

Biosynthesis, characterization, and antimicrobial applications of silver nanoparticles

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Abstract: In the present study, the strain *Brevibacterium frigoritolerans* DC2 was explored for the efficient and extracellular synthesis of silver nanoparticles. These biosynthesized silver nanoparticles were characterized by ultraviolet-visible spectrophotometry, which detected the formation of silver nanoparticles in the reaction mixture and showed a maximum absorbance at 420 nm. In addition, field emission transmission electron microscopy revealed the spherical shape of the nanoparticles. The dynamic light scattering results indicated the average particle size of the product was 97 nm with a 0.191 polydispersity index. Furthermore, the product was analyzed by energy dispersive X-ray spectroscopy, X-ray diffraction, and elemental mapping, which displayed the presence of elemental silver in the product. Moreover, on a medical platform, the product was checked against pathogenic microorganisms including *Vibrio parahaemolyticus*, *Salmonella enterica*, *Bacillus anthracis*, *Bacillus cereus*, *Escherichia coli*, and *Candida albicans*. The nanoparticles demonstrated antimicrobial activity against all of these pathogenic microorganisms. Additionally, the silver nanoparticles were evaluated for their combined effects with the commercial antibiotics lincomycin, oleandomycin, vancomycin, novobiocin, penicillin G, and rifampicin against these pathogenic microorganisms. These results indicated that the combination of antibiotics with biosynthesized silver nanoparticles enhanced the antimicrobial effects of antibiotics. Therefore, the current study is a demonstration of an efficient biological synthesis of silver nanoparticles by *B. frigoritolerans* DC2 and its effect on the enhancement of the antimicrobial efficacy of well-known commercial antibiotics.

Keywords: *Brevibacterium frigoritolerans*, biosynthesis, silver nanoparticles, antimicrobial activity, synergistic effect

Introduction

Currently, the development of a biological methodology for the synthesis of nanoparticles as well as for their characterization and applications on medical and environmental platforms are important aspects of nanotechnology.^{1,2} There is a particularly pressing need for new and efficient methodologies for the synthesis of nanomaterials. Various physical and chemical methods have been reported along with many drawbacks that limit the use of nanoparticles in biological applications; these disadvantages include the use of toxic solvents, the generation of hazardous byproducts, and high energy consumption. The demand for a methodology that would avoid the hazardous byproducts associated with current physicochemical processes leads to the exploration of a biological species capable of synthesizing nanoparticles.³ The concept of exploiting biological diversity for the synthesis of nanoparticles promises to be quite simple and economical. In addition, these species have been proved to be free of the limitations associated with chemical and physical synthesis. Furthermore, the use of these organisms represents new product development and may allow for applications of biological resources in various fields.⁴ In the recent past, nanoparticles have been

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synthesized by using various plant extracts, actinomycetes, fungi, and bacteria.⁵⁻⁸ The microbial resource is a striking opportunity for nanotechnology, as a number of microorganisms are available and can be explored for this action.⁹

From a medical perspective, the development of resistance mechanisms against antibiotics by pathogenic microorganisms has been a subject of major concern.¹⁰ These resistance mechanisms, due to several enzymatic and genetic mutations in the pathogens that cause infectious diseases, has encouraged researchers to design new antimicrobial agents against pathogens to control infections.^{10,11} Therefore, there is always a need to develop new alternatives to control pathogenic organisms.

Metal nanoparticles are an effective way to control many pathogenic and antibiotic-resistant microorganisms.¹² Nanoparticles are applicable in diverse areas such as medicine, diagnostic agents, drug and gene delivery, electronics, cosmetics, coatings, biosensors, imaging, and environmental remediation. Among the many metal nanoparticles, silver nanoparticles have been intensely studied because of the distinct properties of their optical behavior, conductivity, chemical stability, and catalytic activity.¹³ In addition, silver nanoparticles exhibit broad spectrum bactericidal and fungicidal activity. These properties make them suitable for use as a disinfectant in many medical devices and goods. In addition, silver nanoparticles and antibiotic conjugates can be made efficiently and could result in an increase in the efficacy of antibiotics against various pathogenic, antibiotic-resistant microorganisms.¹⁴ Because of the applications of silver nanoparticles in the fields of biology, environment, and technology, there is a growing need for the development of a cost-effective method for the biosynthesis of silver nanoparticles.

The present research was carried out to explore a biological method for the synthesis of silver nanoparticles and to apply these nanoparticles in controlling pathogenic microorganisms. Silver nanoparticles were biosynthesized by *Brevibacterium frigoritolerans* DC2, which was isolated from a fermented food product. These nanoparticles were tested against different pathogenic microorganisms for their antibacterial efficacy both independently and in combination with various conventional antibiotics.

Materials and methods

Materials

All of the media were purchased from MB Cell (Los Angeles, CA, USA). Analytical grade silver nitrate (AgNO_3) and cycloheximide were purchased from Sigma-Aldrich Co.

(St Louis, MO, USA). Standard antibiotic discs with standard concentrations for their action against microorganisms were used for the antimicrobial tests. The antibiotics discs that were used are as follows: 1) vancomycin, 30 $\mu\text{g}/\text{disc}$; 2) rifampicin, 5 $\mu\text{g}/\text{disc}$; 3) oleandomycin, 15 $\mu\text{g}/\text{disc}$; 4) penicillin G, 10 $\mu\text{g}/\text{disc}$; 5) novobiocin, 30 $\mu\text{g}/\text{disc}$; and 6) lincomycin, 15 $\mu\text{g}/\text{disc}$. All of the discs were purchased from Oxoid Ltd. (Basingstoke, England).

The pathogenic bacterial strains *Vibrio parahaemolyticus* ATCC 33844, *Bacillus anthracis* NCTC 10340, *Salmonella enterica* ATCC 13076, *Bacillus cereus* ATCC 14579, *Escherichia coli* ATCC 10798, and *Candida albicans* KACC 30062 were used. The bacterial strains were cultured on nutrient agar media at 37°C and preserved at -70°C in glycerol stock vials for further study. *C. albicans* was cultured on Sabouraud dextrose agar at 28°C and preserved at -70°C in glucose yeast peptone broth glycerol stock vials.

Isolation and molecular identification of bacteria

Fermented skim yogurt was obtained from Indonesia. The food sample was serially diluted in sterile 0.8% NaCl and then spread onto Tryptic soy agar (TSA) media to obtain isolated colonies. To check the metal-tolerating capacity of isolated strains, the strains were further streaked on TSA medium supplemented with 1 mM filter-sterilized AgNO_3 . The plate was then incubated at room temperature for 48 hours and observed for bacterial growth. The isolated colonies were subcultured and obtained in pure form for further experiments.

Molecular identification of the isolated strain was carried out using a 16S ribosomal ribonucleic acid (rRNA) sequencing-based method. The genomic deoxyribonucleic acid (DNA) was extracted by using a commercial genomic DNA extraction kit (Core Bio System, Seoul, Korea). The 16S rRNA gene was amplified from the chromosomal DNA of the isolated strain by using the universal bacterial primer sets 27F, 518F, 800R, and 1512R.^{15,16} The purified polymerase chain reaction products were sequenced by Genotech (Daejeon, Korea). The nearly complete sequence (1485 base pairs) of the 16S rRNA was compiled by using SeqMan software (version 4.1). The 16S rRNA gene sequences of related taxa were obtained from the GenBank database and EzTaxon-e server.¹⁷

Biosynthesis of silver nanoparticles

For biological synthesis of silver nanoparticles, the selected bacterial isolate was inoculated into a 250 mL Erlenmeyer

flask containing 100 mL of sterile Tryptic soy broth. The cultured flasks were incubated in a rotating shaker set at 37°C for 120 rotations per minute (rpm) for 24 hours. After the incubation time, the culture was centrifuged at 8,000 rpm for 10 minutes to remove the bacterial pellet. The supernatant was obtained and was mixed with a filter-sterilized AgNO₃ solution (1 mM final concentration) for the extracellular production of silver nanoparticles^{1,4,7,8}. Furthermore, the culture supernatant with 1 mM AgNO₃ was incubated in an orbital shaker at 200 rpm and 25°C. The synthesis of silver nanoparticles was monitored by visual inspection for a change in the color of the culture medium. After the completion of the incubation period, the mixture was first centrifuged at 2,000 rpm for 5 minutes to remove any medium components, and then the silver nanoparticles were collected by high speed centrifugation at 16,000 rpm for 20 minutes. The obtained product was washed several times by centrifugation and was redispersed in water to remove the unconverted silver ions and any medium components. Finally, the silver nanoparticles were collected in the form of a pellet and were used for characterization.¹⁸

Characterization of silver nanoparticles

To verify the reduction of silver ions, the solution was scanned in the range of 200–800 nm in a ultraviolet-visible (UV-Vis) spectrophotometer (Ultrospec 2100 Pro; GE Healthcare Bio-Sciences Corp., Piscataway, NJ, USA). The size-distribution profile of silver nanoparticles was studied by using dynamic light scattering with a particle size analyzer (Photal, Otsuka Electronics Co., Osaka, Japan). The hydrodynamic diameters and polydispersity index were analyzed at 25°C. As a reference dispersive medium, pure water with a refractive index 1.3328, viscosity 0.8878 and dielectric constant 78.3 was used. The shape, morphology, and elemental distribution of the nanoparticles were analyzed by using field emission transmission electron microscopy (FE-TEM), energy dispersive X-ray spectroscopy (EDX), and elemental mapping with a JEM-2100F (JEOL, Tokyo, Japan) operated at 200 kV. The sample was prepared by placing a drop of collected silver nanoparticles on a carbon-coated copper grid and subsequently drying the sample in an oven at 60°C before transferring it to the microscope. The X-ray diffraction (XRD) analyses were performed on an X-ray diffractometer, D8 Advance (Bruker, Billerica, MA, USA), which was set to these configurations: 1) 40 kV; 2) 40 mA; 3) CuK α radiation; 4) a scanning rate of 6°/min; and 5) step size 0.02, over the 2 θ range of 20°–80°. The silver nanoparticles were collected by centrifugation and several washings with sterile water.

Finally, the particles were recovered by air drying the samples and were obtained in powder form.

The stability of silver nanoparticles was observed by keeping the silver nanoparticles in solution at room temperature for different time intervals. In addition, the effect of change in pH on the stability of the silver nanoparticles was studied. The sodium hydroxide (base) was added in the pH range of 4–10, and then the solution was scanned by using the UV-Vis spectrophotometer to observe the absorbance.

Analysis of antimicrobial activity of silver nanoparticles

The antimicrobial activity of the biologically synthesized silver nanoparticles against pathogenic microorganisms *B. anthracis*, *V. parahaemolyticus*, *S. enterica*, *E. coli*, *B. cereus*, and *C. albicans* was measured on Muller-Hinton agar (MHA) plates by using the well diffusion method. An overnight log culture of each pathogenic strain (100 μ L) was spread evenly on a MHA plate by using a glass spreader. Wells were made on the MHA plates by using a gel puncture. Then, 50 μ L of the silver nanoparticle reaction mixture was added into each well, and the wells were incubated at 37°C for 24 hours. As a control, 50 μ L of 1 mM silver nitrate solution was used, this concentration is the same as the one used to biosynthesize the nanoparticles. After incubation, the zones of inhibition were measured by measuring the diameter of the zone formed around each well. Similarly, the experiment was conducted with *C. albicans* on Sabouraud dextrose agar plates. The study was done in duplicate to check the reproducibility.¹⁹

Simultaneously, the disc diffusion method was used to investigate the synergistic effect of antibiotics with silver nanoparticles for antimicrobial effect against test strains. In this assay, the MHA medium plates were spread evenly with 100 μ L of an overnight log culture of test organisms. Each standard antibiotic disc of lincomycin, oleandomycin, vancomycin, novobiocin, penicillin G, and rifampicin was further impregnated with 30 μ L (100 mg/L) of freshly prepared, partially purified silver nanoparticle solution and was placed onto the agar plates. For each sample, a corresponding control was maintained by using an antibiotic disc without the silver nanoparticle solution. In addition, a control of only silver nanoparticles and silver nitrate at the same concentrations (30 μ L [100 mg/L]) was maintained. Then, the plates were incubated at 37°C for 24 hours. In case of *C. albicans*, cycloheximide (10 μ g/disc) was used as a control. After the incubation period, the zones of inhibition around each disc were measured and compared with the corresponding control.²⁰

Results and discussion

Screening and identification of bacteria

After the incubation period, the bacterial strain DC2 showed growth on the TSA plate supplemented with 1 mM AgNO_3 ; this suggests that the bacterial strain DC2 was capable of tolerating silver metal at a 1 mM concentration. On the basis of the molecular characterization of the bacterial isolate, DC2 showed 99.8% similarity with *B. frigoritolerans*. *B. frigoritolerans* is strictly aerobic chemo-organotroph, and it is not as well-known as the other members of the genus *Brevibacterium*.²¹ *Brevibacterium casei* has been studied for the biosynthesis of silver and gold nanoparticles.²² Biosurfactant-mediated synthesis of silver nanoparticles by marine *B. casei* MSA19 has been studied.²³ The reports suggested that the some of the species of the genus do show activity for the biosynthesis of metal nanoparticles; however, to our knowledge, this is the first report of the biosynthesis of silver nanoparticles by *B. frigoritolerans* DC2 isolated from a fermented food product. The 16S rRNA sequence of the strain *B. frigoritolerans* DC2 has been submitted to National Center for Biotechnology

Information with the accession number KM583447. The strain has been deposited in Korean Collection for Type Culture (KCTC 29680).

Synthesis and characterization of silver nanoparticles

In the present study, silver nanoparticles were successfully synthesized in the culture supernatant of *B. frigoritolerans* DC2. The formation of silver nanoparticles by the reduction of AgNO_3 was indicated by the color change of the reaction mixture (Figure 1A). As the biosynthesis proceeded over 48 hours, the color changed from light yellow to dark brown. This brown color could be due to the excitation of surface plasmon vibrations; if so, this would indicate the formation of silver nanoparticles in the reaction mixture.²⁴ On the other hand, no color change was observed in the control flask that contained medium with only the 1 mM AgNO_3 solution and had been kept under the same conditions as the other flask (Figure 1B). Because the nanoparticle synthesis was extracellular, the need for downstream processing that would have been otherwise essential for

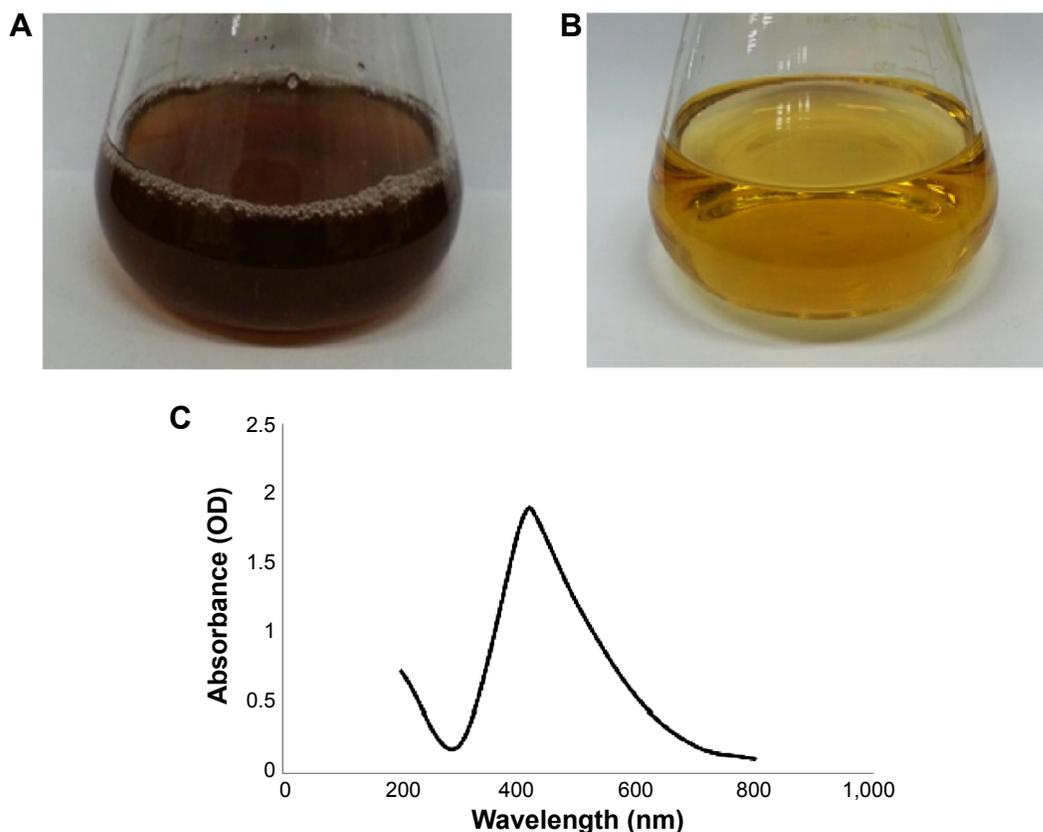


Figure 1 Chromatic properties of culture supernatant of *Brevibacterium frigoritolerans* DC2.

Notes: Culture supernatant of *B. frigoritolerans* DC2 after incubation with AgNO_3 (silver nitrate) (1 mM) (A) and control with medium and AgNO_3 (1 mM) after incubation period (B). UV-Vis spectra of culture supernatant of *B. frigoritolerans* DC2 treated with 1 mM AgNO_3 (C).

Abbreviations: OD, optical density; UV-Vis, ultraviolet-visible.

intracellular synthesis was avoided and thereby made the process simpler and more cost-effective. The exact mechanism behind the extracellular synthesis of silver nanoparticles in the supernatant remains to be elucidated. However, reports have suggested that the extracellular enzyme secreted by microorganism in the culture supernatant is responsible for the reduction of silver ions to silver nanoparticles.^{25,26} The study based on extracellular synthesis of silver nanoparticles by *Bacillus licheniformis* showed that the nitrate reductase enzyme extracellularly secreted by the bacteria in the medium was responsible for the synthesis of silver nanoparticles.⁸

For the characterization of the silver nanoparticles, the reaction mixture was monitored by using UV-Vis spectrophotometer for spectral analysis. After the incubation period, the reaction mixture was scanned in the range of 200–800 nm. In the UV-Vis absorption spectrum, a strong peak at about 420 nm was observed; this is attributed to the surface plasmon resonance band of the silver nanoparticles (Figure 1C). Reports suggest that the band located in this region corresponds to the surface plasmon resonance of silver nanoparticles.^{27,28} Thus, the reaction mixture indicates the formation of silver nanoparticles.

Various studies have characterized the shape and size of silver nanoparticles by TEM.^{22,24,27,29} In the present study, field emission TEM images of silver nanoparticles revealed that the shape of the nanoparticles was spherical (Figure 2A and B). The results showed that particles are uniform in shape with 10 nm to 30 nm in size. The dynamic light scattering particle size analysis results indicated that the hydrodynamic diameter of the particles range was 50–150 nm with a 0.191 polydispersity index (Figure 2C–E). The average particle size was 97 nm.^{30,31} The purity and elemental composition of the biosynthesized silver nanoparticles were determined by EDX, XRD, and elemental mapping. In the EDX spectrum, silver nanocrystallites displayed an optical absorption band peak at approximately 3 keV (Figure 3A); this property is due to the absorption of metallic silver nanocrystallites that correspond to surface plasmon resonance.⁸ Figure 3B shows the XRD pattern of silver nanoparticles, which exhibited intense peaks throughout the whole spectrum of 2θ value, which ranged from 20° to 80° ; this pattern was similar to the Bragg's reflection of silver nanocrystals. Our results are similar to those of previous reports of characterization of silver nanoparticles by XRD.^{32–35}

The elemental mapping results of the biosynthesized silver nanoparticles (Figure 3C and D) indicate that silver was distributed maximally (50.30%); this finding suggests that

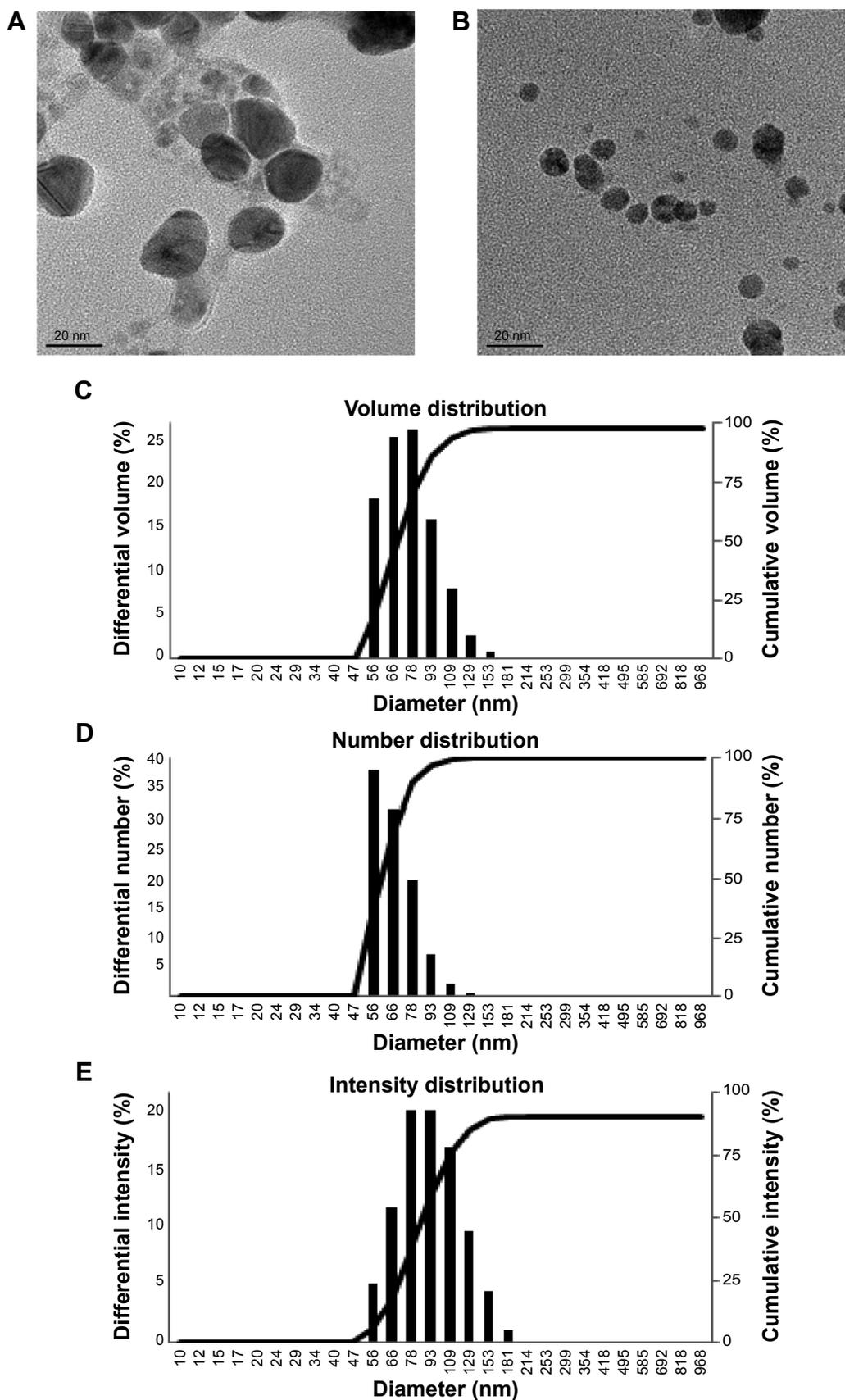
silver was the predominant metal. The other elements, such as copper (26.98%), carbon (14.98%), and sulfur (1.44%) were also observed. The distribution of carbon and copper was due to the use of a TEM grid, and slight contamination by chlorine and sulfur appeared. Thus, the results indicate that the strain *B. frigoritolerans* DC2 was found to be capable of synthesizing spherical silver nanoparticles.

The stability of silver nanoparticles was determined by keeping the nanoparticle solution at room temperature for different time intervals over many days. There was no observable variation in the UV-Vis spectrum of the reaction mixture even after 1 month; this indicated the stable nature of the silver nanoparticles in the reaction mixture. The results proved that the nanoparticles were stable for more than 1 month. In addition, the effect of the change in pH on the stability of the silver nanoparticles in the pH range of 4–10 was studied. The silver nanoparticle solution was observed before and after the addition of sodium hydroxide (base). No major change in the wavelength was observed; this result further confirmed the stable nature of the silver nanoparticles.

Antimicrobial activity of silver nanoparticles

Metal nanoparticles have been shown to be effective against many pathogenic and multidrug-resistant bacteria. For instance, silver nanoparticles have demonstrated activity against methicillin-resistant *Staphylococcus aureus* (MRSA) and methicillin-resistant *Staphylococcus epidermidis* (MRSE).³⁶ Various studies have reported antimicrobial action of silver nanoparticles against microorganisms.^{4,37,38} In our study, the biosynthesized silver nanoparticles displayed antimicrobial activity against a range of pathogenic microorganisms, such as *C. albicans*, *V. parahaemolyticus*, *S. enterica*, *B. anthracis*, *B. cereus*, and *E. coli*.

The antimicrobial activity of the reaction mixture was compared to that of silver nitrate solution after incubation period. Antimicrobial activity against tested microorganisms was measured by measuring the diameter of the zone of inhibition of each well. The results of mean diameters of zones of inhibition are interpreted in Table 1. The results demonstrate that the reaction mixture containing silver nanoparticles exhibited antimicrobial activity against the tested microorganisms in the following manner, *C. albicans* followed by *V. parahaemolyticus*, *B. anthracis* and *B. cereus*, *S. enterica*, and then *E. coli* (Figure 4A–F, respectively). Although the activity of reaction mixture exhibit slightly



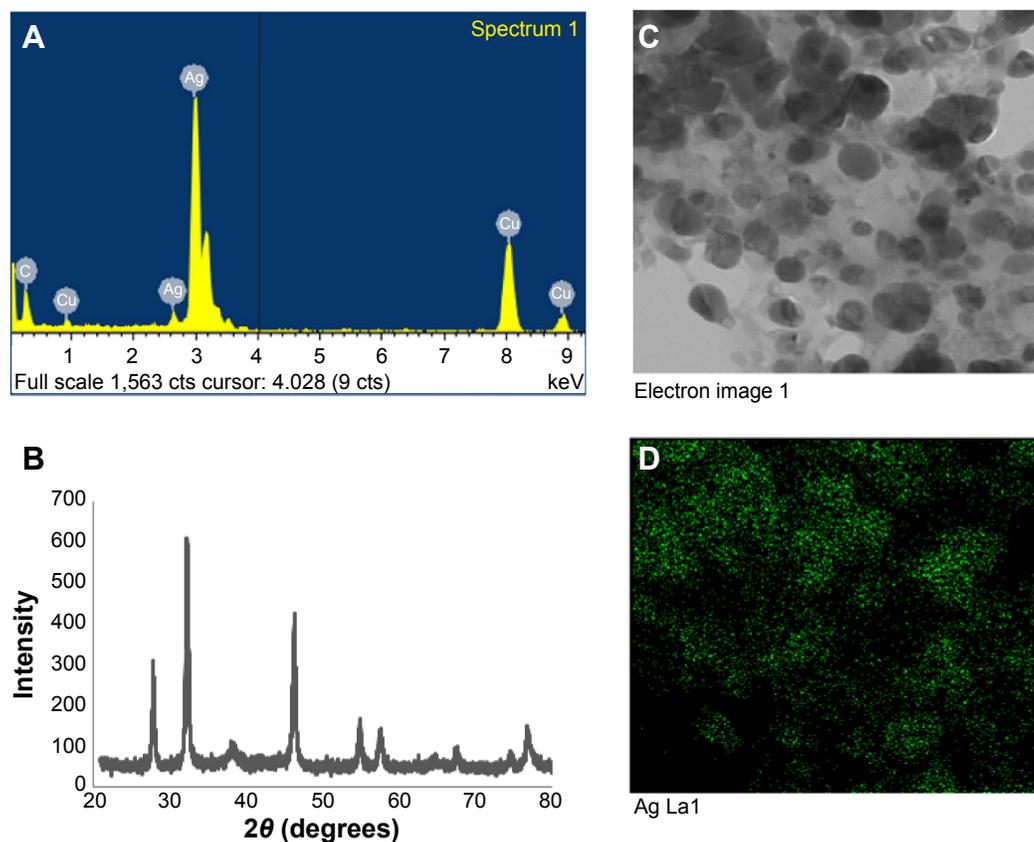


Figure 3 Silver nanoparticles examined under a variety of conditions.

Notes: Energy dispersive X-ray spectroscopy of the whole scan area showing major peak of silver nanoparticles at 3 keV (A); X-ray diffraction patterns of silver nanoparticles obtained by *Brevibacterium frigoritolerans* DC2 (B); transmission electron micrograph of silver nanoparticle pellet solution (C); silver nanoparticles, green (D).

more antimicrobial activity than silver nitrate solution. The activity was further analyzed after the purification of nanoparticles and compared with that of the same concentration of silver nitrate. The exact mechanism of antimicrobial action is not clearly known, but studies suggest that silver nanoparticles have the ability to damage the cell membrane permeability, damage the respiration functions of the cell,

and encourage the formation of free radicals. These factors cause the antimicrobial effect of silver nanoparticles.^{9,39}

Several studies have shown that the combination of silver nanoparticles with antibiotics leads to an enhanced effect of the antibiotics against microorganisms.^{14,20} This action is most probably due to an increase in cell wall penetration by these antibiotics with the nanoparticles. Combinations of antibiotics with nanoparticles have many advantages that minimize the side effects of broad-spectrum antibiotics; these benefits include increasing the concentration of the local antibiotic at the target site and facilitating the binding of antibiotics to microorganisms. Moreover, nanoparticle–antibiotic conjugates lower the amounts of both the drugs and the nanoparticles in the dosage; this property reduces the side effects of the medication while increasing its antimicrobial properties.¹⁴ In our study, the synergistic effect of silver nanoparticles with different commercial antibiotics was investigated against six pathogenic microorganisms by using the disc diffusion method. Figure 5A–E represents the antimicrobial activity of partially purified silver nanoparticles and silver nitrate of the same concentration (1 mM) against *S. enterica*, *E. coli*, *V. parahaemolyticus*, *B. anthracis*, and

Table I Diameters of zones of inhibition (mm) for 50 μ L of reaction mixture containing silver nanoparticles after incubation with various microorganisms

Number	Microorganism	Zone of inhibition (mm)	
		Silver nitrate	Reaction mixture
1	<i>Candida albicans</i> KACC 30062	22 \pm 0.7	24 \pm 1.4
2	<i>Vibrio parahaemolyticus</i> ATCC 33844	23 \pm 1.1	25 \pm 1.0
3	<i>Bacillus anthracis</i> NCTC 10340	14 \pm 0.2	15 \pm 0.8
4	<i>Bacillus cereus</i> ATCC 14579	14 \pm 0.3	15 \pm 0.5
5	<i>Salmonella enterica</i> ATCC 13076	12 \pm 0.4	12 \pm 0.6
6	<i>Escherichia coli</i> ATCC 10798	10 \pm 0.7	11 \pm 0.2

Notes: Each value is the mean of the diameters of two wells. Each well contained 50 μ L of tested samples.

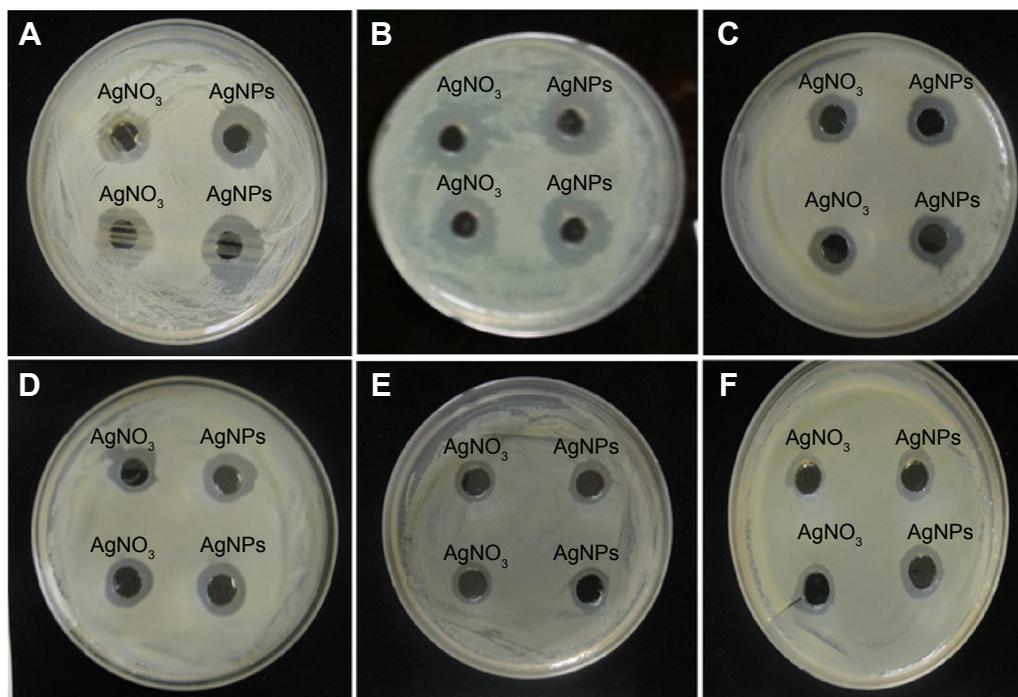


Figure 4 Zones of inhibition for various pathogens.

Notes: Zones of inhibition of 50 μ L of reaction mixture containing silver nanoparticles and 50 μ L of 1mM AgNO_3 (silver nitrate) against the pathogenic strains *Candida albicans* (A), *Vibrio parahaemolyticus* (B), *Bacillus anthracis* (C), *Bacillus cereus* (D), *Salmonella enterica* (E), and *Escherichia coli* (F).

Abbreviation: AgNPs, silver nanoparticles.

B. cereus. The appearance of zones of inhibition indicated that the biosynthesized silver nanoparticles had stronger antimicrobial properties against the tested microorganisms than did silver nitrate when both treatments were used at the same concentration. In addition, the synergistic effect of the antibiotics (lincomycin, oleandomycin, novobiocin, vancomycin, penicillin G, and rifampicin) in conjugation with biosynthesized silver nanoparticles increased the sensitivity of the tested microorganisms. The results show that *S. enterica*, *E. coli*, and *V. parahaemolyticus* (Figure 5F–H) were completely resistant to the antibiotics; however, the addition of the silver nanoparticle solutions to the discs rendered the bacterial strains sensitivity to those discs and thereby resulted in the formation of zones of inhibition (Figure 5K–M). The maximum increase in fold area has been calculated for antibiotics with silver nanoparticles with respect to standard antibiotics.⁴⁰ The average of maximum increase in fold area against the antibiotic-resistant microorganisms *S. enterica*, *E. coli*, and *V. parahaemolyticus* (Figure 6A–C) observed in this study was highest for novobiocin (4.82 fold), then for lincomycin, oleandomycin, and rifampicin (4.23 fold), then for vancomycin (3.98 fold), and finally for penicillin G (3.66 fold). Various researchers have demonstrated the combined effect of biosynthesized silver nanoparticles with commercial antibiotics.^{4,14,20} The

results demonstrate that the silver nanoparticles obtained by *B. frigoritolerans* DC2 enhance the antimicrobial activity of commercial antibiotics against *S. enterica*, *E. coli*, and *V. parahaemolyticus*.

The other pathogenic strains, *B. anthracis* and *B. cereus* (Figure 5I and J), showed a sensitivity pattern to antibiotics. However, when the discs were combined with silver nanoparticles, an increase in the diameters of the zones of inhibition was observed (Figure 5N and O). The results clearly indicate that the addition of silver nanoparticles enhanced the effect of the antibiotics. Thus, an increase in fold area was observed in all of the antibiotics tested against antibiotic-sensitive and antibiotic-resistant pathogenic microorganisms. Similarly, Figure 5P shows the antimicrobial activity of silver nanoparticles against *C. albicans*. Figure 5Q shows that *C. albicans* was completely resistant to cyclohexamide; however, when the silver nanoparticles were added to the cyclohexamide, antimicrobial activity was enhanced and resulted in the formation of a zone of inhibition. The results showed that the average of maximum increase in fold area against antibiotic-sensitive microorganisms *B. anthracis* and *B. cereus* (Figure 6D and E) were, in descending order, as follows: 1) rifampicin (3.69 fold); 2) oleandomycin (1.91 fold); 3) penicillin G (0.36 fold); 4) novobiocin (0.26 fold); 5) vancomycin (0.22 fold); and 6) lincomycin (0.14 fold). Our results are in agreement with

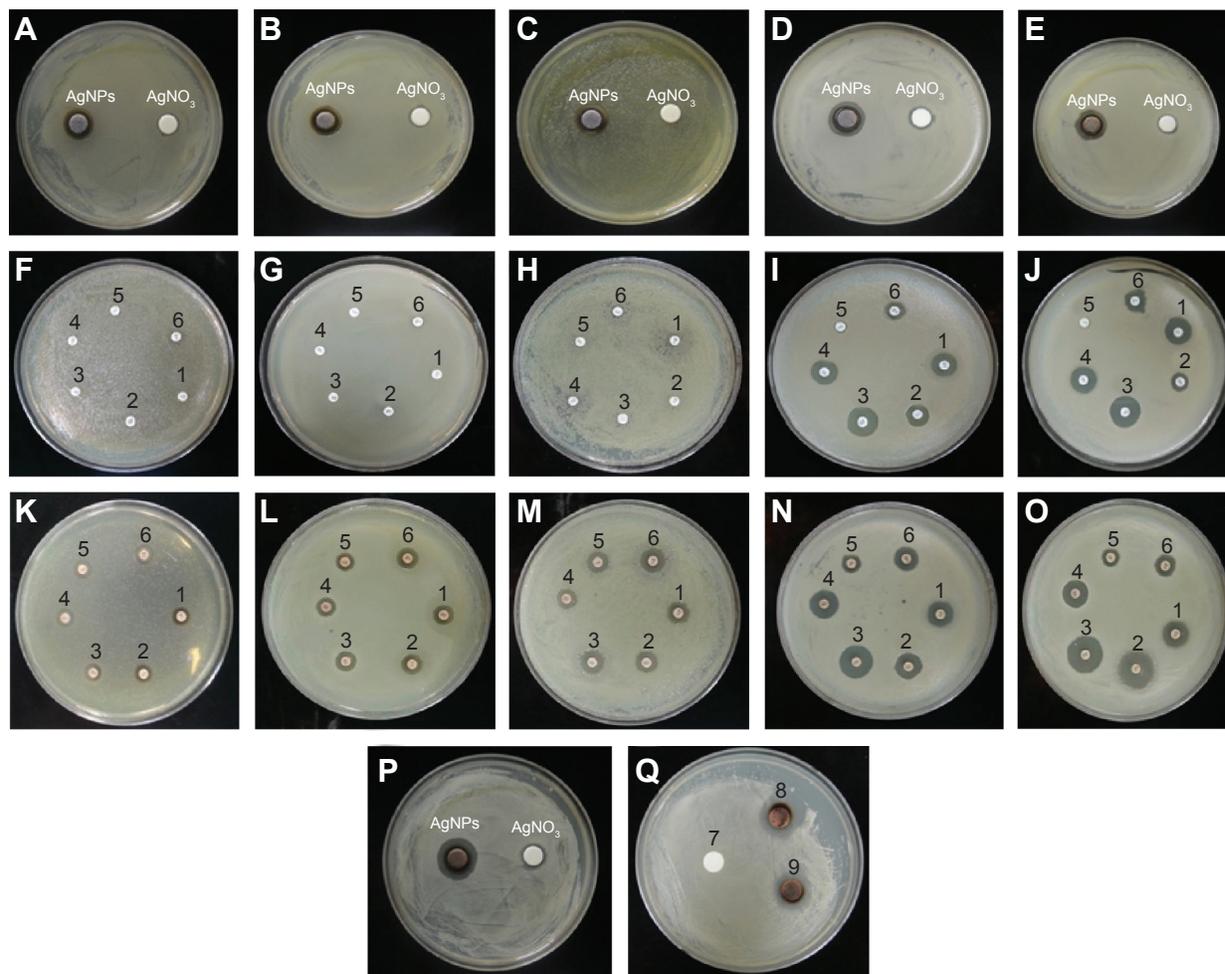


Figure 5 Effects of partially purified silver nanoparticle pellet solution on various pathogens.

Notes: Zones of inhibition of partially purified silver nanoparticle pellet solution and silver nitrate (AgNO_3) (both at same concentration, 30 μL [100 mg/L]) against *Salmonella enterica* (A), *Escherichia coli* (B), *Vibrio parahaemolyticus* (C), *Bacillus anthracis* (D), and *Bacillus cereus* (E). Zones of inhibition of standard antibiotic discs against pathogenic bacteria *Salmonella enterica* (F), *Escherichia coli* (G), *Vibrio parahaemolyticus* (H), *Bacillus anthracis* (I), and *Bacillus cereus* (J). Zones of inhibition of standard antibiotic discs with silver nanoparticles against *Salmonella enterica* (K), *Escherichia coli* (L), *Vibrio parahaemolyticus* (M), *Bacillus anthracis* (N), and *Bacillus cereus* (O). Zones of inhibition of silver nanoparticles and silver nitrate against *Candida albicans* (P), cycloheximide (Q-7), Cycloheximide with AgNPs (Q-8,9). For (A–Q), the numbers, the corresponding antibiotics, and the corresponding concentrations of antibiotics are as follows: 1) lincomycin (MY15), 15 $\mu\text{g}/\text{disc}$; 2) oleandomycin (OL15), 15 $\mu\text{g}/\text{disc}$; 3) novobiocin (NV30), 30 $\mu\text{g}/\text{disc}$; 4) vancomycin (VA30), 30 $\mu\text{g}/\text{disc}$; 5) penicillin G (P10), 10 $\mu\text{g}/\text{disc}$; 6) rifampicin (RD5) 5 $\mu\text{g}/\text{disc}$; 7) cycloheximide, 10 $\mu\text{g}/\text{disc}$; 8) cycloheximide (10 $\mu\text{g}/\text{disc}$) with silver nanoparticles; and 9) cycloheximide (10 $\mu\text{g}/\text{disc}$) with silver nanoparticles.

Abbreviation: AgNPs, silver nanoparticles.

other studies that have shown the antimicrobial activity of silver nanoparticles against many pathogenic and multidrug-resistant microorganisms.^{4,14} In our findings, biosynthesized silver nanoparticles showed antimicrobial activity against pathogenic microorganisms. Moreover, the silver nanoparticles enhanced the antimicrobial activity of commercial antibiotics against pathogenic microorganisms. However, the exact action mechanism still must be explored.

Conclusion

In our study, the method for biological synthesis of silver nanoparticles by *B. frigoritolerans* DC2 was established. This biogenic process will be helpful for minimizing the need for chemical and physical methodologies. In addition

to this, the extracellular synthesis of nanoparticles could be highly advantageous for facilitating downstream processing in large-scale operations. The silver nanoparticles that were obtained showed antimicrobial effects against various pathogenic microorganisms. Furthermore, they enhanced the antimicrobial activity of various commercial antibiotics against pathogenic microorganisms.

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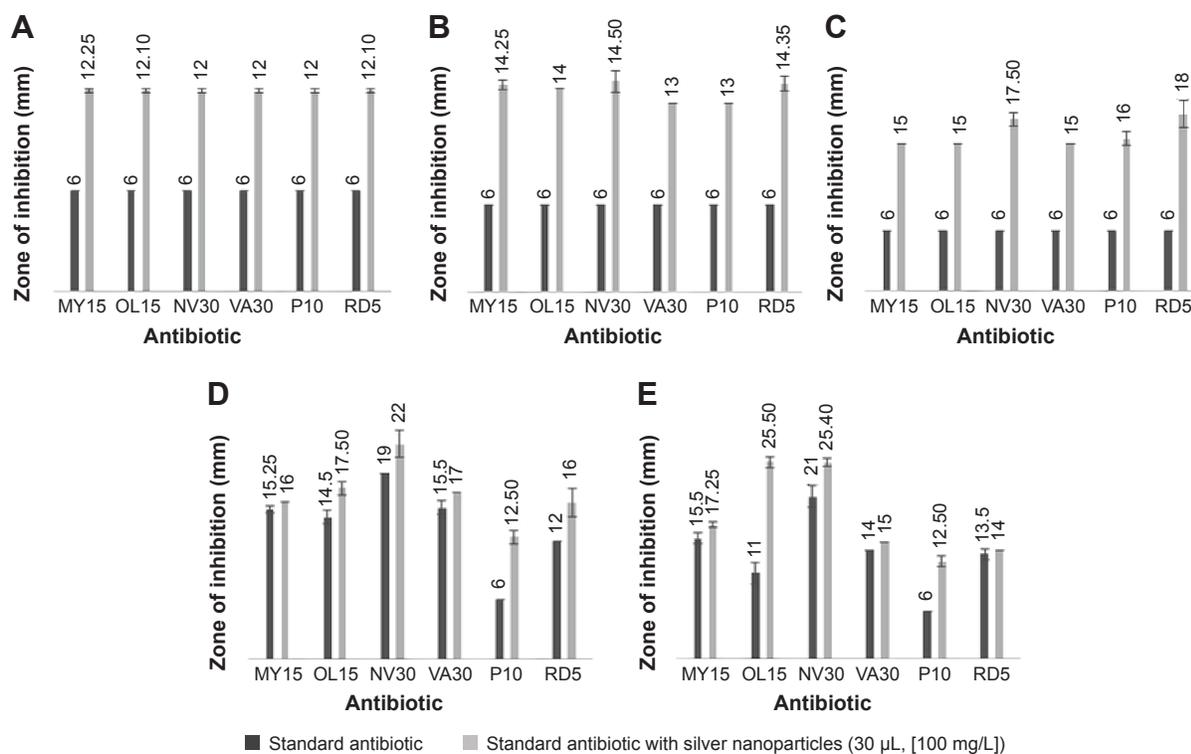


Figure 6 Graphical representation of comparative study between antibiotics and antibiotics with AgNPs.

Notes: *Salmonella enterica* (A), *Escherichia coli* (B), *Vibrio parahaemolyticus* (C), *Bacillus anthracis* (D), and *Bacillus cereus* (E), respectively. Results interpreting the antimicrobial activity of commercial antibiotics alone with that of antibiotics combined with silver nanoparticles.

Disclosure

The authors report no conflicts of interest in this work.

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