Expression of YY1 correlates with progression and metastasis in esophageal squamous cell carcinomas

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Introduction

Esophageal squamous cell carcinoma (ESCC) ranks sixth among all cancers in mortality and is one of the deadliest cancers worldwide because of its highly aggressive nature and poor survival rate.1 The overall 5-year survival rate is only approximately 15%, and most patients die in the first year after diagnosis.2 However, the mechanisms regulating the malignancy and progression of ESCC remain under investigation, and clarifying the biological mechanisms leading to the development of esophageal tumors has become an important question in esophageal cancer research.

Transcription factors (also called sequence-specific DNA-binding factors) are specific proteins that can bind to specific DNA sequences and consequently regulate the flow of the genetic message from DNA to messenger RNA.3,4 Transcription factors have been reported to play important roles in cell growth, development, and differentiation,5 and there is substantial evidence that transcription factor dysfunction leads to detrimental outcomes, even tumorigenesis.6 Massive evidence indicates...
that transcription factors act as important players in human cancer development and progression, and it has been shown that the dysregulation of many transcription factors, such as Oct3/4, Sox2, KLF4, and EGR-1, is involved in the occurrence and development of esophageal cancer. For example, Oct3/4 and Sox2 overexpression leads to a poorer clinical outcome in ESCC patients. Although KLF4 acts as a tumor suppressor, in esophageal cancer, there is a high frequency of loss of heterozygosity of chromosome 9q31, on which KLF4 is located.

Yin Yang 1 (YY1) is a ubiquitous and multifunctional zinc-finger transcription factor that belongs to the Polycomb group protein family. YY1 has been shown to exert important biological functions in mammals, including embryogenesis, growth, differentiation, proliferation, and response to genotoxic stimuli. YY1 regulates gene expression by binding to DNA directly through its C-terminal zinc-finger domain. YY1 activates transcription in the presence of E1A, a protein that activates the AAV P5 promoter; however, the role of YY1 is reversed when E1A is absent, converting to a transcriptional repressor. Furthermore, YY1 can also regulate targets independently of its DNA-binding ability by mediating protein posttranslational modifications and acting as a transcriptional cofactor. YY1 modulates posttranslational modification by interacting with proteins that mediate posttranslational modifications, such as histone deacetylases, p300/CREB, and EZH2, and EZH1. Although YY1 plays an important role in normal growth, many studies have focused on its relationship with cancer. Because YY1 can regulate oncogenes and tumor suppressor expression, there is not yet a complete picture about the role of YY1 in various cancers.

It is reported that YY1 is overexpressed in many tumors, such as prostate, ovarian, breast, colon, liver, lung, cancers, melanoma, and leukemia. In a recent study, we found that YY1 was significantly increased in ESCC tissues compared with normal esophageal tissues or adjacent tumor tissues. However, it remains unclear whether there are differences between various ESCC patients and whether YY1 has a relationship with the development and progression of ESCC. In this study, we investigated the protein levels of YY1 in different ESCC tissue samples, and we discuss the relationship of YY1 expression with clinicopathological factors.

**Materials and methods**

**Patients and tissues**

All of 104 ESCC samples were collected from patients who underwent surgical treatment in Changzhou Tumor Hospital (Changzhou, People’s Republic of China). These patients consisted of 73 males and 31 females; their median age was 59 years, ranging from 32 to 76 years old. None of the patients accepted any chemotherapy or radiotherapy prior to surgery. All patients provided signed, informed consent for their tissues to be used for scientific research, and ethical approval for the study was obtained from Changzhou Tumor Hospital. All diagnoses were based on pathological and/or cytological evidence. The histological features of the samples were evaluated by a senior pathologist according to the classification criteria of the World Health Organization.

**Immunohistochemistry (IHC)**

An IHC analysis was performed as reported previously. The tissue samples were fixed in 10% neutral buffered formalin and embedded in paraffin. Three-micrometer-thick paraffin sections were deparaffinized, dehydrated, and heat-treated with citrate buffer (pH 6.0) for 15 minutes as an epitope retrieval protocol. The endogenous peroxidase activity was blocked with 3% hydrogen peroxide for 30 minutes, and the nonspecific binding sites were blocked with 4% skim milk powder for 30 minutes. After three washes in phosphate-buffered saline, the sections were incubated with a YY1 antibody (Santa Cruz Biotechnology, Santa Cruz, CA, USA; dilution 1:200) overnight at 4°C. The sections were mixed with 2% skim milk powder to reduce nonspecific staining, and a biotinylated secondary antibody was added for 30 minutes. Avidin–biotin–peroxidase complex (Dako LSAB2 system; DAKO Co, Carpenteria, CA, USA) was added, and the color was developed using 3,3′-diaminobenzidine. Counterstaining was performed with hematoxylin. All steps were carried out at room temperature.

The following standards were used to score the stained sections: negative (0), <10% of the entire tissue section stained positive; weakly positive (+1), 10%–25% of the entire tissue section stained positive; moderately positive (+2), 25%–75% of the entire tissue section stained positive; and strongly positive (+3), >75% of the tissue section stained positive.

**Cell culture and transfection**

The human ESCC cell line TE-1 was maintained in Dulbecco’s modified Eagle’s medium (DMEM) supplemented with 10% fetal bovine serum and antibiotics (100 units/mL penicillin G, 100 units/mL streptomycin sulfate; Gibco, Grand Island, NY, USA). The cells were grown in a 37°C incubator with 5% CO2. For transfection, the cells were transfected with plasmids using Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA), as reported previously.
**Table 1** Patient demographics for the immunohistochemistry analysis

<table>
<thead>
<tr>
<th>Age range (years)</th>
<th>Sex</th>
<th>Number</th>
<th>Stage I</th>
<th>Stage II</th>
<th>Stage III</th>
<th>Stage IV</th>
<th>All tumors</th>
</tr>
</thead>
<tbody>
<tr>
<td>55–69</td>
<td>Male</td>
<td>9</td>
<td>37</td>
<td>23</td>
<td>4</td>
<td>1</td>
<td>73</td>
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<tr>
<td></td>
<td>Female</td>
<td>6</td>
<td>15</td>
<td>10</td>
<td>0</td>
<td>1</td>
<td>31</td>
</tr>
</tbody>
</table>

**Tumor cell invasion assay**

TE-1 cell invasion was evaluated in vitro using a Transwell system (Corning, Tewksbury, MA, USA). The Transwell chamber includes a polycarbonate filter membrane with a pore size of 8 μm. Matrigel (12.5 μg; Sigma-Aldrich, St Louis, MO, USA) was diluted with 50 μL phosphate-buffered saline on ice and then added to the filter. After the gel formed a thin layer, TE-1 cells were suspended in 100 μL serum-free DMEM and added to the upper chamber of the Transwell insert. The lower chamber was filled with 500 μL DMEM plus 20% fetal bovine serum. After 24 hours of incubation at 37°C, the cells on the upper surface of the filter were removed using a cotton swab. The cells that adhered to the lower surface of the filter were fixed with methanol and then stained with hematoxylin. The cells were counted under a microscope.

**Statistical analysis**

A correlation analysis between the YY1 IHC staining score and clinicopathological factors was carried out using χ² tests and the Mann–Whitney U-test when only two groups were compared or the Kruskal–Wallis test when more than two groups were compared. The statistical analysis was performed using the SPSS software (Release 19.0, IBM Corporation, Armonk, NY, USA), and a P-value <0.05 was considered to be significant.

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**Results**

Patient demographics for the IHC analysis

To detect the expression of YY1, we collected a total of 104 ESCC tissues. The demographic features of the tissue samples are summarized in Table 1. For the total ESCC group, there were 15 cases at stage I, nine males and six females, with a median age of 62.8 years, ranging from 55 to 69; there were 25 cases at stage II, 37 males and 15 females, with a median age of 59.21 years, ranging from 33 to 76; there were 33 cases at stage III, 23 males and 10 females, with a median age of 56.53 years, ranging from 32 to 75; and there were four cases at stage IV, four males and zero females, with a median age of 66 years, ranging from 52 to 67.

**Association of YY1 protein expression with clinicopathological factors**

To study the relationship between the YY1 expression level and clinicopathological characteristics of the patients, including age, sex, histological grade, lymph node metastasis, TNM stage, and lymphatic invasion, IHC staining was performed on the 104 paraffin-embedded ESCC tissue samples.

In the IHC analysis, 55.7% (58 cases) of samples were positive for YY1 expression, and 44.3% (46 cases) were negative. Representative IHC staining of YY1 in ESCC tissues is shown in Figure 1. The relationship between YY1 expression and clinicopathological features is shown in Table 2. There were no obvious differences between age greater than 64 or less than 64 (P=0.286), and also no notable distinction between the males and females (P=0.255). Conversely, the protein level of YY1 was significantly higher in ESCC tissues with lymph node metastasis than in those without lymph node metastasis (P=0.042, Figure 2). Furthermore, YY1 protein expression...
Table 2 Relationship between clinicopathological parameters and protein expression of YY1 in esophageal squamous cell carcinoma (n=104)

<table>
<thead>
<tr>
<th>Clinicopathological parameters</th>
<th>YY1 expression</th>
<th>P-value</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>Cases (n)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Negative</td>
<td>Positive</td>
</tr>
<tr>
<td>Age (years)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;64</td>
<td>61</td>
<td>27</td>
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<td>≥64</td>
<td>43</td>
<td>19</td>
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<td>Histological grade</td>
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<tr>
<td>Well</td>
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</tr>
<tr>
<td>Moderate</td>
<td>48</td>
<td>18</td>
</tr>
<tr>
<td>Poor</td>
<td>31</td>
<td>16</td>
</tr>
<tr>
<td>pN+</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(+)</td>
<td>42</td>
<td>12</td>
</tr>
<tr>
<td>(-)</td>
<td>62</td>
<td>34</td>
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<td>10</td>
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<tr>
<td>4</td>
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<td>1</td>
</tr>
<tr>
<td>Lymphatic invasion</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(+)</td>
<td>49</td>
<td>16</td>
</tr>
<tr>
<td>(-)</td>
<td>55</td>
<td>30</td>
</tr>
</tbody>
</table>

Note: *pN: lymph node metastasis.
Abbreviations: pN, lymph node metastasis; YY1, Yin Yang 1.

was stronger in the stage III–IV patients than in the stage I–II patients (P=0.002, Figure 3), though the protein levels between the different histological grades (well, moderate, or poor) exhibited no statistical significance (P=0.752). Similarly, the expression of YY1 showed no differences in patients with or without lymphatic invasion (P=0.141). Taken together, YY1 expression was associated with TNM stage and lymph node metastasis but had no statistical correlation with age, sex, histological grade, and lymphatic invasion.

YY1 promotes the invasion of ESCC cells

It was found that the protein level of YY1 was significantly higher in ESCC tissues with lymph node metastasis than those without lymph node metastasis. As it remains unclear whether YY1 affects ESCC cell invasion, a classical Transwell system was used to investigate the effect of YY1 in esophageal cancer cell invasion. The results showed that the number of TE-1 cells migrating through the membrane increased after transfection with a YY1 overexpression vector, and significantly decreased after silencing with small hairpin RNA, indicating that YY1 increased the invasive ability of TE-1 cells and the inhibition of YY1 could reverse this promotion (Figure 4).
Discussion
YY1, a ubiquitously expressed transcription factor, has complex and diverse biological functions. YY1 can either activate or repress gene transcription depending on the stimuli received by the cells and the association with other cellular factors. YY1 belongs to the Polycomb group protein family, a group of homeobox gene receptors that play a key role in hematopoiesis and cell cycle regulation. YY1 was initially found simultaneously by two independent groups in 1991\textsuperscript{32,33} and is located on the telomere region of human chromosome

![Representative images of TE-1 cells transfected with pcDNA3.1 or pcDNA3.1-YY1 and shRNA-NC or shRNA-YY1.](image1)

**Figure 4** YY1 overexpression promoted the invasion of TE-1 cells and the low expression of YY1 suppressed the invasion.

**Notes:** (A) Representative images of TE-1 cells transfected with pcDNA3.1 or pcDNA3.1-YY1 and shRNA-NC or shRNA-YY1. (B) Calculated relative migrated cells after transfection with pcDNA3.1 or pcDNA3.1-YY1 and shRNA-NC or shRNA-YY1. Cells were counted under a microscope (*P<0.05).

**Abbreviations:** pcDNA3.1, pcDNA3.1 vector; pcDNA3.1-YY1, YY1 overexpression vector; shRNA-NC, small hairpin RNA control; shRNA-YY1, small hairpin RNA targeting YY1; YY1, Yin Yang 1.
The human YY1 gene produces eight different transcripts (named a, b, c, d, e, f, g, and h) generated by alternative splicing, encoding eight different putative protein isoforms. However, the functional significance of these different isoforms remains unknown.

YY1 has been reported to be overexpressed in multiple types of tumors, and it is known that YY1 overexpression may affect the clinical behavior of cancers. Thomasen et al found that YY1 is upregulated in metastatic breast cancer, and YY1 overexpression strongly correlates with the malignancy degree of osteosarcoma. Chinnappan et al reported that the YY1 expression level was more marked in poorly differentiated tumors than in moderately or well-differentiated colon carcinomas. To understand whether differences in YY1 expression levels occur in different clinicopathological ESCC, IHC staining was used to detect the protein levels of YY1 in 104 ESCC tissue samples. We found that the level of YY1 protein was higher in ESCC tissues with lymph node metastasis than in those without lymph node metastasis and was also higher in ESCC tissues of stage III–IV versus stage I–II. Thus, YY1 expression is associated with TNM stage and lymph node metastasis.

The role of YY1 in cancer progression is still a matter of heated debate and may depend on the type of tumor or other still-unknown factors. In a previous study, using the human ESCC TE-1 cell line, we found that YY1 could inhibit TE-1 cell proliferation by enhancing the binding of P21 to Cyclin D1 and Cyclin-dependent kinase 4, a protein complex known to mediate cell cycle progression. We also found that YY1 upregulated heme oxygenase 1 expression, which also inhibited ESCC proliferation. This evidence demonstrates that YY1 may act as a tumor suppressor in ESCC, at least in ESCC-derived TE-1 cells. In the present study, we found that YY1 promoted the invasion of ESCC cells and the inhibition of YY1 could reverse this promotion. Esophageal carcinoma normally metastasizes to the lymph nodes, and lymph node metastases and clinical stage are correlated with the prognosis of patients. In our study, which encompassed a relatively large number of ESCC patients, higher YY1 expression was associated with tumors with an advanced lymph node metastatic status ($P=0.042$) and tumors with a later TNM stage ($P=0.002$). Therefore, YY1 may be responsible for human esophageal cancer metastasis.

So, as in other types of cancer, many genes and pathways are involved in ESCC progression. YY1 is one of those genes that can inhibit ESCC proliferation and promotes the metastasis of ESCC. And these data suggest that the examination of YY1 expression might be helpful in guiding clinical management. However, the functional role and mechanisms of YY1 in esophageal cancer are still unclear and require further investigation.

**Conclusion**

In conclusion, the expression of YY1 is correlated with tumor stage and pN, and these findings suggest that YY1 plays a role in ESCC progression and metastasis.

**Acknowledgments**

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**Disclosure**

The authors report no conflicts of interest in this work.

**References**


42. Aurello P, D’Angelo F, Nigri G, et al. Comparison between site-directed mutagenesis of Wnt and YY1 expression correlates with progression and metastasis in ESCC