Cytotoxic effect of magnetic iron oxide nanoparticles synthesized via seaweed aqueous extract

Farideh Namvar¹,², Heshu Sulaiman Rahman³,4, Rosfarizan Mohamad¹,5, Javad Baharara², Mahnaz Mahdavi⁶, Elaheh Amini⁷, Max Stanley Chartrand⁸, Swee Keong Yeap³

¹Institute of Tropical Forestry and Forest Products, Universiti Putra Malaysia, Selangor, Malaysia; ²Research Center for Animal Development Applied Biology, Mashhad Branch, Islamic Azad University, Mashhad, Iran; ³Institute of Bioscience, Department of Microbiology and Pathology, Faculty of Veterinary Medicine, ⁴Department of Bioprocess Technology, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, Selangor, Malaysia; ⁵Department of Chemistry, Faculty of Science, Islamic Azad University, Shiraz Branch, Shiraz, ⁶Kharazmi University, Tehran, Iran; ⁷DigiCare Behavioral Research, Casa Grande, AZ, USA

Abstract: Magnetic iron oxide nanoparticles (Fe₃O₄ MNPs) are among the most useful metal nanoparticles for multiple applications across a broad spectrum in the biomedical field, including the diagnosis and treatment of cancer. In previous work, we synthesized and characterized Fe₃O₄ MNPs using a simple, rapid, safe, efficient, one-step green method involving reduction of ferric chloride solution using brown seaweed (Sargassum muticum) aqueous extract containing hydroxyl, carboxyl, and amino functional groups mainly relevant to polysaccharides, which acts as a potential stabilizer and metal reductant agent. The aim of this study was to evaluate the in vitro cytotoxic activity and cellular effects of these Fe₃O₄ MNPs. Their in vitro anticancer activity was demonstrated in human cell lines for leukemia (Jurkat cells), breast cancer (MCF-7 cells), cervical cancer (HeLa cells), and liver cancer (HepG2 cells). The cancer cells were treated with different concentrations of Fe₃O₄ MNPs, and an MTT (3-[4,5-dimethylthiazol-2-yl]-2,5 diphenyl tetrazolium bromide) assay was used to test for cytotoxicity, resulting in an inhibitory concentration 50 (IC₅₀) value of 23.83±1.1 µg/mL (HeLa), 18.75±2.1 µg/mL (MCF-7), 12.5±1.7 µg/mL (HeLa), and 6.4±2.3 µg/mL (Jurkat) 72 hours after treatment. Therefore, Jurkat cells were selected for further investigation. The representative dot plots from flow cytometric analysis of apoptosis showed that the percentages of cells in early apoptosis and late apoptosis were increased. Cell cycle analysis showed a significant increase in accumulation of Fe₃O₄ MNP-treated cells at sub-G1 phase, confirming induction of apoptosis by Fe₃O₄ MNPs. The Fe₃O₄ MNPs also activated caspase-3 and caspase-9 in a time-response fashion. The nature of the biosynthesis and therapeutic potential of Fe₃O₄ MNPs could pave the way for further research on the green synthesis of therapeutic agents, particularly in nanomedicine, to assist in the treatment of cancer.

Keywords: green synthesis, seaweed water extract, anticancer effect, apoptosis

Introduction

Nanoscience and nanotechnology have elegant potential across a broad spectrum of cancer research, including diagnostic, monitoring, and therapeutic strategies, and provide novel approaches in these areas. Some nanocarriers, like liposomes, dendrimers, micelles, carbon nanotubes, and nanoparticles, have been used to help in the diagnosis and theranostics of certain types of cancers. The green approach to synthesis of nanoparticles using plant materials, such as reducing and capping agents, could be considered attractive in nanobiotechnology. When compared with mechanical strategies, this technology is safe, simple, nontoxic, efficient, and environmentally friendly, and also provides efficacious single-pot reactions without the need for additional surfactants or capping agents.

Due to their superparamagnetic behavior and surface modification properties, iron oxides are considered to be capable candidates in cancer therapy. A novel
approach to biosynthesis of iron oxide magnetic nanoparticles (Fe₃O₄ MNPs) is to make nanoparticles using natural products, such as plant extracts, to reduce metal ions, which are readily scalable and nontoxic compared with physical and chemical methods.⁶

Synthesis of iron oxide in the presence of an oxidant creates several forms of iron oxides including magnetite (Fe₃O₄) and hematite (α-Fe₂O₃). Application of magnetite type iron oxide nanoparticles have been studied extensively in recent years for their potential beneficial effects.⁷

The unique magnetic properties of iron oxide nanoparticles mean decreased drug expenditure and drug administration, and improved efficacy of diagnosis, targeting, and treatment of tumors.⁸

The biocompatibility of Fe₃O₄ MNPs makes them suitable for biomedical application, such as in cellular therapy, tissue repair, drug delivery, and magnetofection.⁹ Recent investigations have focused on developing methods that synthesize magnetic nanoparticles with vast potential including size, shape, stability, and morphology.¹⁰ In general, due to aggregation behavior, colloidal stability, and cytotoxicity, surface coatings of the nanoparticles are considered critical in nano research.¹¹ The biodistribution of these nanoparticles in vivo is greatly affected by their surface coating properties and the influence of an external magnetic field. In addition, a number of biocompatibility assays have been performed for polysaccharide-coated magnetic nanoparticles, due to concerns regarding the risks and benefits of magnetic nanoparticles and related technologies to human health and the environment.

Using Fe₃O₄ MNPs in the treatment of cancer, Ling et al reported on superparamagnetic iron oxide nanocrystals concurrently loaded with docetaxel, a chemotherapeutic anticancer drug, and assessed their efficacy as an anticancer agent. The targeted nanoparticles were seen to have an antiproliferative effect in PC3 prostate cancer cells on cytotoxicity assay. The IC₅₀ value for iron nanoparticles loaded with docetaxel was 1.46-fold and 1.57-fold lower than that of docetaxel after 48 hours and 72 hours of treatment. Furthermore, the nanoparticles alone did not have any significant cytotoxic effect on PC3 cells, suggesting that a drug encapsulated in nanoparticles develops increased cytotoxicity in a time-dependent and dose-dependent manner.¹²

Khan et al evaluated the cytotoxic effects of Fe₃O₄ MNPs in A549 human lung epithelial cancer cells and normal IMR-90 lung fibroblasts. In their study, cancerous and normal cells were exposed to various concentrations of Fe₃O₄ MNPs (1–100 µg/mL) for 24 hours and 48 hours. Their findings showed that Fe₃O₄ MNPs had significant cytotoxic effects on A549 human lung cancer cells, but not on normal IMR-90 human lung fibroblasts.¹³ Recent studies have also confirmed the fact that bioactive compounds obtained from macroalgae can increases the chances of discovery of novel and versatile pharmaceutical agents with promise in cancer research, diagnostics, and treatment.¹⁴

As mentioned in previous reports,¹⁵ seaweed is a subgroup of macroalgae and an available food source in many countries, traditionally those in south-east Asia.¹⁶ Seaweed contains a number of potentially biologically active ingredients, including polysaccharides, proteins, lipids, vitamins, soluble fiber, and minerals, with multiple medical applications in cancer,¹⁷ inflammation,¹⁸ allergy,¹⁹ diabetes,²⁰ thrombosis,²¹ and obesity (by bringing down the caloric value of the diet),²² and may be useful in the reduction of lipid absorption and risk of cardiovascular disease,²³ hypertension,²⁴ and other degenerative diseases.²⁵ These biomedical applications of seaweed are mainly due to its functional groups, which act as capping agents in a green single-step process. Polysaccharides are the main constituent of biopolymers in seaweed water extract, and have been found to be strong stabilizers enabling increased biocompatibility. Further, they confer chemical functionality to nanostructures such as Fe₃O₄ MNPs.²⁶

In a previous study,²⁷ we synthesized and characterized Fe₃O₄ MNPs using a brown seaweed (Sargassum muticum) extract via the green method. The aim of this study was to investigate the cytotoxic effects of Fe₃O₄ MNPs prepared by green biosynthesis on various human cancer cell lines using a number of experimental methods.

### Materials and methods

#### Materials

Specimens of the brown seaweed, Sargassum muticum, were obtained from coastal waters in the Persian Gulf. All aqueous solutions were made using distilled deionized water.

#### Cell cultures

All cancer cell lines were purchased from the American Type Culture Collection (ATCC, Rockville, MD, USA), maintained in Roswell Park Memorial Institute 1,640 medium (ATCC) supplemented with L-glutamine 2 mM and 10% fetal bovine serum (ATCC), with 100 U/mL penicillin and 100 µg/mL streptomycin (Sigma-Aldrich, St Louis, MO, USA) according to the ATCC protocol, and cultured and grown in 75 cm² culture flasks (TPP Techno Plastic Products, Trasadingen, Switzerland) at 37°C in an incubator with a humidified atmosphere of 95% air and 5% CO₂ (Binder
GmbH, Tuttlingen, Germany). The cultures were frequently examined under an inverted microscope (MICROS, Sankt Veit an der Glan, Austria) for confluency and viability.

Methods
Preparation of *S. muticum* extract
As mentioned in our previous study,27 the seaweed specimens were washed and stored at −20°C. To make the extract, seaweed samples (about 1 g) were ground, freeze-dried, and boiled with distilled deionized water (100 mL) in an Erlenmeyer flask, with constant stirring for 15 minutes. The extract was then cooled to room temperature, filtered, and stored at −20°C before use.

Preparation and characterization of Fe$_3$O$_4$ MNPs
Fe$_3$O$_4$ MNPs (Figure 1) have been prepared and well characterized previously by our group.27

Cytotoxicity assay
The antiproliferative effect of Fe$_3$O$_4$ MNPs on various cancer cell lines and a normal Chang liver cell line was quantified using a 3-(4,5-dimethylthiazol-2-yl)-2,5 diphenyl tetrazolium bromide (MTT) kit (Sigma-Aldrich) according to the standard method.28 Briefly, cells were allowed to grow in a 75 cm$^2$ culture flask (TPP Techno Plastic Products, Trasadingen, Switzerland) until 95% confluent. The cells were then seeded into each well of a 96-well microculture plate (TPP Techno Plastic Products) at a concentration of 1×10$^5$ cells/mL and treated with Fe$_3$O$_4$ MNPs at various concentrations. After incubation for 72 hours at 37°C in a 5% CO$_2$ incubator (Binder GmbH), 25 µL of a 5.5 mg/mL MTT solution was added to each well, covered with aluminum foil, and incubated for a further 3 hours in the dark. Immediately afterwards, the medium was aspirated and the remaining purple formazan was lysed with MTT solution. The assay was performed in triplicate. The optical density was then measured at 570 nm using an enzyme-linked immunosorbent assay universal microplate reader (Bio Tek Instruments, Inc., VT, USA). The inhibitory concentration 50 (IC$_{50}$) value was determined from the absorbance versus concentration curve. Dimethyl sulfoxide 0.1% was used as the negative control.

Acridine orange/propidium iodide assay
Fe$_3$O$_4$ MNP-induced death of Jurkat cells was investigated using the acridine orange/propidium iodide double-staining method according to the standard procedure29 followed by examination under a fluorescence microscope (Leica, Tokyo, Japan) with Leica Q Fluoro software (Leica) installed. The Jurkat cells were plated at a concentration of 1×10$^6$ cells/mL in a 25 cm$^2$ culture flask (TPP Techno Plastic Products), exposed to Fe$_3$O$_4$ MNPs at the IC$_{50}$ and incubated at 37°C in a 5% CO$_2$ incubator for 24, 48, and 72 hours. Next, the cells were centrifuged at 200×g (Universal 320R Centrifuge, Andreas Hettich GmbH & Co.KG, Tuttlingen, Germany) for 10 minutes and the supernatant was discarded. The cells were then washed twice with phosphate-buffered saline after centrifuging at 200×g for 10 minutes to remove the remaining medium. About 10 µL of the cell pellets were stained with a 10 µL fluorescent dye mixture containing equal volumes (100 µg/mL) of acridine orange and propidium iodide for 2 minutes. Approximately 10 µL of freshly stained cell suspension was placed onto a glass slide, covered with a cover slip, and examined under a fluorescent microscope.

Annexin V-fluorescein isothiocyanate assay
Jurkat cells (1×10$^6$ cells/mL) were exposed to the nanoparticles for 12, 24, and 48 hours, with untreated cells used as controls. The cells were then collected and centrifuged at 200×g (Universal 320R Centrifuge), (Andreas Hettich
GmbH & Co.KG) for 10 minutes to remove the medium. The cell pellets were washed twice with 1 mL of ice-cold phosphate-buffered saline, recentrifuged, and resuspended in ice-cold 1× binding buffer. Exactly 500 µL of cell suspension was transferred to a 5 mL culture tube (TPP Techno Plastic Products), to which 5 µL of Annexin V-fluorescein isothiocyanate (FITC) conjugate and 10 µL of propidium iodide were added. The cells were incubated for 15 minutes at room temperature in the dark. Finally, flow cytometry was done under laser emitting excitation light at 488 nm using a BD FACSCalibur™ flow cytometer equipped with an argon laser (BD Biosciences, Franklin Lakes, NJ, USA). The data were analyzed using BD CellQuest™ Pro software (BD Biosciences).

**Cell cycle assay**
Flow cytometry was used to investigate the cytotoxicity of Fe₃O₄ MNPs in Jurkat cells. Briefly, Jurkat cells (2.0×10⁶ cells/mL) were cultured with the IC₅₀ of Fe₃O₄ MNPs and incubated for 24, 48, and 72 hours. The cells were harvested by centrifugation at 1,500 rpm for 5 minutes and washed with phosphate-buffered saline. Next, 600 µL of 70% ice-cold ethanol were added to the cell pellets, which were then kept at −20°C overnight. A 1 mL volume of phosphate-buffered saline was added and spun down at 1,500 rpm for 5 minutes to remove the ethanol. The cell pellets were then washed twice with phosphate-buffered saline, stained with staining buffer containing 0.1% Triton X-100 (Sigma-Aldrich), 10 mM ethylenediaminetetraacetic acid (Sigma-Aldrich), 50 µg/mL Ribonuclease A (Sigma-Aldrich), and 5 µg/mL propidium iodide (Sigma-Aldrich), and incubated in the dark on ice for 30 minutes. Flow cytometry was performed with laser emitting excitation light at 488 nm using a FACSCalibur™ flow cytometer equipped with an argon laser (BD Biosciences). The data analysis was performed using CellQuest™ Pro software.

**Caspase-3 and caspase-9 assays**
The activity of caspase-3 and caspase-9 in Jurkat cells treated with Fe₃O₄ MNPs was determined after 24, 48, and 72 hours using a colorimetric assay kit (BD Biosciences) according to the manufacturer’s instructions without modification.

**Statistical analysis**
The experiments were done in triplicate, and the results are expressed as the mean ± standard deviation. The statistical analysis was done using IBM® SPSS® Statistics for Windows version 20.0 software (IBM Corp., Armonk, NY, USA). P-values <0.05 were considered to be statistically significant.

**Results and discussion**
Fe₃O₄ MNPs inhibit proliferation of human cancer cells
Observation of morphological changes in cells indicated that Fe₃O₄ MNPs inhibited proliferation of the various cancer cell lines in a dose-dependent and time-dependent manner (Figure 2A). No toxicity was seen in the normal Chang liver cell line (Figure 2B). The IC₅₀ values calculated for Fe₃O₄ MNPs in the various cancer cell lines were 23.83±1.1 µg/mL (HepG2), 18.75±2.1 µg/mL (MCF-7), 12.5±1.7 µg/mL (HeLa), and 6.4±2.3 µg/mL (Jurkat) after treatment for 72 hours. Therefore, Jurkat cells were selected for further investigation.

It is relevant to the evaluation toxicity of metal nanoparticles against cancer cells as many studies have already published. Coradeghini et al reported that BALB/C-3T3 mouse fibroblasts at higher doses than 50 µM for 72 hours of exposure to gold nanoparticles with a thickness of 5 nm, as compared with 15 nm, induced dose-dependent cell cytotoxicity. They reported that the size of gold nanoparticles is an important indicator of potential application in the biomedical field. Another recent study reported that different types of iron oxide nanoparticles demonstrate diverse biological activity as a result of versatile surface coating. Novotna et al...
examined the effect of superparamagnetic iron oxide nanoparticles on human bone marrow mesenchymal stromal cells from two donors. Their results showed only hBMSCs-2 were sensitive to these nanoparticles; however, increased oxidative injury to lipids, proteins, and DNA was detected in cells from both donors. 

Schweigera et al reported on Fe$_3$O$_4$ MNPs stabilized with poly(ethyleneimine)-g-poly(ethylene glycol) and branched poly(ethyleneimine). They also evaluated the cytotoxic effects of polymer-coated iron oxide nanoparticles in A549 epithelial adenocarcinoma cells. Their findings for cell viability showed that poly(ethyleneimine) layered onto iron oxide nanoparticles produced significant cytotoxicity compared to free poly(ethyleneimine) polymer.

**Quantification of apoptosis using acridine orange/propidium iodide double-staining**

Induction of apoptosis is a useful strategy in the treatment of cancer. Several cellular and molecularbiological features can be demonstrated in apoptotic cells, including cell shrinkage, DNA fragmentation, and activation of the caspase cascade. The cytotoxicity of Fe$_3$O$_4$ MNPs was evaluated by inhibition of cell growth. When the growth-inhibited cells were stained with acridine orange/propidium iodide, apoptotic cell death was observed in a time-dependent and dose-dependent manner in all cultures (Figure 3).

Intact membranes exclude propidium iodide, but allow uptake of acridine orange, which binds to double-stranded DNA and fluoresces green under 488 nm excitation. Untreated cells show diffuse green fluorescence, while apoptotic cells (containing condensed chromatin material) show clumps of intense green fluorescent spots within the cell. Characteristic condensation patterns were observed such as crescent shapes found at the nuclear periphery and an increase in the number of round clumps.

**Annexin V-FITC assay**

The apoptotic effect induced by the Fe$_3$O$_4$ MNPs was investigated further by determining the percentage of apoptotic cells using flow cytometric analysis with Annexin V/propidium iodide double-staining. Annexin V-positive/propidium iodide-negative staining indicated cells in early apoptosis due to the strong affinity of Annexin V-FITC for phosphatidylserine, which transports Annexin V protein from the inner leaflet of the plasma membrane to the outer surface of the membrane during early apoptosis. On the other hand, Annexin V-negative/propidium iodide-positive staining indicates necrotic cells, since propidium iodide, which cannot pass...
through an intact cell membrane, penetrates the compromised membranes of cells that are dead or in late apoptosis, and binds to nucleic acids. Meanwhile, Annexin V-negative/propidium iodide-negative staining indicates viable cells and Annexin V-positive/propidium iodide-positive staining indicates cells in late apoptosis. The representative dot plots for the flow cytometric analysis comparing untreated cells and treated cells (at 12, 24, and 48 hours) showed an increase in the percentages of treated cells in early apoptosis and late apoptosis (Figure 4 and Table 1). In addition, exposure to Fe$_3$O$_4$ MNPs resulted in a slight decrease in the amount of viable cells at 12, 24, and 48 hours. These results suggest that the antiproliferative effect of Fe$_3$O$_4$ MNPs on Jurkat cells results from induction of cell apoptosis.

**Cell cycle assay**

In cells treated with agents that induce apoptosis, a subpopulation of cells appears before the G1 peak and is
Table 1  Flow cytometric analysis of Jurkat cells treated with Fe₃O₄ MNPs

<table>
<thead>
<tr>
<th>Cell Condition</th>
<th>Control 12 hours (Mean ± SD)</th>
<th>Treated 12 hours (Mean ± SD)</th>
<th>Control 24 hours (Mean ± SD)</th>
<th>Treated 24 hours (Mean ± SD)</th>
<th>Control 48 hours (Mean ± SD)</th>
<th>Treated 48 hours (Mean ± SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Viable cells</td>
<td>95.54±0.11</td>
<td>89.59±0.26</td>
<td>93.33±0.55</td>
<td>85.29±0.16</td>
<td>74.98±0.33</td>
<td>93.78±0.26</td>
</tr>
<tr>
<td>Early apoptosis</td>
<td>1.77±0.14</td>
<td>4.75±0.61*</td>
<td>1.98±0.57</td>
<td>8.00±0.38*</td>
<td>3.15±0.41</td>
<td>15.04±0.52*</td>
</tr>
<tr>
<td>Late apoptosis/necrosis</td>
<td>2.69±0.34</td>
<td>5.66±0.95**</td>
<td>4.69±0.50</td>
<td>7.71±0.82**</td>
<td>3.07±0.25</td>
<td>9.98±0.50**</td>
</tr>
</tbody>
</table>

**Notes:** The cells were stained with FITC-conjugated Annexin V and propidium iodide, then incubated at 37°C for 12, 24, and 48 hours. Values are expressed as the mean ± standard deviation of three different experiments. The data were analyzed using post hoc comparison test-one way analysis of variance, and means were compared by Tukey’s B test. *Significant (P<0.05) increase in early apoptotic cells in Fe₃O₄ MNP-treated groups compared with untreated controls. **Significant (P<0.05) increase in late apoptotic/necrotic cells in Fe₃O₄ MNP-treated groups compared with untreated controls.

**Abbreviations:** Fe₃O₄ MNP, magnetic iron oxide nanoparticles; FITC, fluorescein isothiocyanate.

referred to as the sub-G1 peak. This is believed to be the result of endonuclease activation and subsequent leakage of DNA from the cells. Necrotic cells do not show an immediate reduction in DNA content, so a clear distinction could be made between necrotic and apoptotic cells. As shown in Figure 5 and Table 2, the sub-G1 population, indicating apoptotic cells, increased in a time-dependent manner from 10.10±0.28 after 24 hours. The data were analyzed using post hoc comparison test-one way analysis of variance, and means were compared by Tukey’s B test. *Significant (P<0.05) increase in late apoptotic/necrotic cells in Fe₃O₄ MNP-treated groups compared with untreated controls. **Significant (P<0.05) increase in late apoptotic/necrotic cells in Fe₃O₄ MNP-treated groups compared with untreated controls.

**Abbreviations:** Fe₃O₄ MNP, magnetic iron oxide nanoparticles; FITC, fluorescein isothiocyanate.

Figure 5  Cell cycle analysis of Jurkat cells treated with Fe₃O₄ MNPs after staining with propidium iodide. (A1–C1) Untreated Jurkat cells treated for 24, 48, and 72 hours, respectively. (A2–C2) Effects of 24, 48, and 72 hours of exposure of Jurkat cells to Fe₃O₄ MNPs, respectively. G0/G1, G2/M, and S indicate the cell phase, and sub-G0/G1 refers to the proportion of apoptotic cells.

**Abbreviations:** Fe₃O₄ MNP, magnetic iron oxide nanoparticles; DNA, deoxyribonucleic acid.
Table 2 Flow cytometric analysis of Jurkat cells treated with FeO₃ MNPs

<table>
<thead>
<tr>
<th>Cells (%)</th>
<th>Control 24 hours</th>
<th>Treated 24 hours</th>
<th>Control 48 hours</th>
<th>Treated 48 hours</th>
<th>Control 72 hours</th>
<th>Treated 72 hours</th>
</tr>
</thead>
<tbody>
<tr>
<td>G0/G1</td>
<td>56.25±0.06</td>
<td>33.44±0.45</td>
<td>51.13±0.29</td>
<td>42.61±0.52</td>
<td>37.64±0.32</td>
<td>45.68±0.68</td>
</tr>
<tr>
<td>G2/M</td>
<td>17.85±0.76</td>
<td>18.29±0.41</td>
<td>20.16±0.26</td>
<td>12.29±0.35</td>
<td>24.22±0.22</td>
<td>10.06±0.93</td>
</tr>
<tr>
<td>Synthesis</td>
<td>24.91±0.06</td>
<td>38.23±0.33</td>
<td>29.24±0.06</td>
<td>22.93±0.12</td>
<td>38.95±0.61</td>
<td>21.35±0.18</td>
</tr>
<tr>
<td>Sub G0/G1</td>
<td>0.98±0.23</td>
<td>10.10±0.28*</td>
<td>0.20±0.34</td>
<td>20.01±0.20*</td>
<td>0.02±0.46</td>
<td>23.85±0.56*</td>
</tr>
</tbody>
</table>

Notes: The cells were stained with propidium iodide and incubated at 37°C for 24, 48, and 72 hours. Values are expressed as the mean ± standard deviation of three different experiments. The data were analyzed using post hoc comparison test-one way analysis of variance, and means were compared by Tukey's B test. *Significant (P<0.05) increase of cells in sub-G0/G1 phase in FeO₃ MNP-treated groups compared with untreated controls.

Abbreviation: FeO₃ MNP, magnetic iron oxide nanoparticles.

Caspase assay
The apoptotic effect of FeO₃ MNPs was also examined by determining caspase-3 and caspase-9 activity relative to varying concentrations of protein content for cells treated with the IC₅₀ of FeO₃ MNPs. Our results show that enzyme activity increased in a time-dependent manner, as shown in Figure 6 and Table 3.

Apoptosis is orchestrated by a family of cysteine proteases known as caspases. The main effectors of apoptosis encompass proteases from the caspase family, which reside as latent precursors in most nucleated animal cells. Fourteen mammalian caspases have been identified, three of which (caspase-3, caspase-6, and caspase-7) are thought to coordinate the execution phase of apoptosis by cleaving multiple structural and repair proteins. Pathways to caspase-3 activation have been identified, which are either dependent on or independent of release of mitochondrial cytochrome c and caspase-9 function. Caspase-3 activation is the hallmark of apoptosis, and is necessary for apoptotic chromatin condensation and DNA fragmentation in all cell types examined. Thus, caspase-3 is essential for certain processes associated with dismantling of the cell and the formation of apoptotic bodies. However, it may also function before or at the stage when commitment to loss of cell viability is made. As shown in Figure 6, FeO₃ MNPs increased the activity of caspase-9 and caspase-3 in a time-dependent manner. According to studies of caspase activity, several distinct pathways exist, resulting in induction of apoptosis by FeO₃ MNPs. In our study, we noted that FeO₃ MNPs activated caspase-3 and caspase-9 in a time-response manner.

Conclusion
FeO₃ MNPs with different sizes and characteristics have been developed and widely investigated in a number of biomedical applications. Stabilizing and preventing formation of aggregation are critical for controlling chemical reactions, which provide dependable, well-dispersed size and consistency. In this study, we investigated the toxicity of FeO₃ MNPs in human leukemia (Jurkat) cells. Our results show that exposure of Jurkat cells to FeO₃ MNPs results in significant cytotoxicity, with an apoptotic response, but not in a normal Chang liver cell line, providing new opportunities for safe delivery of FeO₃ MNPs and application in anticancer therapy.

Acknowledgment
The authors are grateful to the staff of Universiti Putra Malaysia for their help with this research, and to the Department of
Table 3 Spectrophotometric analysis of caspases in Jurkat cells treated with Fe$_3$O$_4$ MNPs for 24, 48, and 72 hours

<table>
<thead>
<tr>
<th>Caspase</th>
<th>Control 24 hours</th>
<th>Treated 24 hours</th>
<th>Control 48 hours</th>
<th>Treated 48 hours</th>
<th>Control 72 hours</th>
<th>Treated 72 hours</th>
</tr>
</thead>
<tbody>
<tr>
<td>Caspase-3</td>
<td>0.057±0.003</td>
<td>0.095±0.001*</td>
<td>0.068±0.010</td>
<td>0.158±0.025*</td>
<td>0.080±0.095</td>
<td>0.199±0.055*</td>
</tr>
<tr>
<td>Caspase-9</td>
<td>0.059±0.001</td>
<td>0.102±0.009*</td>
<td>0.071±0.081</td>
<td>0.179±0.016*</td>
<td>0.095±0.039</td>
<td>0.207±0.031*</td>
</tr>
</tbody>
</table>

Notes: Values are expressed as the mean ± standard deviation of three different experiments. The data were analyzed using post hoc comparison test-one way analysis of variance, and means were compared by Tukey’s B test. *Significant (P<0.05) increase in apoptotic cells in Fe$_3$O$_4$ MNP-treated groups compared with untreated controls.

Abbreviation: Fe$_3$O$_4$, MNP, magnetic iron oxide nanoparticles.

Chemistry, Faculty of Science, Universiti Putra Malaysia, for allowing us to use its laboratory facilities and providing technical assistance.

Disclosure

The authors report no conflicts of interest in this work.

References


