which protects the cells from external stress,\textsuperscript{96,97} ii) inhibiting apoptosis and senescence of cells, iii) controlling autophagy, p53, and mitochondrial activity.\textsuperscript{94,97} Apart from cellular adaptations influenced by hypoxia, lowered oxygenation of the tumor tissue confers chemoresistance by affecting iv) drug delivery and cellular uptake through associated acidity and drug efflux pump expression such as P-gp,\textsuperscript{98} as well as by the v) lack of oxygen required for the cytotoxicity of a number of chemotherapeutics.\textsuperscript{99}

One of the most important parameters of radio-resistance is oxygenation, the state of cell cycle, and the nature of radiation, whether it is gamma, X-ray, neutron radiation, or linear energy transfer including alpha and beta particles.\textsuperscript{94,96} When oxygen is abundant, the normoxic cells are sensitive to radiation, due to “oxygen fixation” which happens when available molecules of oxygen react with free radicals in DNA generated by ionizing radiation leading to irreversible DNA damage. Whereas, cells irradiated in hypoxic conditions are resistant to death, due to decreased production of DNA radicals (which can be restored “chemical restitution”) caused by reduced generation of ROS and decreased DNA damage.\textsuperscript{95} The cell cycle phase determines the radiosensitivity of the tumors. It was shown that cells exposed to ionizing radiation are the least sensitive at the end of S phase, less sensitive in the G1 phase, and the most radiosensitive in the G2/M phase where DNA repair mechanisms are the most prone to malfunction.\textsuperscript{100} Hence, anticancer treatments, both radio- and chemotherapy, preferentially target the bulk of rapidly proliferating tumor cells. On the other hand, the cells which are most resistant to treatment are quiescent, low-proliferating, stem-cell-like cell fractions residing in the most hypoxic region.\textsuperscript{82,84,97,101} For instance, it was demonstrated that glioblastoma cancer stem cells (CD133\textsuperscript{*}) showed higher DNA repair and decreased apoptosis in response to irradiation, compared to non-stem-cell-like (CD133\textsuperscript{-}) population.\textsuperscript{94,96,102}

Hypoxia causes slow-proliferating stem-cell-like phenotype of cells, decreases senescence, creates chaotic and malfunctioning blood vessels, and augments metastasis, which all together further induces therapy resistance.\textsuperscript{96} Currently, assessment of tumor oxygenation and HIF expression pattern helps determine tumor chemo- and radio-sensitivity.\textsuperscript{94,103} It was reported that head-and-neck cancer samples with high expression of HIF-1α and HIF-2α were more resistant to chemotherapu (carboplatin) compared to biopsies with low HIF-α expression which were chemo-sensitive.\textsuperscript{104} Patients with oropharyngeal cancer demonstrating high expression of HIF-1α had a lower chance to achieve complete remission after irradiation.\textsuperscript{103} In addition, irradiation was shown to induce HIF-1 activity, leading to production of angiogenic molecules such as VEGF which protects ECs from irradiation-induced apoptosis.\textsuperscript{105} Therefore HIF-1 represents a valid predictive marker and therapeutic target for manipulation, in combination with chemotherapy and radiotherapy, in order to sensitize the cells to treatments.\textsuperscript{73}

As demonstrated by others, inactivation of HIF-1α in mouse embryonic fibroblasts increased their susceptibility to carboplatin and etoposide compared to wild-type, both in vitro and in vivo.\textsuperscript{106} Similarly, inhibition of HIF-2α with short hairpin RNA reversed the resistance to doxorubicin and etoposide of human clear cell renal cell cancer cells.
(one of the most resistant tumors) by restoring p53. It was reported that treating tumor-bearing mice with HIF-1 inhibitor (YC-1) induced radiation-induced vessel damage. Similarly, treatment of glioma, squamous and pancreatic cancer cells with the HIF-1α inhibitor (PX-478) radiosensitized hypoxic cells. Silencing of HIF-1α with siRNA in mouse embryonic fibroblasts increased susceptibility to irradiation, and also, HIF-2α inhibition was shown to enhance radiation-induced apoptosis due to HIF-2-mediated increase of p53 activity and accumulation of ROS, thus DNA damage (Figure 2).

**Tumor microenvironment**

The tumor would not thrive without the interaction, cross-talk, and support with the tumor microenvironment including cellular components such as stromal cells, immune cells, ECs as well as non-cellular components including ECM, cytokines, and other mediators. Thus, in hypoxic tumor tissue, not only cancer cells but also the tumor microenvironment is affected by hypoxia-inducible changes.

Hypoxia was shown to induce metabolic and molecular changes in ECs, increasing expression of pro-angiogenic molecules, blood vessel formation, and thus providing more oxygen and nutrients for tumor cells. Hypoxia also regulates inflammatory mediators and growth factors, which then stimulate platelet, leukocyte, and smooth muscle cell activity. One of the most significant changes is increase in adhesive-ness of ECs to neutrophils facilitating NK cell trafficking and local inflammatory reaction. Depending on the duration of oxygen depletion, hypoxia regulates expression of NO synthase expression contributing to vasoconstriction. Since blood vessels nourish tumors, targeting ECs will prevent or reverse tumor growth.

Stromal cells, on the other hand, facilitate tumor growth and tumor dissemination mostly by regulating cancer cell adhesion and contributing to cell proliferation and survival. It was shown that hypoxia induces stromal cells to produce a number of factors including Ang-2, ANGPTL-4, PDGF, VEGF, SDF-1, LOX, and SCF (KIT-ligand), influencing ECs and EPCs thus promoting new blood vessel formation and lymphangiogenesis. Also, stromal-derived SDF-1 attracts cancer cells and thus facilitates metastasis.

It was demonstrated that hypoxia leads to immune-resistance and immune-suppression, which help tumor cells to escape from immune surveillance. Some of the immune-suppressive effects include: 1) shedding of immune-recognition molecules by tumor hypoxia, which results in decreased sensitivity to T- and NK-mediated killing; 2) inhibition of T cells’ and dendritic cells’ maturation and cytokine production; 3) and promotion of suppressive cells such as regulatory T cells and tumor-associated macrophages, which block immune effector cells.

Therefore, there is an increasing importance of the hypoxic phenotype of stromal and immune cells in the tumor microenvironment providing non-cancer cells as potential novel targets in the fight against the tumor.

**Conclusion**

Pathological hypoxia affects both cancer cells and the tumor microenvironment, and plays a pivotal role in the process of cancer progression and dissemination. Hypoxia regulates tumor neovascularization, metabolism, cell survival, and cell death. In addition, hypoxia contributes to EMT-like cancer cell migration and cancer stem-cell-like properties including resistance to treatment, one of the nightmares in the medical field. Each step of the cancer adaptive processes is controlled by hypoxia-activating transcriptional programs involving HIF, NFκB, PI3K, and MAPK pathways.

Since hypoxia signifies increased tumor progression and aggressiveness hampering patients’ survival, direct and indirect methods of measuring hypoxia combined with clinical observations may help to predict patients’ outcome as well as identify patients who could benefit from hypoxia/HIF-targeted treatments. Better understanding of hypoxic phenomenon and dissecting out the hypoxia-inducible responses and signaling pathways will grant numerous novel targets in the near future.

**Disclosure**

Dr AK Azab receives research support from Verastem, Sellex, Karyopharm and Cell Works, and is the founder and owner of Targeted Therapeutics LLC and Cellatrix LLC. No potential conflicts of interest were disclosed by B Muz, P de la Puente and F Azab.

**References**

Hypoxia-induced enrichment and mutagenesis of cells that have lost function of BNIP3 and BNIP3L via their BH3 domains.

A. Hypoxia and angiogenesis, perfusion and metastatic dissemination.

B. Tumors exposed to acute cyclic hypoxic stress show enhanced force in genetic instability.

C. Hypoxia on DNA damage responses and genomic instability.

D. Hypoxia-induced autophagy: cell death or long-term HIF-1 signal transduction activity.

E. Response of tumor: radiobiological effects.

F. DNA damage repair, radioprotection and long-term HIF-1 signal transduction activity.

G. Mitochondria-dependent and -independent pathways.

H. NFkB in Cancer Treatment: Is it a Rational Approach for Controlling Malignant Progression?

I. Regulation of hypoxia-inducible factor-1alpha by NF-kappaB.

J. Mitogen-activated protein kinases phosphorylate hypoxia-inducible factor-1alpha by NF-kappaB.


74. Rohwer N, Lobitz S, Daskalow K, et al. HIF-1alpha determines
73. Semenza GL. Defining the role of hypoxia-inducible factor 1 in cancer
71. Mountzios G, Pentheroudakis G, Carmeliet P. Bevacizumab and
67. Krock BL, Skuli N, Simon MC. Hypoxia-induced angiogenesis: good
66. de la Puente P, Muz B, Azab F, Azab AK. Cell Trafficking of Endothe
64. Conway EM, Collen D, Carmeliet P. Molecular mechanisms of blood
63. Azab AK, Hu J, Quang P, et al. Hypoxia promotes dissemination of
60. Grabmaier K, MC AdW, Verhaegh GW, Schalken JA, Oosterwijk E.
57. Roberts AM, Watson IR, Evans AJ, Foster DA, Irwin MS, Ohh M.
56. An WG, Kanekal M, Simon MC, Maltepe E, Blagosklonny MV,
53. Iliopoulos O, Levy AP, Jiang C, Kaelin WG Jr, Goldberg MA. Negative
47. Hsu M, Andl T, Li G, Meinoth J, Herlyn M. Cadherin repertoire
determines partner-specific gap junctional communication during
46. Horder N, Azab F, Blotta S, et al. RhoA and Rac1 GTPases play
major and differential roles in stromal cell-derived factor-1-induced
cell adhesion and chemotaxis in multiple myeloma. Blood. 2009;
45. Azab AK, Runnels JM, Pitsillides C, et al. CXCR4 inhibitor AMD3100
promotes dissemination and colonization in new bone marrow niches
44. Hu J, Handisides DR, Van Valckenborgh E, et al. Targeting the multiple
myeloma hypoxic niche with TH-302, a hypoxia-activated prodrug. Blood.
2010;116(9):1524–1527.
43. Lou Y, McDonald PC, Oloumi A, et al. Targeting tumor hypoxia:
suppression of breast tumor growth and metastasis by novel carbonic
42. Erler JT, Bennewith KL, Cox TR, et al. Hypoxia-induced lysi oxidase
is a critical mediator of bone marrow cell recruitment to form the
41. Azab AK, Azab F, Blotta S, et al. RhoA and Rac1 GTPases play
major and differential roles in stromal cell-derived factor-1-induced
cell adhesion and chemotaxis in multiple myeloma. Blood. 2009;
40. Azab AK, Rannels JM, Pitsillides C, et al. CXCR4 inhibitor AMD3100
enables the interaction of multiple myeloma cells with the bone marrow
microenvironment and enhances their sensitivity to therapy. Blood.
2009;113(18):4341–4351.
trafficking through modulation of CXCR4 and E-cadherin expression.
37. Munshi NC, Anderson KC. Minimal residual disease in multiple
on the functional interaction of HIFs and cell death pathways. Drug
35. Gray LH, Conger AD, Ebert M, Hornsey S, Scott OC. The concentration
of oxygen dissolved in tissues at the time of irradiation as a factor in
34. Vaupel P, Kelleher DK, Hockel M. Oxygen status of malignant tumors:
pathogenesis of hypoxia and significance for tumor therapy. Semin
33. Das B, Tsuchida R, Malkin D, Koren G, Baruchel S, Yeger H. Hypoxia
enhances tumor stemness by increasing the invasive and tumorigenic
32. Abraham J, Salama NN, Azab AK. The role of P-glycoprotein in
drug resistance in multiple myeloma. Leuk Lymphoma. 2015;56(1):
26–33.
Hypoxia is an international, peer-reviewed, open access journal that aims to improve understanding of the biological response to hypoxia. The journal will publish original research articles, reviews, methodological advances, clinical studies, and expert opinions that identify developments in the regulation of the physiological and pathological responses to hypoxia and in the therapeutic targeting of hypoxia-responsive pathways.

The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit http://www.dovepress.com/testimonials.php to read real quotes from published authors.