Embryonic–maternal cross-talk via exosomes: potential implications

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Abstract: A myriad of locally produced factors into the microenvironment of the reproductive tract is regulated, not one-way but rather, through embryonic–maternal cross-talk. In this mini-review, we focused on the exosomes, which are cell-derived vesicles of 30–100 nm in diameter, as a communicating language facilitating this dialog. These nanovesicles are secreted from pre-implantation embryos, oviduct epithelium, and endometrium as well as from the placenta, and contain proteins, messenger RNA (mRNA), microRNA, and DNA cargoes, and have pleiotropic effects on both embryonic and maternal environments. A better understanding of the molecular mechanisms mediating this cross-talk will lead to the development of new regulating agents, with novel diagnostic, biological, and therapeutic potential for either supporting or hindering the normal reproductive functions.

Keywords: embryo, endometrium, placenta, mRNA, miRNA

Exosomes

An ever growing number of studies worldwide have helped to substantiate the essential functions of the cell-secreted, membrane-derived vesicles, particularly exosomes, and provided new dimensions for the concept of intercellular signaling. Exosomes are nanosized vesicles (30–100 nm in diameter) (Figure 1A and B) that contain, not only proteins, but also, messenger RNAs (mRNAs), microRNAs (miRNAs), and double-strand or genomic DNA.1–6 The molecular cargoes carried by exosomes affect cellular activity via ready-made proteins and miRNA or by translation of transferred mRNAs (Figure 1C). The term “exosomal shuttle RNA (esRNA)” was proposed for those transferred RNAs.3

Exosomes, as cell membrane-derived nanovesicles, are specifically equipped to mediate intercellular communication, via the transfer of genetic information to recipient cells. As a result, exosomes play a fundamental biological role in the regulation of normal physiological as well as aberrant pathological processes, via altered gene regulatory networks and via epigenetic programming.2 For example, exosome-mediated genetic transfer can regulate the maintenance of stem cell plasticity and induce beneficial cell phenotype modulation.7 Alternatively, such vesicles play a role in tumor pathogenesis8 and the spread of neurodegenerative diseases, via transfer of specific miRNAs and pathogenic proteins.9 In addition, these cell membrane-derived vesicles are involved in cell adhesion and signal transfer, and provide an important means of cell communication.10 Evidence of secretion of exosomes has been reported in most cell types, including embryonic stem cells and in vitro–produced embryos.11–16
These exosomes were shown to act as a mediator, carrying proteins and RNAs between embryos that developed to the blastocyst stage and those that failed to develop from the morula to blastocyst stage, deemed degenerate embryos. miR-25, miR-302c, and miR-645 were found to be more highly concentrated in media from failed IVF/ICSI cycles. miR-191, miR-372, and miR-645 were found to be more highly concentrated in media from failed IVF/ICSI cycles. miRNAs have also been found to exist in embryo culture medium, in both human and domestic animals.Embryo-derived exosomes In study of the physical properties, including size and concentration, of in vitro fertilized (IVF) embryos-derived exosomes, it was revealed that their numbers increase with developmental stage and further, that their size correlates with embryo quality and may predict recovery from apparent growth-arrested embryos. Furthermore, in previous work, we showed the uptake of embryo-derived exosomes by cultured embryos. These exosomes were shown to act as a mediator, carrying early reprogramming mRNAs, such as Oct4, Sox2, cMyc, and Klf4, which improved the development of the cocultured embryos in group culture system. We suggest that the continuous transfer of mRNA cargoes among cultured embryos via exosomes is advantageous over the acute transfer of mRNA by the conditioned medium—this confirms the concept of the "dynamic microenvironment" or "niche" among the cultured embryos. It was suggested that the stability of foreign mRNAs in cells is often tightly and intricately regulated with low transcriptional rates. The foreign transferred mRNAs are rapidly turned over with half-lives of 20–40 minutes. In our preliminary work, we found expression of sex determining mRNAs, Xist and Sry, in the conditioned medium of in vitro–derived embryos cultured individually (unpublished data), which could be used for sexing of in vitro–produced embryos.

In equines, a previous in vitro study suggested that exosomes can be secreted by day 8 embryos, which can modulate the functions of the oviduct epithelium through transfer of early pregnancy factor (HSP10) and miRNAs.

miRNAs, small (~22 nucleotides) noncoding RNAs that regulate gene expression, have been implicated in a wide array of biologic processes, including early embryo development and stem cell differentiation. Human blastocysts express miRNAs, which may be important to their survival. Differential miRNA expression between euploid and aneuploid embryos may be an early indicator of their prognosis or a mechanism behind their eventual fate. Interestingly, miRNAs have also been found to exist in embryo culture medium, in both human and domestic animals.

The expression of miRNA in culture medium has been correlated to fertilization method, chromosomal status, and pregnancy outcome, which makes miRNA a potential biomarker for predicting IVF success. Some miRNAs, including miRNA-191, have been shown to be putative noninvasive candidates for predicting aneuploidy, while miR-191, miR-372, and miR-645 were found to be more highly concentrated in media from failed IVF/non–intracytoplasmic sperm injection (ICSI) cycles. Additionally, miRNAs were found to be more highly concentrated in ICSI and day 5 media samples when compared to regularly inseminated and day 4 samples, respectively. Furthermore, it was shown that differential miRNA gene expression analysis could be used to early discriminate between embryos that developed to the blastocyst stage and those that failed to develop from the morula to blastocyst stage, deemed degenerate embryos. miR-25, miR-302c, miR-196a2, and miR-181a expression was found to be higher concentration in ICSI day 5 media samples when compared to regularly inseminated and day 4 samples, respectively. Additionally, miRNAs were found to be more highly concentrated in ICSI and day 5 media samples when compared to regularly inseminated and day 4 samples, respectively. Furthermore, it was shown that differential miRNA gene expression analysis could be used to early discriminate between embryos that developed to the blastocyst stage and those that failed to develop from the morula to blastocyst stage, deemed degenerate embryos. miR-25, miR-302c, miR-196a2, and miR-181a expression was found to be higher
in degenerate embryos compared to blastocyst embryos. Therefore, expression of miRNAs in in vitro culture media could allow for the development of biological markers for selection of better quality embryos and for subsequent successful pregnancy.

Furthermore, embryonic and fetal genomic DNA was also found to circulate in the maternal blood. This raises many questions about the involvement of exosomes/microvesicles in transportation of embryonic and fetal genomic DNA and may mean that these exosomes can act as pot-mers, specifically, for the Y-chromosome specific DNA; for embryo/fetal sexing as early as 6 weeks of gestation or as a noninvasive prenatal genetic diagnostic method to detect aneuploidy in the embryo and fetus.

**Maternal-derived exosomes**

On the other hand, exosomes can be secreted from maternal side; bioinformatic analysis of endometrium-derived exosome miRNAs revealed that these miRNAs have potential targets in biological pathways highly relevant for embryo implantation. Western blot analysis demonstrated that in vivo–derived exosomes contain HSP70 and OVGP, which have roles in fertilization and early pregnancy; however, in vitro–derived exosomes contain only HSPA8 and MYH9. They are also secreted from the endometrium and have been found to contain miRNAs for ovine enJSRV-envelope, HSP70, interleukins, and IFN-regulatory factors, thereby stimulating trophectoderm cells to proliferate and secrete IFNT for successful pregnancy recognition in sheep. These results would explain that free and/or exosomal enJSRV acts on the trophectoderm, via TLRs, to induce the secretion of IFNT.

**Placenta-derived exosomes**

A previous study unraveled the important role of placenta-derived exosomes; the syncytiotrophoblast constitutively and throughout the pregnancy secretes exosomes. More recently, exosomes have been shown to be released by trophoblasts and to carry molecules involved in placental physiology. This involves immunomodulators proteins such as fibronectin and syncytin, Wnt/βcatenin-related molecules, galectin-3, and HLA-G; but also bioactive lipids, such as immunosuppressiive PG_E2; PPARγ ligand 15d-PGJ2; and miRNAs.

Exosomes contain all the ingredients (ie, syncytin, phosphatidic acid, and endosomal phospholipid BMP) to promote intercellular fusion and might behave as initiators of syncytiotrophoblast formation. Moreover, the placenta-derived exosomes are immunosuppressive and carry proteins and RNA molecules that, in a redundant manner, influence a number of mechanisms and promote the fetal allograft survival. Specifically, isolated human placental exosomes carry NKG2D ligands, ULBPs (ULBP1–5), and MIC proteins on their surface by which they induce downregulation of the NKG2D receptor on NK, CD8(+), and gamma delta T cells. This, in turn, leads to reduction of their in vitro cytotoxicity without affecting the perforin-mediated lytic pathway. Moreover, bioactive FasL- and TRAIL-carrying exosomes, which are able to convey apoptosis, are secreted by the placenta and responsible for the immunomodulatory and protective role of the human placenta. Exosomes can also be used for prediction of preeclampsia: Villous syncytiotrophoblast exosomes have been found to be positive for HERV envelope gene proteins and to be rapidly taken up by BeWo choriocarcinoma cells in a syncytin-1- and syncytin-2-dependent manner. Syncytin 2 has also been found to be reduced in women with preeclampsia.

Furthermore, it has been shown that trophoblast-derived exosomes containing miRNAs could mediate cross-talk between the fetoplacental unit and the mother during pregnancy. The function of placenta-derived miRNAs, particularly from the primate-specific chromosome 19 miRNA clusters, which are highly expressed in human placenta and in the serum of pregnant women, has been reviewed.

**Synthetic and semisynthetic exosomes**

The application of synthetic or semisynthetic exosomes for therapeutic drug delivery is still in its infancy. Issues regarding the understanding of exosomes biogenesis, large-scale production, and in vivo interactions need to be addressed to develop successful and cost-effective exosome-based drug delivery systems. Few investigations have explored their potential to serve as a platform for the development of semisynthetic nanovesicles. Given their nanoscale size, potential to express targeting ligands in native conformations, and their deformable structure, exosomes offer a logical biological vesicle platform for adapting and producing semisynthetic vesicles with excellent potential for nanomedicine applications. No doubt, progress in synthetic biology to harnessing exosomes for therapeutic nucleic acid delivery is crucial to mimic the physicochemical properties of naturally occurring exosomes to overcome delivery challenges and to establish robust technology platforms. If successful, the exosomes could then transport the nucleic acid into a subsequent population of cells, including embryos, that manifest the desired effect. This piggy-back mechanism
could be responsible for reported deep-tissue RNA or DNA effects with certain carriers, and offers novel and promising roles in interfering with or supporting the dialog between the embryos and the mother.\(^5^0\)

**Conclusion**

From these novel results, we may suggest a dynamic mutual paracrine communication between both the embryonic and the maternal environments, through the exosomes, at the early stages of preimplantation embryo development. Anticipating the potential implications of the functions of the embryonic- and maternal-derived exosomes is essential. Particularly, the detection and characterization of embryo-derived exosomes could be a noninvasive method for: 1) the judgment of embryo quality and implantation capability; 2) the sexing of the embryos; and 3) the prediction of abnormal genetic constituents of embryos, as an alternative to the invasive prenatal genetic diagnosis (PGD) method. Therefore, advancement in the physicochemical analysis of small quantities of embryo culture medium, including analysis of exosomes size (such as nanoparticle tracking analysis)\(^5^1\) and content of mRNA, miRNA and DNA, is crucial to substantiate these targets.

Finally, much remains to be learned concerning the communicating signals comprising the embryonic–maternal cross-talk through the exosomes, especially considering that their effects are often redundant and pleiotropic. Deepening our understanding of exosomes will be critical in understanding the physiology and the pathophysiology of the early stages of pregnancy recognition. Moreover, this study will enable the development of novel nanodiagnostics and nanotherapeutics, to maximize pregnancy rates during IVF treatment; to improve the treatment of disorders such as infertility, pregnancy loss, and preeclampsia; and possibly even to offer new methods for locally acting contraceptives.

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**Disclosure**

The authors declare that there is no conflict of interest.

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