Clinicopathological significance and potential drug target of CDH1 in breast cancer: a meta-analysis and literature review

Abstract: CDH1, as a tumor suppressor gene, contributes sporadic breast cancer (BC) progression. However, the association between CDH1 hypermethylation and BC, and its clinicopathological significance remains unclear. We conducted a meta-analysis to investigate the relationship between the CDH1 methylation profile and the major clinicopathological features. A detailed literature was searched through the electronic databases PubMed, Web of Science™, and EMBASE™ for related research publications. The data were extracted and assessed by two reviewers independently. Odds ratios (ORs) with corresponding confidence intervals (CIs) were calculated and summarized respectively. The frequency of CDH1 methylation was significantly higher in invasive ductal carcinoma than in normal breast tissues (OR = 5.83, 95% CI 3.76–9.03, P < 0.00001). CDH1 hypermethylation was significantly higher in estrogen receptor (ER)-negative BC than in ER-positive BC (OR = 0.62, 95% CI 0.43–0.87, P = 0.007). In addition, we found that the CDH1 was significantly methylated in HER2-negative BC than in HER2-positive BC (OR = 0.26, 95% CI 0.15–0.44, P < 0.00001). However, CDH1 methylation frequency was not associated with progesterone receptor (PR) status, or with grades, stages, or lymph node metastasis of BC patients. Our results indicate that CDH1 hypermethylation is a potential novel drug target for developing personalized therapy. CDH1 hypermethylation is strongly associated with ER-negative and HER2-negative BC, respectively, suggesting CDH1 methylation status could contribute to the development of novel therapeutic approaches for the treatment of ER-negative or HER2-negative BC with aggressive tumor biology.

Keywords: methylation, estrogen receptor, HER2, triple-negative breast cancer

Introduction

Breast Cancer (BC) is the most common malignancy among women in most western countries.1 The development of BC involves a progression, starting with atypical hyperplasia, followed by intermediate stages until the invasive carcinoma, and finally into metastatic disease. A series of epigenetic and genetic changes contribute to this multistep process of BC onset and progression. Epigenetics refers to heritable changes in gene activity and expression that do not involve changes to underlying DNA sequence and thus have a role in tumorigenesis as a tumor suppressor gene.7–10 Alteration in E-cadherin expression...
has been observed in several types of malignancies, including ovarian cancer, prostate cancer, hepatocellular carcinoma, and BC. Recent evidence indicate that reduced E-cadherin expression is associated with pathological features, such as poor differentiation, infiltrative growth, lymph node metastasis, and poor prognosis. However, the association of between CDH1 promoter hypermethylation and BC, and its clinicopathological significance, remains under investigation. We conducted a meta-analysis to investigate the relationship between CDH1 methylation status in BC and the major clinicopathological features including estrogen receptor (ER), progesterone receptor (PR) and HER2.

Methods

Literature search and selection of studies

We conducted a systematic search of the literature published in English and Chinese prior to November 2014. The electronic databases included PubMed, Web of Science™, and EMBASE™. The search was conducted using the following key words: “CDH1 or E-cadherin methylation” and “breast cancer” or “breast carcinoma” for relative articles. There were 84 articles identified from PubMed, 143 articles from Web of Science, and 101 articles from EMBASE. A total of 328 articles were screened using article titles and abstracts. Reference lists of identified articles were searched manually for further relevant articles (Figure 1).

The following inclusion criteria were applied: 1) studies that reported the association between CDH1 methylation and the clinicopathological significance of BC and 2) studies that investigated the frequency of CDH1 methylation in different ER, PR, and HER2 statuses of BC. After screening by article titles and abstracts, 25 relevant articles were included for full text review. The exclusion criteria were as follows: 1) reviews; 2) studies in which CDH1 protein expression was investigated; 3) studies in which cell lines or mice were utilized; and 4) studies in which same populations or overlapping data were used. Finally, eleven articles were selected for inclusion in this meta-analysis. The variables from eleven relevant studies were listed in Table 1.

Data extraction and study assessment

Two independent reviewers (RH and PD) extracted the following data: first author’s name, year of publication, geographical location, sample size of the different histologic categories of BC, grade of BC, stage of BC, and ER, PR, and HER2 status. Any disagreement was resolved by discussion between two authors (RH and PD). If they could not reach a consensus, a third author (YD) was consulted.

Statistics analysis

The meta-analysis was conducted using Review Manager 5.2 (software update; The Nordic Cochrane Centre, Copenhagen, Denmark). Odds ratios (ORs) with 95% confidence intervals (CIs) were calculated. The assessment of statistical heterogeneity was done by using the Cochran’s Q statistic and I² tests. If the I² value was below 50%, a fixed effects model was used, and if the I² value was 50% or greater, a random effects model was used. The analysis was conducted to evaluate the association of the frequency of CDH1 methylation with invasive ductal carcinoma (IDC). Furthermore, we compared the frequency of CDH1 methylation in different ER, PR, and HER2 statuses as well as in different grades and stages. All P-values were two sided, with P<0.05 considered statistically significant. Publication bias was determined using funnel plot analysis.

Results

CDH1 was more frequently methylated in IDCs than in normal breast tissues (pooled OR =5.83, 95% CI 3.76–9.03,

<table>
<thead>
<tr>
<th>Author</th>
<th>Year</th>
<th>Country</th>
<th>Sample Normal</th>
<th>Stage (I + II/III)</th>
<th>Lymph node (0/+ )</th>
<th>Grade (I + II/III)</th>
<th>ER status (−/+ )</th>
<th>HER2 status (−/+ )</th>
</tr>
</thead>
<tbody>
<tr>
<td>Asiif et al14</td>
<td>2014</td>
<td>India</td>
<td>128</td>
<td>89/39</td>
<td>79/49</td>
<td>108/20</td>
<td>39/89</td>
<td></td>
</tr>
<tr>
<td>Karray-Chouayekh et al</td>
<td>2010</td>
<td>Tunisia</td>
<td>78</td>
<td>21/30</td>
<td>36/42</td>
<td>61/17</td>
<td>31/47</td>
<td>51/13</td>
</tr>
<tr>
<td>Hoque et al15</td>
<td>2009</td>
<td>Italy</td>
<td>12</td>
<td>45</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Zou et al16</td>
<td>2009</td>
<td>New Zealand</td>
<td>19</td>
<td>20</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sunami et al17</td>
<td>2008</td>
<td>USA</td>
<td>130</td>
<td>102/8</td>
<td>76/54</td>
<td>15/35</td>
<td>65/65</td>
<td>78/29</td>
</tr>
<tr>
<td>Prasad et al18</td>
<td>2008</td>
<td>India</td>
<td>50</td>
<td></td>
<td>83/71</td>
<td>27/23</td>
<td>54/134</td>
<td>142/30</td>
</tr>
<tr>
<td>Li et al19</td>
<td>2006</td>
<td>Australia</td>
<td>193</td>
<td></td>
<td>102/43</td>
<td>37/114</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Shinozaki et al20</td>
<td>2005</td>
<td>USA</td>
<td>10</td>
<td>151</td>
<td>53/18</td>
<td>45/26</td>
<td>63/8</td>
<td></td>
</tr>
<tr>
<td>Caldeira et al21</td>
<td>2005</td>
<td>Brazil</td>
<td>71</td>
<td>53/18</td>
<td></td>
<td>15/8</td>
<td>5/10</td>
<td>10/3</td>
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<tr>
<td>Hu et al22</td>
<td>2002</td>
<td>People’s Republic of China</td>
<td>23</td>
<td>18/5</td>
<td></td>
<td>15/8</td>
<td>5/10</td>
<td>10/3</td>
</tr>
</tbody>
</table>

*Abbreviations: ER, estrogen receptor; IDC, invasive ductal carcinoma.*
CDH1 hypermethylation was significantly higher in ER-negative than in ER-positive patients with BC (OR = 0.62, 95% CI 0.43–0.87, \( z = 2.72, P = 0.007, I^2 = 39\%, P = 0.13 \)) (Figure 3). CDH1 promoter was significantly more methylated in HER2-negative patients than in HER2-positive patients with BC (OR = 0.26, 95% CI 0.15–0.44, \( z = 4.97, P < 0.00001, I^2 = 0\%, P = 0.57 \)) (Figure 4). The methylation frequency was similar between PR-positive and -negative patients with BC (OR = 0.76, 95% CI 0.49–1.15, \( z = 1.30, P = 0.19, I^2 = 11\% \)) (Figure 5). The frequency of CDH1 hypermethylation was not significantly changed between late stage and early stage of BC (OR = 1.09, 95% CI 0.86–1.44, \( z = 1.40, P = 0.16, I^2 = 11\% , P = 0.35 \)) (Figure 6).

![Schematic flow diagram for selection of included studies.](image)

**Figure 1** Schematic flow diagram for selection of included studies.

**Figure 2** Forest plot for CDH1 hypermethylation in IDC and normal breast tissue.

**Abbreviations:** CI, confidence interval; IDC, invasive ductal carcinoma; M–H, Mantel–Haenszel.
**Figure 3** Forest plot for CDH1 hypermethylation in ER-positive and -negative BC.

**Abbreviations:** BC, breast cancer; CI, confidence interval; ER, estrogen receptor; M–H, Mantel–Haenszel.

<table>
<thead>
<tr>
<th>Study or subgroup</th>
<th>ER (+) Events</th>
<th>ER (+) Total</th>
<th>ER (-) Events</th>
<th>ER (-) Total</th>
<th>Weight</th>
<th>Odds ratio M–H, fixed, 95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Asif et al(^{19})</td>
<td>50</td>
<td>89</td>
<td>24</td>
<td>39</td>
<td>17.9%</td>
<td>0.80 (0.37, 1.73)</td>
</tr>
<tr>
<td>Caldeira et al(^{24})</td>
<td>15</td>
<td>26</td>
<td>37</td>
<td>45</td>
<td>14.0%</td>
<td>0.29 (0.10, 0.88)</td>
</tr>
<tr>
<td>Hu et al(^{15})</td>
<td>2</td>
<td>10</td>
<td>1</td>
<td>5</td>
<td>1.3%</td>
<td>1.00 (0.07, 14.64)</td>
</tr>
<tr>
<td>Karray-Chouayekh et al(^{27})</td>
<td>23</td>
<td>47</td>
<td>13</td>
<td>31</td>
<td>9.8%</td>
<td>1.33 (0.53, 3.31)</td>
</tr>
<tr>
<td>Li et al(^{13})</td>
<td>103</td>
<td>134</td>
<td>48</td>
<td>54</td>
<td>19.4%</td>
<td>0.42 (0.16, 1.06)</td>
</tr>
<tr>
<td>Shinozaki et al(^{27})</td>
<td>53</td>
<td>114</td>
<td>27</td>
<td>37</td>
<td>26.7%</td>
<td>0.32 (0.14, 0.73)</td>
</tr>
<tr>
<td>Sunami et al(^{22})</td>
<td>54</td>
<td>65</td>
<td>53</td>
<td>65</td>
<td>11.0%</td>
<td>1.11 (0.45, 2.74)</td>
</tr>
<tr>
<td><strong>Total (95% CI)</strong></td>
<td><strong>485</strong></td>
<td><strong>276</strong></td>
<td></td>
<td></td>
<td>100%</td>
<td><strong>0.62 (0.43, 0.87)</strong></td>
</tr>
</tbody>
</table>

Total events 300 203
Heterogeneity: \( \chi^2 = 9.80, df = 6 \) (P = 0.13); I\(^2\) = 39%
Test for overall effect: Z = 2.72 (P = 0.007)

**Figure 4** Forest plot for CDH1 hypermethylation in HER2-positive and -negative BC.

**Abbreviations:** BC, breast cancer; CI, confidence interval; M–H, Mantel–Haenszel.

<table>
<thead>
<tr>
<th>Study or subgroup</th>
<th>HER2 (+) Events</th>
<th>HER2 (+) Total</th>
<th>HER2 (-) Events</th>
<th>HER2 (-) Total</th>
<th>Weight</th>
<th>Odds ratio M–H, fixed, 95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Caldeira et al(^{24})</td>
<td>5</td>
<td>8</td>
<td>47</td>
<td>63</td>
<td>7.9%</td>
<td>0.57 (0.12, 2.65)</td>
</tr>
<tr>
<td>Hu et al(^{15})</td>
<td>0</td>
<td>3</td>
<td>2</td>
<td>10</td>
<td>2.3%</td>
<td>0.49 (0.02, 12.93)</td>
</tr>
<tr>
<td>Karray-Chouayekh et al(^{27})</td>
<td>4</td>
<td>13</td>
<td>30</td>
<td>51</td>
<td>16.8%</td>
<td>0.31 (0.08, 1.16)</td>
</tr>
<tr>
<td>Li et al(^{13})</td>
<td>10</td>
<td>30</td>
<td>109</td>
<td>142</td>
<td>50.3%</td>
<td>0.15 (0.06, 0.36)</td>
</tr>
<tr>
<td>Sunami et al(^{22})</td>
<td>20</td>
<td>29</td>
<td>68</td>
<td>78</td>
<td>22.7%</td>
<td>0.33 (0.12, 0.91)</td>
</tr>
<tr>
<td><strong>Total (95% CI)</strong></td>
<td><strong>83</strong></td>
<td><strong>344</strong></td>
<td></td>
<td></td>
<td>100%</td>
<td><strong>0.26 (0.15, 0.44)</strong></td>
</tr>
</tbody>
</table>

Total events 39 256
Heterogeneity: \( \chi^2 = 2.93, df = 4 \) (P = 0.57); I\(^2\) = 0%
Test for overall effect: Z = 4.97 (P < 0.00001)

**Figure 5** Forest plot for CDH1 hypermethylation in PR-positive and -negative BC.

**Abbreviations:** BC, breast cancer; CI, confidence interval; PR, progesterone receptor; M–H, Mantel–Haenszel.

<table>
<thead>
<tr>
<th>Study or subgroup</th>
<th>PR (+) Events</th>
<th>PR (+) Total</th>
<th>PR (-) Events</th>
<th>PR (-) Total</th>
<th>Weight</th>
<th>Odds ratio M–H, fixed, 95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Asif et al(^{19})</td>
<td>51</td>
<td>85</td>
<td>23</td>
<td>43</td>
<td>24.5%</td>
<td>1.30 (0.62, 2.73)</td>
</tr>
<tr>
<td>Caldeira et al(^{24})</td>
<td>15</td>
<td>23</td>
<td>37</td>
<td>48</td>
<td>16.7%</td>
<td>0.56 (0.19, 1.66)</td>
</tr>
<tr>
<td>Hu et al(^{15})</td>
<td>2</td>
<td>9</td>
<td>1</td>
<td>6</td>
<td>1.9%</td>
<td>1.43 (0.10, 20.44)</td>
</tr>
<tr>
<td>Karray-Chouayekh et al(^{27})</td>
<td>20</td>
<td>47</td>
<td>15</td>
<td>28</td>
<td>21.7%</td>
<td>0.64 (0.25, 1.65)</td>
</tr>
<tr>
<td>Li et al(^{13})</td>
<td>92</td>
<td>120</td>
<td>59</td>
<td>68</td>
<td>35.2%</td>
<td>0.50 (0.22, 1.14)</td>
</tr>
<tr>
<td><strong>Total (95% CI)</strong></td>
<td><strong>284</strong></td>
<td><strong>193</strong></td>
<td></td>
<td></td>
<td>100%</td>
<td><strong>0.76 (0.49, 1.15)</strong></td>
</tr>
</tbody>
</table>

Total events 180 135
Heterogeneity: \( \chi^2 = 3.69, df = 4 \) (P = 0.45); I\(^2\) = 0%
Test for overall effect: Z = 1.30 (P = 0.19)
CI 0.63–1.88, \( z = 0.30, P = 0.77, I^2 = 41\%, P = 0.16 \) (Figure 7).

The frequency of \( CDH1 \) hypermethylation was similar between BC patients with positive lymph node metastasis and those with negative lymph node metastasis (OR = 1.21, 95% CI 0.59–2.47, \( z = 0.53, P = 0.60, I^2 = 71\%, P = 0.007 \) (Figure 8).

The assessment of quality of the included articles was done using the Newcastle–Ottawa Quality Assessment Scale.\(^\text{18}\) This scale allocates a maximum of 9 points for the quality of selection, comparability, exposure, and outcomes for study participants. The Newcastle–Ottawa scale scores ranged from 0 to 9 (a score of 7 or more indicates a good quality). Of the studies, three scored 9 points, three scored 8 points, and five scored 7 points. Hence, the studies were of a relatively high quality (data not shown). We further conducted analyses to determine the result stability in the choice of a single study; the finding suggested no single study had the effect on the pooled ORs, indicating the stability of our analyses. There were no publication biases in the meta-analysis according to the largely symmetric funnel plots (Figure 9).

### Discussion

The \( CDH1 \) gene encodes E-cadherin protein, which is a 120 kDa glycoprotein consisting of an extracellular domain of five tandem repeated domains, a cytoplasmic domain, and a single transmembrane domain.\(^\text{19,20}\) The extracellular domain binds to cadherin on adjacent cells and forms cell–cell adhesion, whereas the cytoplasmic domain binds to catenin and activates the Wnt signaling cascade.\(^\text{21}\) \( CDH1 \) hypermethylation is one of the mechanisms that cause gene silencing and results in reduced E-cadherin expression. Lack of E-cadherin leads to the dysfunction of the cell–cell adhesion system, resulting in a loss of cell–cell adhesion, release of cytoplasmic \( \beta \)-catenin, increase in Wnt signaling, and increased tumor invasiveness.\(^\text{22}\) \( CDH1 \) hypermethylation and its reduced expression have been observed in several of malignancies, including gastric, ovarian, lung, and BC.\(^\text{23–26}\) In the present study, we compared the frequency of \( CDH1 \) hypermethylation between IDC and normal breast tissues. Our results showed \( CDH1 \) promoter hypermethylation was significantly higher in IDC than in normal tissues.
CDH1 is a tumor suppressor gene whose expression is downregulated during epithelial-to-mesenchymal transition (EMT). EMT is an important process in embryonic development and also plays a critical role in tumor progression. Loss of E-cadherin, a protein that mediates cell-cell adhesion, is a hallmark of EMT and is associated with increased metastasis potential and decreased overall survival.

Several studies have investigated the association between CDH1 hypermethylation and clinicopathologic parameters in breast cancer (BC). In a pooled analysis of 11 studies including 940 patients, our data showed that CDH1 hypermethylation was significantly associated with higher frequency of lymph node metastasis in BC, with a high-power sample compared with previous studies.

We investigated CDH1 methylation status in ER-positive versus -negative BC in the present study, showing significantly higher frequency of CDH1 hypermethylation in ER-negative BC than in ER-positive BC. Interestingly, a few studies reported that loss of β-catenin membranous expression and nuclear accumulation of β-catenin are associated with ER-negative status and reduced CDH1 expression, along with aggressive tumor phenotype and poor patient outcome in BC. CDH1 methylation is one of the mechanisms to regulate E-cadherin expression in the ER-negative subtype of BC. In addition, HER2 is a transmembrane tyrosine kinase receptor, located on chromosome 17q21. HER2, encoded by the ERBB2/HER2 oncogene, belongs to a family of epidermal growth factor receptor family members (EGFRs) structurally related to EGFR.

This oncogene is overexpressed in 20%–30% of BC and has been recognized as a marker of poor prognosis, including increased metastasis potential and decreased overall survival. Inhibition of HER2 activity with antibodies has been applied to treat HER2-positive BC. In present study, the frequency of CDH1 hypermethylation was significantly higher in lymph node-positive BC than in lymph node-negative BC, with a high-power sample compared with previous studies.

CDH1, as a suppressor gene, is a potential novel drug target, and its hypermethylation could be reversed through demethylation. Inhibitors of DNA methyltransferase (DNMTs), such as 5-Aza-CdR and 5-fluoro-deoxycytidine, have been applied to human lung cancer and BC cells, and currently 5-fluoro-2′-deoxycytidine is in clinical trials for the treatment of BC and other solid tumors.

The ER is responsible for estrogen-induced mitogenic signaling in epithelial cells in the breast and plays a crucial role in BC onset and progression. About 70% of BCs are ER-positive. The ER is responsible for estrogen-induced mitogenic signaling in epithelial cells in the breast and plays a crucial role in BC onset and progression. When estradiol (E2) binds to ER, the ER undergoes conformational changes and forms dimers, which attract coactivators and corepressors. This complex regulates hundreds of genes involved in a variety of processes, including proliferation, differentiation, survival, invasion, and metastasis. The ER status of breast provides prognostic information but more importantly, is a predictor of response to endocrine therapy. We investigated CDH1 methylation status in ER-positive versus -negative BC in the present study, showing significantly higher frequency of CDH1 hypermethylation in ER-negative BC than in ER-positive BC. Interestingly, a few studies reported that loss of β-catenin membranous expression and nuclear accumulation of β-catenin are associated with ER-negative status and reduced CDH1 expression, along with aggressive tumor phenotype and poor patient outcome in BC. CDH1 methylation is one of the mechanisms to regulate E-cadherin expression in the ER-negative subtype of BC. In addition, HER2 is a transmembrane tyrosine kinase receptor, located on chromosome 17q21. HER2, encoded by the ERBB2/HER2 oncogene, belongs to a family of epidermal growth factor receptors (EGFRs) structurally related to EGFR.

This oncogene is overexpressed in 20%–30% of BC and has been recognized as a marker of poor prognosis, including increased metastasis potential and decreased overall survival. Inhibition of HER2 activity with antibodies has been applied to treat HER2-positive BC. In present study, the frequency of CDH1 hypermethylation was significantly higher in lymph node-positive BC than in lymph node-negative BC, with a high-power sample compared with previous studies.

Abbreviations: BC, breast cancer; CI, confidence interval; M–H, Mantel–Haenszel.
higher in HER2-negative BC than in HER2-positive BC. Our results showed that CDH1 hypermethylation was increased in HER2-negative and ER-negative BCs, respectively, but was not significantly associated with PR status. In the future, CDH1 hypermethylation should be investigated in triple-negative BCs, which are HER2-negative, ER-negative, and PR-negative. As we know, about 85% of triple-negative BCs are basal-like subtypes, which are associated with poor clinical outcome. CDH1 hypermethylation could contribute to the development of a new treatment strategy.

Figure 9 Funnel plots for publication bias.

Notes: (A) CDH1 methylation in IDC and normal breast tissue; (B) CDH1 methylation in ER-positive and -negative BC; (C) CDH1 methylation in HER2-positive and -negative BC; (D) CDH1 methylation in PR-positive and -negative BC; (E) CDH1 methylation in different grades of BC; (F) CDH1 hypermethylation in different stages of BC; and (G) CDH1 hypermethylation in lymph node-positive and -negative metastasis of BC.

Abbreviations: BC, breast cancer; ER, estrogen receptor; IDC, invasive ductal carcinoma; PR, progesterone receptor; OR, odds ratio.
for triple-negative BC, especially in the basal-like subtypes. Finally, our study only selected the published articles and did not include some relevant unpublished papers, which may have resulted in certain publication bias. Therefore, the results should be interpreted carefully.

Conclusion
In conclusion, CDH1 hypermethylation was shown to be significantly higher in IDC than in normal breast tissue, indicating that CDH1 is a potential novel drug target in the development of personalized therapy. CDH1 hypermethylation is strongly associated with ER-negative and HER2-negative BC, respectively, suggesting CDH1 methylation status could contribute to the development of novel therapeutic approaches of ER-negative and or HER2-negative BC, which is a subtype of BC with poor patient outcome.

Disclosure
The authors report no conflicts of interest in this work.

References