

Neural stem cell-derived exosomes mediate viral entry

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Background: Viruses enter host cells through interactions of viral ligands with cellular receptors. Viruses can also enter cells in a receptor-independent fashion. Mechanisms regarding the receptor-independent viral entry into cells have not been fully elucidated. Exosomal trafficking between cells may offer a mechanism by which viruses can enter cells.

Methods: To investigate the role of exosomes on cellular viral entry, we employed neural stem cell-derived exosomes and adenovirus type 5 (Ad5) for the proof-of-principle study.

Results: Exosomes significantly enhanced Ad5 entry in Cocksackie virus and adenovirus receptor (CAR)-deficient cells, in which Ad5 only had very limited entry. The exosomes were shown to contain T-cell immunoglobulin mucin protein 4 (TIM-4), which binds phosphatidylserine. Treatment with anti-TIM-4 antibody significantly blocked the exosome-mediated Ad5 entry.

Conclusion: Neural stem cell-derived exosomes mediated significant cellular entry of Ad5 in a receptor-independent fashion. This mediation may be hampered by an antibody specifically targeting TIM-4 on exosomes. This set of results will benefit further elucidation of virus/exosome pathways, which would contribute to reducing natural viral infection by developing therapeutic agents or vaccines.

Keywords: neural stem cell-derived exosomes, adenovirus type 5, TIM-4, viral entry, phospholipids

Findings

Exosomes released from cells are nanostructures with diameters of 30–200 nm.¹ These microvesicles are actively secreted into all body fluids, including blood, saliva, urine, and breast milk.^{2,3} Exosomes are routinely released at the plasma membranes of different cell types and are part of the intercellular communication network.^{1,3} Exosomes play important roles in adaptive immune responses to pathogens and tumors by transferring proteins, soluble factors, messenger RNAs, and microRNAs to the recipient cells.^{3,4} Notably, exosomes have been shown to transport retroviruses into macrophages⁵ and other immune cells.⁶ However, other particles such as prions can also be transported by exosomes.⁷ In this study, for the purpose of investigating the potential role of exosomes on virus entry, we prepared exosomes from neural stem cells and preliminarily evaluated the mediation of the exosomes in adenovirus type 5 (Ad5) entry into Cocksackie virus and adenovirus receptor (CAR)-deficient cells. CAR is a cellular receptor that specifically mediates the cellular entry of subgroup C adenoviruses (Ads), including Ad5.⁸

Generation of neural stem cell-derived exosomes

Exosomes have gained attention as effectors in the conditioned medium of stem cells. Neural stem cells have been shown to produce large amounts of exosomes.⁹ We

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sought to establish that we could collect a high concentration of neural stem cell-derived exosomes. Mouse neural stem cells were cultured in serum-free culture media for the purposes of extracting exosomes. In brief, culture medium from the cells was collected, filtered, and ultracentrifuged at $110,000\times g$ to pellet exosomes.⁴ In order to confirm the successful purification of exosomes, Coomassie staining and Western blot analysis were applied on sample preparations to detect two key exosomal markers, cluster of differentiation (CD) 81 and CD63. Sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) was performed, followed by Coomassie staining. The Coomassie staining demonstrated the presence of CD81 proteins, with a major band of 26 kDa and a minor band at ~ 37 kDa (Figure 1A). Western blot analysis with an exosome-specific antibody, anti-CD81, further confirmed the CD81 protein bands at both 26 kDa and 37 kDa (Figure 1B). These multiple species of protein

detected in both the Coomassie staining and Western blot analysis may indicate posttranslational modification (eg, glycosylation). In addition, SDS-PAGE and Western blot analysis were performed with an exosome-specific antibody, anti-CD63, further confirming the major, glycosylated CD63 protein band at 40 kDa (Figure 1C). In order to further verify the integrity of the exosomes, electron microscopy was performed on purified pelleted fractions. Figure 1D illustrates an electron micrograph of the mouse neural stem cell-derived exosomes. These validations ensured the integrity of exosomes for subsequent studies.

Mediation of exosomes on Ad transfer

Previous studies have shown that exosomes and other microvesicles increase the ability of viruses such as the human immunodeficiency virus (HIV) to enter the cells.¹⁰ Ad has a similar size as HIV. Therefore, as proof of principle,

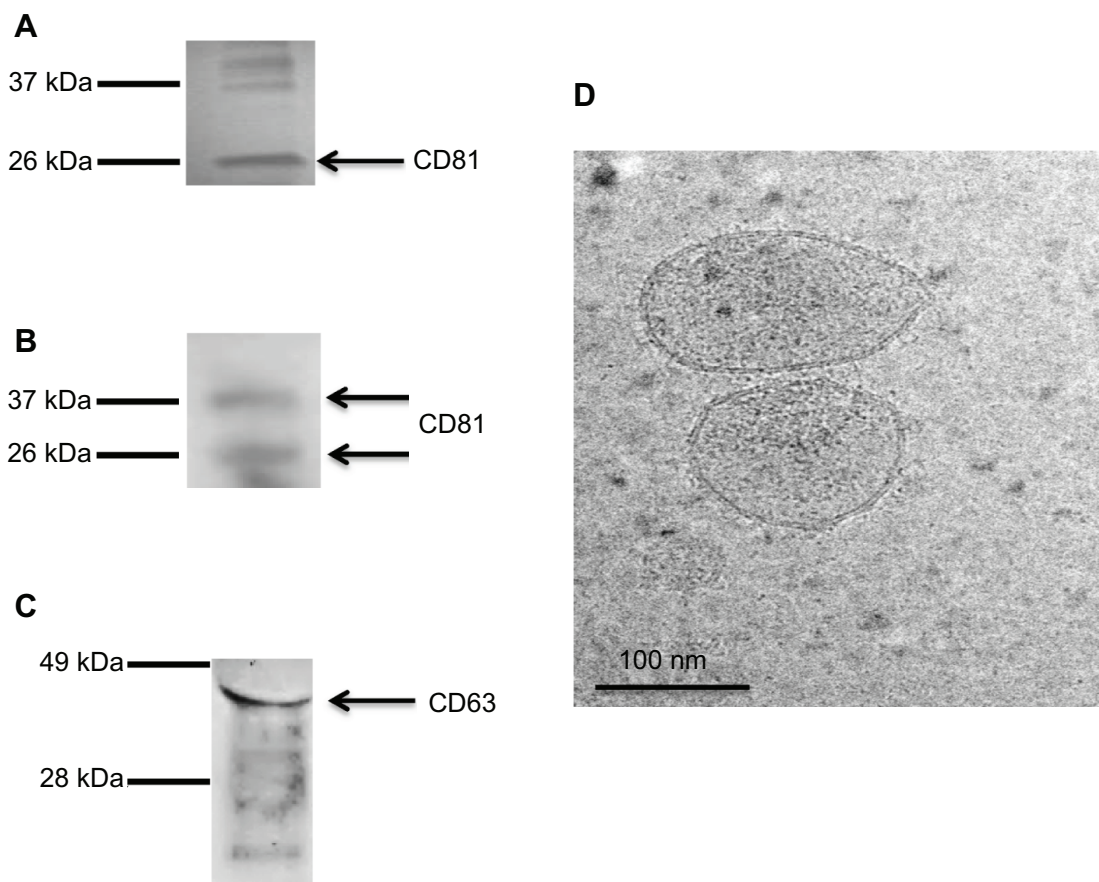


Figure 1 Validation of purified exosomes derived from mouse neural stem cells.

Notes: Five million mouse neural stem cells were cultured, collected, filtered, and ultracentrifuged at $110,000\times g$ to pellet neural stem cell-derived exosomes. Subsequently, $50\ \mu\text{g}/\text{well}$ of exosomes were subjected to SDS-PAGE, followed by (A) Coomassie Brilliant Blue staining for exosomal surface marker cluster of differentiation (CD) 81 and Western blot analyses for exosomal surface markers (B) CD81 and (C) CD63. (D) EM was employed to identify exosomes. The exosomes were prepared in a similar fashion as described. A $10\ \mu\text{g}/\text{mL}$ sample of exosomes were used for the EM analysis. All samples for EM were collected in our laboratory and processed at the University of Alabama at Birmingham Cryo-EM facility. The facility uses a FEI Tecnai F20 200 kV field-emission gun transmission electron microscope. Images were collected using a $4\text{k} \times 4\text{k}$ -pixel Gatan charge-coupled device camera.

Abbreviations: SDS-PAGE, sodium dodecyl sulfate polyacrylamide gel electrophoresis; EM, electron microscopy.

receptor-independent Ad5 entry via neural stem cell-derived exosomes was assessed. Using Ad5 (with a luciferase reporter) and exosomes, we validated exosome-mediated entry of Ad5 into a mouse B cell line, A20, which has deficient levels of CAR.¹¹ Luciferase activity was evaluated in different cell treatment groups (cell-only, exosomes, Ad, and Ad + exosomes groups) to illustrate the Ad5 entry level under different conditions. In brief, in the treatment group of Ad + exosomes, exosomes at three concentrations (0.1 μg , 1.0 μg , and 10 μg) were incubated with Ad5 (100 viral particles per cell) separately for 1 hour at 37°C before being applied to cells. Cells (1×10^6 per group) were then treated (cell-only, exosomes, Ad, and Ad + exosomes groups) for 24 hours. Cell-only, exosomes, and Ad groups were used as assay controls. Each group was washed to eliminate virus/exosome residues in the assay medium, and the cells were subsequently lysed. Lysates were subjected to luciferase analysis. In order for Ad to express the luciferase gene, the Ad/exosome complex would have to be taken up into the cells, followed by transcription and translation of the Ad genome and luciferase gene/protein. The data demonstrated that exosomes mediated the transfer of Ad5 to A20 cells, and there was significance at a quantity of 10 μg of exosomes (Figure 2). These data clearly suggested a significant quantity of receptor-independent transfer of virus into the B cell line via the mediation of exosomes.

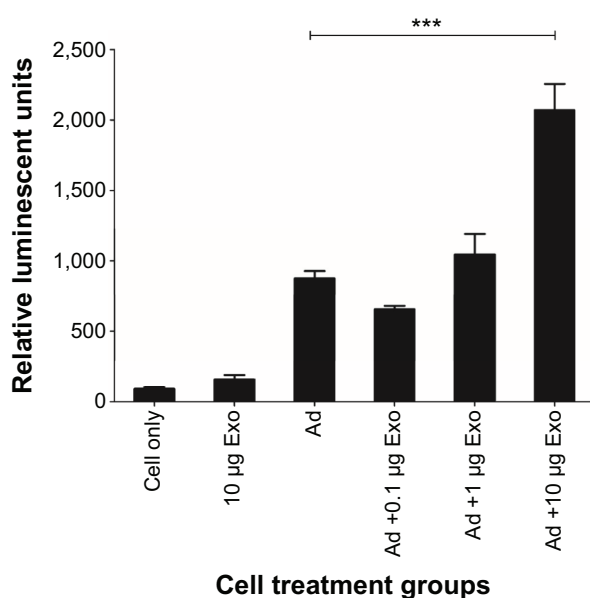


Figure 2 Mouse neural stem cell-derived exosomes mediate Ad5 entry into CAR-deficient A20 cells.

Notes: A20 cells were subjected to various treatments and subsequent detection of luciferase expression signals, which reflect the entry levels of Ad. Coincubation of 10 μg exosomes with Ad significantly facilitates the virus entry, as shown by the value $P < 0.001$ (***). Data in each treatment group represent means and standard deviations from three independent replicates.

Abbreviations: Ad, adenovirus; Ad5, adenovirus type 5; CAR, Coxsackie virus and adenovirus receptor; Exo, exosomes.

Blockage of exosome-mediated Ad entry by interfering with T-cell immunoglobulin mucin protein-4 interactions

In order to elucidate the mechanisms that facilitate exosome interactions, we focused on T-cell immunoglobulin mucin protein 4 (TIM-4)-mediated exosome interactions. It is well known that TIM-4 acts as a phosphatidylserine (PS) receptor. Exosomes derived from various cell types have been shown to contain PS-binding proteins.¹² We evaluated the presence of TIM-4 on neural stem cell-derived exosomes by Western blot (Figure 3A) and enzyme-linked immunosorbent assay (ELISA) analyses (Figure 3B). Purified exosomes were obtained as described in the section “Generation of neural stem cell-derived exosomes” (Figure 1). Exosomes were subjected to SDS-PAGE and Western blot analysis, using anti-TIM-4 antibody and donkey anti-goat secondary antibody conjugated with horseradish peroxidase. The data showed a protein band that resolved at ~64 kDa, which is likely to be glycosylated TIM-4 associated in exosomal preparations (Figure 3A). As further confirmation of the association of TIM-4 with exosomes, we performed an ELISA. Samples containing 0.1 μg , 0.5 μg , 50 μg , and 100 μg of exosomes were bound to an ELISA plate. The plate was then washed, blocked, and treated with anti-TIM-4 antibody. Subsequently, the plate was incubated with donkey anti-goat secondary antibody conjugated with horseradish peroxidase and subjected to plate reading at optical density of 450 nm. These data demonstrated that TIM-4 was present on neural stem cell-derived exosomes (Figure 3B). Because exosomes contain PS, Ad5 could potentially bind exosomes through interaction with anionic phospholipids,¹³ followed by entry of the Ad/exosome complex by means of interaction with anionic phospholipids. Of important note, we demonstrated that exosome-mediated uptake of Ad5 could be blocked with coincubation of anti-TIM-4 antibody (Figure 3C). As a negative control, we used a nonspecific antibody in this assay, and this antibody could not block exosome-mediated virus entry (data not shown). Typical PS-binding proteins can also bind phosphatidylcholine and phosphatidylethanolamine. Therefore, TIM-4 could potentially be associating with one of these phospholipids as well.

Virus trafficking is a complicated biological process. This process can be facilitated by nonclassic receptor-independent pathways such as exosomal trafficking. This pathway also allows virus to be delivered to a wide variety of cell types and organs that may not be a natural target for the viruses such as those described herein. This pathway is extremely important for retroviruses, which can be dormant and can be reactivated

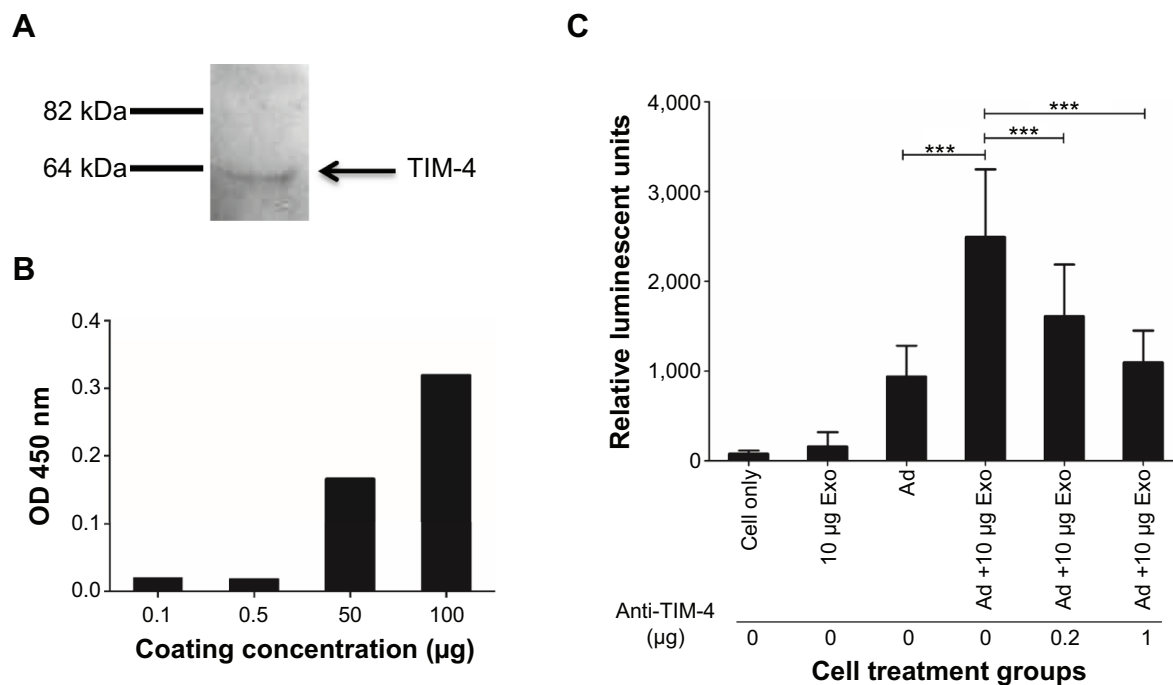


Figure 3 Exosome-mediated Ad5 entry was blocked by interfering with TIM-4 interactions.

Notes: (A) Western blot analysis and (B) ELISA were carried out to demonstrate the presence of TIM-4 on exosomes. A sample of exosomes (50 µg/well) was subjected to SDS-PAGE, followed by blotting with anti-TIM-4 antibody and appropriate secondary antibody. For ELISA experiments, increasing concentrations of exosomes were bound to an ELISA plate, washed, and treated with anti-TIM-4 antibody and appropriate secondary antibody. (C) A20 cells were subjected to various treatments and subsequent detection of luciferase expression signals, which reflect the entry levels of Ad. In the treatments of anti-TIM-4 + Ad + 10 µg Exo, anti-TIM-4, at different concentrations, was coincubated with Ad and 10 µg Exo simultaneously for 1 hour at 37°C before the three part mixtures were applied to cells. The luciferase signals, expressed as RLU, indicated that coincubation of anti-TIM-4 with exosomes and Ad significantly decreased exosome-mediated virus entry, with value of $P < 0.001$ (***). Data in each treatment group represent the mean and standard deviation from three independent replicates.

Abbreviations: Ad, adenovirus; ELISA, enzyme-linked immunosorbent assay; OD, optical density; RLU, relative luminescent units; SDS-PAGE, sodium dodecyl sulfate polyacrylamide gel electrophoresis; TIM-4, T-cell immunoglobulin mucin protein 4; Exo, exosomes.

in various cell types and organs. The interaction of Ad5 with neural stem cell-derived exosomes in Figure 2 is possibly through interaction of the Ad protein, hexon, with the anionic phospholipids.¹³ Subsequent entry of the Ad/exosome complex is possible by means of interaction with anionic phospholipid. This interaction allows receptor-independent entry of Ad virus into a CAR-deficient cell line. Data in Figure 3 highlight that TIM-4 interactions are involved in exosome-mediated transport. Further identification aimed at elucidating virus/exosome pathways may be important to reduce natural virus infection. In addition, elucidation of exosome trafficking can be a potentially therapeutic target or avenue for vaccine application.

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Author contributions

BS, LLG, and QLM designed the study; BS, LLG, and AK performed the experiments; BS, LLG, and QLM analyzed the

data; BS, LLG, and QLM wrote the manuscript; BS, LLG, AK, and QLM critically revised the manuscript. All authors gave final approval of the article to be published and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

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