Upconversion nanoparticles conjugated with curcumin as a photosensitizer to inhibit methicillin-resistant Staphylococcus aureus in lung under near infrared light

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Abstract: Curcumin has phototoxic effects on bacteria under <450 nm irradiation, but it is unstable in vivo and cannot exert effects on deep tissues. Near infrared light (NIR) is harmless to the body and has stronger penetration than visible light. In order to improve the effects of curcumin, upconversion nanoparticles conjugated with curcumin (UCNPs-curcumin) are designed to upconvert NIR to the excitation wavelength of curcumin. UCNPs-curcumin were synthesized using polyethyleneimine to combine curcumin and UCNPs, based on typical composition of lanthanide nitrates \( \text{Re(NO}_3)_3 \) (Y:Yb:Er = 78.20:2%) linked by ethylenediaminetetraacetic acid in sodium fluoride (NaF) matrix, to upconvert NIR to 432 nm light. The product was characterized by size distribution, thermogravimetric analysis, differential scanning calorimetry, and scanning electron microscopy. Growth inhibition of methicillin-resistant Staphylococcus aureus (MRSA) was not only measured in vitro but also investigated on MRSA-induced pneumonia in mice. The results showed that curcumin was covered by UCNPs to form stable nanoparticles whose average size was 179.5 nm and zeta potential was ~33.7 mV in normal saline. The UCNPs-curcumin produced singlet oxygen, which reaches a stable level after 30 minutes of irradiation, and took effect on MRSA through bacterial cytoplasm leakage. They alleviated MRSA-induced pneumonia and reduced bacterial counts in lungs with 980 nm irradiation (0.5 W/cm\(^2\)) on chests of mice. It is confirmed that the UCNPs-curcumin in lungs are activated under NIR irradiation and strengthen their antibacterial effects on MRSA. This research provides a new type of NIR photosensitizer, which plays an important role in phototoxic effects of curcumin in deep tissues under NIR.

Keywords: curcumin, upconversion nanoparticle, near infrared light, methicillin-resistant Staphylococcus aureus, phototoxicity

Introduction
Curcumin (diferuloylmethane) is a well-known polyphenol from Curcuma longa L., which is used as a colorant in Asian food and has anti-inflammatory, antimicrobial, and anticarcinogenic activities.\(^1\) It mainly takes effect by photoxicity through production of singlet oxygen \((^{1}\text{O}_2)\) upon irradiation at <450 nm to kill bacteria and tumors.\(^2,3\) Because the penetration of the laser light depends on its wavelength,\(^4\) \(\leq450\) nm has weaker penetration than red light and near infrared light (NIR), leading to feeble action on deep lesions in vivo.

NIR falls in the region of the spectrum with the lowest absorption in tissue and therefore enables the deepest tissue penetration.\(^5\) It is also safe and causes minimal photo damage to the biological specimen involved, but it cannot activate curcumin.
In order to bridge this gap, a transducer that is capable of converting NIR light to excitating light of curcumin is required. Upconversion nanoparticles (UCNPs) can convert two or more low-energy pump photons from NIR to a higher-energy output photon with a shorter wavelength. In recent years, UCNPs have been developed for applications in biological labeling, sensing, and imaging. For most UCNPs, lanthanide ion codoped sodium yttrium fluoride (NaYF₄) nanoparticles possess one of the highest upconversion efficiencies, with tunable photoluminescence, controlled size, and low toxicity. To convert NIR to <450 nm light, we prepared NaYF₄ UCNPs to conjugate with curcumin as a photosensitizer to take effect in deep tissues. The physicochemical properties of the UCNPs, including particle morphology, size distribution, differential scanning calorimetry (DSC), and \( ^{1}O_2 \) production, were analyzed.

*Staphylococcus aureus* is a major pathogenic bacterium for pulmonary nosocomial infection. The disease is currently treated by antibiotics but easily acquires drug resistance, resulting in the deficiency of effective treatment. Methicillin-resistant *S. aureus* (MRSA), in particular, is resistant to most antibiotics and is responsible for significantly higher morbidity and mortality and increased health care costs. Even though curcumin shows the effect of photokilling MRSA in vitro, it is uncertain to be effective in deep tissue lesions such as pneumonia infected by MRSA.

We not only evaluated the inhibitory effect of UCNPs conjugated with curcumin on bacterial growth in vitro but also investigated their effect on MRSA-induced pneumonia in mice. The scheme is illustrated in Figure 1. Bacterial count and the expression of inflammatory cytokines tumor necrosis factor (TNF)-\( \alpha \) and interleukin (IL)-1\( \beta \) in lungs were analyzed after NIR radiation. Bacterial cytoplasmic leakage was measured to discuss the possible mechanism.

### Materials and methods

#### Chemicals
Curcumin (purity 98%) was purchased from Guangzhou Qiyun Biotech Company (Guangzhou, People’s Republic of China). MRSA (ATCC 25923) was obtained from the Chinese National Institute for Food and Drug Control. Yttrium oxide (Y₂O₃, 99.99%), ytterbium oxide (Yb₂O₃, 99.99%), erbium oxide (Er₂O₃, 99.99%), and sodium fluoride (NaF, 98%) were purchased from Shanghai Chemical Industrial Company (Shanghai, People’s Republic of China) and used as starting materials without further purification. Amoxicillin, TNF-\( \alpha \), and IL-1\( \beta \) kits were bought from Jiancheng Biotech Company (Nanjing, People’s Republic of China). Sodium ethylenediaminetetraacetic acid (EDTA-Na), polyethyleneimine (PEI), and other chemicals were of analytical purity and were purchased from Qianhui Reagent Company (Guangzhou, People’s Republic of China).

#### Animals
Sixty-four Kunming mice (half male and half female) weighing 20–30 g and aged 6–8 weeks were used for the experiment. They were kept in plastic cages at 25°C±2°C.
with 50%±10% humidity and on a 12-hour light–dark cycle with free access to pellet food and water. The animals were acclimatized to laboratory conditions for 1 week prior to the experiments. All procedures performed were reviewed and approved by the South China University of Technology and conformed to internationally accepted principles. All efforts were made to minimize animal suffering and to reduce the number of animals to be used.

Synthesis of UCNPs conjugated with curcumin

The UCNPs were synthesized by hydrothermal reaction following literature protocol with slight modifications. All the lanthanide nitrates Re(NO)₃ were prepared by dissolving the respective rare-earth oxides in nitric acid. Typically, 1 mmol of Re(NO)₃ (Y:Yb:Er=78%:20%:2%), 20 mmol of NaF, and 2 mmol of EDTA-Na were mixed in 50 mL water and heated to 180°C in autoclave and kept at this temperature for 20 hours. After cooling down to room temperature, the products were precipitated by ethanol, washed three times with water, and dried in vacuum, and 1.2 g of product was collected. Upconversion of the UCNPs was detected by F4500 Fluorescence Spectrometer (Hitachi Instrument Co, Ltd, Japan) with excitation wavelength of 980 nm.

The conjugation of UCNPs with curcumin (UCNPs-curcumin) was by means of molecular interaction and was carried out with the following procedure. One hundred and fifty milligrams of UCNPs, 100 mg curcumin, and 50 mg PEI were dissolved in dichloromethane and stirred at room temperature for 5 hours. The product (0.3 g) was obtained by evaporation of dichloromethane and washing with ethanol.

Particle size distribution and zeta potential

Particle size distribution and zeta potential of the UCNPs-curcumin were determined by Nano-2S MDT-2 Malvern Particle Size Analyzer (Malvern Instruments Ltd, Malvern, UK). Before testing, the appropriate sample was dispersed in normal saline with a refractive index of 1.33.

Morphological observation

Morphology of the UCNPs-curcumin was investigated by scanning electron microscopy (SEM, LEO 1530 VP, Germany). Ten milligrams of sample was suspended in 1 mL water, and 2 µL of the suspension was placed on a glass surface and dried in air. The samples were gold sputtered under high vacuum before analysis at an accelerating voltage of 20 kV.

Thermogravimetric analysis and differential scanning calorimetry analysis

Thermogravimetric (TG) analysis and DSC were carried out simultaneously using a Q6000-SDT Thermogravimetric Apparatus (TA Instrument Ltd, DE, USA) with a heating rate of 20°C/min from 30°C to 800°C under nitrogen atmosphere.

Determination of ¹⁸O₂

¹⁸O₂ was measured by the bleaching of p-nitroso-dimethylanilaine (RNO) according to the reference. The UCNPs-curcumin (0.5 mg/mL) were mixed with 50 mmol of RNO and 100 mmol of imidazole in 20 mmol of phosphate buffer (pH 7.4) and then irradiated by a 980 nm laser (0.5 W/cm², Threetop Optical Electric Technology Company, Dongguan, People’s Republic of China) for 10, 20, 30, 40, and 60 minutes. Power density of 0.5 W/cm² was applied on the basis of its safety and wide use in photodynamic therapy in vivo. The RNO absorption at 440 nm was determined by UV-3010 ultraviolet (UV) spectrometer (Hitachi Instrument Co, Ltd, Japan). The reduction of optical density (OD) at 440 nm can reflect the production of ¹⁸O₂. UCNPs and curcumin were treated as the controls in the same method.

Phototoxic assay against MRSA

The phototoxic activity was tested as reported. Aliquots of 100 µL of MRSA suspension (1×10⁶ colony-forming units [CFU]/mL) were individually transferred to separate wells of 96-well microtiter plates. An equal volume of the UCNPs-curcumin suspension in Mueller–Hinton broth (MH) was added to each well to give final concentrations of 10, 20, 40, 80, 160, and 320 µg/mL. After dark incubation at 37°C for 20 minutes (preirradiation time), each plate was placed under 0.5 W/cm² irradiation at 980 nm for 30 minutes. The incubation was kept in dark for 24 hours, and then OD value at 490 nm was measured with water as blank to calculate the bacterial growth inhibition rate. Minimal inhibitory concentration (MIC) was determined as the lowest concentration with clear wells. The phototoxic activities of UCNPs and curcumin were measured in the same way. UCNPs were directly dispersed in MH broth, and curcumin was dissolved in dimethyl sulfoxide to 30 mg/mL and then diluted to final concentration with MH broth.

Bacterial growth inhibition rate (%) = (Negative OD₄₉₀ – Drug OD₄₉₀)/Negative OD₄₉₀ × 100% (1)
OD$_{490}$, OD at 490 nm, reflects bacterial concentration. The wells with bacteria and solvent were used as negative controls.

**Cytoplasmic leakage of MRSA**

MRSA was monitored over time for cytoplasmic leakage via absorbance at 260 nm ($\Delta_{260}$) measurements according to the reference. $\Delta_{260}$ can reflect the content of amino acids and nucleic acids leaked out of the cell. $\Delta_{260}$ is the increase in the absorbance of bacterial supernatant at 260 nm compared with untreated controls. Bacteria were treated the same as phototoxic assay and incubated for 24 hours. Sample aliquots were centrifuged for 5 minutes at 10,000× g and room temperature to remove cellular debris. The supernatant was collected and the absorbance was detected at 260 nm by UV spectrometer to deduce cytoplasmic leakage.

**Antibacterial test in pneumonia mice induced by MRSA**

The pneumonia model was described by the reference with a little modification. Mice were divided into eight groups with eight mice in each group: normal control, negative control, amoxicillin group, and five medicated groups (UCNPs, curcumin, and UCNPs-curcumin at high, middle, and low doses) with irradiation. Mice, except the normal group, were intranasally infected with 50 μL inoculums of MRSA (1×10$^6$ CFU/mL) suspension in saline applied dropwise to the nares. The mice were allowed to aspirate the inoculums for 10 minutes before being returned to cages to recover. The mice were administered with vein injection of the UCNPs-curcumin at low dose (5 mg/kg), middle dose (15 mg/kg) and high dose (45 mg/kg) with 980 nm irradiation (0.5 W/cm$^2$) on chest for 30 min. The control mice were administered respectively with 45 mg/kg amoxicillin, 45 mg/kg UCNPs, and 45 mg/kg curcumin instead. The drugs were suspended in normal saline, and the dosage used was based on safety and effectiveness according to our previous research and the reference. Three days later, the mice were euthanized and lung tissue was removed and divided into two aliquots. One aliquot was plated onto Tryptose Soya Agar plates for bacterial colony count, and the bacterial density was expressed as the number of CFU per lung. Another was used for determination of TNF-α and IL-1β by enzyme-linked immunosorbent assay according to kit description. Briefly, lung tissue was weighed and homogenized in normal saline, then centrifuged at 1,000× g for 20 minutes. TNF-α and IL-1β in the supernatant were labeled by biotin antibody and horseradish peroxidase Avidin and detected at 450 nm.

**Statistical analysis**

Data were expressed as mean ± standard deviation ($\bar{x}$ ± s) and analyzed with SPSS Version 13.0 software (IBM, Armonk, NY, USA). Significant tests among the groups were based on one-way analysis of variance and Student–Newman–Keuls test.

**Results and discussion**

**Characters of the UCNPs-curcumin**

There have been several reports on the synthesis of UCNPs. However, most of these nanoparticles are synthesized at high temperatures. In recent years, a hydrothermal synthesis method at lower temperature has been found. Different constitutions of nanoparticles have various upconverting effects. Yb/Er codoped NaYF$_4$ nanoparticles have been reported as the most efficient infrared-to-visible upconversion fluorescent. Therefore, the NaYF$_4$ UCNPs were synthesized by hydrothermal method in our research.

Curcumin shows many pharmacological effects in vitro, but it has disadvantages such as limited absorption, poor bioavailability, and rapid metabolism and excretion. Linkage of curcumin with UCNPs may improve its stability in vivo. We conjugated UCNPs with curcumin to produce the new nanoparticles (UCNPs-curcumin). The results showed that the average size of the nanoparticles was 179.5 nm and the size distributed in the range 142–263 nm. The zeta potential was −33.7 mV, suggesting that UCNPs-curcumin are stable in normal saline because of negative charge repulsion. Plain nanoparticles (UCNPs) have an average size of 6.5 nm and zeta potential of −5.4 mV, showing smaller size but easier precipitation. Morphology of the UCNPs-curcumin was observed and photographed with SEM to identify microstructures of the nanoparticles. The SEM images of the UCNPs-curcumin displayed an oval shape with UCNPs binding together (Figure 2). Based on molecular structure, more than three UCNPs can be chelated by ion bonds with one molecule of PEI, which also binds to two molecules of curcumin by hydrogen bonds at branched chain (Figure 3). Many UCNPs are connected by PEI and curcumin and become a large particle.

In order to further evaluate the composition of the nanoparticles, DSC and TG are applied as a simple method to verify it. DSC and TG curves of the UCNPs-curcumin are shown in Figure 4. It exhibits three single endothermic peaks at 59°C, 183°C, and 250°C, respectively, representing the melting points of PEI, curcumin, and EDTA. TG patterns of the nanocapsules showed gradual loss of weight because of slow decomposition of PEI, EDTA, and curcumin.
Upon calculation of the weight loss by TG curves of the UCNPs and curcumin, the UCNPs-curcumin are made up of 55.5% UCNPs, 27.8% curcumin, and 16.7% PEI.

**Production of $^{1}$O$_2$**

The production of $^{1}$O$_2$ was evaluated by the change of RNO absorbance at 440 nm. RNO is a scavenger of $^{1}$O$_2$ and bleached by it. A$_{440}$ decreased significantly ($P<0.01$) after the treatment of the UCNPs-curcumin with 980 nm irradiation, and reached stability at a time interval of 30 minutes (Figure 5). The UCNPs and curcumin had no obvious change in A$_{440}$ alone. It suggests that the UCNPs-curcumin produce $^{1}$O$_2$ under irradiation, and 30 minutes of illumination are appropriate for their stable effect at 0.5 mg/mL concentration with 0.5 W/cm$^2$ irradiation.

Curcumin can be excited by 450 nm laser, but it cannot be activated by 980 nm laser. The UCNPs can upconvert 980 nm light to shorter wavelength. Upconversion tests showed that

![Figure 2 Scanning electron micrograph of the upconversion nanoparticles conjugated with curcumin at 5,000× (A) and 20,000× (B) amplification.](image)

![Figure 3 Possible structure of the upconversion nanoparticles conjugated with curcumin by polyethyleneimine (PEI).](image)
the UCNPs emitted four peaks (432 nm, 576 nm, 655 nm, and 860 nm) of light in the spectrum after excitation at 980 nm (Figure 6), and the 432 nm peak completely overlaps with absorption spectra. Among the four emission peaks, only light at 432 nm can activate curcumin to produce $^{1}O_{2}$. Others are too weak to excite curcumin. Because 432 nm light is about one-quarter of upconversion efficiency, how to increase the upconversion ratio of 980 nm to 432 nm is worth further research.

Phototoxicity of the UCNPs-curcumin on MRSA

The UCNPs-curcumin had inhibitory effects on MRSA growth at dose dependence accompanying 30 minutes of irradiation at 980 nm (Figure 7). MIC was 30 µg/mL less than the curcumin (130 µg/mL), and the MIC of curcumin was similar, as reported. It suggests that the UCNPs-curcumin had stronger phototoxicity on MRSA than curcumin.

Cytoplasmic leakage of MRSA was measured by the increase in the absorbance of bacterial supernatant at 260 nm compared with untreated controls. $\Delta A_{260}$ increased significantly ($P<0.01$) after treatment of the UCNPs-curcumin under 980 nm irradiation at concentration dependence (Figure 8). It suggests that the UCNPs-curcumin destroy MRSA cell membrane leading to leakage of cytoplasm. This is the possible mechanism of the UCNPs-curcumin inhibiting MRSA under irradiation.

Inhibitory effects of the UCNPs-curcumin on pneumonia in mice

Even though it is proven that curcumin can inhibit MRSA in vitro, it is difficult to play a role in pneumonia induced by MRSA in vivo because of its disadvantages, such as instability and low bioavailability. The UCNPs-curcumin improve the stability of curcumin and will have better effects in vivo. The lung, an internal organ, cannot be affected by visible light, since its penetrating depth is $<10$ mm; only NIR can reach the lung without harm and become an excellent light source to activate the UCNPs-curcumin in the lung.

Animal tests showed that the infectious lungs of mice had more colonies by bacterial counting. TNF-$\alpha$ and IL-1$\beta$ in the lungs of mice increased significantly when compared with normal mice, indicating pneumonia caused by MRSA in mice. Amoxicillin had little improvement on bacterial quantity and cytokines expression in the lungs of mice, suggesting...
antibiotic resistance of the bacteria. The UCNPs-curcumin with irradiation significantly ($P<0.01$) decreased bacterial counts in lungs with dose dependence (Figure 9), proving their effectiveness on inhibition of MRSA.

The concentration of UCNPs-curcumin of mouse lung was directly determined by fluorescence intensity at 860 nm after excitation at 980 nm. The results showed that there were, respectively, 3.2 µg/g, 16.5 µg/g, and 58.3 µg/g UCNPs-curcumin in lungs 2 hours after intravenous (IV) injection of UCNPs-curcumin at 5 mg/kg, 15 mg/kg, and 45 mg/kg. That is to say, about 0.3%–0.6% UCNPs-curcumin could reach the lungs and take effect after IV administration, indicating the lower effective dose.

In order to compare the effects of different treatments, the same IV administration was applied in this research. Even though inhalation is direct to deliver drugs to the lung, and may be a good way to increase drug concentration in lungs, like other drugs, it is difficult to measure the dose accurately in mice. This needs further investigation on different routes of administration for our next research.

TNF-α and IL-1β are key inflammatory cytokines and cause phosphorylation and degradation of inhibitor of nuclear factor kappa-B kinase, leading to translocation of nuclear factor-κB into the nucleus to promote expression of inflammatory genes. This experiment shows that MRSA increases the expression of TNF-α and IL-1β, triggering...
inflammation in the lung. Injection of the UCNPs or curcumin alone did not inhibit the bacterial growth and the expression of TNF-α and IL-1β in the lung, but they were reduced by the treatments of UCNPs-curcumin at dose dependence under 980 nm irradiation for 30 minutes. It confirms that MRSA-induced pneumonia is controlled by UCNPs-curcumin through activation by NIR to directly inhibit the bacteria in the lung.

Curcumin is a safe food additive with a maximum tolerance dose of 12 g/kg.33 UCNPs-curcumin are also safe at a dose of 5–45 mg/kg after vein injection. But irradiation excites the UCNPs-curcumin to produce $^{1}\text{O}_2$, which is harmful to cell membrane. The damage is relevant to irradiation power, time, and position. No injury to lungs has been found in our research with 0.5 W/cm² irradiation on the chest for 30 minutes. These parameters can be good references for future clinical application.

**Conclusion**

UCNPs-curcumin are synthesized by upconversion NaYF₄ nanoparticles conjugated with curcumin by PEI. The UCNPs are based on typical composition of Re(NO)₃(Y:Yb:Er=78%:20%:2%) linked by EDTA in NaF matrix, and can upconvert NIR to 432 nm light. The product is characterized by size distribution, DSC, TG, and SEM, confirming that curcumin is covered by the UCNPs to make stable nanoparticles. UCNPs-curcumin can produce $^{1}\text{O}_2$ after irradiation at 980 nm and inhibit MRSA growth by destroying cell membrane leading to bacterial cytoplasmic leakage. The antibacterial effect of UCNPs-curcumin is also proven by the control on MRSA-induced pneumonia in mice. It is mainly attributed to direct bacteria inhibition by phototoxicity through activation of the UCNPs-curcumin under NIR irradiation. The dosage, irradiation power, and time are crucial to protect normal tissue from injury. This research provides a new type of NIR photosensitizer, which improves the effects of curcumin in vivo. It is effective to inhibit MRSA-induced infections in deep organs, and has prospects in clinical application.

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**Disclosure**

The authors declare no conflicts of interest.

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