

# Multidrug resistance protein 1 localization in lipid raft domains and prostasomes in prostate cancer cell lines

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**Background:** One of the problems in prostate cancer (CaP) treatment is the appearance of the multidrug resistance phenotype, in which ATP-binding cassette transporters such as multidrug resistance protein 1 (MRP1) play a role. Different localizations of the transporter have been reported, some of them related to the chemoresistant phenotype.

**Aim:** This study aimed to compare the localization of MRP1 in three prostate cell lines (normal, androgen-sensitive, and androgen-independent) in order to understand its possible role in CaP chemoresistance.

**Methods:** MRP1 and caveolae protein markers were detected using confocal microscopy, performing colocalization techniques. Lipid raft isolation made it possible to detect these proteins by Western blot analysis. Caveolae and prostasomes were identified by electron microscopy.

**Results:** We show that MRP1 is found in lipid raft fractions of tumor cells and that the number of caveolae increases with malignancy acquisition. MRP1 is found not only in the plasma membrane associated with lipid rafts but also in cytoplasmic accumulations colocalizing with the prostatic markers Caveolin-1 and CD59, suggesting that in CaP cells, MRP1 is localized in prostasomes.

**Conclusion:** We hypothesize that the presence of MRP1 in prostasomes could serve as a reservoir of MRP1; thus, taking advantage of the release of their content, MRP1 could be translocated to the plasma membrane contributing to the chemoresistant phenotype. The presence of MRP1 in prostasomes could serve as a predictor of malignancy in CaP.

**Keywords:** prostate cancer, MRP1, Caveolin-1, lipid rafts, caveolae, prostasomes

## Introduction

In the developed world, prostate cancer (CaP) is the most common non-skin cancer in men (899,000 new cases each year) and the fifth most common cancer overall. Nearly three-quarters of registered cases occur in developed countries (64,000 cases), and it is estimated to be the second leading male cancer in Spain in 2015.<sup>1</sup>

Localized CaP is the most commonly diagnosed stage. However, metastatic CaP still remains a major oncological problem in its follow-up. Despite initial response, most tumors relapse within 2 years to an incurable androgen-independent state.<sup>2</sup>

Chemotherapy is the main treatment to deal with hormonoresistant metastatic CaP. Unfortunately, CaP is also resistant to a broad range of antineoplastic agents, a phenomenon known as multidrug resistance (MDR) phenotype,<sup>3,4</sup> which plays an important role in the progressive resistant phenotype observed in CaP.

One of the mechanisms leading to the MDR phenotype is the activation of efflux proteins belonging to the ATP-binding cassette (ABC) transporter superfamily;<sup>5</sup>

P-glycoprotein (Pgp)<sup>6</sup> and the multidrug resistant protein 1 (MRP1) are members of this family with a putative role in CaP chemoresistance.<sup>7</sup> Although ABC transporters are mainly localized in the plasma membrane,<sup>8</sup> some of them have also been described in other cellular localizations such as lysosomes,<sup>9</sup> cytoplasmic vesicles,<sup>10,11</sup> and secretory vesicles putatively derived from the Golgi apparatus.<sup>5,12,13</sup> A more recent localization of ABC transporters is in specific regions of the plasma membrane rich in cholesterol and sphingolipids, called lipid rafts. Several functions have been attributed to lipid rafts such as signal transduction,<sup>14</sup> vesicle trafficking,<sup>15,16</sup> and cell adhesion and motility.<sup>17</sup> There is strong evidence that the lipid composition of biological membranes is closely related to the presence of ABC transporters.<sup>6,18–20</sup> In the case of MRP1, there are some controversies about the presence of this transporter in lipid rafts.<sup>21–23</sup>

A subset of lipid rafts is found in cell surface invaginations known as caveolae. Caveolae are formed from lipid rafts by polymerization of caveolins, integral membrane proteins that tightly bind cholesterol.<sup>24</sup> Elevated expression of caveolin-1 (Cav-1) is accompanied by the acquisition of MDR in various chemotherapy-resistant prostate tumor cells<sup>25</sup> and has been found in late stages of CaP progression.<sup>26</sup> Cav-1 is secreted by CaP cells, and the results of recent studies showed that secreted Cav-1 can stimulate cell survival and angiogenic activities, defining a role for Cav-1 in the CaP microenvironment.<sup>27–30</sup>

Additionally, Cav-1 has also been related to prostasomes, secretory granules stored and released by the glandular epithelial prostate cells. Cav-1 has been observed in prostasomes of the prostate cancer cell 3 line (PC3) human CaP cell line together with CD59,<sup>31</sup> a specific marker of these vesicles. Prostasomes have also been related to cell transformation, proliferation, and angiogenesis.<sup>32–35</sup>

In addition, there are studies that suggest that prostasomes may be implicated in CaP, and they are proposed as a prognostic indicator of tumor progression.<sup>36</sup>

Our studies have been focused on the analysis of a new localization for MRP1. We show that MRP1 is localized both in cytoplasmic accumulations and in plasma membranes of cell lines colocalizing with Cav-1. We also found MRP1 together with Cav-1 in lipid rafts, and this localization is more obvious in the cancer cell lines. The presence of caveolae is more abundant in the cancer cell lines compared with the normal cell line. Finally, we have demonstrated that MRP1 colocalizes with Cav-1 and CD59 in the accumulations found in the cytoplasm, concluding that MRP1 is found in prostasomes. The function of MRP1 in prostasomes needs to be

studied in more detail, but one of the hypotheses we propose is that prostasomes could serve as a reservoir/storage of MRP1 and, taking advantage of the fusion of prostasomes to the plasma membrane, MRP1 could be transported to the plasma membrane, participating in the development of CaP.

## Methods

### Cell culture

The normal human prostate epithelial cell line (PNT2) was provided by Dr Maitland (YCR Cancer Research Unit, University of York). The two tumor cell lines used were the androgen-dependent cell line lymph node carcinoma of the prostate (LNCaP) and the androgen-independent cell line PC3, both provided by The Prostate Center (Vancouver, Canada). All three cell lines were maintained at 37°C in a 5% CO<sub>2</sub> atmosphere and cultured in Roswell Park Memorial Institute (RPMI) 1640 supplemented with inactivated 10% fetal bovine serum, 1% L-glutamine, and 1% penicillin–streptomycin (Biological Industries, Israel Beit Haemek LTD). All cell cultures were mycoplasma free. Cancer cell lines LNCaP and PC3 derive from metastasis to lymph nodes and bone, respectively (American Type Culture Collection [ATCC], Manassas, VA, USA). They present intrinsic MDR phenotype, expressing MRP1, and acquired MDR when exposed to increasing doses of chemotherapeutics.

### Isolation of lipid rafts

Lipid rafts were isolated from 5×10<sup>7</sup> cells of each cell line. Pelleted cells were disrupted with 1 mL of raft II buffer (50 mM Tris HCl pH =8.0, 10 mM MgCl<sub>2</sub>, 0.15 M NaCl, 1% Triton X-100, 5% Glycerol, 50 mM PMSF, 25× cocktail of protease inhibitors [Complete™ EDTA-free; Biological Industries, Israel Beit Haemek LTD] [to a final 1× concentration] and 0.03% of β-mercaptoethanol). The disrupted pellet was centrifuged at 500×g for 5 minutes at 4°C. The supernatant was transferred into a 15 mL falcon tube and rotated for 1 hour at 4°C. Afterward, 1 mL of raft I buffer (50 mM Tris HCl pH =8, 10 mM MgCl<sub>2</sub>, 0.15 M NaCl) containing 80% sucrose was added to achieve a final concentration of 40% sucrose, followed by 7.5 mL of the same buffer containing 38% sucrose and 2 mL of buffer with 15% sucrose, thereby achieving a discontinuous sucrose gradient.

Finally, the gradient was ultracentrifuged at 100,000×g for 18 hours at 4°C in a SW41 rotor (Ti70 Beckman rotor). Fractions of 1 mL (12 fractions) were collected from the top of the gradient to the bottom (F1–F12 respectively). Immunoblot analysis was performed to confirm the fractions that contained lipid raft microdomains.

The isolation of lipid rafts was performed at least three times from three independent biological replicates to ensure reproducibility.

## Western blot analysis

Fractions obtained from the sucrose gradient were analyzed by SDS/PAGE on 7.5% or 12% (w/v) gel, transferred onto a nitrocellulose membrane, blocked in 5% nonfat dry milk and 0.05% Tween-20 in phosphate buffered saline (PBS). The primary antibodies used were Cav-1 (BD Transduction Laboratories), MRP1 (provided by Dr G Sheffer and Dr R Skepper of Free University Medical Center, Amsterdam, the Netherlands), and Flotillin-1 (BD Transduction Laboratories). Secondary antibodies were HRP-conjugated goat antimouse and goat antirat immunoglobulin, and the blots were developed using the ECL Western blotting kit (Amersham Biosciences) and visualized using LAS-3000 Fujifilm.

## Confocal microscopy

Cells were grown in 24-well plates on sterilized coverslips. Fixation and permeabilization was with methanol ( $-20^{\circ}\text{C}$  for 3 minutes), followed by two washes with PBS and blocked with 0.2% bovine serum albumin. The cells were incubated for 1 hour at room temperature (RT) with Cav-1 (1/500), MRP1 (1/200), or CD59 (1/1,000) (Exbio antibodies). After three washes with PBS, cells were incubated with: Alexa Fluor<sup>®</sup> 488 goat antirat IgG (molecular probes; Invitrogen) (1/400), Cys<sup>TM</sup>3 conjugated Affini Pure, fragment donkey antimouse IgG (Jackson ImmunoResearch, Laboratories, Inc, West Grove, PA, USA) (1/500), for 45 minutes at RT. The nuclei were stained with Topro III (Calbiochem) for 30 minutes at RT. Controls were done incubating the fixed cells with only the secondary antibodies. Finally, the cells were mounted on Mowiol (Calbiochem-Novabiochem) and visualized in a Leica TCS-SL confocal microscope.

## MRP1 quantification

To quantify the presence of MRP1 in both the membrane and the cytoplasm, the images from the confocal microscopy were examined and assessed by two investigators independently. By prior agreement, the intensity of MRP1 antibody was scored depending on the staining of cells in strongly positive (+++), moderate (++), or weak (+) staining. To obtain a numerical value with which to do the statistical analysis, a constant =1 has been added to the number of + and has been multiplied by the percentage of stained cells. For instance: a sample that has stained 30% of the cells with

moderate intensity (++) would be:  $(2 + 1) \times 30 = 90$ . Statistical analyses were performed using the Mann–Whitney *U* test.

## Electron microscopy

Cell monolayers were fixed with 2.5% glutaraldehyde in 0.1 M PBS for 2 hours at  $4^{\circ}\text{C}$ . Cells were gently scraped, collected, and pelleted by centrifugation (5 minutes at  $500\times g$ ). After three rinses in 0.1 M PBS, pellets were postfixed in 1%  $\text{OsO}_4$  –0.8%  $\text{FeCNk}$  for 90 minutes at RT. Finally, the three cell lines were embedded in Spurr resins (Sigma-Aldrich Co, St Louis, MO, USA), and ultrathin sections were analyzed with a Jeol1010 electron microscope. Grids were systematically screened, and the number of both caveolae and prostasomes was determined in random fields of sections from images acquired at the same magnification ( $25,000\times$ ). The images were examined and assessed by three investigators independently. For each cell line, over ten cells were examined with an estimated total linear surface of  $500\text{ }\mu\text{m}$ . Finally, for each cell line an average number of caveolae/ $\mu\text{m}^2$  and prostasomes/ $\mu\text{m}^2$  perimeter was calculated. Statistical analyses were performed using the Mann–Whitney *U* test.

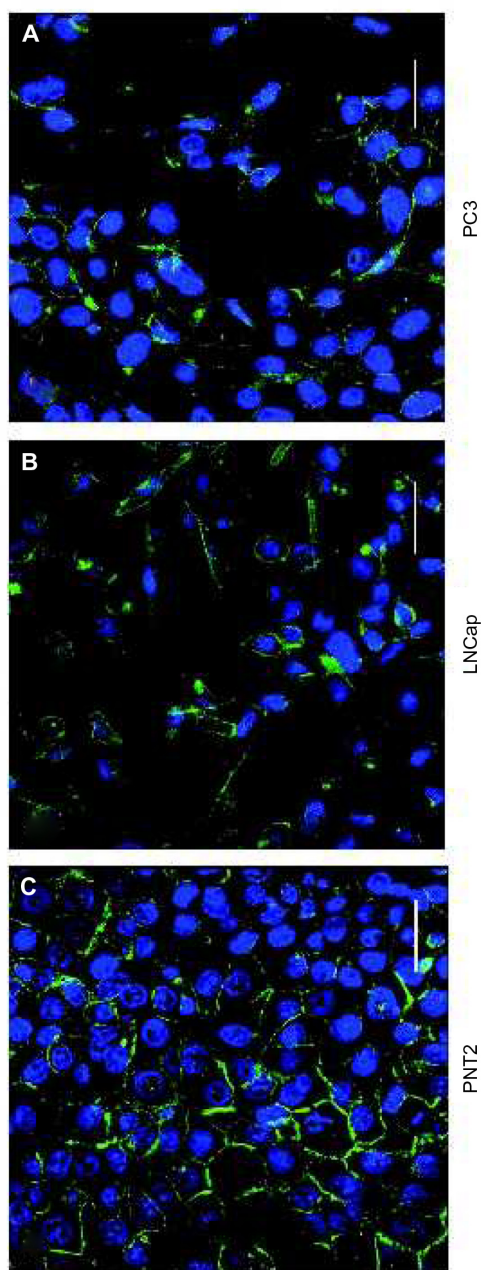
## Results

### Localization of MRP1 in prostate cancer cell lines

MRP1 was localized through immunocytochemistry techniques in two cellular compartments, with different distributions in normal versus cancer cell lines. In the normal PNT2 cell line, MRP1 was mainly localized in plasma membrane, more in some areas than others (Figures 1A, 2A, and 3A). On the other hand, in the cancer cell lines, MRP1 was observed not only in plasma membrane but also in cytoplasm regions (Figures 1B, C, 2D, G, 3D and J). These results were confirmed when the staining of MRP1 was quantified both in the membrane and in the cytoplasm of the three cell lines: the PNT2 cell line had more staining of MRP1 in the membrane ( $190.2\pm 46.7$ ) than in the cytoplasm ( $0.25\pm 0.5$ ). In addition, compared with malignant prostate cell lines, membrane MRP1 staining was higher in PNT2 than in LNCaP and PC3 ( $118.75\pm 14.6$  [ $P=0.029$ ] and  $98.7\pm 12.7$  [ $P=0.020$ ], respectively), whereas in the cytoplasm, MRP1 expression was higher in LNCaP and PC3 cells ( $33\pm 12.3$  [ $P=0.019$ ] and  $22\pm 6.9$  [ $P=0.017$ ], respectively).

### Localization of MRP1 in lipid raft domains

We analyzed the presence of MRP1 in lipid raft regions of the three cell lines. To that purpose, confocal microscopy was applied using Cav-1 as a typical lipid raft marker.



**Figure 1** MRPI localization in prostate cell lines.

**Notes:** (A) The ABC transporter was mainly localized in plasma membrane in the normal prostate cell line PNT2. Prostate cancer cell lines (B) LNCaP and (C) PC3 also showed cytoplasmic MRPI accumulations cell lines.

**Abbreviations:** MRPI, multidrug resistance protein 1; ABC, ATP-binding cassette; PNT2, normal human prostatic epithelial cell line; LNCaP, lymph node carcinoma of the prostate; PC3, prostate cancer cell 3 line.

Cav-1 expression was found in plasma membrane of the three cell lines, and also in cytoplasm accumulations in the case of LNCaP and PC3 (Figure 2B, E, and H; Figure 4A, D, G, J, and N). When a merging of MRPI and Cav-1 images was done, the two proteins colocalized both in the plasma membrane, especially in PNT2 (Figure 2C), and cytoplasm accumulations in LNCaP (Figure 2F) and PC3

(Figure 2I). Similar results were obtained when performing colocalization assays with Flotillin-1, another lipid raft marker (data not shown).

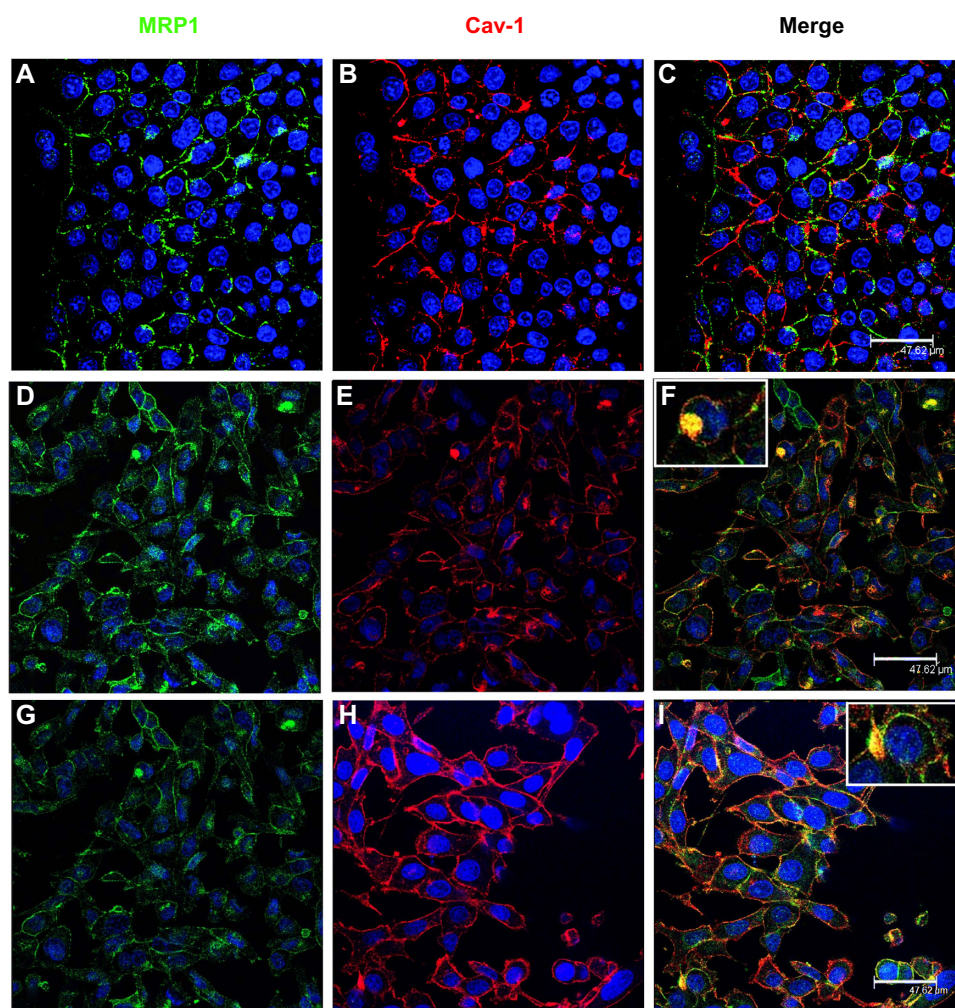
To verify the presence of MRPI in lipid raft domains together with Cav-1, lipid rafts were isolated followed by MRPI and Cav-1 detection in the different fractions (F) obtained (Figure 5). In all cell lines, Cav-1 localized in a different grade in the fractions that corresponded to lipid raft (F1–3), where Flotillin-1 (Flot-1), a protein also found in lipid rafts, was present. In PNT2, most of the MRPI was observed in the soluble fractions (F5–12) and also in small amounts in the last fraction (slightly in F2, and mainly in F3) (Figure 5A). In LNCaP, MRPI started to be detected in early insoluble fractions corresponding to lipid rafts (mainly in F3), and, similarly to PNT2, most MRPI was found in more soluble fractions (F3 and on) (Figure 5B). By contrast, MRPI entirely localized in the lipid raft fractions of PC3 (F1–2) and, to a lesser extent, in the soluble fractions (Figure 5C).

### Presence of lipid raft-related structures: caveolae and prostasomes

The presence of caveolae was detected in PNT2 (Figure 6A and B), LNCaP (Figure 6D and E), and PC3 (Figure 6G and H). The number was counted, and the ratio between the number and the perimeter of the cells was obtained. CaP cell lines contained twice as many caveolae as the normal prostate cell line (Table 1).

Due to the fact that MRPI and Cav-1 also colocalized in cytoplasm accumulations (Figure 2), and that lipid rafts are related to prostasomes,<sup>31</sup> the presence of prostasomes was also analyzed by electron microscopy. The three cell lines showed prostasomes, structures formed of vesicles approximately 0.5  $\mu\text{m}$  containing an electron-lucent substance and small vesicles and dense bodies in the inside (Figure 6C, F, and I). The shape of the storage vesicles was mostly circular. Two kinds of prostasomes were observed: “dense” prostasomes often filled with small dark particles or inclusions (Figure 6C and F), and “clear” prostasomes filled with less electro dense content. Prostasomes had thin flexible walls, which rendered them irregular in outline in those cellular areas where the vesicles were crowded (Figure 6I). When the number of prostasomes was counted, the androgen-independent and thus more aggressive PC3 cell line displayed a higher number of prostasomes compared with the other two cell lines (the ratio of prostasomes/ $\mu\text{m}^2$  was 2.77, 2.75, and 3.76 in the PNT2, LNCaP, and PC3 cell lines, respectively), the differences between the PNT2 and the PC3 cell lines being statistically significant ( $P < 0.01$ ).





**Figure 2** Colocalization of MRP1 and Cav-1 in the three cell lines.

**Notes:** (A–C) PNT2, (D–F) LNCaP, and (G–I) PC3 using confocal microscopy. MRP1 and Cav-1 colocalized in CaP cells in some regions of the plasma membrane and in cytoplasmic accumulations (F and I). A few colocalizations were found in the plasma membrane of normal prostate cells (C).

**Abbreviations:** MRP1, multidrug resistance protein 1; Cav-1, Caveolin-1; CaP, prostate cancer; PNT2, normal human prostatic epithelial cell line; LNCaP, lymph node carcinoma of the prostate; PC3, prostate cancer cell 3 line.

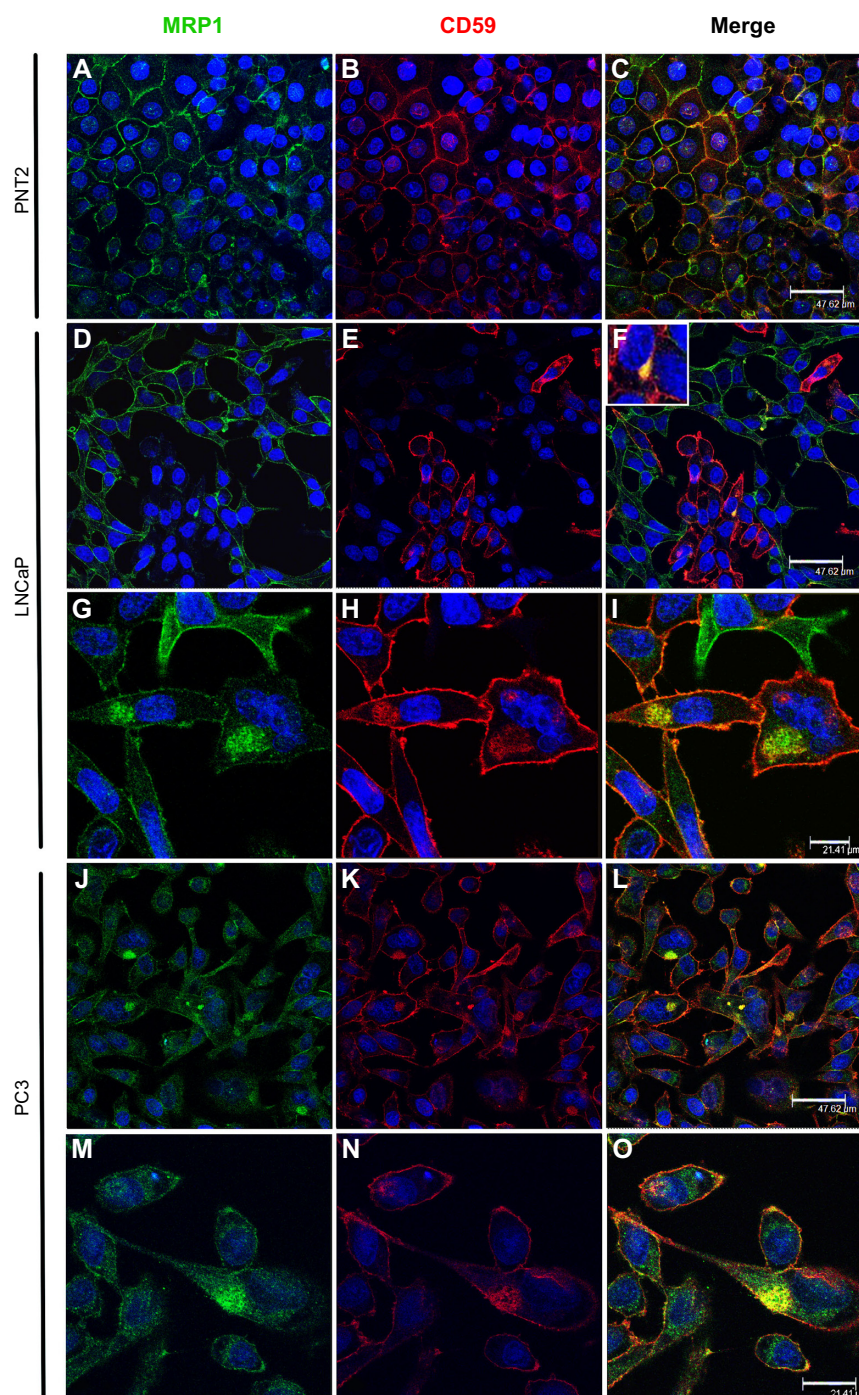
### Localization of MRP1 in prostasomes

To verify the presence of Cav-1 in prostasomes of tumor cell lines, colocalization studies of Cav-1 and CD59 (a positive marker of prostasomes) were performed in PNT2, LNCaP, and PC3 (Figure 4). As already observed, Cav-1 was localized in the plasma membrane of PNT2 (Figure 2), where some portions of the membrane presented stronger staining (Figure 4A). CD59 was also present in the plasma membrane, and some cells had some specific dots in the cytoplasm (Figure 4B). But the merge showed that in general, there was no colocalization (Figure 4C). In LNCaP and PC3, Cav-1 was found along the plasma membrane, stronger in specific regions than in others (Figure 4D, G and J, N, respectively) and, in contrast to PNT2, it was also found in cytoplasmic regions (Figure 4A). CD59 was found along the plasma membrane, also stronger in some regions and in

cytoplasmic accumulations (Figure 4E, H, L, and O). Cav-1 and CD59 colocalized in both cancer cell lines, in regions of the membrane where the individual staining for each of the proteins was strongest (Figure 4I and P). The cytoplasmic accumulations where Cav-1 and CD59 colocalized were, in most cases, close to the plasma membrane, although in a few cases these accumulations were not close to it (Figure 4F and upper left M).

The next step was to analyze if the cytoplasmic accumulations where Cav-1 and MRP1 colocalized were prostasomes, and for that purpose, the prostatic marker CD59 was used.

In PNT2, MRP1 and CD59 were localized only in the plasma membrane, with stronger staining in specific membrane regions (Figure 3A and B). CD59 was expressed only in some cells as nonhomogeneous cytoplasmic accumulations, and in these localizations, Cav-1 did not colocalize



**Figure 3** Presence of MRP1 in prostasomes.

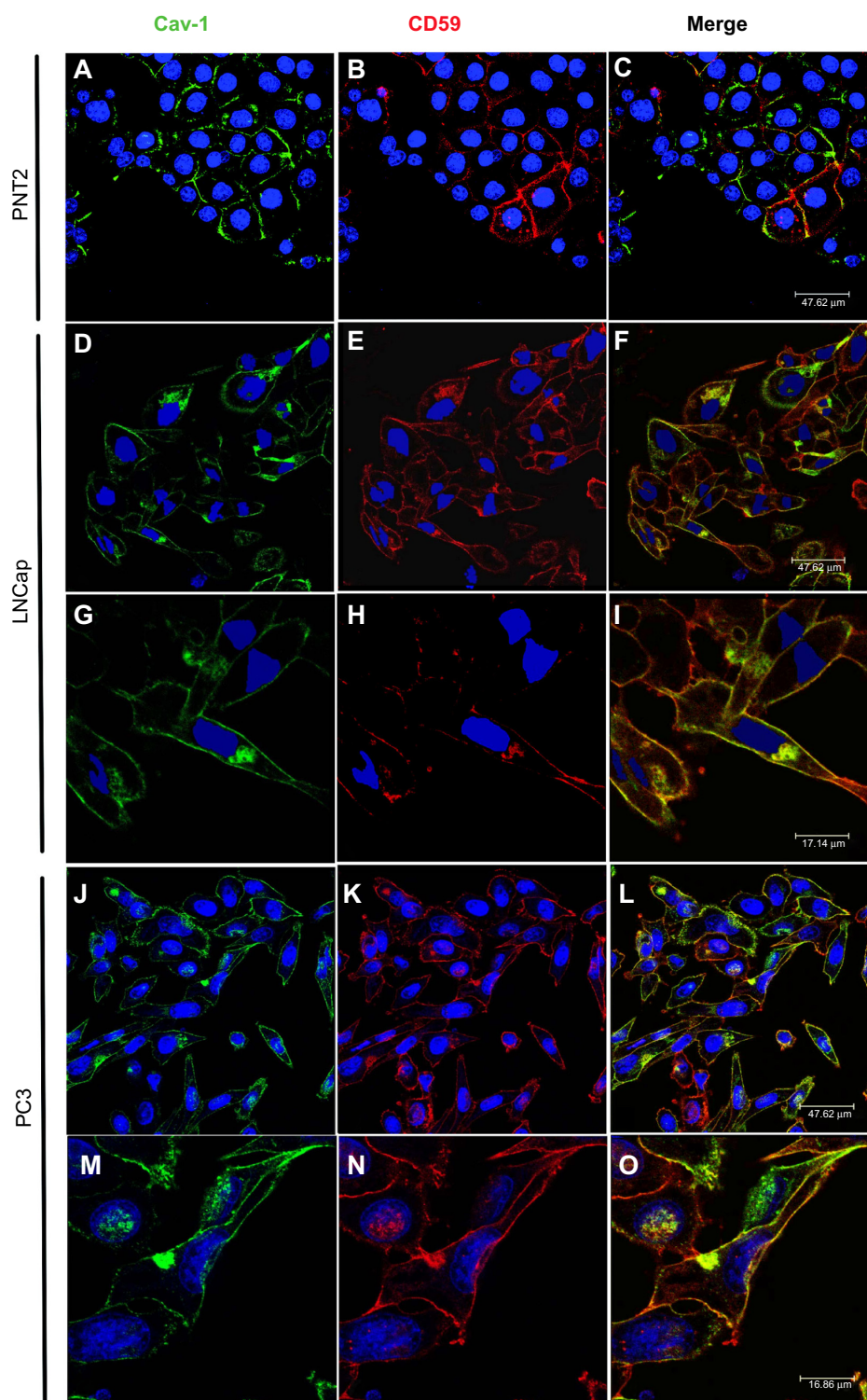
**Notes:** Colocalizations of MRP1 with the prostasome marker CD59 were performed to detect the ABC-transporter in these structures. Localization of MRP1 (green) in PNT2 (A), LNCaP (D and G), and PC3 (J and M) cell lines; localization of CD59 (red) in PNT2 (B), LNCaP (E and H), and PC3 (K and N) cell lines. MRP1 colocalized with CD59 in cytoplasmic accumulations in the cancer cell lines LNCaP (F and I) and PC3 (L and O) whereas it did not colocalize in the normal cell line PNT2 (C).

**Abbreviations:** MRP1, multidrug resistance protein 1; ABC, ATP-binding cassette; LNCaP, lymph node carcinoma of the prostate; PC3, prostate cancer cell 3 line; PNT2, normal human prostatic epithelial cell line.

with MRP1 (Figure 3C). Thus, in PNT2, MRP1 was not found in prostasomes. In LNCaP, MRP1 and CD59 were found in the plasma membrane (Figure 3D, E, G, and H) but not in a continuous way; curiously, the regions positive for MRP1 within a cell were negative for CD59, and all the way around.

There were also some cytoplasmic accumulations where both proteins colocalized (Figure 3F and I). This colocalization was very specific and observed only in a few cells. Finally, in PC3, MRP1 and CD59 were found along the plasma membrane in continuous staining in all cells (Figure 3J, K, M, and N).

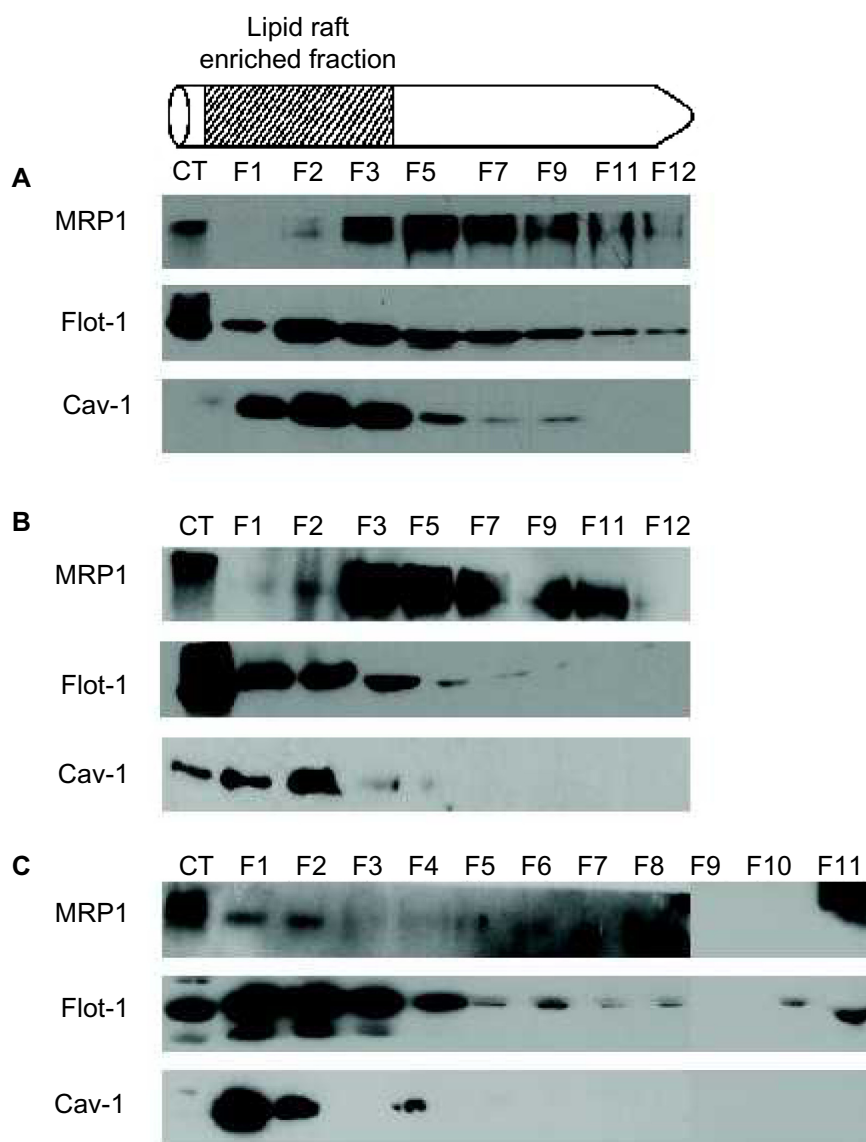




**Figure 4** Presence of Cav-1 in prostatosomes.

**Notes:** Colocalization studies of Cav-1 were performed in the three prostate cell lines using CD59, a prostatosome marker. Cav-1 (green) was present in normal prostate cell line PNT2 (A) and in the cancer cell lines LNCaP (D and G) and PC3 (J and M). CD59 (red) was also present in PNT2 (B), LNCaP (E and H) and PC3 (K and N) cell lines. Neither Cav-1 nor CD59 was found in prostatosomes in the PNT2 cell line (C), while a colocalization of Cav-1 and CD59 was found in CaP LNCaP cells (F and I) and in greater proportion in the androgen independent CaP cell line PC3 (L and O), indicating that in CaP cells Cav-1 is localized in CD59 positive structures, known as prostatosomes.

**Abbreviations:** Cav-1, Caveolin-1; CaP, prostate cancer; PNT2, normal human prostatic epithelial cell line; LNCaP, lymph node carcinoma of the prostate; PC3, prostate cancer cell 3 line.



**Figure 5** Lipid raft isolation from the three prostate cell lines.

**Notes:** (A) Lipid raft enriched fractions were isolated from the normal cell line PNT2 and showed MRP1 was present only in the last lipid raft enriched fraction, enriched with Cav-1. (B) When lipid raft fractions were isolated from the androgen-sensitive prostate cancer cell line LNCaP, MRP1 was already detected in early lipid raft enriched fractions together with Cav-1 (F1–F2) and in greater amounts in a later lipid raft enriched fraction (F3). Both PNT2 and LNCaP cells presented soluble fractions of MRP1 (F5–F11). (C) Lipid raft enriched fractions isolated from the androgen-independent prostate cancer cell line PC3 showed the presence of MRP1 mainly in the first two fractions (F1–F2). Flotillin-1 (Flot-1) was used as a control of the lipid raft isolation.

**Abbreviations:** PNT2, normal human prostatic epithelial cell line; MRP1, multidrug resistance protein 1; Cav-1, Caveolin-1; LNCaP, lymph node carcinoma of the prostate; PC3, prostate cancer cell 3 line.

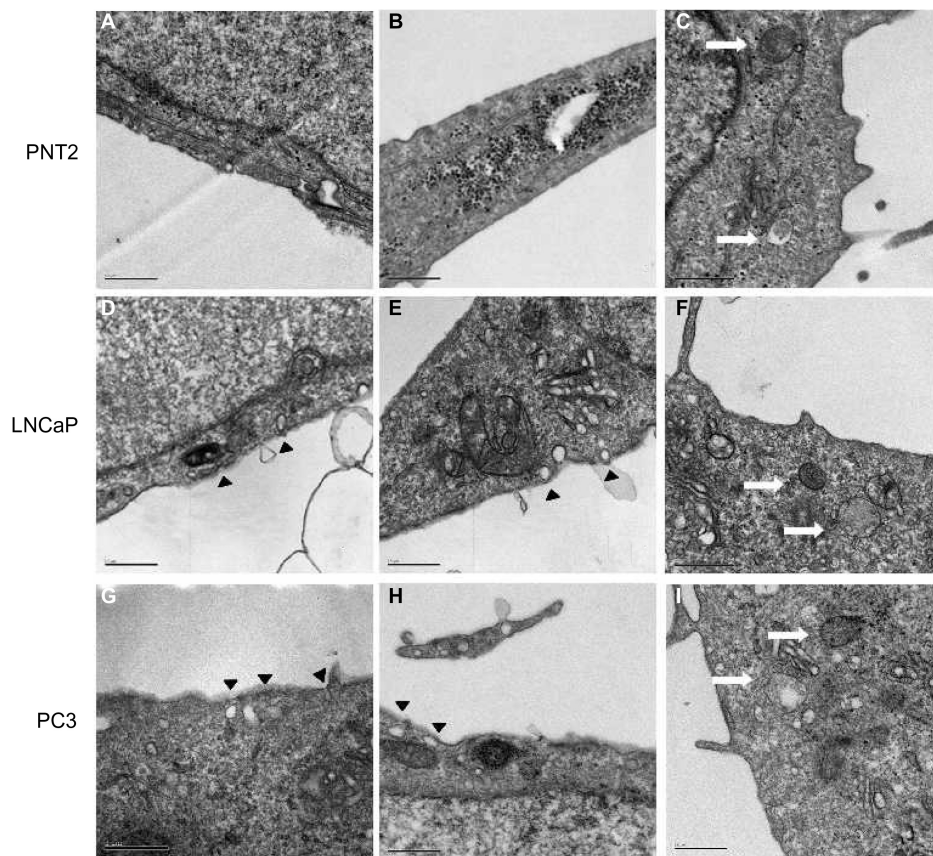
In a quarter of the cells observed, both proteins were also found in cytoplasmic accumulations where both colocalized (Figure 3L and O). Thus, in LNCaP and PC3, MRP1 colocalized in CD59 positive cytoplasmic regions, also positive for Cav-1, markers typically used to describe prostasomes.

## Discussion

The present study is focused on the cellular localization of MRP1 in CaP cancer cells with particular attention to lipid rafts. Lipid rafts lead to plasma membrane invaginations called

caveolae, and cytoplasmic vesicles called prostasomes. MRP1 was found in the three prostate cell lines, both in the plasma membrane and also in the cytoplasm. The fact that MRP1 had stronger staining in specific regions of the plasma membrane and that previous studies have reported the presence of other ABC transporters in lipid raft microdomains, led us to investigate the presence of MRP1 in these membrane regions. Lipid rafts have been implicated in many cellular functions, including protein and lipid transport and signal transduction. MRP1 and





**Figure 6** Representative electron micrographs of caveolae and prostasomes in the three prostate cell lines.

**Notes:** The presence of caveolae (black arrows) was observed in the normal PNT2 cell line (**A** and **B**), androgen-sensitive LNCaP cell line (**D** and **E**), and the androgen-independent PC3 cell line (**G** and **H**). Prostasomes were also localized in PNT2 (**C**), LNCaP (**F**), and PC3 cell lines (**I**) (white arrows).

**Abbreviations:** PNT2, normal human prostatic epithelial cell line; LNCaP, lymph node carcinoma of the prostate; PC3, prostate cancer cell 3 line.

glucosylceramide have been described to be overexpressed and enriched in lipid rafts during the acquisition of the MDR phenotype in colon cancer cells.<sup>22</sup> We found colocalization of Cav-1 and MRP1 in the plasma membrane, suggesting that MRP1 could also be localized in lipid rafts in CaP cell lines. Lipid raft isolation showed the presence of MRP1 since MRP1 was found in Cav-1 (and Flot-1) enriched fractions. In addition, MRP1 colocalized with Cav-1 not only in the plasma membrane but also in cytoplasmic accumulations. This phenomenon was observed only in CaP cell lines and not in PNT2, normal prostate cell line. Furthermore, the number of caveolae was counted and it was found to be double in LNCaP and PC3 compared with PNT-2, as Quest et al. observed in 2004.<sup>26</sup> MRP1 is known to have a major role in CaP.<sup>37</sup> Although previous studies described the presence of MRP1 in lipid rafts domains<sup>38</sup> an extended work needed to be done in CaP.

Studies by Llorente et al<sup>31</sup> showed that Cav-1 colocalized in cytoplasmic accumulations with the prostatic marker CD59, in PC3, and concluded that Cav-1 was localized in

prostasomes. Prostasomes are vesicles/granules secreted by prostate epithelial cells to the prostatic fluid that release Cav-1, contributing to cell survival and metastasis.<sup>25,28,31,39,40</sup> Our results showed that MRP1 colocalized with Cav-1 mostly in cytoplasmic accumulations where CD59 was also present. Confocal microscopy results of LNCaP and PC3 with Cav-1 and CD59 showed that the two proteins colocalized. Parallel experiments with MRP1 and CD59 also showed a colocalization between the two proteins, suggesting that MRP1 was in prostasomes. Our results showed a higher colocalization of MRP1 and Cav-1 in the androgen-independent PC3 cell line compared with the androgen-sensitive LNCaP cell line. Thus, PC3 malignancy seems to be in accordance with higher colocalization/expression of the two proteins.

When performing the colocalizations of MRP1 and CD59, in LNCaP there was a sort of “mirror effect”: most of the cells that expressed MRP1 did not express CD59, and those who expressed CD59, did not express MRP1, in LNCaP, a small percentage of cells presented colocalization of both proteins. This could be explained by the fact that perhaps there might be

**Table I** Counting of caveolae in the surface of the plasma membrane of the three prostate cell lines: PNT2 (normal), LNCaP (androgen-sensitive), and PC3 (androgen-independent)

Cell type	Perimeter of analyzed membrane surface ( $\mu\text{m}$ )	Number of caveolae	Ratio (caveolae/perimeter)
PNT2	213.63 <sup>a</sup>	45.66 $\pm$ 4.00	0.21 $\pm$ 0.02
LNCaP	325.62 <sup>b</sup>	145.66 $\pm$ 5.50	0.45 $\pm$ 0.02
PC3	294.67 <sup>c</sup>	103.33 $\pm$ 2.52	0.35 $\pm$ 0.01

**Notes:** Different sections of ten cells were counted ( $n=3$  times from the same sample) for caveolae, and the ratio of the number of caveolae per perimeter of analyzed surface was obtained. Prostate cancer cell lines have approximately double the number of caveolae that normal prostate cells do. <sup>a</sup> $P<0.001$  PNT2 compared with LNCaP; <sup>b</sup> $P=0.033$  PNT2 compared with PC3; <sup>c</sup> $P=0.123$  LNCaP compared with PC3.

**Abbreviations:** PNT2, normal human prostatic epithelial cell line; LNCaP, lymph node carcinoma of the prostate; PC3, prostate cancer cell 3 line; ns, not significant.

two different populations of cells within LNCaP: those cells less aggressive that do not express MRP1 in prostasomes, and those cells that have MRP1 in prostasomes colocalizing with MRP1, these being more aggressive cells. We could conclude that both CD59 and Cav-1 could act as prostasome markers when the cells are acquiring malignancy.

The fusion of prostasomes to the plasma membrane could enrich the plasma membrane with MRP1 while releasing Cav-1, contributing to the chemoresistant phenotype.<sup>29,41</sup> The presence of MRP1 in prostasomes could serve as a reservoir of the protein. Cancer cells that have more prostasomes would have less MRP1 in the plasma membrane, and all the way around.

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## Disclosure

The authors report no conflicts of interest in this work.

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