Effects of exendin-4, a glucagon like peptide-1 receptor agonist, on neutrophil count and inflammatory cytokines in a rat model of endotoxemia

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Background: Sepsis remains a major cause of morbidity and mortality. A variety of strategies targeting modulation of the pro-inflammatory response associated with early sepsis have been reported without clinical success. GLP-1 enhances glucose-stimulated insulin secretion. In addition, it was shown to have anti-inflammatory effects. We hypothesized that treatment with exendin-4, a GLP-1 receptor agonist, would attenuate inflammation and improve glucose control in a lipopolysaccharide (LPS) rat model of inflammation.

Methods: Two-month-old male Wistar rats were randomly assigned to one of the following four groups: 1) treatment: intraperitoneal (IP) injection of LPS 10 mg/kg followed by exendin-4, 30 μg/kg, 10 minutes later; 2) control-1: IP injection of LPS 10 mg/kg, followed by normal saline (NS); 3) control-2: IP NS injection followed by exendin-4; 4) sham: IP injection of NS followed by another NS injection. Glucose concentration, total white blood count with absolute neutrophil count, and pro- and anti-inflammatory cytokine concentrations were measured at 0, 3, 6, and 10 hours following LPS injection.

Results: At 3 hours, rats injected with LPS developed neutropenia, elevated pro- and anti-inflammatory cytokines, and mild hypoglycemia. Treatment with exendin-4 significantly modulated neutropenia, and decreased pro-inflammatory cytokine concentrations (IL-1α, IL-1β, IL-6, TNFα, and IFNγ). However, exendin-4 had no effect on IL-10 concentrations. LPS injection led to mild hypoglycemia, that was not observed in rats treated with exendin-4. Sham animals exhibited no significant change from baseline in all parameters.

Conclusion: In this LPS model of acute early phase inflammation, treatment with exendin-4 decreased pro-inflammatory cytokine concentrations without changing IL-10 blood levels and improved neutropenia. Following LPS injection, rats developed a tendency toward hypoglycemia that improved with exendin-4. Overall our data suggest that exogenous exendin-4 mediates anti-inflammatory effects early in this rat model of endotoxin-induced inflammation.

Keywords: glucagon like peptide-1, exendin-4, sepsis, endotoxemia, inflammation, neutrophils

Background

Sepsis remains a significant health problem and a major cause of morbidity and mortality.1,2 Although both pro-inflammatory and anti-inflammatory responses begin rapidly with sepsis, the initial response in patients with severe sepsis is typified by an overwhelming hyper-inflammatory phase mediated by a variety of pro-inflammatory cytokines.3,4

Over the last 15 years an increasingly robust body of literature has described the deleterious effects of hyperglycemia in a variety of clinical contexts including sepsis.
Glycemic control using insulin infusion has provided conflicting results with earlier studies showing benefits with decreased mortality in subgroups of adult patients, but subsequent studies showing no benefit and potential harm associated with complications, most importantly hypoglycemia.5–7

GLP-1 is a tissue-specific post-translational proteolytic product of the proglinucagon gene, released from intestinal L-cells in response to nutrient ingestion that enhances glucose-stimulated insulin secretion.8 GLP-1 is one of the major incretin hormones in humans and has been studied extensively in association with type 2 diabetes.9,10

The biologic effects of GLP-1 and GLP-1 receptor agonists (GLP-1RA) in association with glucose control are well studied and include: 1) stimulation of glucose-dependent insulin secretion; 2) inhibition of glucagon and stimulation of somatostatin secretion; 3) stimulation of β-cell proliferation and neogenesis and inhibition of β-cell apoptosis, thereby increasing β-cell mass; 4) activation of the expression of immediate early genes encoding transcription factors that regulate islet cell proliferation and differentiation.11 GLP-1 was shown to be an effective treatment for type 2 diabetic patients, and its analogs are currently in clinical use in this group of patients.10,12

The GLP-1 receptor (GLP-1R) is well characterized, cloned, and sequenced. It is expressed in a wide range of organs including α-, β-, and δ-cells of the pancreatic islets, lung, heart, kidney, stomach, intestine, pituitary, skin, nodose ganglion neurons of the vagus nerve, and several regions of the central nervous system including the hypothalamus and brainstem.13,14 With this wide organ distribution it is likely that GLP-1 plays a role in organs apart from the pancreas.

Anti-inflammatory properties of GLP-1 have been studied in vitro, with promising results in cells derived from a variety of organs (lung, myocardium, neurons).15–17

In vivo animal studies showed a wide range of anti-inflammatory effects of GLP-1RA in the cardiovascular system, endothelium, intestine, kidneys, and the brain.18–23

More recently, a study among humans with obesity and type 2 diabetes mellitus reported decreased levels of pro-inflammatory cytokines in patients treated with exenatide (another GLP-1RA).24

Exendin-4 (Ex-4) is a specific GLP-1RA purified from Heloderma suspectum venom.25,26

The current study was designed to evaluate the effects of Ex-4, on inflammation and glycemic control in a rat model of endotoxin (lipopolysaccharide [LPS])-induced inflammation. We hypothesized that treatment with Ex-4, would reduce inflammation and improve glycemic control without adverse effect of hypoglycemia.

Materials and methods

Animals

Two-month-old male Wistar rats (Harlan Laboratories, Indianapolis, IN, USA) weighing 200–250 g, were used in all experiments. Rats were housed under specific pathogen free conditions using a standard 12:12 hour light/dark cycle and received regular diet and water ad libitum. Experiments were conducted at least 5 days following transfer to allow acclimatization.

Inflammation protocol

Rats were divided into four groups: the treatment group received a 10 mg/kg intraperitoneal (IP) injection of Escherichia coli LPS (from E. coli, EH100, Sigma-Aldrich Co, St Louis, MO, USA), followed by a 30 μg/kg IP injection of Ex-4 (Sigma-Aldrich Co) 10 minutes later (LPS/Ex-4 group, n=11). One control group, received the IP LPS injection followed by a normal saline (NS) injection 10 minutes later (LPS/NS, n=6). To rule out any effects associated with the Ex-4, another control group received an IP NS injection followed by a 30 μg/kg injection of Ex-4 (NS/Ex-4, n=6). A sham group received an IP NS injection followed by another NS injection 10 minutes later (NS/NS, n=4). Blood samples for all tests were collected from the lateral vein tail at 0, 3, 6, and 10 hours following initial injection. Ten hours after LPS or NS injection, rats were euthanized using pentobarbital overdose IP injection. All animal studies were approved by the Institutional Animal Care and Use Committee at the University of Washington.

Biochemical analyses

Plasma glucose

Plasma glucose concentrations were measured with a glucometer (Ascensia Contour, Bayer AG, Leverkusen, Germany).

Complete blood count

Complete blood count was obtained using a Coulter counter (Hemavet 950FS; Drew Scientific Inc., Oxford, CT, USA). Leukocyte differential was confirmed with blood smears and manual counting.

GLP-1 assays

Blood samples for GLP-1 were collected in a tube with ethylenediaminetetraacetic acid (EDTA) and dipeptidyl peptidase IV inhibitor (EMD Millipore, Billerica, MA, USA).
Samples were cooled on ice, fractionated by centrifugation at 10,000×g for 5 minutes, and the plasma stored at −20°C. GLP-1 concentrations were measured using total active GLP-1 assay kit (Meso Scale Discovery, Gaithersburg, MD, USA).

Cytokine assays
Blood for cytokine analyses was maintained at room temperature for 10 minutes to allow clotting. Subsequently it was fractionated by centrifugation at 10,000×g for 5 minutes. Resultant serum was stored until analysis at −20°C. Cytokines’ concentrations were measured using the multiplex beads – Discovery assay (Eve Technologies Corporation, Calgary, CA, USA).

Ex-4
Serum Ex-4 concentrations were measured using an enzyme-linked immunosorbent assay (ELISA) kit (Phoenix Pharmaceuticals, Burlingame, CA, USA).

Statistics
All numerical data are presented as group means with standard error of the mean (SEM). P-values for glucose levels and absolute neutrophil count (ANC) were determined using Student’s two-tailed t-test at the stated time points. To derive P-values for cytokine levels we used a Student’s t-test and two-way analysis of variance using Bonferroni’s multiple comparison test. Statistical analysis was performed using a program supplied by GraphPad Software Inc., La Jolla, CA, USA.

Results
Clinical changes
Rats in the LPS/NS group developed behavior changes including lack of grooming, reduced mobility, eating bedding, and attempted to hide. Treatment group (LPS/Ex-4) exhibited minimal changes (mostly lack of grooming and reduced mobility). Control groups had no behavioral changes.

Glucose concentrations
Surprisingly rats in the LPS/NS group, showed no signs of hyperglycemia, a common feature of human sepsis associated with stress mediated gluconeogenesis. In fact, the LPS/NS group, exhibited a tendency toward hypoglycemia. Treatment with Ex-4 (LPS/Ex-4 group) provided no statistically significant impact on glucose concentrations. However, there was a trend toward normoglycemia (glucose concentration of 70–130 mg/dL) in the treatment group especially at 3 and 6 hours. Interestingly, rats treated with Ex-4 only (NS/Ex-4), exhibited a tendency toward mild hyperglycemia. Sham group (NS/NS) showed no significant change in glucose concentrations from baseline (Figure 1A).

Neutrophil counts
Mean ANC was 3.8 (range 2.38–4.97)×10³ cells/µL at baseline with no significant differences between groups. Sham (NS/NS) rats and rats treated with Ex-4 only (NS/Ex-4) exhibited no significant changes from baseline throughout the experiment. Rats injected with LPS/NS developed significant neutropenia with minimal recovery at 10 hours, when compared to sham rats (NS/NS) (P<0.001 at 3 and 6 hours, P<0.01 at 10 hours). Treatment with Ex-4 (LPS/Ex-4 group) significantly improved neutrophil

Figure 1 Longitudinal changes in glucose levels and ANCs.
Notes: (A) Longitudinal changes in serum glucose concentrations in the rat LPS model of sepsis. (B) Longitudinal changes in absolute neutrophil count (ANC) in the rat LPS model of sepsis. Values are group means ± SD. Comparison done at each given time point (0, 3, 6, and 10 hours). # refers to LPS/NS (blue line) versus LPS/Ex-4 (red line); † refers to LPS/NS (blue line) versus NS/NS (black line); * and ‡; P<0.05; ‡‡; P<0.01; ‡‡‡; P<0.001.
Abbreviations: LPS, lipopolysaccharide; NS, normal saline; Ex-4, Exendin-4; SD, standard deviation.
counts at 3, 6, and 10 hours post-injection ($P<0.05$ at 3 hours). ANC in the treatment group (LPS/Ex-4) was not statistically different from the sham rats (NS/NS) (Figure 1B).

**Inflammatory cytokines**

**IL-1α**

IL-1α was elevated at 6 hours following LPS injection in the control group (LPS/NS) compared to sham rats. Treatment with Ex-4 decreased IL-1α significantly ($P<0.05$).

**IL-1β**

IL-1β was elevated at 3, 6, and 10 hours following LPS injection compared to the sham group. Treatment with Ex-4 reduced IL-1β concentrations. However, it reached statistical significance only at 6 hours ($P<0.05$) mostly due to wide standard deviation in the LPS group.

**IL-6**

LPS injection led to elevated IL-6 blood levels at 3 hours with a trend back toward baseline at 10 hours. Levels were significantly greater than the sham group at 3 hours ($P<0.001$) and at 6 and 10 hours ($P<0.01$). Treatment with Ex-4 decreased IL-6 levels at 3 and 10 hours ($P<0.05$), and at 6 hours ($P<0.01$).

**TNFα**

TNFα levels peaked at 3 hours post-LPS injection with return to baseline at 10 hours. Treatment with Ex-4 decreased TNFα levels at 3 hours ($P<0.001$), and at 6 hours ($P<0.05$). Sham and NS/Ex-4 groups demonstrated no change in TNFα levels throughout the experiment.

**IFNγ**

IFNγ levels peaked at 6 hours post-LPS injection. Treatment with Ex-4 decreased IFNγ levels significantly ($P<0.05$). IFNγ remained undetectable in the sham (NS/NS) and Ex-4 only (NS/Ex-4) groups.

**IL-10**

In contrast to a variety of pro-inflammatory cytokines, IL-10 levels were significantly increased at 3, 6, and 10 hours, in both control and treatment groups compared to sham and NS/Ex-4 groups ($P<0.01–0.05$). However, there was no difference between control and treatment groups (Figure 2).

**GLP-1 concentrations**

Plasma GLP-1 levels ranged from 15 to 68 pg/mL with no significant change in levels between groups and time course. Of note, rats had free access to food, but experiments took place during light hours, a time they typically fast.

**Ex-4 concentrations**

Ex-4 levels were non-detectable at time 0 in all groups. Levels remained undetectable at all time points in the sham and LPS/NS groups. Rats treated with Ex-4 (LPS/Ex-4 or NS/Ex-4) had mean levels of 2–5 ng/mL at 3 hours returning to 0 at 6 and 10 hours (data not shown).

**Discussion**

In this study, we report that treatment with Ex-4 decreases pro-inflammatory cytokine levels, with no effects on IL-10 levels in the early phase of LPS-induced endotoxemia. We also show attenuation of neutropenia which was prominent in this rat model of early sepsis. To our best knowledge, this is the first study examining the effect of a GLP-1RA in a rat model of LPS-induced inflammation. Elevated pro-inflammatory markers, especially in early sepsis, indicate excessive inflammatory host response and are correlated with severe sepsis and worse outcome.27,28

Systemic inflammatory response syndrome is defined as systemic inflammatory response to a variety of severe clinical insults. The response is manifested by two or more of the following conditions: hypo/hyperthermia, tachycardia, tachypnea, and leukocytosis or neutropenia.25 IL-1 and TNFα are strong pro-inflammatory cytokines, released shortly after the inciting event, and are early regulators of the immune response. They are also strong pyrogenic cytokines.4 IL-6 is produced in response to stimulation with LPS, IL-1, and TNFα. It has a variety of biological effects including activation of T and B lymphocytes and the coagulation system. It is also a strong pyrogenic cytokine.4 In our study, Ex-4 attenuated levels of these pro-inflammatory cytokines at the very early phase of inflammation. There is likely a role for drugs that block inflammatory cytokines in sepsis; however, such agents should be short acting, applied early in sepsis, and used only in patients who have substantially elevated pro-inflammatory cytokines.3

Leukocytosis is the usual response to bacterial infection. Neutropenia in septic animals and humans, may be secondary to enhanced leukosequestration at the site of infection (site of LPS injection in our model) or within the lungs, as an antecedent to sepsis-associated acute lung injury, and is associated with worse outcome. In our rat model, treatment with Ex-4 improved ANC throughout the experiment period, indicating an additional favorable effect of GLP-1RA in this rat model.
GLP-1RA have been approved by the US Food and Drug Administration (FDA) for treatment for type 2 diabetic patients. A recent study demonstrated the anti-inflammatory effects of GLP-1RA in obese humans with type 2 diabetes. The benefits of better glycemic control without the risk of hypoglycemia and the potential to reduce inflammation led us to assess the effects of treatment with Ex-4, a GLP-1RA, in this in vivo rat model of LPS-induced inflammation. One of our goals was to evaluate the effects of Ex-4 on hyperglycemia, a common manifestation of the stress response in critically ill patients. Glycemic control can be achieved with insulin infusion, but this intervention has the real risk of hypoglycemia. GLP-1RA has been shown to reduce insulin needs and decrease glucose levels in diabetic patients following major surgery. In this study, all rats administered with LPS, developed hypoglycemia or remained in the lower range of normoglycemia. Although no statistically significant differences were found between the groups, rats treated with Ex-4 exhibited a tendency toward higher (more normal) glucose levels. Hypoglycemia was reported in rat models of endotoxemia, thought to be associated with mitochondrial phosphoenolpyruvate carboxykinase. Lower levels of IL-1 and TNFα associated with treatment with Ex-4, may have helped to prevent hypoglycemia seen in untreated rats in our study but further investigation is needed to better understand the underlying mechanism. Rats treated with NS/Ex-4 demonstrated an increase in glucose levels at 1 hour with marginally high glucose levels at 3 and 6 hours. This phenomenon was described before. It is reasonable to suggest that these effects of Ex-4 are not dependent on its interaction with GLP-1R only.
GLP-1RA might offer the potential for safe glucose control without risks for hypoglycemia among critically ill patients, including those with sepsis, however, this rat model of endotoxemia failed to demonstrate this phenomenon frequently seen in humans. There are currently two FDA-approved GLP-1RA analogs, liraglutide and exenatide (synthetic Ex-4), being used for type 2 diabetes. Additionally, there are many other GLP-1RA in advanced stages of development (albiglutide, taspoglutide, lixisenatide, CJC-1134) likely to be approved in the near future.

These drugs are potential candidates for glycemic control in septic human patients with low risk for hypoglycemia.

We recognize several limitations in this experimental study. This LPS model is a sterile form of systemic inflammatory response syndrome rather than full clinical model of sepsis. Results should be interpreted accordingly and further studies using cecal ligation and puncture or other clinical models of sepsis are needed. Our study was limited to 10 hours follow-up and focused on the initial phase of sepsis. We did not explore potential mechanisms for Ex-4 effects. We did not compare different doses of Ex-4. In contrast to humans, rats did not demonstrate significant hyperglycemia in this model. Hence our ability to assess the effects of Ex-4 on glucose homeostasis in this sepsis model is limited.

**Conclusion**

Taken together, the results of our study, indicate that Ex-4 a GLP-1RA reduces inflammation and improves neutropenia during early phase of endotoxemia. Additional animal in vivo studies using a bacterial model of sepsis are needed to establish effects of this treatment on mortality. Further experiments are needed to define mechanisms by which Ex-4 is operating to modulate the inflammatory response. A different animal model, or direct human application will be needed to assess effects of GLP-1RA on glycemic control in sepsis.

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**Disclosure**

The authors have no conflicts of interest to disclose in this work.

**References**


