Genomic imprinting is an epigenetically regulated process determining parental-origin allele-specific expression. Beckwith–Wiedemann syndrome (BWS) is an imprinting disease related to 11p15.5 genetic and epigenetic alterations, among them loss-of-function CDKN1C mutations. Intriguing is that CDKN1C gain-of-function variations were recently found in patients with IMAGe syndrome (intrauterine growth restriction, metaphyseal dysplasia, congenital adrenal hypoplasia, and genital anomalies). BWS and IMAGe share an imprinted mode of inheritance; familial analysis demonstrated the presence of the phenotype exclusively when the mutant CDKN1C allele is inherited from the mother. Interestingly, both IMAGe and BWS are characterized by growth disturbances, although with opposite clinical phenotypes; IMAGe patients display growth restriction whereas BWS patients display overgrowth. CDKN1C codifies for CDKN1C/KIP2, a nuclear protein and potent tight-binding inhibitor of several cyclin/Cdk complexes, playing a role in maintenance of the nonproliferative state of cells. The mirror phenotype of BWS and IMAGe can be, at least in part, explained by the effect of mutations on protein functions. All the IMAGe-associated mutations are clustered in the proliferating cell nuclear antigen-binding domain of CDKN1C and cause a dramatic increase in the stability of the protein, which probably results in a functional gain of growth inhibition properties. In contrast, BWS mutations are not clustered within a single domain, are loss-of-function, and promote cell proliferation. CDKN1C is an example of allelic heterogeneity associated with opposite syndromes.

**Keywords:** Beckwith–Wiedemann syndrome, IMAGe syndrome, CDKN1C, genomic imprinting, growth disturbances

**Introduction**

Genomic imprinting is an epigenetically regulated process determining parental-origin allele-specific expression. Aberrant expression of imprinted genes characterizes imprinting diseases, including Beckwith–Wiedemann syndrome (BWS, OMIM 130650, chromosome 11p15.5). To date, eight imprinting diseases have been reported: Prader-Willi (OMIM 176270) and Angelman (OMIM 105830) syndromes (chromosome 15q11-q13), transient diabetes mellitus (OMIM 601410; chromosome 6q24), maternal/paternal uniparental disomy 14 syndromes (OMIM 608149; chromosome 14), Silver-Russell syndrome (SRS, OMIM 180860; chromosome 7 and 11p15), pseudohypoparathyroidism type IB syndrome (OMIM 603233; 20q13) and BWS. Generally, the chromosomal regions subjected to imprinting contain genes that are involved in growth and development (eg, *IGF2* and *H19*). This statement is in accordance with the model of the “struggle of the sexes”, based on the observation...
that paternally expressed genes are often growth promoting, while maternally expressed ones are growth inhibiting.3

Genetic and epigenetic alterations in BWS involve the imprinted chromosomal region 11p15.5 containing genes with expression controlled by two neighboring imprinted domains, ie, ICR1 (imprinting control region 1, regulating IGF2/H19 genes) and ICR2 (regulating KCNQ1/CDKN1C genes). ICR1 is imprinted in the male germline and operates as an insulator; ICR2 is imprinted in the female germline and acts as a promoter for the regulatory noncoding RNA KCNQ1OT1.4 The pathogenic alterations recognized in BWS (Table 1) comprise CDKN1C loss-of-function mutations present in the maternally derived allele.

The 11p15.5 region is also involved in another two syndromes with imprinting defects: SRS and the IMAGe (intrauterine growth retardation, metaphyseal dysplasia, adrenal hypoplasia congenita, and genital anomalies syndrome, OMIM 614732). IMAGe syndrome is not due to primary imprinting defects such as BWS and SRS, and is exclusively related to mutations of CDKN1C.5 This gene contains three exons and encodes p57(KIP2), a potent tight-binding inhibitor of several G1 cyclin/Cdk complexes (cyclin E-CDK2, cyclin D2-CDK4, and cyclin A-CDK2).6 It is a negative regulator of cell proliferation, playing a role in the maintenance of the nonproliferative state throughout life, probably acting as a tumor suppressor gene. CDKN1C is expressed in the placenta, heart, brain, lung, skeletal muscle, kidney, pancreas, and testis, in the eye, and in the subcapsular or developing definitive zone of the adrenal gland. It is paternally imprinted, with a preferential expression of the maternal allele;7 however, the imprint is not absolute, as the paternal allele is also expressed at levels comparable with the maternal one in the fetal brain.8

The codified protein consists of three distinct domains, ie, a cyclin-dependent kinase inhibitory domain, a proline and alanine repeat domain, and a QT domain (Figure 1). The cyclin-dependent kinase inhibitory domain contains a cyclin-binding region; the proline and alanine repeats interact with the LIM domain kinase 1 and regulate actin dynamics; the proliferating cell nuclear antigen (PCNA) binding domain, able to prevent DNA replication in vitro and S-phase entry in vivo.4 As in the RET gene,9 different mutations in CDKN1C can give rise to different phenotypes.

**Beckwith–Wiedemann syndrome**

The Beckwith–Wiedemann syndrome, described for the first time in 1963 by Beckwith, and again in 1964 by Wiedemann, is the most common overgrowth syndrome, with an incidence of about one in 13,700 live births.10,11 This occurrence is probably underestimated as milder phenotypes may not be ascertained.12 Males and females are affected equally, with the exception of monozygotic twins who show a female predominance.13 BWS occurs sporadically in most cases (85%), with the remaining cases having autosomal dominant inheritance.14

**Clinical diagnosis**

Clinical findings of BWS include macrosomia, macroglossia, abdominal wall defects (diastasis recti, omphalocele, or umbilical hernia), visceromegaly, hemihyperplasia, anterior ear creases and posterior helical pits, kidney abnormalities (medullary dysplasia, later development of medullary sponge kidney), cytomegaly of the adrenal fetal cortex, a positive family history of BWS, and, rarely, cleft palate. Height and weight are typically around the 97th percentile in children,

### Table 1 Molecular heterogeneity of BWS

<table>
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<tr>
<th>Genetic mechanisms</th>
<th>Occurrence (%)</th>
<th>Methods of detection</th>
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<tr>
<td>Paternal UPD</td>
<td>10–20</td>
<td>Microsatellite/SNP analysis, Southern blot, MS-MLPA, pyrosequencing, mass spectrometry</td>
<td>Hemihyperplasia, high risk of Wilms’ tumor and hepatoblastoma, severe phenotype (high level of somatic mosaicism UPD)</td>
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<tr>
<td>ID at ICR1 (gain of methylation)</td>
<td>2–8</td>
<td>Southern blot, MS-MLPA, pyrosequencing, mass spectrometry</td>
<td>Macroglosia, macroglossia, hemihyperplasia, high risk of Wilms’ tumor and hepatoblastoma</td>
</tr>
<tr>
<td>ID at ICR2 (loss of methylation)</td>
<td>50–60</td>
<td>Southern blot, MS-MLPA, pyrosequencing, mass spectrometry</td>
<td>Hemi hyperplasia, omphalocele</td>
</tr>
<tr>
<td>Paternal 11p structural rearrangements</td>
<td>1–2</td>
<td>FISH, aCGH</td>
<td>Developmental delay</td>
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<tr>
<td>Mutations of the maternal CDKN1C allele</td>
<td>5–10 (sporadic cases)</td>
<td>Sequencing</td>
<td>Cleft palate, omphalocele, genital anomalies, neuroblastoma</td>
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<td>40 (AD trait)</td>
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**Notes:** In column 4, the clinical features associated with the different genetic mechanisms are reported. The description is not exhaustive of the whole clinical presentation of BWS.

**Abbreviations:** BWS, Beckwith–Wiedemann syndrome; UPD, uniparental disomy; ICR, imprinting control region; ID, imprinting defects; AD, autosomal dominant; FISH, fluorescent in situ hybridization; aCGH, array comparative genomic hybridization; MS-MLPA, methylation specific-multiplex ligation-dependent probe amplification; SNP, single nucleotide polymorphism.
with head size closer to the 50th percentile, while adult height generally settles in the normal range. Additional findings may include neonatal hypoglycemia, nevus flammeus, cardiomegaly, structural cardiac defects, cardiomyopathy, advanced bone age, and a characteristic facial appearance. Most BWS patients have normal psychomotor development, but mental retardation has been noted in cases with chromosomal abnormalities and/or perinatal complications.

Pregnancies with fetuses affected with BWS may be complicated by polyhydramnios, large/dysplastic placenta, a long and thickened umbilical cord, and an increased risk for premature delivery. Children conceived by in vitro fertilization are at increased risk of developing BWS, and the overall risk of BWS with in vitro fertilization is about one in 4,000.

Although consensus diagnostic criteria for BWS have not been defined, the presence of three major features (eg, prenatal and postnatal overgrowth, macroglossia, and abdominal wall defects) or two major features and one minor feature (eg, ear anomalies, neonatal hypoglycemia, nephromegaly, and hemihyperplasia) is required for the postnatal clinical diagnosis of BWS.

Early diagnosis of BWS is crucial because of the well documented increased risk of cancer, most commonly Wilms’ tumor and hepatoblastoma, but also adrenocortical carcinoma, rhabdomyosarcoma, and neuroblastoma. Isolated hemihyperplasia (OMIM 235000) is a congenital overgrowth disorder related to BWS that displays asymmetric involvement of the body. It may manifest few of the abnormal features associated with BWS; however, patients with isolated hemihyperplasia have an increased risk for embryonal tumors, primarily Wilms’ tumor and hepatoblastoma, which are usually diagnosed before 10 years of age. Furthermore, BWS patients with hemihyperplasia have a four-fold increased tumor risk compared with BWS patients without hemihyperplasia, emphasizing the increased risk of tumors in isolated hemihyperplasia. Isolated hemihyperplasia cases should also be included in the specific follow-up created for BWS patients.

Molecular genetic testing

Molecular diagnosis is important to confirm the provisional BWS clinical diagnosis and to identify BWS patients with cancer susceptibility. An etiologic molecular heterogeneity is clearly evident in BWS, due to various genetic and/or epigenetic alterations in growth regulatory genes located on the chromosome 11p15 region.

The genetic and epigenetic abnormalities associated with BWS are listed in Table 1 and include loss of methylation on the maternal chromosome at IC2, gain of methylation on the maternal chromosome at IC1, paternal uniparental disomy for chromosome 11p15 (usually as mosaicism), mutations in the CDKN1C gene, and cytogenetically detectable or submicroscopic genomic alteration within 11p15.5. CDKN1C mutations are found in about 5%–10% of sporadic cases, whereas dominant maternal transmission of germline mutations are found in 40% of familial BWS cases.

It is largely accepted that the variability of oncologic risk in BWS mostly depends on genetic heterogeneity (Figure 2) and duration of the observation. It is now well established that patients with a molecular abnormality...
of the telomeric domain (ICR1 hypermethylation) and 11p15 paternal uniparental disomy have an increased risk of embryonal tumors (especially Wilms' tumor), whereas patients with defects of the centromeric domain only (ICR2 hypomethylation and CDKN1C mutations) are at lower risk of such tumors.  

CDKN1C mutations in BWS: genotype-phenotype correlations

Focusing on CDKN1C mutations, subfertility in males, predisposition to aneurysmal arterial dilation, renal abnormalities, hearing loss, and, perhaps, increased risk for adult-onset malignancy are known. Romanelli et al reviewed the CDKN1C-related phenotypic features in BWS patients, and suggested that these patients show a peculiar pattern of clinical malformations in comparison with those with other molecular defects, and therefore the peculiar clinical presentation of the patients can be useful to address the molecular analysis. Genotype-phenotype studies indicate that mutations in CDKN1C are more frequently associated with omphalocele, clefting, and genital anomalies and less frequently with development of tumors, with the exception of rare cases with neuroblastoma. Focusing on cleft palate, although this finding has been rarely reported in patients affected by BWS (2.5%), it has usually been associated with CDKN1C alterations; instead, cleft lip has never been documented in these patients.

In addition, CDKN1C seems to be highly expressed in the placenta, and therefore may have importance in the pathophysiology of pre-eclampsia/HELLP syndrome during pregnancy. Furthermore, a few patients with other rare malformations such as polydactyly and supernumerary nipples have been reported.

IMAGe syndrome

IMAGe syndrome is a rare developmental disorder, mainly characterized by growth restriction and described for the first time in 1999 by Vilain et al. To date, there are only 12 familial or isolated cases reported in the literature, but this condition could be underestimated as the causative gene and the mode of inheritance have only recently been identified by Arboleda et al.

Clinical features

The acronym IMAGe indicates the presence of Intrauterine growth restriction, Metaphyseal dysplasia, congenital Adrenal hypoplasia, and Genital anomalies. The first sign that becomes evident after birth is adrenal insufficiency, which can be severe and life-threatening if not recognized and treated early; it usually manifests in the first days of life with adrenal crises, but a case of late onset has been reported. Radiologic identification of metaphyseal dysplasia is often crucial for the diagnosis, but this could be very mild and identifiable only in late infancy or in childhood and then progress with age. A more precocious sign, ie, delayed endochondral ossification associated with osteopenia, hypercalcemia, and/or hypercalciuria, can be present at birth and normalizes later in infancy. Growth hormone deficiency has also been described in some patients, and it has been shown that an early substitutive therapy assessment could help to improve the linear growth.

Dysmorphic craniofacial features in IMAGe syndrome include nonspecific signs, such as prominent forehead,
low-set ears, flat nasal bridge, and short nose, while genital abnormalities seem to be confined to males and include micropenis, undescended testes, and varying severity of hypospadias.42

Molecular genetic testing
The IMAGe causative gene and inheritance model have been recently identified.545-47 To date, six missense mutations, all of them occurring in the PCNA-binding domain in the carboxy-terminal region of CDKN1C, were reported (Figure 1).5,45 Recently, Hamajima et al48 demonstrated that the IMAGe-associated mutations cause a dramatically increased instability of the CDKN1C proteins that probably results in a functional gain. This hypothesis is also supported by evidence of CDKN1C truncation mutants lacking PCNA binding (ie, F276X10; Figure 1) identified in BWS patients showing the loss-of-function phenotype of CDKN1C.49

For molecular diagnosis of IMAGe syndrome, CDKN1C sequencing is recommended. Notably, familial analysis demonstrated de novo mutations or an imprinted mode of inheritance, exclusively with maternal transmission of the mutation. This means that each child of a woman heterozygous for the CDKN1C mutation has a 50% chance of inheriting the pathogenic variant and being affected, whereas each child of a man with the mutation has a 50% chance of inheriting the CDKN1C pathogenic variant but is expected to be unaffected.50 This pattern of transmission suggests that IMAGe can be considered as an imprinting disease, albeit characterized by point gene mutations.

Same gene, mirror phenotypes
The mechanisms underlying genomic imprinting are very complex, in particular regarding the 11p15.5 imprinted region. Indeed, different mutations (or epimutations) in the same gene (CDKN1C) or imprinted region can result in opposite phenotypes. CDKN1C can be associated with BWS, SRS, and IMAGe syndrome, and IC1 opposite imprinting defects with BWS (IC1 hypermethylation) and SRS (IC1 hypomethylation). Among these syndromes, BWS is mainly characterized by overgrowth, and IMAGe and SRS by growth restriction.46 A detailed description of the mirror phenotype in BWS and IMAGe is reported in Table 2.

BWS-associated CDKN1C mutations are most often truncating mutations distributed throughout the gene, or missense mutations in the aminoterminal CDK inhibitor domain, and those associated with IMAGe are missense variants in the PCNA-binding domain (Figure 1), suggesting that the mutations causing these two imprinting disorders have different biologic consequences.

CDKN1C mutations in BWS act as a “double-negative”, promoting proliferation due to loss of cell cycle inhibition, and then predisposition to cancer. In contrast, IMAGe syndrome is caused by mutations leading to a gain-of-function of CDKN1C protein, most probably related to increased stability of the protein.5 IMAGe-associated mutations inhibit binding to PCNA and ubiquitination of CDKN1C. Evidence for a gain-of-function effect was detected in Drosophila melanogaster, in which expression of the mutant but not the wild-type human protein reduced eye and wing size.5 It is conceivable that CDKN1C mutations in IMAGe patients can trigger an excess of inhibition of growth and differentiation.

A good example of the effects of these mutations on the single organ has been provided by Arboleda et al,5 who showed how CDKN1C was strongly expressed during the embryonic development of the adrenal glands in mice. In BWS, the decreased inhibition of G1 CDKs due to loss-of-function mutations causes adrenal cytomegaly (one of the

<table>
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<th>Table 2 Mirror phenotypes in BWS and IMAGe syndrome</th>
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<tr>
<td><strong>BWS</strong></td>
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<td>Macrosmia/hemihyperplasia</td>
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<td>Abdominal wall defects</td>
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<td>Visceromegaly, macroglossia, increased risk of cancer</td>
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<td>Adrenal hyperplasia</td>
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<td>Anterior ear creases, posterior helical pits, cleft palate, nevus flammeus</td>
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<tr>
<td>Kidney abnormalities (cytomegaly</td>
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<td>of the adrenal fetal cortex, medullary dysplasia, delayed development of medullary sponge kidney), increased risk of cancer</td>
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<td>Neonatal hypoglycemia</td>
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<td>Structural cardiac defects</td>
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<td>Advanced bone age</td>
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<tr>
<td>Fetal macrosomia (LGA), polyhydramnios, large/dysplastic placenta, long and thickened umbilical cord, increased risk for premature delivery</td>
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<tr>
<td>Metaphyseal dysplasia</td>
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<td>Micro penis, undescended testes, and varying severity of hypospadias</td>
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<td><strong>IMAGe syndrome</strong></td>
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**Abbreviations:** BWS, Beckwith–Wiedemann syndrome; IMAGe, intrauterine growth restriction, metaphyseal dysplasia, congenital adrenal hypoplasia, and genital anomalies; LGA, large for gestational age; SGA, small for gestational age.
most common signs in BWS fetuses) and adrenocortical tumors (also sporadic adrenocortical tumors can display CDKN1C inactivation), while in IMAGe syndrome the increased inhibition of the cell cycle due to augmented stability of CDKN1C results in congenital adrenal hypoplasia (Table 2).

Finally, in the fetal brain, as mentioned before, CDKN1C is biparentally expressed. This observation could explain why the occipitofrontal circumference in IMAGe syndrome is relatively well preserved in comparison with length/height and weight that are severely compromised.\(^{41}\)

The link between the nature of CDKN1C mutation and the mirror phenotype of BWS and IMAGe can be explained by the importance of p57Kip2 in embryogenesis, which is also highlighted by the finding that knockout mice exhibit gastrointestinal tract defects, omphalocele, abnormal endochondral ossification, cleft palate, adrenal cortex enlargement, renal medullary dysplasia, increased body weight, and placental abnormalities. p57Kip2 excess in mice increases embryonic lethality and decreases body size, suggesting that embryonic growth requires accurate control of p57Kip2 dosage.\(^{41}\)

In conclusion, after the discovery that SRS is characterized by the opposite growth defect compared with BWS, IMAGe syndrome represents an additional imprinting disorder with a mirror phenotype of BWS, strengthening the primary role of the 11p15.5 imprinted region in prenatal and postnatal growth regulation.

**Disclosure**

The authors report no conflicts of interest in this work.

**References**