Synthesis, antimicrobial and in vitro antitumor activities of a series of 1,2,3-thiadiazole and 1,2,3-selenadiazole derivatives

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Abstract: Three derivatives of substituted 1,2,3-thia- or 1,2,3-selenadiazole (4a–c) were prepared and characterized by different chemical techniques. These compounds were evaluated for their antimicrobial and antitumor activities. Compounds 4a (propenoxide derivative), 4b (carbaldehyde derivative), and 4c (benzene derivative) were active against the yeast-like fungi Candida albicans. Compound 4a was active against gram-negative Escherichia coli, and compound 4c was active against the gram-positive Staphylococcus aureus. For the antitumor activity, both compounds 4b and 4c were active against all tested tumor cell lines, namely, SW480, HCT116, C32, MV3, HMT3522, and MCF-7. The activity of compound 4c was greater than that of compound 4b and more than that of the reference antitumor 5-flourouracil against the SW480, HCT116, and MCF-7 tumor cell lines. In conclusion, a number of the prepared 1,2,3-thia- or 1,2,3-selenadiazole compounds showed promising antifungal, antibacterial, and in vitro antitumor activities. Further investigations are required to explore the mechanism by which active compound are inducing their cytotoxicity.

Keywords: thiadiazole, selenadiazole, cell lines, antimicrobial activity

Introduction

Microbes are highly diverse, and their genetic makeup is subjected to continuous wavering in response to environmental settings.1 Human pathogens, both nosocomial and enterobacter species, are the most common infectious pathogens.2,3 The induction of highly resistant pathogens urges researchers to develop new antimicrobial agents from natural sources.3 Heterocyclic compounds containing sulfur and selenium are attractive due to their interesting biological and synthetic applications. 1,2,3-Thia- and 1,2,3-selenadiazole derivatives are well-known, and the organic synthesis of their intermediates have attracted the attention of researchers.4 Numerous substituted 1,2,3-thia- and 1,2,3-selenadiazole derivatives have been prepared, and most of them have showed noticeable antibacterial activities against Bacillus subtilis and Escherichia coli. The antifungal activity of 1,2,3-thia- and 1,2,3-selenadiazole against Aspergillus niger, Cryptococcus neoformans, and Candida albicans has also been determined.5–7 It has been found that the introduction of the 1,2,3-selenadiazole ring to known molecules with biological activity will modify and even improve their biological activity. Other heterocyclic compounds containing triazole, oxazole, benzoazole, quinazoline, pyridazoline, pyridazine, pyrazole, and thiazole have been found to be biologically active substances.8–10

Heterocyclic ring systems are present in numerous antiparasitic, antifungal, anthelmintic, and anti-inflammatory drugs.11 The beta-lactam antibiotics derivatized with a 1,2,3-thiadiazole-5-mercapto moiety have been found to be active against...
Materials and methods

Chemistry

The melting points (mps) were determined on an electrothermal digital melting point apparatus. The solvents were purified by standard procedures. Infrared (IR) spectra were recorded (in cm⁻¹) using a Nicolet Impact 410 FTIR Spectrometer (Nicolet Instrument Corp, Madison, WI, USA).

The IR spectra of pure substances were measured as KBr pellets. Proton nuclear magnetic resonance (¹H-NMR) spectra were recorded on a 400 MHz Fourier transform nuclear magnetic resonance (FT-NMR) system (JNM-ECP400; JEOL, Ltd, Peabody, MA, USA) with a 300 MHz Bruker AC 200 spectrometer Bruker Corporation, Billerica, MA, USA). Tetramethylsilane was used as internal reference. The spectral data were reported in delta (δ) units relative to the tetramethylsilane reference line. Carbon-13 nuclear magnetic resonance (¹³C-NMR) spectra were recorded on the 100 MHz JEOL FT-NMR system, (75 MHz). Mass spectrometry was carried out by using the Finnigan MAT95XP mass spectrometer (Thermo Fisher Scientific Inc, Waltham, MA, USA) (field desorption [FD]: 5 kV ionizing energy).

4-((1,2-Propanoic acid-3-yl) acetophenone (2a)

A solution of (3.20 mmol) 4-(2-propenoic acid) acetophenone (1a) in (12 mL) dry dichloromethane cooled in an ice bath to 0°C was added dropwise into a solution of (4.32 mmol) m-chloroperoxybenzoic acid in (12 mL dry dichloromethane) at 0°C under vigorous stirring. After complete addition, the whole solution was left stirring at room temperature for 6 hours and under reflux for further 3 hours. The precipitated m-chlorobenzoic acid was removed from solution by simple filtration. The filtrate was washed many times with 10% NaOH followed by distilled water and then dried over magnesium sulfate. After removing the solvent, through evaporation, to dryness, the residue was purified on silica gel column chromatography using ethanoll/dichloromethane (1:30) as eluent. This compound was obtained as light brown viscous oil in 53% yield (2.1 g); ¹H NMR (CDCl₃): δ = 2.27 (s, 3H, CH₃), 2.61 (q, ³JHH=4.3 Hz, 1H, CH₂-epoxide), 2.92
1-(1,2,3-Selenadiazole-4-yl)carbaldehyde (3a)

A solution of 4-(1,2-propenoxide-3-yl)acetophenone (2a) (2.50 mmol) and ethoxycarbonyl hydrazine (2.60 mmol) in ethanol (20 mL) was added to the product solution. This mixture was refluxed for 20 minutes under reflux. The product was then added to the reaction mixture and then heated for 20 minutes under reflux. The product was then heated for 20 minutes under reflux. The product was then added to the reaction mixture and then heated for 20 minutes under reflux. The product was then added to the reaction mixture and then heated for 20 minutes under reflux. The product was then added to the reaction mixture and then heated for 20 minutes under reflux. The product was then added to the reaction mixture and then heated for 20 minutes under reflux. The product was then added to the reaction mixture and then heated for 20 minutes under reflux. The product was then added to the reaction mixture and then heated for 20 minutes under reflux. 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vigorously stirring. The solution was kept in dark, and the reaction progress was followed up with TLC that showed reaction completion in approximately 20 hours. Then, 1M H₂SO₄ (10 mL) was added and the solution was gently heated to 40°C for 5 hours. After reducing the volume of the reaction mixture to two-thirds of its original volume under vacuum, distilled water (30 mL) was added, and chloroform (3x50 mL) was used for extraction. The combined chloroform layers were dried using magnesium sulfate. The crude product was obtained after solvent removal under vacuum. This compound was obtained as dark brown solid in 85% yield with mp 227°C (decomposition); ¹H NMR (CDCl₃): δ=9.83 (s, 1H, CHO), 8.98 (s, 1H, CHSe) ppm; ¹³C NMR (CDCl₃): δ=136.7.2 (1C, C-Se), 158.3 (1C, C=O), 192.3 (1C, CHO) ppm; IR (KBr): ν =3,057, 1,628, 1,476, 1,412, 1,263, 1,218, 959 cm⁻¹; MS: (5 kV, FD) m/z (%) 161 (100%). Anal Calcd For C₃H₂N₂OSe: C, 22.37; H, 1.24; N, 17.39; Se, 49.04. Found: C, 22.27; H, 1.75; N, 17.41.

1,3,5-Tris(1,2,3-thiadiazole-4-yl)benzene (4c)

This compound (Figure 3) was prepared according to the procedure described, and obtained as pale yellow powder in 93% yield with mp 227°C (decomposition); ¹H NMR (CDCl₃): δ=8.92 (s, 3H, CH-phenyl), 9.83 (s, 3H, CHS) ppm.

Microbiology

All bacterial strains were obtained from American Type Culture Collection (ATCC) (Manassas, VA, USA). The filter paper disk and the well diffusion methods were applied to measure the inhibitory activity as indicated by the diameter of the inhibition zone. The well method was carried out as follows: a 100 µL of the heterocyclic compounds at each concentration were placed in 5 mm-diameter compounds with nutrient agar that was inoculated spontaneously with the tested pathogens. The plates were incubated at 37°C for 48 hours. The clear zones around the wells were measured as inhibition zones. No clear zone around the well was indicated as a negative activity. The filter paper disk method was carried out as follows: Whatman® III filter paper puncture disks (5 mm) were saturated with different concentrations of the tested heterocyclic compounds. They were dried in an oven at 60°C for 12 hours. The dried disks were placed on already cultured nutrient agar plates with the different pathogens. The diameter of clear zone around the disks was measured after 48 hours of incubation at 37°C. No clear zone around the disks indicated negative activity. In both methods, dimethyl sulfoxide (DMSO) was used as a solvent to prepare the heterocyclic compound solutions.

Well diffusion assay

Antibacterial activity of the chemical compounds was tested against several gram-positive and gram-negative bacteria, and Candida by well diffusion method. The antibacterial susceptibility tests were performed on Muller–Hinton agar medium. Briefly, 20 mL agar medium was poured into the plates to obtain uniform depth. The standard inoculum suspensions (10⁶ cfu/mL) were streaked over the surface of the media using a sterile cotton swab to ensure the confluent growth of the organism. Each compound was dissolved in 5% DMSO to obtain stock solution concentrations. Then, 5 mm-diameter wells were prepared in the medium and filled with 100 µL of each compound. Chloramphenicol (100 µg/mL) and cyclohexamide (100 µg/mL) were used as positive references against the bacteria and Candida, respectively, and 5% DMSO was used as negative control. Finally, the inoculated plates were incubated at 37°C for 24 hours, and the inhibition zones were observed, including the diameter of the well (5 mm). All the experiments were carried out in triplicate.

Determination of minimum inhibitory concentration (MIC)

The MIC values were determined for the bacterial strains that were sensitive to the compounds in the well diffusion assay. The MIC of the compounds was tested in Mueller–Hinton broth by broth macrodilution method. The inoculation of the bacterial strains was prepared from 4-hour-old broth cultures, and the suspensions were adjusted to standard turbidity (10⁶ cfu/mL). The compounds were dissolved in 5% DMSO to obtain 20 mg/mL stock solution. The stock solution

Figure 3 Compound 4c. Chemical structure of 1,3,5-Tris(1,2,3-thiadiazole-4-yl)benzene.

Abbreviation: 4c, benzene derivative.
was diluted with Muller–Hinton broth to give different concentrations (12, 25, and 50 µg/mL). Finally, bacteria were incorporated with the chemical compounds and Muller–Hinton broth to give a final concentration of 10⁷ cfu/mL. The control tubes contained bacteria with chloramphenicol (30 µg/mL), cyclohexamide (10 µg/mL), and 5% DMSO were used as control. The culture tubes were incubated at 37°C for 24 hours. The lowest concentrations that did not show any growth of the test organism on the culture medium after macroscopic evaluation was determined as MIC.

### Antitumor activity

The acute cytotoxic effects of the compounds on a panel of cancer cell lines, namely, breast (HMT3522, MCF7), colorectal (SW480, HCT116), and melanoma (MV3, C32) were determined using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay.²¹ Briefly, cells were cultured in phenol red–free Dulbecco’s Modified Eagle’s Medium (DMEM) containing 10% fetal bovine serum (Bio Whittaker, Verviers, Belgium) containing the antimicrobial agents amphotericin B and penicillin/streptomycin (PAA Laboratories, GmbH, Pasching, Austria). Cells were seeded at 5,000 per well onto flat-bottomed 96-well culture plates and allowed to grow for 24 hours before the desired treatment. Cells were then incubated with a wide range of the compounds concentrations (0–250 µg/mL) for 72 hours. The antimitabolite anticancer agent 5-fluorouracil (5-FU), at 50 µg/mL, was used as a control. Cells were then labeled with MTT (at a final concentration of 1.2 mM) from the Vybrant MTT Cell Proliferation Assay Kit (Molecular Probes; Grand Island, NY, USA) according to the manufacturer’s instruction, and resulting formazan was solubilized with DMSO. Absorbance was read on a microplate reader (ELx800, Bio-tek instruments, plate reader, Highland Park, VT, USA) at 540 nm.

### Statistical analysis

Data were analyzed using GraphPad PRISM version 5 (La Jolla, CA, USA). All data points were presented as (mean ± standard deviation) of three independent experiments. One-way analysis of variance (ANOVA) followed by Tukey’s post hoc test were used for statistical analysis. P<0.05 was considered significant.

### Results

1,2,3-Thiadiazole derivative 4a was prepared from the corresponding ketone 1a, as described previously but with a different methodology, as shown in Figure 1. The synthetic procedure started from ketone 1a that was prepared following a specific procedure. Ketone 1a was transformed into the corresponding epoxide derivative 2a through oxidation with m-chloroperbenzoic acid. The hydrazone derivative 3a was obtained from the condensation reaction between compound 2a and ethyl carbazate. The 1,2,3-thiadiazole derivative 4a was obtained from the reaction of the hydrazone 3a with thionyl chloride, as described by Li.²² 1,2,3-Selenadiazole derivative 4b was prepared from the corresponding methylglyoxal 1b, which was transformed into the corresponding methylglyoxal bis(semicarbazone) (2b) through condensation with semicarbazide. The 1,2,3-selenadiazole derivative 3b was obtained from the reaction of the methylglyoxal bis(semicarbazone) (2b) with selenium dioxide in glacial acetic acid, as previously described²³ and shown in Figure 2.

The antimicrobial activity of the three synthetic compounds was evaluated against gram-positive Enterococcus faecalis and Staphylococcus aureus, and gram-negative Salmonella typhi, E. coli, and P. aeruginosa, as well as against C. albicans, a fungal pathogen. The sensitivity test of the three compounds indicated their highly antagonistic activity against the selected microbes except P. aeruginosa. At 10 µg/mL concentration of each of the three compounds,

### Table 1 Antimicrobial activity of compounds 4a–c, represented by the diameter of inhibition zone

<table>
<thead>
<tr>
<th>Synthetic compound</th>
<th>Microorganism</th>
<th>Zone diameter (mm)</th>
<th>MIC (µg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>5 µg</td>
<td>10 µg</td>
</tr>
<tr>
<td>4a</td>
<td>Candida albicans</td>
<td>15</td>
<td>16</td>
</tr>
<tr>
<td></td>
<td>Escherichia coli</td>
<td>0</td>
<td>11</td>
</tr>
<tr>
<td></td>
<td>Enterococcus faecalis</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>Staphylococcus aureus</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>4b</td>
<td>C. albicans</td>
<td>19</td>
<td>25</td>
</tr>
<tr>
<td></td>
<td>E. coli</td>
<td>14</td>
<td>21</td>
</tr>
<tr>
<td></td>
<td>E. faecalis</td>
<td>0</td>
<td>16</td>
</tr>
<tr>
<td></td>
<td>S. typhi</td>
<td>10</td>
<td>12</td>
</tr>
<tr>
<td></td>
<td>S. aureus</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>4c</td>
<td>C. albicans</td>
<td>0</td>
<td>12</td>
</tr>
<tr>
<td></td>
<td>E. coli</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>E. faecalis</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>S. typhi</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>S. aureus</td>
<td>0</td>
<td>10</td>
</tr>
<tr>
<td>Chloramphenicol</td>
<td>E. coli</td>
<td>0</td>
<td>25</td>
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<tr>
<td></td>
<td>E. faecalis</td>
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<td>18</td>
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<tr>
<td></td>
<td>S. typhi</td>
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</tr>
<tr>
<td></td>
<td>S. aureus</td>
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<td>23</td>
</tr>
<tr>
<td>Cycloheximide</td>
<td>C. albicans</td>
<td>22</td>
<td>25</td>
</tr>
</tbody>
</table>

**Abbreviations:** MIC, minimum inhibitory concentration; 4a, propenoxide derivative; 4b, carbaldehyde derivative; 4c, benzene derivative; NA, not applicable.
the antimicrobial activity was higher than 5 µg/mL, as indicated in Table 1. At 10 µg/mL, compound 4B inhibited the growth of C. albicans, E. coli, E. faecalis, and S. typhi, with 25, 21, 16 and 12 mm inhibition zones, respectively; S. aureus was resistant to compound 4b. Compound 4c had inhibitory activity only against S. aureus. Compound 4a was active against C. albicans, and E. Coli with 11 and 16 mm inhibition zones, respectively. The antimicrobial activity of these compounds at 10 µg/mL was similar to antimicrobial activity of 5 µg/mL of chloramphenicol or 5 µg/mL of cyclohexamide and the antifungal reference compounds. These results proved the promise of these synthetic compounds as antimicrobial agents. The MIC of the three synthetic compounds against reference microbes varied from 0.625 to 6.25 µg/mL, as shown in Table 1 and Figure 4.

The antitumor activity of the three synthetic compounds against SW480, HCT116, C32, MV3, HMT3522, and MCF-7 cell lines showed that compound 4b was active against all tumor cell lines, with variable half-maximal inhibitory concentration (IC₅₀) values ranging from 52.17 to 114.79 µg/mL. Compound 4c was more potent than compound 4b against all tumor cell lines, as shown in Table 2. The IC₅₀ of compound 4c varied from 31.91 to 528.10 µg/mL. These results showed that compound 4c had higher antitumor activity than did compound 4b. Compound 4a had no antitumor activity against any of the selected tumor cell lines. Comparing the IC₅₀ of compound 4c with the reference antitumor compound 5-FU, the IC₅₀ against SW480, HCT116, and MCF-7 was lower, indicating its higher potential to inhibit tumor cell proliferation. In addition, results showed that HMT3522 breast cancer cells were highly resistant to both compounds 4b and 4c. The rest of the tumor cell lines were more sensitive to compound 4c than to compound 4b.

**Discussion**

The results of the present study showed the antimicrobial and antitumor activities of the synthetic derivatives (4a, 4b, and 4c), with promising potential in comparison with the active reference chemicals. Variation in antimicrobial activity, as indicated by the diameters of the inhibition zones of the three compounds, could have been due to variation in the genetic makeup of the microbial reference strains used in this study, variation in compounds metabolic activity, or compounds with different resistance or different sensitivity.
Compound 4b showed higher antimicrobial activity of over compound 4c but lower antitumor or cytotoxicity in comparison with compound 4c. This could have been due to their active sites on the living cells, especially the genetic makeup, or could have been due to the different molecular weight and different solubility in water and hence, differences in diffusion rate in the culture medium.

In summary, new derivatives of substituted 1,2,3-thia- or 1,2,3-selenadiazole (4a–c) were prepared and characterized. These compounds were evaluated for their antimicrobial and antitumor activity. Compounds 4a–c were found to be active against the yeast-like fungi *C. albicans*. Compound 4a was active against gram-negative *E. coli*, and compound 4c was very active against the gram-positive *S. aureus*. The antitumor activity of the three synthetic compounds against different tumor cell lines (SW480, HCT116, C32, MV3, HMT3522, and MCF-7) showed that both compounds 4b and 4c had an antitumor activity against all the tumor cell lines. Furthermore, the activity of compound 4c was greater than those of compound 4b and the reference antitumor 5-FU, against SW480, HCT116, and MCF-7 tumor cell lines.

Notably, investigations of the specific mechanism of cytotoxicity of the tested compounds (4a–c) are beyond the scope of the current study, and it is certainly the subject of our future work. This study was carried out with the rationale of investigating the antibacterial and antitumor activities of synthesized 1,2,3-thia- and 1,2,3-selenadiazole derivatives. The results have shown various activities of the tested compounds, as described earlier.

### Acknowledgments

We are grateful to the Deanship of Scientific Research of the Jordan University of Science and Technology, for financial support (grant number 226/2009).

### Disclosure

The authors report no conflicts of interest in this work.

### Table 2 Antiproliferative activities of compounds 4a–c against different cancer cell lines

<table>
<thead>
<tr>
<th>Synthetic compound</th>
<th>SW480</th>
<th>HCT116</th>
<th>C32</th>
<th>MV3</th>
<th>HMT3522</th>
<th>MCF-7</th>
</tr>
</thead>
<tbody>
<tr>
<td>5-FU</td>
<td>48.01±3.6</td>
<td>42.61±3.9</td>
<td>41.71±3.9</td>
<td>53.41±4.8</td>
<td>58.87±4.9</td>
<td>49.31±4.1</td>
</tr>
<tr>
<td>4a</td>
<td>&gt;2.000</td>
<td>&gt;2.000</td>
<td>&gt;2.000</td>
<td>&gt;2.000</td>
<td>&gt;2.000</td>
<td>&gt;2.000</td>
</tr>
<tr>
<td>4b</td>
<td>52.17±4.2*</td>
<td>55.28±4.1*</td>
<td>75.39±5.8*</td>
<td>114.79±9.6*</td>
<td>495.87±29*</td>
<td>96.91±6.3*</td>
</tr>
<tr>
<td>4c</td>
<td>31.91±2.8*</td>
<td>33.65±3.1*</td>
<td>68.56±6.3*</td>
<td>70.86±8.1*</td>
<td>528.10±32*</td>
<td>35.71±4.2*</td>
</tr>
</tbody>
</table>

**Notes:** Data shown represent the mean ± SE of three independent experiments. One-way ANOVA followed by Tukey’s post hoc. *Significant difference from 5-FU (P<0.05); $Significant difference from 4b (P<0.05).

**Abbreviations:** 5-FU, 5-fluorouracil; ANOVA, analysis of variance; IC₅₀, half-maximal inhibitory concentration; SE, standard error; 4a, propenoxide derivative; 4b, carbamoyl aldehyde derivative; 4c, benzene derivative.

### References