

PLGA-PLL-PEG-Tf-based targeted nanoparticles drug delivery system enhance antitumor efficacy via intrinsic apoptosis pathway

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Abstract: Chemotherapy offers a systemic cancer treatment; however, it is limited in clinical administration due to its serious side effects. In cancer medicine, the use of nanoparticles (NPs) drug delivery system (DDS) can sustainedly release anticancer drug at the specific site and reduce the incidence of toxicity in normal tissues. In the present study, we aimed to evaluate the benefit of a novel chemotherapeutic DDS and its underlying mechanisms. Daunorubicin (DNR) was loaded into poly (lactic-co-glycolic acid) (PLGA)-poly-L-lysine (PLL)-polyethylene glycol (PEG)-transferrin (Tf) NPs to construct DNR-PLGA-PLL-PEG-Tf-NPs (DNR-loaded NPs) as a DDS. After incubating with PLGA-PLL-PEG-Tf-NPs, DNR, and DNR-loaded NPs, the leukemia K562 cells were collected and the intracellular concentration of DNR was detected by flow cytometry, respectively. Furthermore, the effect of drugs on the growth of tumors in K562 xenografts was observed and the relevant toxicity of therapeutic drugs on organs was investigated *in vivo*. Meanwhile, cell apoptosis in the excised xenografts was measured by transferase-mediated dUTP nick-end labeling assay, and the expression of apoptosis-related proteins, including Bcl-2, Bax, Caspase-9, Caspase-3, and cleaved-PARP, was determined by Western blotting analysis. Results showed that DNR-loaded NPs increased intracellular concentration of DNR in K562 cells *in vitro* and induced a remarkable improvement in anticancer activity in the xenografts *in vivo*. The expression of Bcl-2 protein was downregulated and that of Bax, Caspase-9, Caspase-3, and cleaved-PARP proteins were obviously upregulated in the DNR-loaded NPs group than that in other ones. Interestingly, pathological assessment showed no apparent damage to the main organs. In summary, the results obtained from this study showed that the novel NPs DDS could improve the efficacy of DNR in the treatment of leukemia and induce apoptosis via intrinsic pathway. Thus, it can be inferred that the new drug delivery may be a useful clinical tool.

Keywords: PLGA-PLL-PEG, daunorubicin, transferrin, K562 cells, apoptosis

Introduction

Cancer is a major public health problem in the world. Frequent challenges encountered by current cancer therapies include nonspecific systemic distribution of chemotherapeutic agents, inadequate drug concentrations reaching the tumor, relatively short circulating half-lives, poor solubility, and limited ability to monitor therapeutic responses.¹ Current advances in biotechnology and related areas are aiding the discovery and rational design of many new classes of drugs, in which it is crucial to improve specific drug delivery methods.²

As we know, nanoparticles (NPs) have been widely evaluated as carriers for the delivery of drugs and genes because of their small size, sustained-release characteristics,³ ability to offer physical protection against ribonuclease (RNase)

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activity,⁴ and ability of specifically targeting tumors for either drugs or genes.^{5,6} Owing to both active and passive targeting, NPs are a promising carrier system for the targeted delivery of chemotherapeutic agents, and serious systemic toxicity or adverse effects can be avoided.⁷ Poly (lactic-co-glycolic acid) (PLGA) is one of the most effective biodegradable polymeric NPs, and it has been approved by the Food and Drug Administration for drug delivery and has been frequently used.⁸ Poly-L-lysine (PLL) has an advantage of being water soluble, hydrodegradable, biodegradable, and biocompatible,⁹ and so it is suitable for the backbone as a drug delivery carrier. Recently, it has been reported that PLL-modified PLGA NPs showed significantly higher amounts than unmodified PLGA NPs.¹⁰ PLGA has a relatively rapid rate of hydrolysis,¹¹ and it could reduce systemic clearance rates and prolong circulation half-life in vivo when being coated with biocompatible polymers such as polyethylene glycol (PEG).¹²

It is well known that the transferrin receptor (TfR) was highly expressed in metastatic and drug-resistant tumors, while it was lowly expressed in normal cells.¹³ Given the differential expression of TfR between the normal cells and tumor tissue, targeted chemotherapeutic agents can efficiently differentiate them and deliver antineoplastic drugs to the tumor cells.^{14,15} Nano-based drug delivery, modified with targeted moieties, including transferrins (Tfs), can enhance selective cellular uptake through Tf-mediated processes, and increase therapeutic benefit through the ability to encapsulate high concentrations of drug to the tumor site.¹⁶

Daunorubicin (DNR) is widely used in the treatment of leukemia. However, the undesirable properties including cumulative cardiotoxicity, which is one of the most serious side effects, lead to a very narrow therapeutic index;¹⁷ there is a need to explore alternate formulations. In this study, we aimed to test a novel PLGA-PLL-PEG-Tf-based targeted NPs drug delivery system (DDS) in enhancing antitumor of a classical chemotherapeutic agent, DNR, on K562 cells in vitro and in vivo.

Materials and methods

Main materials

DNR was purchased from Main Luck Pharmaceuticals Inc (Shenzhen, People's Republic of China), RPMI-1640 from Gibco Chemical Co. (Carlsbad, CA, USA), 10% heat-inactivated fetal bovine serum from Sigma-Aldrich Co (St Louis, MO, USA), Hematoxylin–Eosin Staining Kit from Beyotime Institute of Biotechnology (Shanghai, People's Republic of China), In Situ Cell Death Detection

Kit from Nanjing KeyGen Biotech Co Ltd (Nanjing, People's Republic of China), and monoclonal for Western blotting from Santa Cruz Biotechnology Inc. (Dallas, TX, USA). All other reagents were of analytical grade.

Preparation and characterization of DNR-loaded NPs

PLGA-PLL-PEG-NPs¹⁸ were provided by College of Pharmacy, Nanjing University of Technology. The conjugation was analyzed with ¹H NMR, and then the PLGA-PLL-PEG-NPs-loaded DNR was prepared using a modified double-emulsion solvent evaporation/diffusion method¹⁹ and separated using ultracentrifuge (GL-20G-H; Shanghai Anting Scientific Instrument Factory, Shanghai, People's Republic of China) for 30 minutes (4°C, 1,500 rpm). Then, PLGA-PLL-PEG-NPs-loaded DNR was well dissolved in phosphate buffered saline (PBS) using ultrasound treatment for 30 seconds and then Tf was added (Tf/PLGA-PLL-PEG molar ratio 8:1) with stirring at room temperature for 3 hours to conjugate with the surface of DNR-PLGA-PLL-PEG via CDI (*N,N*-carbonyldiimidazole) of PEG. Thereafter, DNR-PLGA-PLL-PEG-Tf-NPs (DNR-loaded NPs) were extracted using ultracentrifuge for 20 minutes (4°C, 1,500 rpm).

Mean particle size and ZP value of DNR-loaded NPs were determined by a Zetasizer 3000HS system (Malvern Instruments, Malvern, UK), the diameters and width of particle distribution were calculated using a volume distribution from the Gaussian distribution, and morphological evaluation was observed under transmission electron microscope (Hitachi Ltd, Tokyo, Japan). Meanwhile, drug loading efficiency was calculated as the percent ratio of actual amount of DNR encapsulated into PLGA-PLL-PEG-Tf-NPs to total amounts of DNR-PLGA-PLL-PEG-Tf-NPs. The drug entrapping efficiency of DNR was calculated as the actual amount of DNR encapsulated into PLGA-PLL-PEG-Tf-NPs vs the theoretical of DNR in DNR-PLGA-PLL-PEG-Tf-NPs, respectively.¹⁹

Cell line and cell culture

Leukemia K562 cells were obtained from Institute of Hematology, Chinese Academy of Medical Sciences (Beijing, People's Republic of China). The cells were cultured in RPMI-1640 media supplemented with 10% (v/v) fetal bovine serum, 100 U/mL penicillin, and 100 µg/mL streptomycin at 37°C in a humidified atmosphere containing 5% carbon dioxide. The cells used in experiments were in logarithmic growth phase.

Cellular accumulation of DNR

Briefly, K562 cells were incubated with PLGA-PLL-PEG-Tf-NPs, DNR, and DNR-loaded NPs with equivalent concentration of DNR for 48 hours. Meanwhile, those were exposed to RPMI-1640 medium as control. Then cells were harvested and washed three times with phosphate buffered saline (PBS). Thereafter, cells were suspended in 500 μ L PBS, and the cellular accumulation of DNR in each sample was directly detected by FACSCalibur flow cytometry (Becton, Dickinson and Company, Franklin Lakes, NJ, USA) at excitation/emission wavelengths of 488/575 nm. The relative fluorescence intensity (RFI) of DNR was calculated as $FI_{\text{treated group}}/FI_{\text{control group}}$.

Animals

Six-week age BALB/c nude mice, weighing 18–22 g, from the Shanghai National Center for Laboratory Animals were housed in cages under specific pathogen-free conditions where temperature was maintained at 22°C and humidity was in the range of 40%–50%, and sterile water and food were fed. All experiments involving the mice were carried out following the Guidelines for the Care and Use of Laboratory Animals of the National Institutes of Health.

Experimental design and animal treatments

Mice were subcutaneously inoculated into the left thigh with an injection of 1×10^7 K562 cells suspended in 0.2 mL of RPMI-1640 medium. Tumor sizes were measured with a digital caliper, and the tumor volumes were calculated according to the following formula: tumor volume V (mm^3) = $(1/2) \times a \times b^2$, where a is the largest diameter (length) and b is the smallest diameter (width) of the tumor. When tumors reached an average volume of approximately 50 mm^3 , animals were divided into four groups with six animals each for treatment as follows: Group 1, saline water (control); Group 2, PLGA-PLL-PEG-Tf-NPs; Group 3, free DNR (1 mg/kg for the single injection); and Group 4, DNR-loaded NPs (containing 1 mg/kg DNR for the single injection).

The intravenous treatments were done once every other day for 9 times for 18 days, and antitumor activity was determined by calculating the tumor volume as described earlier twice a week before sacrifice. After intravenous treatment for 18 days, mice were sacrificed and their xenograft tumors were harvested and processed for histopathologic analysis and determination of protein expression. During the whole

experiment, the trial animals were monitored for signs of toxicity.

Histopathologic examination

After the mice were sacrificed, the lung, heart, liver, kidney, and spleen were quickly removed and immediately fixed in 4% paraformaldehyde, dehydrated in a graded series of alcohol, and then embedded in paraffin. Tissue sections (4 μ m) were prepared and stained with hematoxylin and eosin. Thereafter, the sections were examined and microphotographed by using Olympus IX51 microscope (400 \times ; Olympus Corporation, Tokyo, Japan).

Transferase-mediated dUTP nick-end labeling

Based on the detection of single- and double-stranded DNA breaks by the terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling (TUNEL) assay, analysis of apoptosis was performed using an In Situ Cell Death Detection Kit according to the manufacturer's instructions. Tissue sections were analyzed to detect the localized green fluorescence of apoptotic cells and blue fluorescence of cell nuclei by fluorescence microscope; fluorescent images were acquired and photographed under the Olympus IX51 microscope (400 \times).

Western blot assays

In brief, the tissues were extracted with Triton X-100, and then the proteins were subjected to 10% sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) and transferred to a polyvinylidene difluoride membrane. The membrane was blocked with buffer containing 5% skim milk. Thereafter, Western blotting was performed with the primary antibodies for Bcl-2, Bax, c-PARP, Caspase-3, Caspase-9, and GAPDH overnight at 4°C; and then with horseradish peroxidase-conjugated secondary antibody for 2 hours at room temperature. The protein bands were detected by enhanced chemiluminescence (ECL system; Amersham, UK) and analyzed by Gel-Pro32 software.

Statistical analysis

All experiments were performed three times. The results were presented as mean \pm standard deviation, and data were analyzed with SPSS software (version 13.0; SPSS Inc, Chicago, IL, USA). The statistical significance of differences was determined by one-way analysis of variance among multiple groups, and differences with $P < 0.05$ were considered statistically significant.

Results

Characterization of DNR-loaded NPs

DNR-loaded NPs had a spherical shape and dispersed uniformly under transmission electron microscope (Figure 1A); the mean size and ZP of DNR-loaded NPs were 176.4 ± 11.0 nm (Figure 1B Size distribution by intensity) and -19.24 ± 0.12 mV, respectively. Figure 2 shows the schematic representation of DNR loading into NPs. The drug entrapping efficiency was $75.12\% \pm 1.68\%$, and the loading efficiency was $5.32\% \pm 0.15\%$. All these results demonstrated that DNR-loaded NPs were relatively homogenous in size and stably stored.

Intracellular DNR

The relative fluorescence intensity of intracellular DNR ($FI_{\text{treated group}}/FI_{\text{control group}}$) in K562 cells was lower in DNR group than that in DNR-loaded NPs (equivalent DNR concentration) group (8.14 ± 0.29 vs 14.26 ± 0.39) (Figure 3A), and there was a significant difference between them ($P < 0.05$, Figure 3B). Otherwise, there was no significant difference between the PLGA-PLL-PEG-Tf-NPs group and the control groups ($P > 0.05$, Figure 3B). These results suggested that DNR modified by PLGA-PLL-PEG-Tf-NPs could be targeted successfully to leukemia cells.

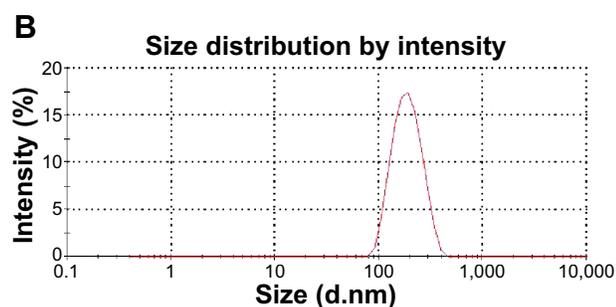
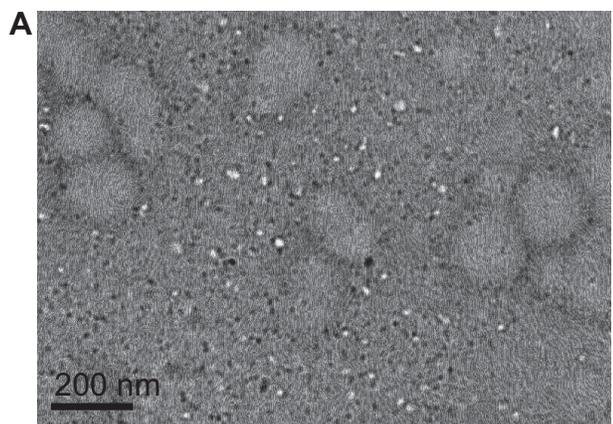


Figure 1 Physical image under transmission electron microscope (A) and particle size distribution (B) of DNR-loaded NPs.

Abbreviations: DNR, daunorubicin; NPs, nanoparticles.

Drug inhibits the growth of tumor tissue

After intravenous treatment for 18 days, the mean tumor volume was $1,099 \pm 238$ mm³ for the saline water group, $1,034 \pm 178$ mm³ for the PLGA-PLL-PEG-Tf-NPs group, 536 ± 137 mm³ for the DNR group, and 237 ± 85 mm³ for the DNR-loaded NPs group (Figure 4); and the inhibition of tumor growth in the DNR or DNR-loaded NPs groups was significantly higher than that in the PLGA-PLL-PEG-Tf-NPs group and the control group ($P < 0.05$). Interestingly, the mean tumor volume in the DNR-loaded NPs group was smaller than that in the DNR group ($P < 0.05$), while there was no significant difference between the PLGA-PLL-PEG-Tf-NPs group and control group ($P > 0.05$). These results were further demonstrated by gross-observation (Figure 5A). Moreover, the weight of the tumors from the animals treated with either DNR or DNR-loaded NPs decreased significantly when compared with the PLGA-PLL-PEG-Tf-NPs group and control groups ($P < 0.05$; Figure 5B). Thus, we inferred that the DNR-loaded NPs had a greater antitumor efficacy than DNR ($P < 0.05$).

Toxicity in vivo

All the mice were alive, and they were weighed during the experiment period of 18 days. It is noteworthy that no significant differences in body weight ($P > 0.05$; Figure 6) and no abnormal findings in any group were observed.

Next, we carried out histological bioanalysis of organs to evaluate the potential side effects of NPs on the main organs of mice in vivo. There were no apparent histopathologic changes in the tissues, including lung, heart, liver, kidney, and spleen (Figure 7). These results indicate that DNR-loaded NPs are safe and have no obvious toxic effects on the main organs of mice.

Analysis of cell apoptosis in K562 xenograft tumors

Samples were analyzed to detect the localized green fluorescence of apoptotic cells and blue fluorescence of cell nuclei using fluorescence microscope (Figure 8A). The percentage of apoptotic cells compared to the average number of cells per field was determined and graphically represented (Figure 8B). The apoptotic rate in DNR-loaded NPs group increased significantly compared with that in DNR group ($P < 0.05$). However, there was no significant difference between the PLGA-PLL-PEG-Tf-NPs group and the control group ($P > 0.05$), indicating that DNR-loaded NPs are a more effective drug treatment to achieve apoptosis than DNR ($P < 0.05$).

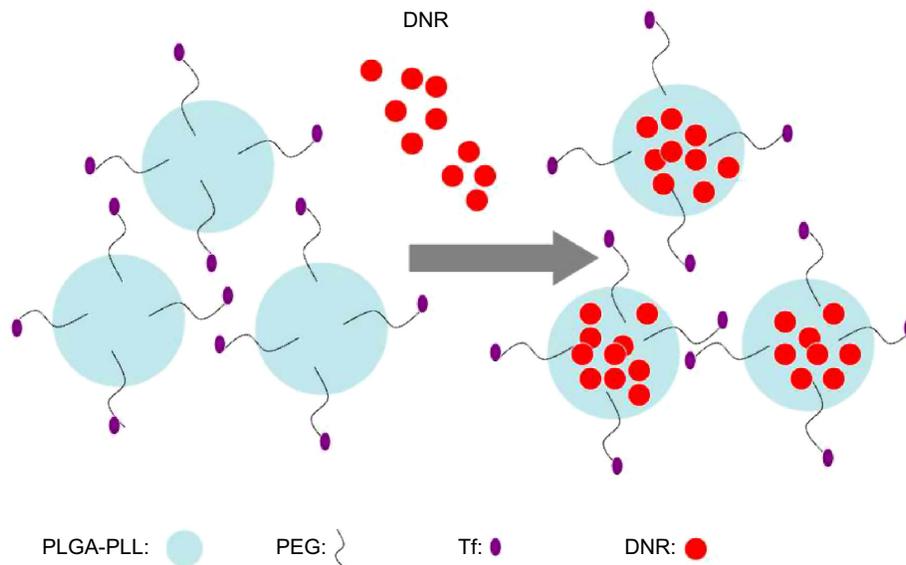


Figure 2 Schematic representation of DNR loaded nanoparticles.

Abbreviations: DNR, daunorubicin; PEG, polyethylene glycol; Tf, transferrin; PLGA, poly (lactic-co-glycolic acid); PLL, poly-L-lysine.

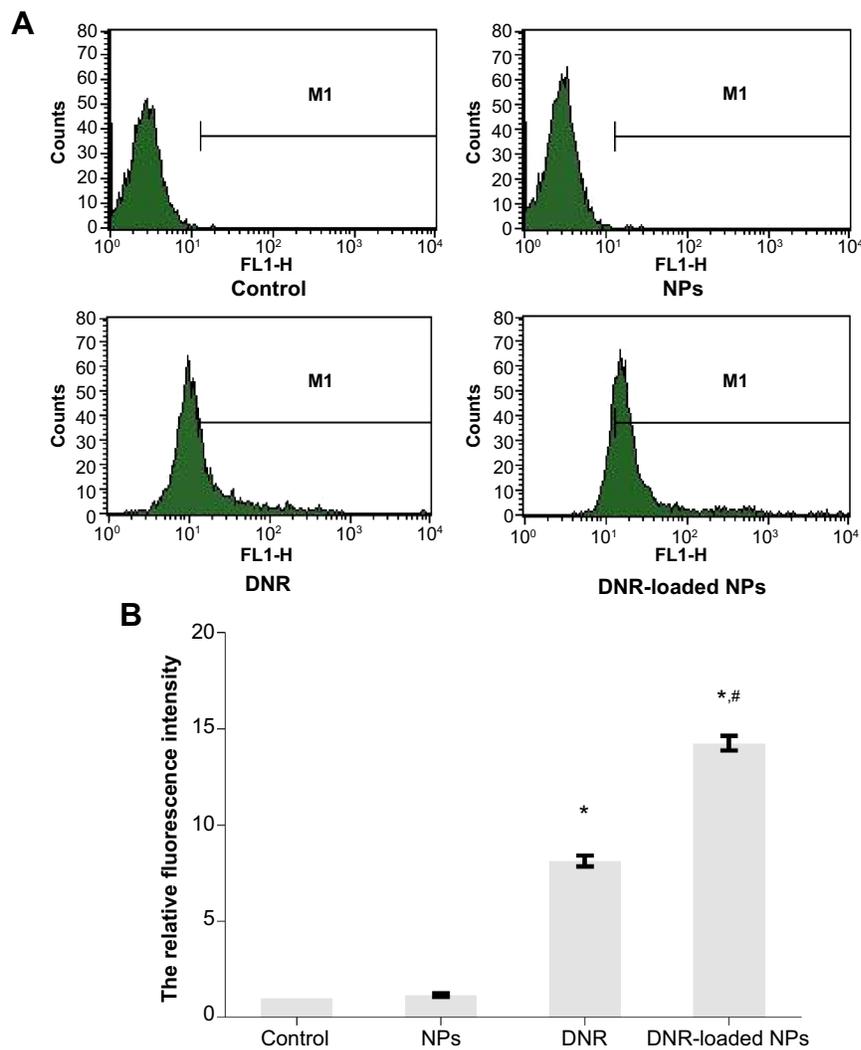


Figure 3 Intracellular accumulation of DNR in K562 cells incubated for 48 hours by FCM assay (A). Intracellular relative fluorescence intensity associated with DNR (B).

Notes: * $P < 0.05$ when compared with the control group and # $P < 0.05$ when compared with the DNR group.

Abbreviations: DNR, daunorubicin; FCM, flow cytometry; NPs, nanoparticles.

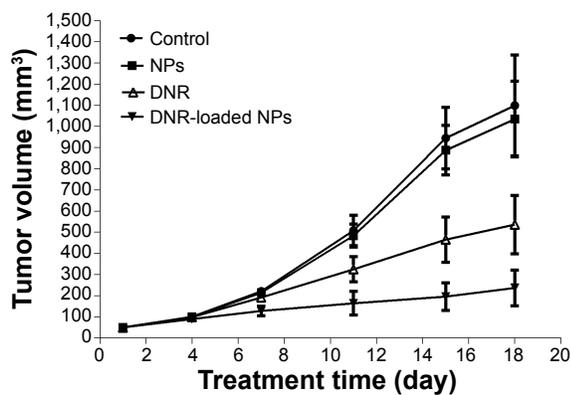


Figure 4 Tumor volume of mice after treatment for various times. **Abbreviations:** DNR, daunorubicin; NPs, nanoparticles.

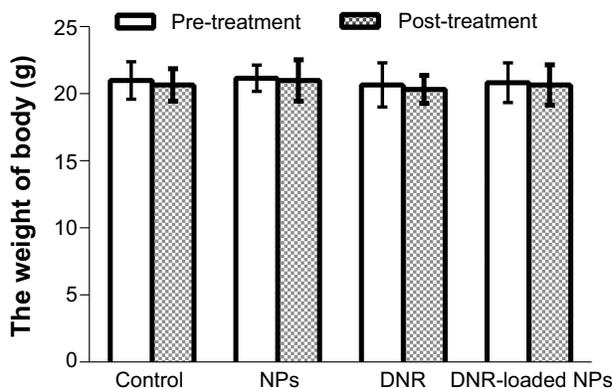


Figure 6 The body weight of mice pre- and post-treatment for various times. **Abbreviations:** DNR, daunorubicin; NPs, nanoparticles.

Expression of Bcl-2, Bax, cleaved-PARP, Caspase-3, and Caspase-9

To explore the possible signaling pathways through which DNR-loaded NPs induced more anticancer activity, we examined the changes in the expression levels of apoptosis-related proteins by Western blotting. In our study, the levels of apoptosis regulating proteins Bcl-2 in both DNR group and DNR-loaded NPs group were downregulated while that of Bax was upregulated compared with the control group ($P < 0.05$). In addition, the expression of Bcl-2 and Bax

proteins in DNR-loaded NPs group was more obviously regulated than that in DNR group ($P < 0.05$). The expression of Caspase-9 was upregulated in the DNR or DNR-loaded NPs group compared with the control group ($P < 0.05$), and the increase was much more dramatic in the DNR-loaded NPs group than that in the DNR group ($P < 0.05$; Figure 9). Similar results for Caspase-3 and cleaved-PARP were detected. Notably, the protein expression in the PLGA-PLL-PEG-Tf-NPs group was not obviously changed when compared with the control group ($P > 0.05$).

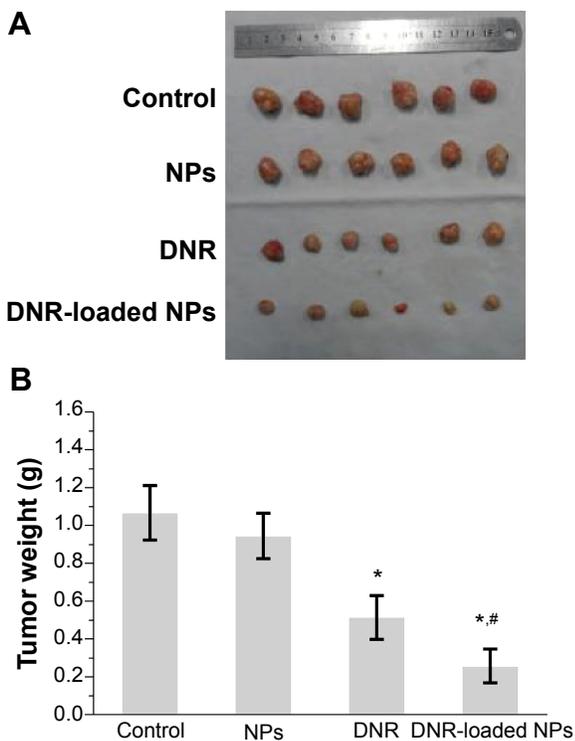


Figure 5 Appearance (A) and weight (B) of tumor at the end of the observation. **Notes:** * $P < 0.05$, when compared with control group, # $P < 0.05$, when compared with DNR group. **Abbreviations:** DNR, daunorubicin; NPs, nanoparticles.

Discussion

Current cancer treatments are restricted to chemotherapy, radiation, and surgery. Radiotherapy and surgery are largely localized in nature and fail to remove occult metastatic disease that can lead to eventual recurrence and poor patient survival. Nowadays, chemotherapy, particularly with respect to drug delivery, has evolved from traditional methodology.²⁰ DDS is an intriguing field of research that has captured the interest of researchers. Many studies focus on the safe and effective chemotherapeutic DDS, which can directly target tumor cells while sparing normal cells.¹ Targeted DDS can convey drugs more effectively and conveniently than chemotherapy in the past.²¹

As we know, nanotechnology has provided a wide field of research among chemistry, biology, medicine, material science, engineering, and mathematics. These innovations are referred to as nanomedicines by the National Institutes of Health and have a significant impact on drug delivery. NPs as carrier can reduce the cytotoxic side effects and achieve targeting chemotherapy.^{22,23} We had previously evaluated the application of NPs and found that it would provide a promising strategy for chemotherapy.²⁴

Tfs are a family of homologous iron-binding glycoproteins. A growing number of studies have demonstrated that

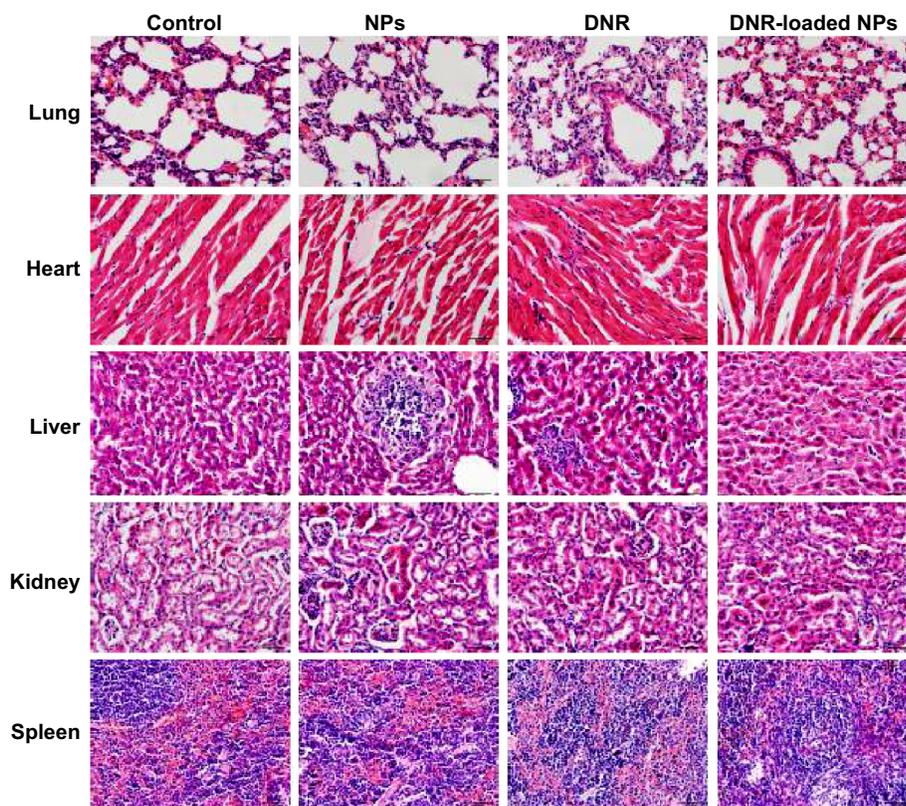


Figure 7 Histopathologic examination to determine the toxicity of therapeutic drugs on lungs, hearts, livers, kidneys, and spleens at the end of the experiments (H&E $\times 400$). **Abbreviations:** DNR, daunorubicin; NPs, nanoparticles; H&E, hematoxylin and eosin stain.

use of the Tf uptake pathway is highly effective in cancer therapy.²⁵ The TfR is a cell membrane-associated glycoprotein involved in the cellular uptake of iron and in the regulation of cell growth.²⁶ Many studies have shown that the level of TfR expressed in malignant cells is many times higher than their normal counterparts. Its expression can be correlated with tumor stage or cancer progression.^{26,27} Because of the elevated levels of TfR in malignancies, it can be targeted by directly conjugating with its ligand Tf or by monoclonal antibodies specific for the extracellular domain. Compared with free drugs, the combination of Tf targeting was highly effective in increasing levels of DNA double-stranded breaks and apoptosis in leukemia cells without accumulation of conjugate in normal cells, retaining the conjugate for longer periods of time, and declining the rate of chemotherapeutic agent efflux.²⁸ In an *in vivo* experiment, artemisinin-Tf conjugate demonstrated its ability to significantly retard the growth of breast cancer tumors in a rat model.²⁹ Tf-based DDS for cancer therapy has been tested to produce tumor responses without systemic toxicity in patients.¹⁵ On the other hand, a possible limitation of Tf-based therapeutics is the short period that Tf remains within the cells to deliver its payload. Recently, nano-based drug delivery has been demonstrated

to enhance the circulation time of targeted therapeutic drug.¹⁵ In accordance with these findings, we aim to explore whether the targeted delivery of a therapeutic drug DNR-loaded NPs can improve therapeutic benefit in leukemia. Currently, 100–200 nm is accepted as the optimal size for DDS because nanocarriers take advantage of the EPR effect in tumors and avoid filtration in the spleen.³ In the present study, spherical DNR-loaded NPs with diameters of 176.4 ± 11.0 nm could be produced with a remarkably high DNR entrapping efficiency ($75.12\% \pm 1.68\%$) and loading efficiency ($5.32\% \pm 0.15\%$), which is suitable for drug delivery.

Drug targeting can be classified into two types: passive and active targeting. PEG-modification of NPs is useful to passively deliver the NPs to cancer tissues via an enhanced permeability and retention, avoiding the reticuloendothelial system.^{30,31} Active targeting allows the increased accumulation of drug in cancer tissue through specific interactions, such as antigen–antibody and ligand–receptor binding. Endocytosis plays a major role in active targeting.¹⁵ The formulation of DNR-PEG-PLGA-PLL-Tf-NPs in our study was based on either passive or active processes. *In vitro*, we found that DNR-loaded NPs showed higher DNR accumulation than the DNR alone (Figure 3). *In vivo*, the tumor volume

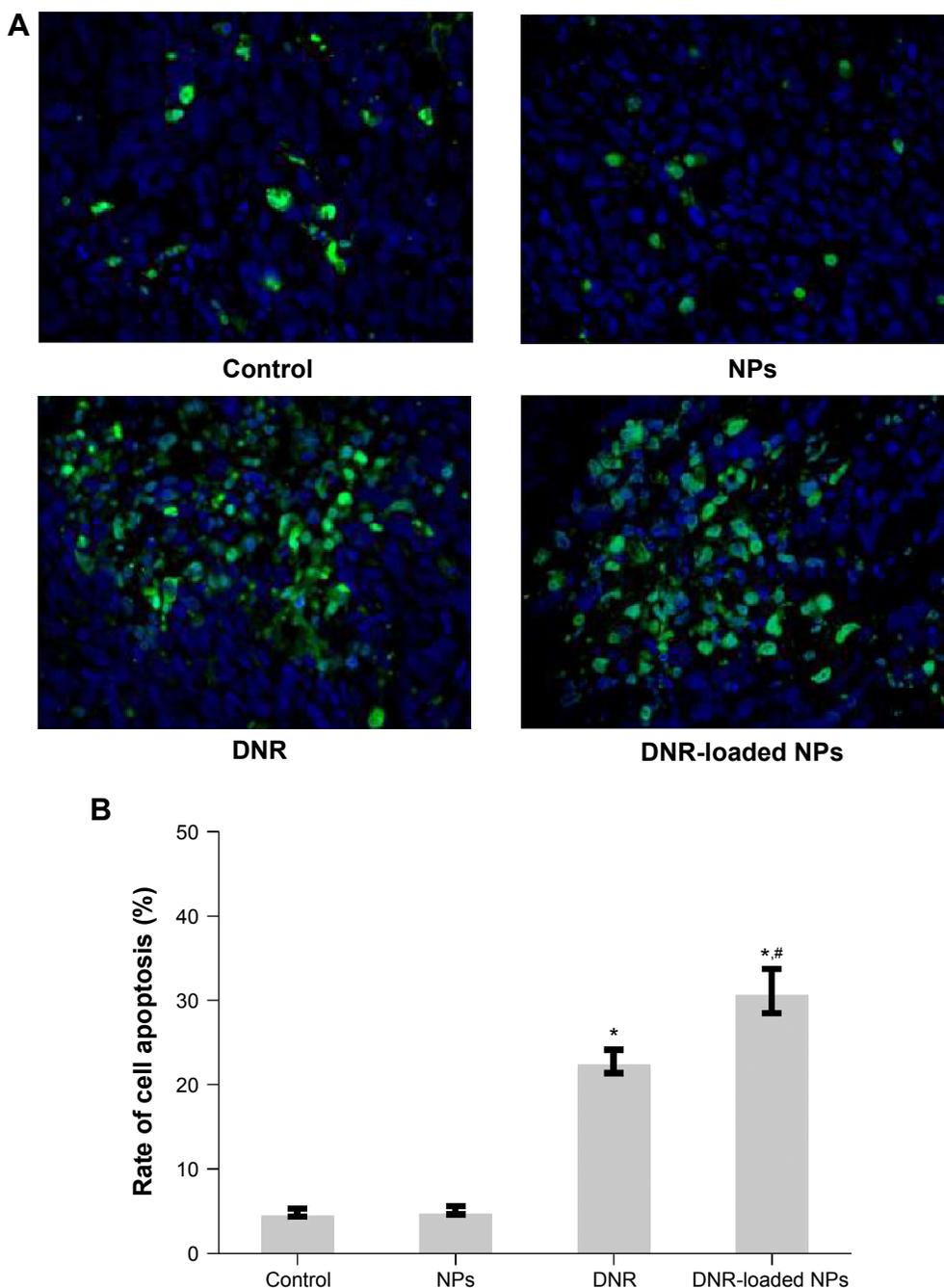


Figure 8 Apoptotic cells in K562 xenograft tumors.

Notes: (A) Representative images of the TUNEL assay (green fluorescence of apoptotic cells and blue fluorescence of cell nuclei were detected using fluorescence microscope $\times 400$); (B) rate apoptosis of each group. * $P < 0.05$ when compared with control group, # $P < 0.05$ when compared with DNR group.

Abbreviations: DNR, daunorubicin; NPs, nanoparticles.

of the mice injected with DNR-loaded NPs was smaller than that of the mice treated with DNR, and PLGA-PLL-PEG-Tf-NPs without DNR did not obviously affect the tumor volume in mice (Figures 4 and 5). These results indicated that DNR-loaded NPs showed more significant antitumor activity, and the intravenous delivery of DNR using NPs amplified the DNR antitumor that possibly elicited systemic targeting in leukemia. We infer that DNR-loaded NPs can

be directly delivered into tumor cells by receptor-mediated endocytosis and drug-sustained release from NPs surface with the NPs skeleton degradation. Thereby, DNR-loaded NPs may render longer half-life of DNR in blood, and DNR concentration must be increased in the tumor tissue. Besides, there were no apparent histopathologic damages to the main organs of mice due to the size of the NPs.³² Based on mice body weights, the tolerability of the NPs formulation was

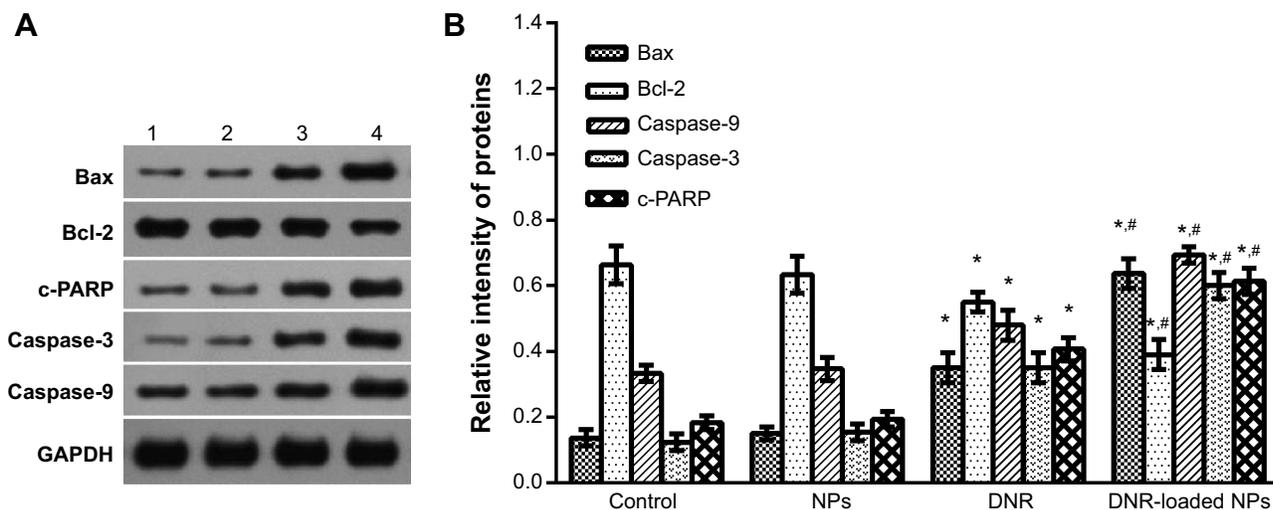


Figure 9 Expression of Bax, Bcl-2, Caspase-9, Caspase-3, and c-PARP after treatment.

Notes: Lane 1, control; Lane 2, NPs; Lane 3, DNR; Lane 4, DNR-loaded NPs. * $P < 0.05$ when compared with control group, # $P < 0.05$ when compared with DNR group.

Abbreviations: DNR, daunorubicin; NPs, nanoparticles.

acceptable. These results may advocate the feasibility of reducing the dose and intensifying the tumor-tissue targeting ability of DNR.

Apoptosis is a physiological process of programmed cell death that is essential for normal tissue development, and resistance to apoptosis is a hallmark feature of cancer; therefore, apoptosis induction of cancer cells is one of the most important and direct ways to control the development of cancer and eliminate tumors. In our present study, extensive cell apoptosis by TUNEL assay was observed in tumor tissues of xenografts treated with DNR-loaded NPs compared to the control one. Studies have shown that apoptosis can be triggered via two distinct pathways: the extrinsic and the intrinsic or mitochondria pathway.³³ Once cellular stress signals arise, the intrinsic pathway is activated and then causes permeabilization of mitochondrial outer membrane, which allows Caspase-9-mediated mitochondrial damage, thereby activating the downstream member Caspase-3.³⁴ To further explore the possible apoptosis pathways through which DNR-loaded NPs induced targeted chemotherapy, we assayed the expressions of some key apoptosis-associated proteins, including Caspase-3, Caspase-9, and c-PARP. In our study, the level of Caspase-9, Caspase-3, and c-PARP proteins was remarkably elevated in DNR-loaded NPs group compared to other groups. As we know, one of the most important functions of PARP is to help repair single-stranded DNA nicks, and the cleaved-PARP is critically involved in the intrinsic apoptosis pathway and considered to be a marker of apoptosis.^{35,36} In the present study, we observed the expression of proapoptotic and antiapoptotic members of the Bcl-2 family, which is the mitochondrial-related death switch. The members of

Bcl-2 homology are divided into three main groups based on function and regions of the domains: multi-domain anti-apoptotic proteins (Bcl-2, Bcl-xL, Bcl-w, Mcl-1, and A1), multi-domain proapoptotic proteins (Bax and Bak), and BH3-only proapoptotic proteins (Bid, PUMA, Bim, and NOXA), trigger the activation of Bak and Bax.³⁷ Our present results showed that the level of Bcl-2 protein was downregulated by the treatment of DNR-loaded NPs, compared to DNR group, whereas that of Bax was upregulated, and the ratio of Bax to Bcl-2 was increased. Taken together, these data indicate that DNR-loaded NPs can activate the intrinsic signaling pathway to trigger leukemia cells apoptosis.

Conclusion

In summary, the results obtained from the current study demonstrate that the novel NPs DDS enhances the activity of traditional anticancer drug DNR against leukemia cells through activation of the intrinsic apoptosis by both active and passive targeting pathways. This would ensure improved efficacy and minimize toxicity. Thus, novel NPs DDS may be a useful clinical tool.

Acknowledgments

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Disclosure

The authors report no conflicts of interest in this work.

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