Functionalized magnetic iron oxide/alginate core-shell nanoparticles for targeting hyperthermia

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Abstract: Hyperthermia is one of the promising treatments for cancer therapy. However, the development of a magnetic fluid agent that can selectively target a tumor and efficiently elevate temperature while exhibiting excellent biocompatibility still remains challenging. Here a new core-shell nanostructure consisting of inorganic iron oxide (Fe₃O₄) nanoparticles as the core, organic alginate as the shell, and cell-targeting ligands (ie, D-galactosamine) decorated on the outer surface (denoted as Fe₃O₄/Alg-GA nanoparticles) was prepared using a combination of a pre-gel method and coprecipitation in aqueous solution. After treatment with an AC magnetic field, the results indicate that Fe₃O₄/Alg-GA nanoparticles had excellent hyperthermic efficacy in a human hepatocellular carcinoma cell line (HepG2) owing to enhanced cellular uptake, and show great potential as therapeutic agents for future in vivo drug delivery systems.

Keywords: hyperthermia, iron oxide, alginate, pre-gel, targeting

Introduction
Superparamagnetic iron oxide nanoparticles (SPIONs), including magnetite (Fe₃O₄) and maghemite (γ-Fe₂O₃), have been extensively utilized in biomedical applications, including as magnetic resonance imaging contrast agents,1 for biocatalysis,2 for biological separation,3 for biosensing,4 diagnostic medical devices,5 and hyperthermia. Among these applications, hyperthermia6 is a physical therapy in which SPIONs generate heat and thus kill cancer cells when they are exposed to an external magnetic field.7 The hyperthermia of SPIONs has been considered as one of the most promising cancer therapies because it can effectively destroy cancer cells while keeping normal cells intact. The reason is that cancer cells are more sensitive to heat than normal cells.8,9 When a high temperature (usually around 42.5°C) is produced locally, tumor tissue cannot release heat energy via the bloodstream due to dysfunction of blood vessels.10 In addition, studies have also shown that cancer cells produce heat shock proteins, resulting in an increase in major histocompatibility complex class I antigens. Thus, this increases the possibility of recognition of cancer cells by cytotoxic T lymphocytes.11 Consequently, hyperthermia is an effective and safe cancer therapy.

One of the challenges for effective cancer therapy is how to deliver therapeutic agents to tumor sites. Most anticancer drugs cannot distinguish between cancer cells and healthy cells, which results in undesirable side effects and low efficacy. The fabrication of nanoscale drug-loading carriers has been widely studied because these nanoparticles can accumulate in tumor sites via the enhanced permeability and retention effect.12 However, such passive targeting-based drug delivery systems sometimes cause unwanted accumulation of anticancer agents in the liver, kidney, and spleen.13,14
In contrast with passive targeting, active targeting has attracted more attention in terms of further enhancing delivery efficacy and cancer specificity. Specific ligands (eg, folate) that can be recognized by cancer cells have been widely studied and linked to the surfaces of anticancer agents so that receptor-mediated endocytosis can occur. For example, asialoglycoprotein receptors are located on liver cancer cells and can specifically bind with glycoprotein on/in galactose residues, leading to internalization and subsequent degradation of glycoprotein inside the cells. Such active targeting strategies can not only effectively deliver drugs into cancer cells, but also reduce the amounts of anticancer agents used.

The combination of hyperthermia and active targeting has been regarded as one of the most promising therapies for improving anticancer efficacy. Synthesis of SPIONs and further functionalization of SPIONs with specific targeting ligands is therefore of intense research interest. So far, SPIONs have been synthesized by a coprecipitation method, hydrothermal treatment, and an organic solvent-based method. For example, Hyeon et al synthesized uniform and well suspended SPIONs in organic solvent systems using Fe(CO)₅ as the precursor. Daou et al reported synthesis of SPIONs with a uniform particle size of 39 nm by hydrothermal treatment. Despite these pioneering works, the coprecipitation method is more acceptable because of its simple synthesis (ie, relatively low temperature and a water system). However, this method usually results in aggregated SPIONs, which are difficult to use in biomedical applications. In addition, in order for SPIONs to be functionalized with targeting ligands, the synthesized SPIONs must be reacted with the targeting ligands under the desired conditions, which is usually a complicated and time-consuming process. Therefore, simultaneous synthesis and functionalization of SPIONs in aqueous systems is necessary.

In this study, we developed a new synthetic approach to prepare carboxylic acid-functionalized SPIONs (ie, Fe₃O₄) by combination of coprecipitation of iron oxides and a pre-gel method of alginate (denoted as Fe₃O₄@Alg nanoparticles). The COOH-functionalized Fe₃O₄ nanoparticles were further linked with a liver cancer cell-targeting ligand (ie, galactose), denoted as Fe₃O₄@Alg-GA nanoparticles, and used as targeted hyperthermia therapy. This new approach has several advantages: alginate has excellent biocompatibility and surface functionality (COOH group); coprecipitation of iron oxides takes place within the alginate polymer, preventing aggregation of the synthesized Fe₃O₄ nanoparticles; targeting ligands can be successfully linked on the external surface of Fe₃O₄@Alg nanoparticles, thereby ensuring targeting capability; and Fe₃O₄ nanoparticles with well dispersed, controllable particle sizes provide the capability for hyperthermia. We then characterized the synthesized Fe₃O₄@Alg nanoparticles with X-ray diffraction, scanning electron microscopy (SEM), Fourier transform infrared spectroscopy (FTIR), and thermogravimetric analysis. The Fe₃O₄@Alg-GA nanoparticles showed enhanced hyperthermic efficacy when compared with Fe₃O₄@Alg nanoparticles, which was confirmed to be the result of increased uptake of Fe₃O₄@Alg-GA by human liver cancer (HepG2) cells (ie, the amount of Fe₃O₄@Alg-GA nanoparticles taken up was 20-fold greater than the amount of Fe₃O₄@Alg nanoparticles).

Materials and methods

Chemicals

Iron(II) chloride tetrahydrate (FeCl₂·4H₂O, 98%) was purchased from Alfa Aesar (Ward Hill, MA, USA). Iron(III) chloride hexahydrate (FeCl₃·6H₂O), 1-(3-dimethylaminopropyl)-3-ethylcarbodimide (EDC), N-hydroxysuccinimide (NHS), 3-(4,5-dimethylthiazol-2- yl)-2,5-diphenyltetrazolium bromide (MTT), potassium ferrocyanide (K₃[Fe(CN)₆]·3H₂O), 10% neutral buffered formalin, nuclear fast red counterstain, and sodium alginate were purchased from Sigma-Aldrich (St Louis, MO, USA). D-(-)-galactosamine hydrochloride was provided by Acros Organics (Thermo Fisher Scientific, Bridgewater, NJ, USA). Phosphate-buffered saline (PBS, 10x), fetal bovine serum, sodium bicarbonate (7.5 wt%), L-glutamine (200 mM in saline), trypsin-ethylenediaminetetraacetic acid (10x), and Pen-Strep solution (penicillin 10,000 U/L and streptomycin 10 mg/mL) were provided by Biological Industries Israel Beit-Haemek Ltd (Beit-Haemek, Israel). CellTiter 96 Aqueous MTS Reagent Power was purchased from Promega (Madison, WI, USA). Phenazine methosulfate was purchased from AppliChem GmbH (Darmstadt, Germany).

Synthesis and functionalization of Fe₃O₄@Alg nanoparticles

The synthesis process for Fe₃O₄@Alg nanoparticles is described as follows. Alginate 10 mg was dissolved in 55 mL of deionized (DI) water with stirring until the alginate was dissolved completely. An aqueous solution containing FeCl₂·4H₂O (50 mM, 10 mL) was added to this system at a rate of 0.5 mL per minute and kept under stirring for 1 hour. Subsequently, another solution containing FeCl₃·6H₂O (62.4 mM, 10 mL) was also added to this system by the same method, and the whole system was stirred for 1 hour.
The Fe cations (Fe²⁺ and Fe³⁺) are selectively adsorbed onto the negatively charged COOH groups of the alginate polymer, which is a so-called pre-gel method. Next, we used an aqueous ammonia solution (1.5 M) to adjust the pH of the system to 10, in order to obtain magnetic Fe₃O₄ nanoparticles, which is a so-called coprecipitation method. After 30 minutes of stirring, the solid product was separated from the solution by a magnetic stone, washed with DI water several times, and then resuspended in DI water by sonication. We report for the first time simple and reproducible synthesis of polymer-coated magnetic nanoparticles by the combination of a pre-gel method and a coprecipitation method. Several factors, including the amount of alginate, pH value, and reaction time, were studied in order to control the particle size of the synthesized Fe₃O₄@Alg nanocomposites (see discussion below).

To functionalize Fe₃O₄@Alg nanoparticles with ligands that can specifically target cancer cells, we reacted the synthesized Fe₃O₄@Alg nanoparticles with galactosamine via the EDC/NHS reaction, because alginate has carboxylic acid groups and galactosamine has amine groups. Typically, the Fe₃O₄@Alg nanoparticle suspension was first diluted with PBS (pH 5) buffer to a concentration of 1.25 mg/mL. Next, EDC 50 mg and NHS 75 mg were separately dissolved in 1 mL of PBS (pH 5) and the two solutions were added into 8 mL of Fe₃O₄@Alg nanoparticle suspension. After the whole mixture had been stirred for 15 minutes, the EDC/NHS-functionalized Fe₃O₄@Alg nanoparticles were separated by a magnetic stone, washed with PBS buffer, and resuspended in 5 mL of PBS buffer (pH 7.5). A PBS (pH 7.5) solution containing galactosamine hydrochloride (52 mM, 5 mL) was added to this solution. After 4 hours of stirring, the galactosamine-functionalized Fe₃O₄@Alg nanoparticles (denoted as Fe₃O₄@Alg-GA nanoparticles) were separated by a magnetic stone, washed with PBS buffer (pH >9) several times (the basic buffer was used here to deprotonate the carboxylic acid groups of alginate), and resuspended in DI water by sonication.

Characterization

The hydrodynamic diameter and zeta potential of the synthesized Fe₃O₄@Alg nanoparticles were measured using a Zetasizer Nano ZS system (Malvern Instruments Ltd, Malvern, UK) at 25°C with DI H₂O as the solvent. Samples were sonicated for 1 hour before measuring. The average particle size was evaluated based on dynamic light scattering. The morphology of the samples was observed using an SEM (Nova NanoSEM™, FEI, Hillsboro, NJ, USA). The Fe₃O₄@Alg nanoparticle suspension was sonicated for 1 hour, dropped onto copper grids (200 mesh, carbon-coated), dried under vacuum, and subjected to platinum coating before observation under SEM. Functional groups of the samples were examined by FTIR (Spectrum 100, PerkinElmer, Waltham, MA, USA) at a resolution of 4 cm⁻¹. Samples used for FTIR measurements were prepared by mixing the vacuum-dried samples with KBr (KBr to sample, 100:1). The mixture was then ground extensively and pressed into a translucent disc. The structural properties of the samples were analyzed using an X-ray diffraction system (Ultima IV, Rigaku, Tokyo, Japan) with Cu Kα radiation (λ =1.5418 Å, 40 kV, 40 mA). Thermogravimetric analysis curves were recorded on a differential scanning calorimeter (Pyris 1, PerkinElmer). Samples (5 mg) were maintained at 50°C for 10 minutes, then heated to 800°C at 10°C per minute, and maintained at 800°C for 1 minute. Nitrogen flow was fixed at 20.0 mL per minute throughout the measurement process. The amount of iron element inside the cells was determined using an inductively coupled plasma mass spectrometer (ICP-MS, Elan-6000, PerkinElmer) after lysis of the cells. The cells were cultured as mentioned before. The cells were transplanted into 24 wells with 5×10⁴ per well for 1 day. After washing with PBS several times, 0.5 mL of solution containing Fe₃O₄@Alg nanoparticles or Fe₃O₄@Alg-GA nanoparticles (1 mg/mL) was added into each well and the cells were cultured for another 4 hours. Each sample of HCl (12 M, 0.6 mL) was added into each well for 2 hours to dissolve the Fe₃O₄ nanoparticles. Next, 600 µL of HCl (6 M) was used to wash the wells. In total, 6 mL of solution (five wells for each sample) was collected into a vial for measurement; one well in which cells cultured without nanoparticles was used to count the number of cells. The weight of Fe₃O₄ in each well as measured by ICP-MS and was divided by the number of cells to get the average weight of Fe₃O₄ nanoparticles inside each cell.

Cell culture, viability test, and observation

The liver cancer (HepG2) cell line was purchased from the National Health Research Institutes, Miaoli, Taiwan. HepG2 cells were incubated in flasks with Dulbecco’s Modified Eagle’s Medium at 37°C and 5% CO₂, in a 95% humidified atmosphere, and were subcultured every 3 days. Every 100 mL of Dulbecco’s Modified Eagle’s Medium was supplemented with 10 mL of fetal bovine serum, 2 mL of NaHCO₃, 1 mL of L-glutamine, and 1 mL of Pen–Strep solution (penicillin 10,000 U/mL and streptomycin 10 mg/mL). Cell viability was measured using an MTT assay. HepG2
cells were seeded onto 96-well plates at a density of $2 \times 10^4$ cells per well and allowed to attach overnight. The medium was then removed, and each well was washed twice with 200 µL of PBS. Media containing samples with various concentrations were added to each well, and the cells were incubated at 37°C for 24 hours. The medium was then removed, and the wells were washed twice with 0.2 mL of PBS. To each well, 20 µL of MTT solution (5 mg/mL in the DI water) was added, and the cells were incubated for an additional 4 hours. The medium was then replaced with 150 µL of dimethyl sulfoxide. The plates were left stationary for 4 hours to dissolve the blue crystals, and the absorbance was recorded by a microplate reader at a wavelength of 570 nm.

Cell viability was expressed as the average absorbance of treated samples relative to untreated ones. HepG2 cells were seeded onto four-well Lab-Tek slides at a density of $1 \times 10^5$ cells per well. After incubation at 37°C with 5% CO$_2$ overnight, the medium was removed, and the slides were washed twice with PBS. To each well, 0.25 mL of DI water containing Fe$_3$O$_4$@Alg samples (1 mg/mL) and 0.25 mL of Dulbecco’s Modified Eagle’s Medium were added. After incubation for an additional 4 hours, the supernatant was removed, and each well was washed twice with 0.2 mL of Dulbecco’s Modified Eagle’s Medium. The in vitro effects of the hyperthermia treatments were evaluated by a proliferation assay. HepG2 cells were first cultured as described in the section on cell culture, viability test, and observation. A total of $1 \times 10^5$ cells were harvested in 24 wells for 1 day; 500 µL of suspension containing Fe$_3$O$_4$@Alg nanoparticles or Fe$_3$O$_4$@Alg-GA nanoparticles at a concentration of 0.5 mg/mL was then added to each well. The cells were cultured for another 4 hours, and then trypsinized and moved to a centrifuge tube. Fresh culture medium was added into the centrifuge tube to achieve 0.4 mL as the total volume. Three centrifuge tubes, including one control sample (without Fe$_3$O$_4$@Alg nanoparticles) and two experimental samples (with Fe$_3$O$_4$@Alg and Fe$_3$O$_4$@Alg-GA nanoparticles), were put in the copper coil at the same time, and the heating time was set to 20 minutes. In order to compare the effects of the magnetic field, cells (both with and without Fe$_3$O$_4$@Alg nanoparticles and Fe$_3$O$_4$@Alg-GA nanoparticles) without treatment by a magnetic field were also prepared. After treatment, 100 µL of each sample was taken from the centrifuge tube and added to 96-well plates. The cells were further cultured at 37°C with 5% CO$_2$ for 18 hours. Next, 20 µL of MTS solution was added to each well, and the cells were incubated for an additional 4 hours. Absorbance at 450 nm was recorded using an enzyme-linked immunosorbent assay plate reader. Cell viability was expressed as the average absorbance of treated samples relative to untreated ones.

**Plasma clotting time and red blood cell hemolysis**

The anticoagulant activity of the synthesized Fe$_3$O$_4$@Alg was evaluated by testing clotting time in human plasma. A 160 µL volume of 100% platelet-poor plasma solution was mixed
with each solution containing Fe$_3$O$_4$@Alg nanoparticles (0.5 mg/mL, 46 µL) in a 96-well plate. The solution was then recalcified by addition of calcium chloride solution (1 M, 4 µL) and shaken for 30 seconds at 37°C. Plasma clotting time was determined as the time at which the onset of the absorbance transition occurred by reading the absorbance at 660 nm using a PowerWave™ microplate spectrophotometer (Bio-Tek, Winooski, VT, USA). Each clotting time is reported as the average value of repeated measurements of six samples.

Disruption of red blood cell membranes was estimated to determine the non-fouling nature of the synthesized Fe$_3$O$_4$@Alg nanoparticles using a hemolysis assay. Red blood cells were isolated by blood centrifugation and washed three times with a 0.15 M saline solution. In each hemolysis experiment, 10$^8$ red blood cells were suspended in 500 µL of PBS. Preparation of 500 µL of red blood cell solution was followed by addition of 500 µL of solution containing Fe$_3$O$_4$@Alg, which was prepared in PBS at a concentration of 20 mg/mL. The Fe$_3$O$_4$@Alg nanoparticles and red blood cell mixtures were incubated in a 37°C water bath for 1 hour. The mixed solution was then centrifuged for 5 minutes at 2,000 rpm to separate intact red blood cells and disrupted membranes from the solution. Absorbance of the supernatant containing the released hemoglobin was then measured at 541 nm using the PowerWave microplate spectrophotometer. One hundred percent hemolysis was determined by measuring the absorbance of 1×10$^8$ red blood cells with complete lysis by suspending them in DI water. The control was 1×10$^8$ red blood cells in PBS. Each hemolysis is reported as the average value of three repeated measurements with 18 samples.

**Results and discussion**

**Synthesis and characterization of Fe$_3$O$_4$@Alg nanoparticles**

The synthesis process is shown in Figure 1. Both FeCl$_2$ and FeCl$_3$ solutions were added slowly to an alginate solution at various concentrations to induce the pre-gel state of alginate (ie, Fe$^{2+}$ and Fe$^{3+}$ ions interact with the carboxylate groups of alginate). An aqueous ammonia solution was then added to increase the pH value for formation of iron oxides (ie, coprecipitation) in alginate polymer (denoted as Fe$_3$O$_4$@Alg nanoparticles). Because alginate consists of d-mannuronic acid (M) and l-guluronic acid (G) with highly negative charges that can interact strongly with cations such as Fe$^{2+}$ and Fe$^{3+}$, the pre-gel state of alginate represents the egg-box structure of alginate-Fe$^{2+}$/Fe$^{3+}$, resulting in formation of nanoparticles. Complete precipitation of Fe$_3$O$_4$ is expected at basic pH values. Here we studied the effects of different pH values and amounts of alginate on the hydrodynamic diameter of the synthesized Fe$_3$O$_4$@Alg nanoparticles. As shown in Figure 2A, Figure S1, and Table S1, as the amount of alginate increased (from 10 to 40 mg), the hydrodynamic diameter of the synthesized Fe$_3$O$_4$@Alg nanoparticles also increased for all three different pH conditions. In addition, the pH in the final solution also influenced the hydrodynamic diameter, ie, particle sizes were decreased when pH increased. This is

![Figure 1 Synthesis of Fe$_3$O$_4$@Alg-GA nanoparticles for magnetic fluid hyperthermia. Abbreviations: Fe$_3$O$_4$, iron oxide; Alg, alginate; GA, galactosamine.](https://www.dovepress.com/)

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because $\text{Fe}_3\text{O}_4$ with smaller particle sizes can be synthesized at higher pH values, as shown in Figure 2B. Here we chose the experiment parameters of 10 mg of alginate and pH 10 for obtaining the optimal particle size for intravenous injection. The role of alginate is to homogeneously distribute the synthesized $\text{Fe}_3\text{O}_4$ nanoparticles. The SEM images in Figure 2C clearly show the different morphology of samples prepared without (left) and with (right) alginate. In the presence of alginate 10 mg, $\text{Fe}_3\text{O}_4$ displayed excellent separation owing to the abundant COOH groups of alginate, in contrast with the
marked aggregation of $\text{Fe}_3\text{O}_4$ nanoparticles when they were synthesized in the absence of alginate. In Table 1, the sizes of $\text{Fe}_3\text{O}_4$ with and without alginate were 5.629±2.257 nm and 122±29.5 nm, respectively, which again indicates that alginate can prevent aggregation of $\text{Fe}_3\text{O}_4$. On the other hand, body temperature is about 37°C and the temperature of hyperthermia is 42.5°C. The hydrodynamic diameters of nanoparticles at both body temperature and hyperthermia temperature are also important. The hydrodynamic diameters of $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles are 129±31.5 nm at 37°C and 275±77.2 nm at 42.5°C. It seems that $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles would remain at a similar particle size at 37°C but aggregate slightly at 42.5°C. Slight aggregation of particles could be accepted for the in vitro experiment; however, the problem of aggregation at 42.5°C should be overcome in order to further apply our samples for in vivo experiment. TEM images clearly show successful alginate coating on $\text{Fe}_3\text{O}_4$ nanoparticles (Figure 2D). After coating with alginate, the surface charge of the samples was changed. For $\text{Fe}_3\text{O}_4$ nanoparticles, the surface potential was 16.1 mV and for $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles was –60.1 mV. The negative charge on the $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles resulted from the abundant COOH groups on alginate. The abundance of carboxylic groups provided a functional group for post-modification. Galactosamine is an asialoglycorprotein receptor containing an amine group which can react with the carboxylic group from alginate via EDC and NHS. At first the carboxylic group of alginate was reacted with EDC to form a highly unstable O-acylisourea, and then the NHS molecule was added to stabilize this compound. Finally, NHS-O-acylisourea, which has high reactivity toward primary amines, reacted with the amine group from galactosamine forming an amide bond between alginate and galactosamine. Post-modification of $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles with galactosamine reduced the negative potential significantly; however, the particle sizes of $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles and $\text{Fe}_3\text{O}_4@\text{Alg-GA}$ nanoparticles were similar with each other, meaning that the particle size of $\text{Fe}_3\text{O}_4@\text{Alg}$ would not be affected by modification of galactosamine.

The synthesized $\text{Fe}_3\text{O}_4$, $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles, and GA-functionalized $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles were characterized with X-ray diffraction, FTIR, thermogravimetric analysis, and SQUID. As shown in Figure 3A, the X-ray diffraction patterns for $\text{Fe}_3\text{O}_4$ and alginate indicated a crystalline phase and an amorphous phase for $\text{Fe}_3\text{O}_4$ and alginate, respectively. The characteristic peaks at 2θ=20–70 were the (200), (310), (400), (511), and (440) of $\text{Fe}_3\text{O}_4$ cubic spinel structure phase. It is worth noting that the synthesized $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles exhibited the same characteristic peaks as the above-mentioned $\text{Fe}_3\text{O}_4$, indicating that the synthetic method used in this study (ie, the combined coprecipitation and pre-gel methods) did not alter the crystalline phase of iron oxide. Further evidence of the formation of $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles was provided by FTIR. As shown in Figure 3B, the FTIR spectrum of $\text{Fe}_3\text{O}_4@\text{Alg}$ showed both of the characteristic bands from $\text{Fe}_3\text{O}_4$ and alginate. The stretch vibrations of Fe–O at 577 cm⁻¹ and 648 cm⁻¹ were observed for $\text{Fe}_3\text{O}_4$, and the stretch vibrations of C–O at 1,046 cm⁻¹ and 1,112 cm⁻¹ were representative of C–O groups for alginate, the band at 1,316 cm⁻¹ was the C–O stretch vibration of the COOH group, and the peaks at 1,628 cm⁻¹ and 1,429 cm⁻¹ were the asymmetry and symmetry stretching vibrations, respectively, of the COOH group for pure alginate. In order to calculate the amount of alginate in the $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles, we used thermogravimetric analysis for all three samples. As shown in Figure 3C, samples were heated up to 800°C, and the remaining weight percentages of $\text{Fe}_3\text{O}_4$, alginate, and $\text{Fe}_3\text{O}_4@\text{Alg}$ were 96.41%, 30.97%, and 91.45%, respectively. Thereafter, we calculated the percentage of alginate as 7.58 wt% per mg of $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles. Finally, to ensure the efficiency of hyperthermia, the magnetic properties of the samples were analyzed with SQUID. As shown in Figure 3D, at a temperature of 310 K, the intensity of magnetization (emu) changes with the magnetic field (Oe). All three samples, ie, $\text{Fe}_3\text{O}_4$, $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles, and $\text{Fe}_3\text{O}_4@\text{Alg-GA}$ nanoparticles, showed similar saturation magnetization (emu/g), ie, 66.2, 69.2, and 71.6, respectively (Table 1), indicating their superparamagnetic properties.

### Magnetic thermal properties and cytotoxicity of $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles

The heating properties of the synthesized $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles were examined by measuring the increased temperature

| Table 1 Summary of the properties of the synthesized $\text{Fe}_3\text{O}_4$, $\text{Fe}_3\text{O}_4@\text{Alg}$ nanoparticles and $\text{Fe}_3\text{O}_4@\text{Alg-GA}$ nanoparticles |
|-------------------------------------------------|----------------|-------------|----------------|------------------|
| $\text{Fe}_3\text{O}_4$                       | 5.629±2.257   | 0.401       | +16.1          | 66.2             |
| $\text{Fe}_3\text{O}_4@\text{Alg}$            | 122±29.5      | 0.242       | –60.1          | 69.2             |
| $\text{Fe}_3\text{O}_4@\text{Alg-GA}$         | 128±18.9      | 0.148       | –29.7          | 71.6             |

**Abbreviations:** Alg, alginate; $\text{Fe}_3\text{O}_4$, iron oxide; GA, galactosamine; PDI, polydispersity index.
upon applying a high-frequency heating instrument (780 kHz, 19 kA/m). As shown in Figure 4A, Fe$_3$O$_4$@Alg nanoparticles could increase the temperature of water up to 10°C when they were exposed to a high frequency for 10 minutes. For a single-domain particle, such as the superparamagnetic particle, the external magnetic field would induce Brown relaxation and Neel relaxation to warm up the surrounding environment. Brown relaxation is described as the random rotation of the momentum through the movement of particles and Neel relaxation is described as the flipping motion of the magnetic moment relative to the particle.  

Enpuku et al showed that Brown relaxation would be affected by aggregation of nanoparticles. The Fe$_3$O$_4$ nanoparticles (without alginate) had worse heating efficiency because they aggregated easily, thereby reducing the Brownian motion. The SAR equation was used to determine the heat efficiency of the samples, and the SAR value for Fe$_3$O$_4$, Fe$_3$O$_4$@Alg, and Fe$_3$O$_4$@Alg-GA was 192.8, 212.0, and 308.4 W/g Fe, respectively. Ma et al showed that SAR values of magnetite particles were size-dependent. The SAR values increased as particle size decreased. It is always important to ensure biocompatibility of newly synthesized nanomaterials. Therefore, we examined the cytotoxicity of Fe$_3$O$_4$@Alg nanoparticles and Fe$_3$O$_4$@Alg-GA nanoparticles by MTT assay. As shown in Figure 4B, different concentrations of Fe$_3$O$_4$@Alg nanoparticles and Fe$_3$O$_4$@Alg-GA nanoparticles were cocultured with HepG2 cells for 4 hours and cell viability was measured using an ultraviolet spectrometer at 570 nm. Cell viability was not significantly decreased even when the concentration was up to 1,000 µg/mL, indicating excellent biocompatibility of the synthesized Fe$_3$O$_4$@Alg-based nanoparticles and implying that they are good candidates for intracellular drug delivery. It is well known that plasma membranes have a large negative charge, meaning that the membranes would repel a nanoparticle with a negative charge. Although cationic sites are scarcer on the plasma membrane, many researchers have pointed out that the cationic sites can be used for interacting with anionic nanoparticles. The process of cell uptake could be separated into two steps, ie, binding on the membrane and internalization. Wilhelm et al hypothesized that electrostatic interactions governed the adsorption of anionic nanoparticles.
onto the membrane, and interacted strongly and nonspecifically with the plasma membrane. This adsorption step precedes the internalization step and governs the overall cell uptake.

We consider that the synthesized Fe\textsubscript{3}O\textsubscript{4}@Alg nanoparticles and Fe\textsubscript{3}O\textsubscript{4}@Alg-GA nanoparticles could be applied for in vivo hyperthermia through intravenous injection, so the hemocompatibility of the samples, including anticoagulant activity and red blood cell hemolysis, was also examined. When nanomaterials interact with plasma, plasma proteins such as fibrinogen would adsorb onto the surface of the materials, resulting in plasma clotting.\textsuperscript{1} Measurement of anticoagulant activity based on plasma clotting has already become a recognized test to estimate the compatibility between blood and a nanomaterial. The synthesized Fe\textsubscript{3}O\textsubscript{4}@Alg nanoparticles and Fe\textsubscript{3}O\textsubscript{4}@Alg-GA nanoparticles were directly incubated with 100% human plasma to assess the effects of direct-contact activation on plasma clotting induced by the nanomaterials as evaluated by their recalcified plasma clotting time. As shown in Figure 4C, plasma clotting time for the recalcified plasma solutions in blank polystyrene (PS) wells was determined to have an upper limit of around 11 minutes at 37°C. The Fe\textsubscript{3}O\textsubscript{4}@Alg nanoparticles and Fe\textsubscript{3}O\textsubscript{4}@Alg-GA nanoparticles showed slightly prolonged plasma clotting times as well as anticoagulant activity in comparison with PS. The results indicate that the synthesized materials do not activate plasma clotting through the intrinsic coagulation pathway. A red blood cell hemolysis assay was performed to evaluate the antimicrobial activity and blood compatibility of the synthesized Fe\textsubscript{3}O\textsubscript{4}@Alg-based nanoparticles. The hemolysis values for red blood cells obtained in DI water and PBS solution at 37°C were used as positive and negative controls, respectively. Released hemoglobin that was destroyed by the nanomaterials could be detected by measuring absorbance of 541 nm. As shown in Figure 4D, both Fe\textsubscript{3}O\textsubscript{4}@Alg nanoparticles and Fe\textsubscript{3}O\textsubscript{4}@Alg-GA nanoparticles showed excellent blood compatibility like the PBS solution, indicating that our Fe\textsubscript{3}O\textsubscript{4}@Alg and Fe\textsubscript{3}O\textsubscript{4}@Alg-GA nanoparticles did not have any hemolytic activity (ie, they did not cause disruption of biological membranes). The results obtained here indicate that the synthesized Fe\textsubscript{3}O\textsubscript{4}@Alg and Fe\textsubscript{3}O\textsubscript{4}@Alg-GA nanoparticles are suitable for in vivo intravenous injection.
In vitro hyperthermia

The morphology and viability of HepG2 cells with and without treatment with Fe₃O₄@Alg nanoparticles and Fe₃O₄@Alg-GA nanoparticles are shown in Figure 5A and D. Prussian blue and nuclear fast red were used to investigate the interaction between the Fe₃O₄@Alg samples and HepG2 cells, where Prussian blue can interact with iron oxide, resulting in blue precipitates, while nuclear fast red can stain the cytoplasm and cell nucleus. As shown in Figure 5A1 and A2, intact HepG2 cells appeared to have a clear cytoplasm and a red cell nucleus. In contrast, the HepG2 cells cultured with Fe₃O₄@Alg contained several blue precipitates present in whole cells, indicating that the accumulation of Fe₃O₄@Alg nanoparticles could be greatly enhanced by functionalization of D-galactosamine on the surface of the Fe₃O₄@Alg nanoparticles, as shown in Figure 5C1 and C2. However, cells that had taken up Fe₃O₄@Alg nanoparticles still showed an elongated shape, indicating that the Fe₃O₄@Alg nanoparticles did not harm cells in the absence of a magnetic field. This conclusion can be further supported by the high cell viability in the case of Fe₃O₄@Alg and Fe₃O₄@Alg-GA nanoparticles, as shown in Figure 5D. The efficacy of hyperthermia with Fe₃O₄@Alg nanoparticles was evaluated by comparing cell viability with and without magnetic field treatment, as shown in Figure 5D. First, when applying a magnetic field to intact cells, cell viability only decreased slightly, as compared with the case without applying a magnetic field (ie, cell [MF]). We suggest that the anticancer effect from the magnetic field can be ignored. For the case of Fe₃O₄@Alg nanoparticles, cell viability decreased to around 60% after the magnetic field treatment (ie, Fe₃O₄@Alg [MF]), successfully demonstrating the hyperthermia effect. This effect was significantly enhanced when the cells were treated with Fe₃O₄@Alg-GA nanoparticles. Almost all the cells were killed (viability was only around 5%) for the case of Fe₃O₄@Alg-GA [MF]). We propose that the hyperthermia effect results from the appearance of Fe₃O₄ nanoparticles inside the cells, and its efficacy should be proportional to the amount of Fe₃O₄ nanoparticles inside the cells. To verify this, we collected the cells that had taken up Fe₃O₄@Alg nanoparticles or Fe₃O₄@Alg-GA nanoparticles, lysed these cells, and then analyzed the amount of Fe₃O₄ inside them by ICP-MS. As shown in Table 1, the amounts of Fe₃O₄ inside the cells were 18.5 pg and 364.43 pg for the Fe₃O₄@Alg nanoparticle and Fe₃O₄@Alg-GA nanoparticle groups, respectively.

Figure 5 (A–C) Cell morphology of HepG2 cells (A1, A2) treated with Fe₃O₄@Alg nanoparticles (B1, B2) and Fe₃O₄@Alg-GA nanoparticles (C1, C2). (D) Viability of HepG2 cancer cells incubated with Fe₃O₄@Alg and Fe₃O₄@Alg-GA nanoparticles with and without a magnetic field. The * and *** indicate the apparent difference of the results.

Abbreviations: Alg, alginate; DI, deionized; Fe₃O₄, iron oxide; GA, galactosamine; MF, magnetic field.
This result indeed supports our hypothesis, and indicates that efficient hyperthermia could be achieved by enhanced internalization of \( \text{Fe}_3\text{O}_4 \) nanoparticles.

**Conclusion**

This study reports the synthesis of \( \text{Fe}_3\text{O}_4@\text{Alg} \) nanocomposites with controllable particle sizes through the combination of coprecipitation and pre-gel methods under desired synthesis conditions. The synthesized \( \text{Fe}_3\text{O}_4@\text{Alg} \) nanoparticles demonstrated excellent cell and blood compatibility, along with hyperthermic activity toward HepG2 cells. The abundant COOH groups on the external surface afford the ability to functionalize \( \text{Fe}_3\text{O}_4@\text{Alg} \) nanoparticles with a liver cancer cell-targeting ligand (i.e., d-galactosamine). \( \text{Fe}_3\text{O}_4@\text{Alg-GA} \) nanoparticles show enhanced hyperthermia efficacy owing to their increased internalization, which shows significant potential as effective and visually observable transmembrane heat nanogenerator. Further development of these \( \text{Fe}_3\text{O}_4@\text{Alg} \) nanoparticles with controlled drug delivery can lead to a new generation of nanodevices for biomedical applications.

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**Disclosure**

The authors report no conflicts of interest in this work.

**References**


Supplementary materials

Figure S1 Size distribution of Fe₃O₄ (A), Fe₃O₄@Alg (B), Fe₃O₄@Alg-GA (C).

Table S1 Size and PDI of Fe₃O₄@Alg nanoparticles according to pH value and amount of alginate included during synthesis

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Abbreviation: PDI, polydispersity index