S1pping fire: Sphingosine-1-phosphate signaling as an emerging target in inflammatory bowel disease and colitis-associated cancer

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Abstract: Inflammatory bowel disease (IBD) is a complex disease that involves unpredictable and destructive inflammation in the gastrointestinal tract resulting in gastrointestinal symptoms, infection, and tissue destruction, and which can be associated with an increased risk of colon cancer. The underlying cause of IBD involves disruption of the innate and adaptive immune mechanisms that maintain homeostasis between the gut mucosa and its environment. Elucidating how the homeostatic mechanisms controlling gut mucosal immunity and inflammation are disrupted in IBD represents the first steps to identifying novel therapeutic targets. Sphingosine-1-phosphate (S1P) is a bioactive sphingolipid that is enriched in the blood and lymph, and functions in innate and adaptive immunity. S1P signaling regulates inflammation via its impact on the trafficking, differentiation, and effector functions of bone marrow-derived immune cells. S1P also activates nuclear factor kappa B and signal transducer and activator of transcription 3 inflammatory pathways. S1P is generated by the ubiquitously expressed lipid kinase, sphingosine kinase (SphK)1 and its tissue-restricted homolog, SphK2. S1P is irreversibly degraded by S1P lyase, which is highly expressed in enterocytes. Recent studies targeting S1P metabolism and signaling have shown promise in preclinical models of IBD and have shed light on the mechanisms by which S1P signaling impacts IBD. The evidence suggests that targeting S1P signaling and metabolism may represent a novel strategy in treating IBD and it may reduce colon cancer risk by interrupting the progression from inflammation to carcinogenesis.

Keywords: sphingolipids, sphingadienes, sphingosine phosphate lyase, sphingosine kinase, STAT3, S1PR

Introduction

Inflammatory bowel disease (IBD) is a complex disease comprised of Crohn’s Disease or Crohn Disease (CD) and ulcerative colitis (UC), two conditions that involve unpredictable and destructive inflammatory processes resulting in weight loss, diarrhea, abdominal pain, and fever. UC is restricted to the colon, whereas CD is characterized by skip lesions and can involve any site in the gastrointestinal tract. Symptoms include abdominal pain, ulcers, bloody or nonbloody diarrhea, cramping, weight loss, anorexia, and fatigue. Affected children can also exhibit stunted growth and delayed puberty. Pathological features include ulceration, abscesses, inflammatory infiltrates, edema, mucin depletion, and loss of the normal crypt architecture. IBD leads to fistulas, perforations, infection, abscesses, and increases the risk of intestinal dysplasia and colon cancer in proportion to its duration.

IBD is caused by dysregulation of the immune mechanisms that maintain homeostasis between the intestinal epithelium and commensal and pathogenic gut flora.
Genetic analyses have implicated the human genes ATG16L, NOD2/CARD15, IBD5, CTLA4, TNFSF15, JAK2, STAT3, IL23R, and ORMDL3 as risk factors in IBD. These clues, combined with basic research, have revealed that antimicrobial peptides, autophagy, endoplasmic reticulum stress, innate and adaptive immune cell function, T-helper (Th)17 cells, regulatory T-cells, and cytokines (tumor necrosis factor (TNF)-α, interleukin (IL)-17, IL-23/IL-12, IL-22, and IL-6) are contributing factors in IBD. These mediators initiate signaling pathways that activate key inflammatory transcription factors including nuclear factor kappa B (NFκB) and signal transducer and activator of transcription (STAT)3, which integrate and amplify signals from a wide range of intrinsic and environmental stimuli. Many cell compartments of the gut including enterocytes, Paneth cells, T-cells, mature and immature myeloid cells, and vascular cells contribute to the regulation of NFκB, STAT3, and the inflammatory milieu. Elucidating the complex interactions between intestinal cells, secreted proteins, and transcription factors, their modulation by factors in the gut mucosa and its environment, and how these interactions are disrupted in IBD are the necessary first steps to identifying new targets and curing IBD. Targeted therapy and dietary chemoprevention strategies hold the promise of reducing the toxicities and risks associated with global immunosuppressive regimens that are currently being employed to treat IBD.

Sphingosine-1-phosphate (SIP) is a signaling lipid found in the circulation and in most tissues. SIP is derived from the recycling of endogenous human sphingolipids and the metabolism of sphingolipids found in dietary animal products that, like human tissues, contain sphingolipids, which are built upon a sphingosine structural backbone. SIP has many functions in angiogenesis, development, innate and adaptive immunity, and is a regulator of lymphocyte trafficking. A majority of SIP's biological functions have been linked to its ability to activate a family of five G protein-coupled receptors, SIP receptors 1–5 (SIPR1–5). However, SIP exerts some actions that have not yet been definitively or completely attributed to SIPRs. For example, SIP serves as a major activator of the IL-6/STAT3 pathway implicated in the pathophysiology and genetic basis of IBD, as well as the pathogenesis of colon cancer. In fact, SIP production appears to be oncogenic in colon cancer. SIP is also the cofactor for the TNF receptor associated factor 2 E3 ubiquitin ligase required for activation of NFκB downstream of TNF-α and nucleotide-binding oligomerization domain-containing protein 2. Nuclear actions of SIP have also been linked to SIPR functions.

SIP is generated from sphingosine through the actions of sphingosine kinase (SphK) enzymes, as shown in Figure 1. There are two isoforms of SK: the ubiquitously expressed major SK, SphK1; and the more tissue-restricted isoform, SphK2. S1P can be dephosphorylated by specific and nonspecific lipid phosphatases or irreversibly cleaved by S1P lyase into phosphoethanolamine and trans-2-hexadecenal.

Abbreviation: SIP, sphingosine-1-phosphate.
SIP signaling as a target in autoimmune disease

In 2004 and 2005, several publications from one laboratory demonstrated that SIP signaling through S1PR1 and metabolism by SPL control lymphocyte egress from the thymus into the circulatory system. These studies showed that the pharmacological inhibition of SPL (which disrupts the SIP gradient between the blood and tissues) or treatment with an S1PR1 modulator (which induces downregulation and attenuates signaling through this receptor) induced lymphopenia in mice. The role of SIP signaling and metabolism in the regulation of lymphocyte trafficking has been further confirmed through genetic approaches. Most exciting was the fact that targeting the SIP signaling pathway blocked lymphocyte trafficking without necessarily reducing global immune functions as other immunosuppressive agents do. The strategy of targeting S1PR1 to induce lymphopenia was later shown to be therapeutically effective in the treatment of the human autoimmune disorder, multiple sclerosis.

Other lines of evidence demonstrate that SIP signaling is linked to inflammation, autoimmune disease, and cancer. In 2010, the research group of Lee et al demonstrated that the STAT3 signaling pathway and the SIP/S1PR1 pathway coactivate one another in an inflammatory loop that promotes carcinogenesis. In an atherosclerosis mouse model, S1PR2 activation increased NFκB activity in macrophages, one of the cell types implicated in the development of atherosclerosis. In another study, S1PR1 was found to directly activate the STAT3 pathway via IL-6 in a mouse model of autoimmune encephalomyelitis. These studies implicate the SIP axis in the pathophysiology of autoimmune disease and highlight the need to explore SIP targeting approaches in other autoimmune and inflammatory conditions.

SIP signaling as a target in IBD

The results from preclinical studies employing various mouse models of colitis suggest that targeting SIP signaling and other sphingolipids may also be useful in the treatment of IBD. FTY720, KRP-203, and W-061 are S1PR agonist/functional antagonists that have a high affinity for S1PR1, causing its internalization and downmodulation, as shown in Figure 2A. These agents have been shown to reduce murine models of colitis produced by IL-10 deficiency, dextran sodium sulfate (DSS) treatment, and adoptive T-cell transfer. In contrast to CD, which is characterized by a Th1 cytokine profile, UC is defined by a Th2 cytokine profile. Th2 cytokines are necessary for the differentiation of cluster of differentiation (CD)4+ T-cells into natural killer T (NKT) cells. NKT cells produce IL-13 following activation by glycolipid antigens that are found on the bacteria of the gut or in epithelial cells infected by bacteria. The lamina propria of UC patients is characterized by high expression of IL-13 and IL-13-producing CD4+ T-cells. NKT cells have been shown in vitro to cause cytotoxicity via IL-13 and to negatively affect the epithelial barrier function of a monolayer of HT29. In the oxazolone mouse model of UC, an ex vivo culture of CD4+ T-cells isolated from FTY720-treated mice showed reduced expression of proinflammatory cytokines IL-13, IL-5, and IL-4, produced by Th2 cells. This suggests that FTY720 may have the potential to correct fundamental immune imbalances in the UC microenvironment. In a Th1-driven 2,4,6-trinitrobenzenesulfonic acid (TNBS) mouse model of colitis, FTY720 also showed efficacy in the reduction of colitis. Treg cells are thought to play a key role in IBD by preventing the proliferation and activation of T lymphocytes. In this study, FTY720 was found to directly affect the functional activity of CD4+CD25+ regulatory T-cells. Administration of FTY720 to TNBS-treated mice resulted in the decreased expression of Th1 cytokines, namely IL-12p70 and TNF-α, and an upregulation of the immunosuppressive Treg cytokines IL-10 and transforming growth factor-β. This was accompanied by an increase in the expression of cytotoxic T-lymphocyte antigen 4 and FoxP3, both of which are markers of Treg cells. The ability of FTY720 to attenuate colitis in both Th1- and Th2-driven models of colitis makes it an attractive candidate for the treatment of IBD patients.

In addition to the effects on colitis afforded by agents targeting S1PRs, it was observed that mice lacking SphK1 were less susceptible than littermate controls to colitis induced by DSS. This suggests that SphK1 could be targeted to reduce SIP signaling (Figure 2B). Compared to control mice, SphK1 null mice showed a similar expression of TNF-α, but decreased expression of cyclooxygenase (COX)-2 and no systemic inflammatory response. These findings pinpoint SphK1’s action downstream of TNF-α, but upstream of COX-2. The evidence suggests that targeting SphK1 with a specific inhibitor could attenuate inflammatory responses in IBD patients. In support of this notion, two inhibitors of SK,
ABC747080 and ABC294640, prevented TNF-α activation of NFκB, prostaglandin E₂ production, vascular cell adhesion molecule and intercellular adhesion molecule expression, and reduced clinical and histological signs of DSS-induced colitis in C57Bl/6 mice. The effects of the S1P axis on adaptive immunity are shown in Figure 3.

SPL is highly expressed throughout the intestine, especially in the differentiated epithelial cells of the duodenum and jejunum and, to a lesser degree, in the ileum and colon. In a recent study, an inverse correlation between the colonic concentrations of S1P and SPL’s cofactor, pyridoxal-5-phosphate was found. Of note, plasma pyridoxal-5-phosphate concentration was inversely linked with the concentration of inflammatory markers. These results suggest that SPL expression, as well as its cofactor availability, may be important in preventing or reducing inflammation. Patients suffering from IBD are at a higher risk of developing colon cancer the longer they suffer from unresolved inflammation. Interestingly, we found that SPL expression and activity were downregulated in adenomatous lesions of ApcMin/+ mice. Moreover, human colon cancer tissues expressed less SPL than the normal adjacent tissue. It was demonstrated that SPL expression is required for p38- and p53-induced cell death via caspase 2 activation. The results of this study raise the possibility that changes in SPL expression or enzyme activity could contribute to the initiation of intestinal tumorigenesis.

**SIP in colitis-associated cancer**

Liang et al have provided new evidence supporting a role for SphK2 in the pathogenesis of colitis and colitis-associated cancer (CAC). In their study, the azoxymethane (AOM)/DSS mouse model of CAC was implemented in both wild type and SphK2 knockout (KO) mice. They found that the tumor number, size, and load were higher in SphK2 KO mice compared to wild type mice. The SphK2 KO mice also exhibited elevated disease activity, suffering from severe diarrhea, as well as blood and weight loss. In addition, the KO mice exhibited shorter colons (indicating inflammation-induced
S1P in IBD and colon cancer

**Figure 3** The role of the S1P axis in adaptive immunity.

**Notes:** Dietary or bacterial S1P from the intestinal lumen bind to S1PRs and activate the STAT3 pathway in intestinal epithelial cells. Mucosal NFκB is activated by intracellular S1P produced by the phosphorylation of sphingosine by SphK. These two transcription factors transactivate numerous proinflammatory cytokine genes, thereby contributing to the recruitment of immune cells to the inflamed site. Moreover, STAT3 can amplify S1P/S1PR signaling by increasing the expression of S1PR1. Adaptive immune cells, namely Treg, Th1, and Th2 cells, produce a variety of cytokines in response to intestinal inflammation. Most notably, in response to NFκB and STAT3 pathway activation, Th1 and Th2 cells can produce an array of proinflammatory cytokines, including IL-1β, TNF-α, IL-6, and so on. The S1PR1 superagonist FTY720 increases Treg production of IL-10 and TGF-β, but inhibits proinflammatory cytokine production by Th1 and Th2 cells, possibly by acting through S1PRs.

**Abbreviations:** S1P, sphingosine-1-phosphate; S1PR, sphingosine-1-phosphate receptor; SphK, sphingosine kinase; NFκB, nuclear factor kappa-light-chain-enhancer of activated B cells; STAT3, signal transducer and activator of transcription 3; Treg, regulatory T-cells; IL, interleukin; Th, T-helper; iNOS, inducible nitric oxide synthase; TNF, tumor necrosis factor; TGF, transforming growth factor.

stricture and fibrosis), extensive colon architectural damage, and immune cell infiltration.

In performing biochemical analyses of the two groups, the authors made a surprising discovery. They found increased circulating and colonic S1P levels in SphK2 KO mice compared to controls. Moreover, SphK1 protein and messenger ribonucleic acid levels were higher in the colons of SphK2 KO mice. The authors suggested that the upregulation of SphK1 could be the result of reduced nuclear SphK2. SphK2 in the nucleus produces a source of S1P that inhibits histone deacetylase 1/2. Importantly, one of the transcriptional regulators of SphK1 expression, c-Jun, is regulated by histone acetylation.

Therefore, in the colons of SphK2 KO mice, the authors suggest that increased histone deacetylase activity results in increased activation of c-Jun, thereby driving increased SphK1 expression.

S1P activates NFκB, a transcription factor involved in the expression of many proinflammatory cytokines that contribute to the pathogenesis of IBD and CAC. IL-6, which is a NFκB target gene, can activate STAT3, a regulator of S1PR1 expression. Accordingly, NFκB activation, IL-6 expression and S1PR1 expression were all increased in the colons of SphK2 KO mice following DSS treatment to a greater extent than in controls. To determine the cell type responsible for IL-6 secretion, the authors generated bone marrow chimeras by transferring bone marrow cells of either wild type or SphK2 KO mice to an irradiated recipient. SphK2 disruption in bone marrow cells alone was sufficient to recapitulate the SphK2 global KO mouse phenotype following DSS treatment. Flow cytometry analysis identified macrophages and dendritic cells as the major producers of IL-6. However, intestinal
epithelial cells were at least in part responsible for the increased colonic expression of IL-6 in global SphK2 KO mice, as shown by immunohistochemistry.48 The authors proposed a model involving "a feed-forward amplification loop" in which the SK/S1P/S1PR1 axis leads to the activation of NFκB and persistent STAT3 activation in CAC, as shown in Figure 4.48 To disrupt the feed-forward cycle, the authors administered FTY720, an S1PR1 functional antagonist that can also inhibit SphK1 by causing its proteasomal degradation (Figure 2B).45–47 Administration of FTY720 in SphK2 global KO mice treated with DSS reduced disease activity, SphK1, IL-6, and S1PR1 expression, and NFκB and STAT3 activity.48 FTY720 could prevent CAC in SphK2 KO mice if administered at the beginning of the AOM/DSS regimen, but not when it was administered at a later time point. This was true even though FTY720 administration could still reduce STAT3 activation and S1PR1 expression.48 The authors concluded that FTY720 should be explored as a drug to treat CAC in IBD patients, since it reduces both the expression of S1PR1 and SphK1, thereby disrupting the S1P/SphK1/S1PR1 signaling loop.

**Dietary sphingolipids**

Each day, we consume between 0.3–0.4 g of sphingolipids though our diet.49 In the intestinal tract, these sphingolipids are digested to form sphingoid bases and ceramides.49 In the past three decades, the potential of sphingolipids as dietary bioactive molecules has been studied intensively in different mouse models of IBD and colon cancer. 1,2 dimethylhydrazine (DMH) is a carcinogenic compound used in mouse and rat models of CAC.49,50 DMH promotes colon tumorigenesis by causing oxidative stress and inflammation.51 One report of a study employing the DMH mouse model found that dietary sphingomyelin can reduce the number of premalignant lesions and tumors by 20%.52 In another study, the plant sphingolipid soy glucosylceramide added to the diet of ApcMin/+ or DMH-treated mice decreased the number of adenomas and aberrant crypt foci arising in these two respective models of colon cancer.53 The authors confirmed that the expression levels of two transcription factors involved in tumorigenesis, hypoxia-inducible factor 1α and transcription factor 4, were decreased in response to sphingolipid treatment.53 Mazzei et al54 explored the effects of dietary sphingomyelin in the AOM/DSS mouse model of CAC. They found that adding 1 g/kg of sphingomyelin in the mouse diet decreased both the disease activity index and the tumor number.54 They found that sphingomyelin attenuated the expression of proinflammatory cytokines (interferon-γ, IL-17, and IL-23), but induced antiinflammatory cytokines and receptors implicated in Th2 and Treg cell differentiation (IL-4, IL-3, IL-13ra2, and IL-10rb).54 In contrast, in an acute DSS colitis model, gavage of 4 mg or 8 mg of sphingomyelin/day exacerbated the disease activity by causing increased apoptosis of the intestinal epithelial cells.55 This finding was associated with an increased activity of cathepsin D that mediated caspase-3- and caspase-9-induced apoptosis.56 Using the IL-10 KO mouse model of spontaneous colitis, the authors showed that sphingomyelin aggravated mucosal inflammation.57 These findings – using a variety of mouse models, different sphingomyelin doses, and measuring different disease-related endpoints – raise the possibility that dietary sphingolipids may have untoward effects depending

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**Figure 4** SphK1/S1P/S1PR1 feed-forward loop sustain signals promoting colon tumorigenesis. **Notes:** SphK1 produces S1P, which is kept in the cell and/or is exported in the extracellular milieu. Extracellular S1P activates S1PR1 which, in turn, activates the JAK2/STAT3 pathway. STAT3 is one of the transcription factors involved in the regulation of IL-6 and S1PR1 expression. Intracellular S1P can activate NFκB, a transcriptional regulator of SphK1. All these pathways are interrelated in an amplification loop, allowing for the sustained activation of STAT3, which is involved in cell transformation and tumor progression. Bent arrows indicate transcriptional regulation. **Abbreviations:** SphK, sphingosine kinase; S1P, sphingosine-1-phosphate; S1PR, sphingosine-1-phosphate receptor; STAT3, signal transducer and activator of transcription 3; IL, interleukin; NFκB, nuclear factor kappa-light-chain-enhancer of activated B cells; JAK2, Janus kinase 2; SPNS2, spinster homolog 2; ABC, ATP Binding Cassette; ATP, adenosine triphosphate.
on the state of the target tissue, and possibly the source and structure of the sphingolipid. They also suggest that there may be a tight range of sphingomyelin concentrations that will afford therapeutic benefit in CAC.

Sphingadienes are orally available sphingolipids containing a sphingolipid backbone harboring conjugated trans double bonds. They are found in soy products and other natural sources. Sphingadienes containing double bonds at C4,5 and C8,9 were shown to induce apoptosis in HCT116, DLD1, and SW480 colon cancer cell lines and to decrease tumor development in an ApcMin/+ mouse model of colon cancer. Sphingadienes act, at least in part, by blocking the translocation of Akt from the cytoplasm to the membrane, thereby inactivating Akt. This effect of sphingadienes leads to the induction of apoptosis and autophagy, processes that are repressed by Akt. Sphingadiene treatment also inactivated GSK-3β and, as a consequence, downregulated growth-activating Wnt signaling. Sphingadienes are being explored for their potential efficacy as anti-inflammatory agents in IBD and in their ability to prevent the development of CAC (Degagne, unpublished data, 2014). Importantly, unlike mammalian sphingolipids, sphingadienes cannot be metabolized to S1P. Further, since sphingadienes are not produced in appreciable amounts in humans, their metabolism by the enzymes present in human colonic epithelial cells is limited, their uptake into the bloodstream is minimal, and their action is local, making them appealing candidates for the long-term chemoprevention of CAC.

**Risks, controversies, and remaining questions**

The studies described above highlight the possibility that strategies involving S1PR1 downregulation or inhibition that have proven useful in other autoimmune diseases may afford promise in the context of IBD. However, chemically targeting S1PRs may be complicated by their diverse functions. For example, S1PR1 activity could prove critical in maintaining vascular integrity in the colonic mucosa. This concern is exemplified by results from preclinical studies testing S1PR1 disruption in mice suffering from experimental colitis. However, treatment of C57BL/6J mice with FTY720 did not cause more bleeding than the control group in an experimental colitis model. Of additional concern, administration of FTY720 to mice in an experimental colitis model impaired mucosal immunity such that the mice were then unable to eliminate an enteric bacterial infection. FTY720 treatment was associated with a shift in the immune cell populations present in the colon during the infection, decreasing the number of T and B lymphocytes and macrophages, and resulting in the decreased expression of Th1 cytokines (IL-1β, inducible nitric oxide synthase [iNOS], IL-17a, IL-6, and TNF-α). Even when the treatment was discontinued prior to infection, the mice were still unable to clear the bacterial infection. These findings suggest that the safety of strategies that modulate S1PR signaling in IBD patients must be investigated further. Specifically, it will be critical to understand how S1PR modulation affects the innate immune response to a wide range of bacteria and viruses to which IBD patients are particularly susceptible.

The recent article by Liang et al has established the importance of SphK2 in inflammatory signaling through STAT3. Interestingly, others have shown that CD4+ T-cell SphK2 is required for suppressing IL-2 responses that aggravate IBD via a STAT5-dependent mechanism. Sphk2 null mouse CD4+ T-cells have a hyperactive phenotype, increased proliferation, and cytokine secretions in response to IL-2 signaling. Moreover, these Sphk2 null mouse CD4+ T-cells were not responsive to Treg cell-mediated suppression. This study raises concerns regarding the use of SK inhibitors in treating IBD patients, as they could have adverse effects on immune cells by directly increasing cytokine secretions through their effects on CD4+ T-cells and also by counteracting the immunomodulatory role of Treg cells. Thus, the S1P signaling pathway may be affecting multiple aspects of the innate and adaptive immune response relevant to IBD.

Additional questions remained unanswered. Is the decreased expression of SphK2 a characteristic of or a precursor state to the development of IBD or colon cancer? What are the key mechanisms by which S1P signaling (and downstream NFκB and STAT3 inflammatory nodes) promotes the switch from a state of inflammation to a preneoplastic state and, ultimately, to the development of CAC? At this time, the exact role of S1P in IBD remains incompletely understood. Further, the potential for S1P to be used as a biomarker of disease activity has not been explored. In that regard, there is sparse information regarding the lipid metabolic profiles of IBD colons compared to controls. How lipid metabolomics, S1P degradation, and the enzymes of sphingolipid metabolism that are enriched in the gut epithelium may influence gut immunity and IBD still remain to be determined.

**Conclusion**

The mechanisms that underlie the pathophysiology of IBD are complex and involve numerous signal transduction pathways that regulate innate and adaptive immunity in the
As sphingolipids are involved in all pathways centered on the sphingolipid metabolite S1P, as well as on the NFκB and STAT3 inflammatory signaling pathways with which it interacts, S1P functions as a critical regulator of lymphocyte trafficking and a cofactor in inflammatory signaling. Further, many S1P-related targets, including S1PRs and the enzymes involved in S1P biosynthesis or degradation, are located in the epithelium of the small and large intestine wherein IBD arises. As such, these targets are of interest in the treatment of IBD and in the prevention of its progression to CAC. In addition, dietary sphingolipids may represent another strategy for reducing colitis and preventing colon carcinogenesis. While some safety concerns remain, the preclinical evidence supports further investigation of these strategies in the context of IBD.

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Disclosure

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