Combining rimonabant and fentanyl in a single entity: preparation and pharmacological results

Abstract: Based on numerous pharmacological studies that have revealed an interaction between cannabinoid and opioid systems at the molecular, neurochemical, and behavioral levels, a new series of hybrid molecules has been prepared by coupling the molecular features of two well-known drugs, ie, rimonabant and fentanyl. The new compounds have been tested for their affinity and functionality regarding CB₁ and CB₂ cannabinoid and μ opioid receptors. In [35S]-GTPγS (guanosine 5’-O-[γ-thio]triphosphate) binding assays from the post-mortem human frontal cortex, they proved to be CB₁ cannabinoid antagonists and μ opioid antagonists. Interestingly, in vivo, the new compounds exhibited a significant dual antagonist action on the endocannabinoid and opioid systems.

Keywords: fentanyl, rimonabant, cannabinoid, opioid, behavioral assays

Introduction

Pharmacological data have revealed interactions between the cannabinoid and opioid systems at the molecular, neurochemical, and behavioral levels. Anatomical studies have shown that such pharmacological interactions are likely due to the close vicinity of CB₁ and opioid receptors in the central nervous system structures involved in the control of nociception, motor activity, and brain reward. In this context, the development of a single molecule able to act on both the cannabinoid and opioid systems seems an interesting approach. Multivalent or multifunctional ligands constitute promising pharmacological tools and new targets for drug development. Moreover, targeting G protein-coupled receptors with hybrid molecules has been explored in different domains. The concept of hybridization has become of increasing interest for medicinal chemists. This strategy has resulted in promising leads that show significant therapeutic advantages. The main benefit compared with the administration of two drugs concerns the pharmacokinetic properties of a single molecule. In order to focus our efforts on an efficient designed multiple ligand approach, two well validated drugs were selected, ie, fentanyl and rimonabant (SR141716, Figure 1). Fentanyl is a synthetic μ opioid agonist widely used in clinical practice and rimonabant was the first selective CB₁ cannabinoid receptor inverse agonist/antagonist to be approved for clinical use, even though its use was discontinued due to unwanted effects.

Therefore, our aim was to explore the hybrid molecules represented in Figure 1 (4a–k) based on previous structural modification studies carried out on fentanyl and rimonabant. Fentanyl derivatives (Figure 2), in which a phenyl was replaced by an aliphatic chain bearing a guanidine moiety, have shown high affinity for the μ opioid receptor, with agonistic properties. Concerning the cannabinoid counterpart of the proposed hybrid molecules,
good affinities for the CB₁ receptor have been reported for diarylcarboxamides bearing alkyl chains (Figure 2).

Herein, we describe the synthesis, pharmacological profile, and behavioral effects in vivo of a series of hybrid molecules derived from fentanyl and rimonabant.

Materials and methods

General

All commercially available reagents were purchased from Sigma-Aldrich (Madrid, Spain) and used without further purification. Dry CH₂Cl₂ was obtained by distillation over calcium

**Figure 1** The designed multiple ligands are based on the CB₁ cannabinoid receptor inverse agonist/antagonist rimonabant (SR141716) (Sanofi, Paris, France) and the µ opioid agonist fentanyl. X represents an n-alkyl chain from propyl to dodecyl, 1,3-phenyl, 1,3-benzyl, or bicyclohex-4-ylmethane.

**Figure 2** Examples of fentanyl derivatives with selective µ opioid agonistic properties and rimonabant derivatives with CB₁ receptor affinity.

Abbreviation: Kᵢ, affinity constant.
chloride. Flash column chromatography was performed using silica gel 60 (230–400 mesh) or on a medium pressure flash system with a prepacked silica gel cartridge (Biotage Flash 40 [SYMTA, Madrid, Spain], cartridges KP-Sil 40S [4 × 7 cm] or 4M [4 × 15 cm] with a particle size of 32–63 μm and 60 Å; FlashMaster Personal [SYMTA] with prepacked cartridges FlashPack of 2, 10, 20 or 50 g). Starting products 1 and 2 were prepared as previously described.17 N-[1-phenethyl-4-piperidyl]propanamide 3a–3k were prepared following a procedure previously described by our group.13,18

Chemistry
Compounds 4a–4k were prepared following the synthetic route depicted in Figure 3. Diaryl-1H-pyrazole-3-carboxylic acid 1 was previously prepared according to Krishnamurthy’s procedure.19 Next, pyrazole 1 was converted to the acid chloride 2 using the following procedure. A solution of the appropriate propanamide 3a–3k and triethylamine in dry CHCl3 was added dropwise to pyrazole-3-carboxylic acid chloride 2 dissolved in dry CHCl3 (5 mL). The reaction mixture was stirred at room temperature for 16 hours. The solvent was then removed under reduced pressure. The crude product was purified on silica gel by flash chromatography eluting with CHCl3/(MeOH/NH4Cl) (100:0–100:2) to provide the corresponding pyrazole-3-carboxamides 4a–4k. The yields of the preparation and the structural assignments of 4a–4k are reported in the Supplementary material section (1H-NMR [nuclear magnetic resonance], 13C-NMR, MS [mass spectroscopy], and elemental analysis).

Pharmacology
Post-mortem human brain CB2 and μ opioid binding assays
The ability of the new compounds to bind to the CB2 cannabinoid receptors and to the μ opioid receptor was evaluated in competitive displacement assays using, respectively, [3H]-CP55,940 (PerkinElmer, Boston, MA, USA) and [3H]-DAMGO ([DAla2, N-Me-Phe4, Gly5-ol]-enkephalin; American Radiolabeled Chemicals Inc, St Louis, MO, USA) as radioligands. Prefrontal cortex membranes from post-mortem human brain were used as a source of receptors, given that CB2 receptors20 and μ opioid receptors are present in large amounts in this region. Neural membranes (P3 fractions) were prepared from the prefrontal cortex of human brains obtained at autopsy (Instituto Vasco de Medicina Legal, Bilbao, Spain) as previously reported by our research group.16 These samples belong to the Brain Collection of the University of the Basque Country registered with the National Biobank Register of the Spanish Health Department number (C 0000035). Specific [3H]-CP55,940 or [3H]-DAMGO binding was measured in 0.55 mL aliquots (50 mM Tris-HCl [Tris(hydroxymethyl) aminomethane hydrochloride], pH 7.5) of the neural membranes. The membranes were incubated with [3H]-CP55,940 (1 nM) for 60 minutes at 30°C or [3H]-DAMGO (2 nM) for 60 minutes at 25°C in the absence or presence of the competing compounds (10−12 M to 10−3 M, ten concentrations). Incubations were terminated by diluting the samples with 5 mL of ice-cold Tris-HCl incubation buffer (4°C). Membrane-bound radioligand was separated by vacuum filtration through Whatman GF/C glass fiber filters (GE Healthcare, Buckinghamshire, UK). The filters were then rinsed twice with 5 mL of incubation buffer and transferred to mini-vials containing 3 mL of OptiPhase HiSafe® II cocktail (PerkinElmer) and counted for radioactivity by liquid scintillation spectrometry. Specific binding was determined and plotted as a function of the compound concentration. Nonspecific binding was determined in the presence of WIN 55,212-2 (10−5 M) or naloxone (10−5 M), respectively. Analysis of competition experiments to obtain the inhibition constant (Ki) were performed by nonlinear regression using the GraphPad Prism program (GraphPad Software, Inc, San Diego, CA, USA). All experiments were analyzed assuming a one-site model of radioligand binding.

CB2 binding evaluation
CB2 receptor affinity was evaluated from membrane fractions of human embryonic kidney (HEK293 EBNA) cells transfected with human CB2 receptors (RBXCB2M400UA; Analytical Sciences, Boston, MA, USA).21 The CB2 receptor membrane protein concentration was 5.20 pmol/mg or 6.20 pmol/mg and the protein concentration was 4.0 mg/mL or 3.6 mg/mL depending on the batch. The commercial membranes were diluted (approximately 1:20) with the binding buffer (50 mM Tris-HCl, 5 mM MgCl2⋅H2O, 2.5 mM ethylene glycol tetraacetic acid, 1 mg/mL bovine serum albumin, pH 7.5). The final membrane protein concentration was 0.2 mg/mL of incubation volume. The radioligand used was [3H]-CP55,940 at a concentration of membrane Kd (dissociation constant) ×0.8 nm, and the final volume was 600 mL. The 96-well plates and tubes necessary for the experiment were previously siliconized with Sigmacote (Sigma-Aldrich, St Louis, MO, USA). The membranes were resuspended in the corresponding buffer and were incubated with the radioligand at each compound (10−4–10−11 M) for 90 minutes at 30°C. Nonspecific binding was determined with 10 μM WIN 55,212-2, and 100% binding of the radioligand to the
membrane was determined by its incubation with membrane without any compound. Filtration was performed by a Harvester® FilterMate (PerkinElmer) with Filtermat A GF/C filters pretreated with polyethylenimine 0.05%. After filtering, the filter was washed nine times with binding buffer, dried, and a melt-on scintillation sheet (MultiLex™; PerkinElmer) was melted onto it. Radioactivity was then quantified by a liquid scintillation spectrophotometer (Wallac MicroBeta® TriLux; PerkinElmer). Competition binding data were analyzed using the GraphPad Prism program, and $K_i$ values are expressed as the mean ± standard error of the mean (SEM) of at least three experiments performed in triplicate for each point.

Functionality binding assays
To determine the ability of the selected compounds to activate the CB$_2$ and/or µ opioid receptors, [³⁵S]-GTPγS (guanosine S′-O-[gamma-thio]triphosphate) binding assays were performed in cortical membranes from post-mortem human brain. In this tissue, [³⁵S]-GTPγS binds with high affinity to G$_i$/G$_o$ proteins.²² Thereby, agonists, inverse agonists, and antagonists can modulate this binding acting on a specific receptor, increasing (agonists) or decreasing (inverse agonists) the nucleotide binding or blocking the effect of an agonist (antagonists). The incubation buffer for measuring [³⁵S]GTPγS binding to brain membranes contained 1 mM ethylene glycol tetracetic acid, 3 mM MgCl$_2$, 100 mM NaCl, 50 mM GDP (guanosine diphosphate), 50 mM Tris–HCl at pH 7.4, and 0.5 nM [³⁵S]GTPγS (DuPont NEN, Brussels, Belgium) in a total volume of 500 µL. Protein aliquots were thawed and resuspended in the same buffer. The incubation was started by addition of the membrane suspension (40 µg of membrane proteins) to the previous mixture and was performed at 30°C for 120 minutes with shaking. In order to evaluate the influence of the compounds on [³⁵S]GTPγS binding, ten concentrations (10⁻¹²–10⁻³ M) of the different compounds were added to the assay. Incubations were terminated by adding 3 mL of ice-cold resuspension buffer followed by rapid filtration through Whatman GF/C filters presoaked in the same buffer. The filters were rinsed twice with 3 mL of ice-cold resuspension buffer, transferred to vials containing 5 mL of OptiPhase HiSafe II cocktail, and the radioactivity trapped was determined by liquid scintillation spectrometry (Packard 2200CA; Packard Instrument Company, Meriden, CT, USA). The [³⁵S]GTPγS bound was about 7%–14% of the total [³⁵S]GTPγS added. Nonspecific binding of the radioligand was defined as the remaining [³⁵S]GTPγS binding in the presence of 10 µM unlabeled GTPγS.

In vivo cannabinoid tetrad assays
Male imprinted control region mice weighing 25–30 g were used. Spontaneous behavior was always observed in the cage before treatment and/or performance of the different tests. Animals showing spontaneous behavioral modifications were discarded. To evaluate agonist effects, reference drugs and new compounds were administered 15 minutes (for the cannabinoid tetrad) and 30 minutes (for the opioid hot plate test) before starting the behavioral tests. When the compounds were tested as antagonists, they were administered 20 minutes before the reference agonists (WIN 55,212-2 or morphine). All drugs were given intraperitoneally. Separate groups of mice (n = 8–10 each) were given the following treatments: saline solution or vehicle (controls); WIN 55,212-2 1.5 mg/kg; 4d 10 mg/kg; 4e 5 mg/kg; rimonabant 1 mg/kg; rimonabant 1 mg/kg + WIN 55,212-2 1.5 mg/kg; 4d 2 mg/kg + WIN 55,212-2 1.5 mg/kg; 4d 4 mg/kg + WIN 55,212-2 1.5 mg/kg; 4d 8 mg/kg + WIN 55,212-2 1.5 mg/kg; and 4e 5 mg/kg + WIN 55,212-2 1.5 mg/kg. The tests were conducted consecutively at 5-minute intervals.

Hypothermia
Core mouse temperatures were measured using a lubricated thermometer inserted into the rectum to a constant depth of 1 cm. Temperature was evaluated twice in each animal, ie, before and after every treatment.

Locomotor activity
Spontaneous locomotor activity was evaluated using individual photocell activity chambers (Cibertec®, San Jose, Costa Rica). The mouse was placed in a chamber and, starting 10 minutes later, the number of interruptions of photocell beams was recorded over a 30-minute period. The mean number of crossings was compared with that obtained from a mouse control group that had received vehicle.

Nociception
The hot plate test was carried out using a hot plate at 55°C as the nociceptive stimulus. The latency time of licking of the front paw was taken as an index of nociception. The latency was measured before treatment (control latency) and after every treatment (latency after treatment). The cut-off time was 30 seconds and analgesia was quantified with the formula of the maximum possible effect (MPE), expressed as a percentage:

\[
\% \text{ MPE} = \frac{(\text{Latency after treatment} - \text{Control latency})}{(\text{Cut-off time} - \text{Control latency})} \times 100
\]
Catalepsy was measured using a modified “ring test” as originally described by Pertwee. The mice were placed on a rubber-coated metal ring (6 cm in diameter) fixed horizontally at a height of 30 cm. CB₁ cannabinoid agonists cause animals to become cataleptic, and the sum of all times during which the mice were immobile was registered for a 5-minute period and compared with the time registered in control animals. The criterion for immobility was the absence of all voluntary movement.

To confirm the duration of the effect of the new compounds, they were tested as antagonists on the hot plate test. Next, 4d (4 mg/kg, intraperitoneally) and 4e (5 mg/kg intraperitoneally) were administered 20 minutes before WIN 55,212-2 (1.5 mg/kg intraperitoneally) and the test was performed 1 hour after injection of this agonist.

In vivo opioid response on hot plate test
To assess the opioid activity of the new compounds 4d and 4e, the hot plate test was carried out as described above and separate groups of mice (n = 8–10 each) were treated with: saline solution or vehicle (controls); 4d 10 mg/kg; 4e 10 mg/kg; naloxone 1 mg/kg; rimonabant 1 mg/kg; morphine 10 mg/kg; 4d 10 mg/kg + morphine 10 mg/kg; 4e 10 mg/kg + morphine 10 mg/kg; naloxone 1 mg/kg + morphine 10 mg/kg; and rimonabant 1 mg/kg + morphine 10 mg/kg. The data are expressed as the mean ± SEM. For the in vitro assays, values for EC₅₀ (half-maximal effective concentration) and 95% confidence limits of these values for agonists were calculated by nonlinear regression analysis using the equation for a sigmoid concentration-response curve (GraphPad Prism). For the effects of drugs on in vitro and in vivo tests, a one-way analysis of variance was used for statistical analysis of multiple comparisons within each group. When a significant difference was detected by one-way analysis of variance, the data were further analyzed using Newman–Keuls test. In each test, a P-value less than 0.05 was considered to indicate statistical significance. Drugs (WIN 55,212-2 mesylate and rimonabant) were obtained from Tocris (Biogen Científica SL, Madrid, Spain) and Sanofi, respectively. DAMGO, DL-dithiothreitol, GDP, GTP, GTPγS, fentanyl, and naloxone were purchased from Sigma-Aldrich. Morphine HCl was obtained from Alcaliber SA (Madrid, Spain). Morphine sulfate salt solution and naloxone hydrochloride dihydrate were obtained from Sigma-Aldrich (Madrid, Spain). All other chemicals were of the highest commercially available purity. Cannabinoids were dissolved in ethanol 1 mg:1 mL and subsequently in ethanol and Tween 80 (1:2), after which the ethanol was evaporated. Other drugs were dissolved in saline solution (0.9%).

Alcohol intake assays
The operant ethanol self-administration was examined following a protocol essentially based on a previously published alcohol relapse model in Wistar rats.

Administration, distribution, metabolism, excretion (ADME) properties in silico
ADME properties were calculated using QikProp version 3.5 integrated in Maestro (Schrodinger, LLC, New York, NY, USA). The global minimum energy conformer used as input was generated using the program Spartan ‘08 (Wave Function, Inc, Irvine, CA, USA). Ab initio energy minimizations were performed at the Hartree-Fock 6-31G* level.

Results and discussion
Receptor affinities and structure-activity relationship
Concerning the CB₁ cannabinoid binding, Table 1 displays the Kᵢ values obtained for 4a–4k and for AM251 (Tocris, Bristol, UK) and rimonabant as reference CB₁ ligands. Some of the new compounds (4a, 4b, 4d–4f, 4k) showed low to medium affinity (Kᵢ = 0.19–3.99 µM), while others (4c, 4g–4j) did not show any affinity for the CB₁ receptor in this assay (Kᵢ >10 µM). The ligands showing the highest affinity were compounds 4b (Kᵢ = 0.57 µM), and 4e (Kᵢ = 0.70 µM), containing a butyl and a heptyl chain linker, respectively. Now, if we refer to our recent published binding data on bivalent cannabinoid ligands, it is interesting to note that the bivalent molecule with the heptyl linker also showed the best CB₁ affinity. Actually, the alkyl chain length does not correlate with CB₁ receptor affinity. However, the values obtained for 4g–4j indicate that aromatic spacers and longer alkyl chains lead to a loss of CB₁ cannabinoid receptor affinity. Interestingly, replacing the linear alkyl chain of nine methylene units (4g; Kᵢ =108 µM) with a bicyclohexylene (4k; Kᵢ =2.06 µM) as a conformation constrainer, resulted in increased affinity for the CB₁ receptor.

The µ opioid binding component is reflected in Table 1. Excluding 4a, 4e, and 4h, which showed a Kᵢ higher than 3.8 µM, the Kᵢ values of 4b–d, 4f–g, and 4i–k, that ranged from 205 nM to 1.3 µM, indicate moderate affinity for the µ opioid receptor. Compounds 4g, 4e, and 4d showed the best Kᵢ values (108, 166, and 295 nM, respectively).
These CB₁ and opioid competitive displacement assays were performed in the post-mortem human brain, where CB₁ and opioid receptors are present in large amounts and where the presence of dimers and heteromers has been proposed.

The fact that the new compounds with affinity for both receptors have the shortest linkers suggests that they bind to CB₁ and to opioid monomers. Spacers ranging from 10 to 20 atoms are necessary for the molecule to bridge heteromer receptors. It is unlikely that these hybrid molecules target a CB₁ opioid heteromer. However, they bind individually to the respective receptors. Regarding CB₂ receptor binding assays, compounds 4a–4j did not show any significant affinity (>40 μM), thus indicating CB₁ selectivity. CB₂ binding assay for 4k could not be carried out for solubility reasons. The new compounds have a CB₁ activity profile.

**Functionality**

Although these new compounds showed modest affinity, evaluation of their functionality has yielded interesting results. We selected 4b and 4e for their CB₁ cannabinoid affinity and 4d for its µ opioid affinity along with a micromolar CB₁ affinity constant value. [35S]-GTPγS binding assays have been performed in cortical membranes from the post-mortem human brain to determine the ability of the selected compounds to activate the CB₁ and/or µ opioid receptors.

We first evaluated the direct effect of compounds 4b, 4d, and 4e. Compound 4b produced an inhibition of basal [35S]-GTPγS binding (EC₅₀ 25±4 μM; Figure 4), demonstrating its

![Figure 3 Synthesis of the pyrazole carboxamide derivatives 4a–4k. Reagents and conditions: (i) SOCl₂, toluene, Δ (reflex temperature); (ii) CH₂Cl₂, Et₃N, room temperature.](Image)
inverse agonist effect. This effect is similar to that induced by the well known CB₁ inverse agonist/antagonist rimonabant, used as reference (EC₅₀ 19±4 μM; Figure 4).²⁶ Compounds 4d, 4e, and the opioid-selective antagonist naloxone did not induce any significant change in basal [³⁵S]-GTPγS binding values, indicating a lack of agonist or inverse agonist properties (Figure 4).

The next step was to characterize their antagonist effect on cannabinoid receptors. The maximum agonist effects (Eₘₐₓ) of WIN 55,212-2 (Eₘₐₓ 184±4%; EC₅₀ 1.1±0.22 μM) were reverted by each of the new compounds (4b, 4d, and 4e), as well as by rimonabant, at a single concentration (10⁻⁵ M, Figure 5A). They produced a rightward displacement of the WIN 55,212-2 dose-response curve (EC₅₀ for rimonabant, 21±3 μM; EC₅₀ for 4b, 33±8 μM; EC₅₀ for 4d, 21±2 μM; EC₅₀ for 4e, 16±2 μM). These data show that 4b, 4d, and 4e behave as antagonists of the CB₁ cannabinoid receptor with potency similar to that of rimonabant. Conversely, the opioid antagonist naloxone (10⁻⁵ M) did not change the WIN 55,212-2 stimulation (EC₅₀ 1.3±0.17 μM; Eₘₐₓ 187±4%).

Finally, compounds 4b, 4d, and 4e have been evaluated for their antagonism at the μ opioid receptor. The agonist effects of the opioid fentanyl (Eₘₐₓ 146±2%; EC₅₀ 0.28±0.04 μM) were blocked by 4b, 4d, 4e, and naloxone at a single concentration (10⁻⁵ M, Figure 5B). All of these caused a rightward displacement of the fentanyl dose-response (EC₅₀=456±60 μM for naloxone; EC₅₀=24±5 μM for 4b; EC₅₀=33±2 μM for 4d; EC₅₀=3±1 μM for 4e). These data confirm that 4b, 4d, and 4e behave as opioid antagonists.

With the above functionality studies, we clearly demonstrate that 4b, 4d, and 4e have dual functionality, i.e., CB₁ receptor antagonism with potency similar to that of rimonabant and μ opioid receptor antagonism with potency superior to that of naloxone. Since previous studies suggested therapeutic interest in coadministration of a cannabinoid antagonist and an opioid antagonist,²⁷-³⁰ we considered that it would be interesting to follow on with in vivo studies.

**Behavioral properties**

The in vivo cannabinoid properties of 4d and 4e were characterized on the basis of their behavioral effects in mice. Cannabinoid agonists have been shown to induce a drop in body temperature, catalepsy on an elevated ring, acute analgesia on a hot plate, and a decrease of spontaneous activity in an open field.³¹ Compound 4d, at the three tested doses (intraperitoneal administration; Figure 6) antagonized all the effects produced by the cannabinoid agonist WIN 55,212-2 (1.5 mg/kg), the difference being statistically significant for rectal temperature, catalepsy, and analgesia. Three (catalepsy, analgesia, and reduction of the spontaneous activity) of the four effects induced by WIN 55,212-2 (1.5 mg/kg) on mice were significantly prevented by 4e (5 mg/kg, intraperitoneal administration; Figure 7). These results support the antagonistic CB₁ cannabinoid properties of 4d and 4e. It is unlikely that this antagonism could be due to loss of CB₁ receptors as a consequence of a downregulation process since it is normally the result of chronic rather than acute administration, as in this case.

To assess the duration of effect of 4d and 4e, the hot plate test was carried out after intraperitoneal injection of WIN 55,212-2 (1.5 mg/kg). Both compounds (4d at 4 mg/kg and 4e at 5 mg/kg, administered intraperitoneally) completely

![Figure 4](https://www.dovepress.com/...)

**Figure 4** Concentration-response curves for stimulation of [³⁵S]-GTPγS binding by 4b, 4d, 4e, rimonabant, and naloxone to cortical membranes in the post-mortem human brain. The data are expressed as the mean (± SEM) values for 4–6 experiments performed in different brain samples.

**Notes:** Rimonabant (Sanofi, Paris, France); naloxone (Sigma-Aldrich, Madrid, Spain).

**Abbreviations:** SR, rimonabant; Nx, naloxone; SEM, standard error of the mean; GTPγS, guanosine 5'-O-[γ-thio]triphosphate.
and significantly \(P<0.001\), one-way analysis of variance) reversed the effect of the cannabinoid agonist WIN 55,212-2 (percent of MPE for 4d =3.5±2.5 and percent MPE for 4e =5.7±4.7; values shown as the mean ± SEM \([n=6]\)).

The in vivo opioid properties of 4d and 4e were evaluated using a hot plate test at 55°C as the nociceptive stimulus (Figure 8). Treatment with the opioid agonist morphine induced antinociception that was reversed by the opioid antagonist naloxone but not by the cannabinoid antagonist rimonabant. The data described above show that intraperitoneal administration of 4d (10 mg/kg) or 4e (10 mg/kg) did not induce any analgesic effect. However, when 4d (10 mg/kg, intraperitoneally) or 4e (10 mg/kg, intraperitoneally) were administered 20 minutes before injection of morphine (10 mg/kg), the antinociceptive effect of morphine was partially antagonized. This effect reached statistical significance in the case of 4e. From these findings, it can be concluded that 4d and 4e exhibit a \(\mu\) opioid antagonist profile.

An extensive literature search suggests that the endocannabinoid and \(\mu\) opioid systems are involved in the development of alcohol dependence. In rodent models, rimonabant was found to decrease alcohol consumption.\(^{24}\) Treatment with the nonselective opioid antagonist naltrexone was effective in modulating aspects of alcohol-seeking behavior.\(^{32}\) Due to the \(\mu\) opioid and CB\(_1\) cannabinoid dual antagonist activity on behavioral evaluations of 4e, the effects of this compound on operant ethanol self-administration were examined following a protocol essentially based on the alcohol relapse model in Wistar rats. For this purpose, the alcohol deprivation effect model previously reported by our group was used.\(^{33}\) The animals did not show significant changes in number of alcohol responses compared with the vehicle group during treatment with 4e at doses of 0.5 mg/kg, 2.0 mg/kg, and 8.0 mg/kg (data not shown).

**ADME parameters in silico**

A computational approach is considered to be an appropriate tool in early-stage drug discovery. Therefore, in silico ADME predictions can provide significant insights into drug-like properties.\(^{34}\) Calculation of ADME properties of 4e was performed on the conformer of global minimum energy using QikProp version 3.5 integrated in Maestro with a set of 34 physicochemical descriptors. Based on our predicted data, Lipinski\(^{35}\) and Jorgensen\(^{16}\) pharmacokinetic rules are followed. With regard to blood–brain barrier penetration (logBB =−0.8; =−3.0/1.2 for 95% of drugs), compound 4e was predicted to penetrate the brain. This prediction is consistent with its high lipophilicity (logS =12.6; =6.5/0.5 for 95% of drugs). However, the predicted human oral absorption is good (100%). Finally, in silico studies indicate unfavorable hERG (human Ether-a-go-go Related Gene) K+ channel blockade, with a logIC\(_{50}\) (inhibitory concentration 50) of =−9.671 (range 95% of drugs; logIC\(_{50}\) <−5). This in silico approach suggests that 4e has an acceptable ADME and cardiotoxicity profile.

**Conclusion**

Using the CB\(_1\) cannabinoid receptor inverse agonist/antagonist rimonabant (SR141716) as a scaffold, a series of pyrazole carboxamides containing part of the opioid fentanyl structure were prepared (4a–4k). The structural
**Figure 6** Effects of 4d on the cannabinoid tetrad. Bars show modifications induced by treatment with WIN 55,212-2 1.5 mg/kg, 4d 10 mg/kg, rimonabant 1 mg/kg, and WIN 55,212-2 1.5 mg/kg after treatment with rimonabant 1 mg/kg or 4d 2, 4, or 8 mg/kg. °C represents body temperature; Seconds of immobility represents catalepsy on an elevated ring; Cross number represents the number of interruptions of photocell beams.

**Notes:** Values show the mean ± SEM (n=10). *P<0.05; **P<0.01; and ***P<0.001 indicate a statistically significant difference versus vehicle; *P<0.05, **P<0.01, and ***P<0.001 indicate a statistically significant difference versus WIN (one-way analysis of variance, Newman–Keuls post hoc test). WIN55,212-2 (Tocris/Biogen Científica SL, Madrid, Spain).

**Abbreviations:** W, WIN 55,212-2; SR, rimonabant; SEM, standard error of the mean; MPE, maximum possible effect; C, control; V, vehicle.

**Figure 7** Effects of 4e on the cannabinoid tetrad. Bars show modifications induced by treatment with WIN 55,212-2 1.5 mg/kg, 4e 5 mg/kg, rimonabant 1 mg/kg, and WIN 55,212-2 1.5 mg/kg after treatment with rimonabant 1 mg/kg or 4e 5 mg/kg. °C represents body temperature; Seconds of immobility represents catalepsy on an elevated ring; Cross number represents the number of interruptions of photocell beams.

**Notes:** Values show the mean ± SEM (n=10). *P<0.05; **P<0.01; and ***P<0.001 indicate a statistically significant difference versus vehicle; *P<0.05, **P<0.01, and ***P<0.001 indicate a statistically significant difference versus WIN (one-way analysis of variance, Newman–Keuls post hoc test). Rimonabant (Sanofi, Paris, France).

**Abbreviations:** W, WIN 55,212-2; SR, rimonabant; SEM, standard error of the mean; MPE, maximum possible effect.
variations were focused on the linker separating the two pharmacophores, ie, aliphatic chains, cycloalkanes, and aromatic rings. Competitive binding assays and $^{35}$S-GTP$\gamma$S functionality tests were carried out in post-mortem human prefrontal cortex membrane preparations because receptors for both CB$_1$ and $\mu$ opioids are expressed in this tissue, with the CB$_1$ receptor being expressed in higher proportions. Selected compounds (4b, 4d, and 4e) displayed opioid and cannabinoid antagonism in these assays. Although the new compounds have modest affinity for both CB1 and $\mu$ receptors, an interesting finding is that 4b, 4d, and 4e behaved as CB$_1$ cannabinoid receptor antagonists with a potency similar to that of rimonabant. Subsequently, the in vivo cannabinoid antagonism of 4d and 4e, based on behavioral effects in mice, support the $^{35}$S-GTP$\gamma$S bioassay results. With regard to their opioid properties in vivo, the hot plate test confirmed that 4d and 4e had a $\mu$ opioid antagonist profile. In an alcohol relapse model in Wistar rats, treatment with 4e was not significantly effective in modulating relapse-like behavior. Although these new compounds were designed as analogs of the opioid agonist fentanyl, the fact that they behave as opioid antagonists can be accounted for by subtle alteration of ligand structures that may lead to differences in affinity or intrinsic activity. However, these alterations occur more commonly with a morphine-like structure than with a fentanyl structure. While this paper was being prepared, Le Naour et al$^{36}$ published on hybrid compounds based on rimonabant and morphine molecules. Unlike our case, the reported structural modification of morphine did not modify its opioid agonist nature. The in silico ADME predictions for compound 4e suggest that it is likely to penetrate the blood–brain barrier, and this is consistent with the in vivo data. In conclusion, this is the first description, to our knowledge, of hybrid compounds with cannabinoid and opioid antagonist properties in vitro and/or in vivo. It has been suggested in the literature that coadministration of cannabinoid antagonists and $\mu$ opioid antagonists could offer therapeutic advantages.$^{27–29}$ Opioid-cannabinoid interactions in the regulation of appetite are of particular interest. Very recently, Wright and Rodgers$^{30}$ reported a study showing that the pruritic effect of rimonabant can be attenuated by the opioid receptor antagonist naloxone. In this context, the promising dual activity of the cannabinoid/opioid compounds presented here offers an attractive starting point for future therapeutic applications.

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Disclosure
The authors report no conflicts of interest in this work.

References


Supplementary material
Synthesis procedures
and characterization of new compounds

General
Melting points were determined with a Reichert Jung Thermovar apparatus (Reichert Optische Werke, Vienna, Austria). Mass spectra (MS) were recorded using electrospray positive mode. Elemental analysis was performed on a Heraeus CHN-O rapid analyzer (Foss Heraeus GmbH, Hanau, Germany). Analyses indicated by symbols of the elements or functions were within ±0.4% of theoretical values. Analytical high-performance liquid chromatography (HPLC) was run on a Waters 6000 with a Delta Pak C 18.5 µm (Waters, Cerdanyola del Vallès, Spain), 300 Å, 3.9×150 mm column, using CH3CN/H2O 95:5 (0.05% trifluoroacetic acid [TFA]) as eluent; flow rate 1 mL per minute; 254 nm. Nuclear magnetic resonance (NMR, 1H, 13C) spectra were recorded on Varian 300 (Agilent, Barcelona, Spain) and 400 unity spectrometers. All chemical shifts are reported in ppm. s: singlet; t: triplet; m: multiplet; br: broad triplet; dp: double quadruplet; brm: broad multiplet; q: quadruplet; brt: broad triplet; dpd: double quadruplet; brd: broad doublet; brs: broad signal.

5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1
propyl)-1H-pyrazole-3-carboxamide (4a)
5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxyl chloride (2) (60 mg, 0.15 mmol), TEA (triethylamine) (26 µL, 0.19 mmol) and N-(3-aminopropyl)-N-(1-phenethylpiperidin-4-yl)propionamide (3a) (82 mg, 0.11 mmol), affording 4a as a yellow oil (44%): melting point 69°C–71°C; 1H NMR (399.93 MHz, CDCl3, δ 7.42–7.05 (m, 12H), 4.51 (m, 1H), 3.43 (m, 2H), 3.23 (m, 2H), 2.89 (m, 2H), 2.78 (m, 2H), 2.61 (m, 2H), 2.37 (s, 3H), 2.32 (m, 2H), 2.10 (m, 2H), 1.92 (m, 2H), 1.64 (m, 2H), 1.12 (t, 3H, J=7.3 Hz); 13C NMR (99.98 MHz, CDCl3) δ 173.9, 162.7, 145.0, 143.0, 139.9, 135.9, 135.8, 134.9, 132.9, 130.7, 130.5, 130.2, 129.4, 128.8, 128.6, 128.4, 127.8, 126.1, 117.5, 60.3, 55.3, 53.2, 51.5, 41.1, 39.4, 36.9, 33.8, 30.1, 26.9, 26.8, 9.7, 9.4; MS (ES+) m/z (%): 696 (100) [M+H]+; HPLC 99% purity; anal. C37H42Cl3N5O2 (C, H, N, O).

5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-N-(5-(1-phenethyl piperidin-4-yl)propionamido) pentyl)-1H-pyrazole-3-carboxamide (4c)
5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carbonyl chloride (2) (49 mg, 0.12 mmol), TEA (21 µL, 0.15 mmol) and N-(5-(aminopentyl))-N-(1-phenethylpiperidin-4-yl)propionamide (3c) (52 mg, 0.15 mmol), affording 4c as a yellow solid (73%): 1H NMR (399.93 MHz, CDCl3) δ 7.40–6.92 (m, 12H), 5.80 (m, 0.4H), 4.55 (m, 0.3H), 3.83 (m, 0.6H), 3.70 (m, 0.7H), 3.44 (m, 2H), 3.10 (m, 2H), 2.74 (m, 2H), 2.56 (m, 2H), 2.35 (s, 3H), 2.30 (m, 2H), 2.07 (m, 2H), 1.80 (m, 2H), 1.65–1.30 (m, 8H), 1.12 (m, 3H); 13C NMR (99.98 MHz, CDCl3) δ 173.8, 173.0, 162.7, 144.9, 138.8, 136.0, 134.9, 133.0, 130.8, 130.6, 130.5, 128.9, 128.6, 128.4, 127.9, 126.1, 117.7, 60.4, 55.3, 53.3, 53.0, 43.3, 42.6, 38.7, 33.6, 30.7, 31.3, 29.5, 29.2, 9.8, 9.6, 9.4; MS (ES+) m/z (%): 710 (100) [M+H]+; HPLC 99% purity; anal. C38H44Cl3N5O2 (C, H, N, O).

5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-N-(6-(1-phenethyl piperidin-4-yl)propionamido) hexyl)-1H-pyrazole-3-carboxamide (4d)
5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carbonyl chloride (2) (60 mg, 0.15 mmol), TEA (26 µL, 0.19 mmol) and N-(6-aminohexyl)-N-(1-phenethylpiperidin-4-yl)propionamide (3d) (68 mg, 0.19 mmol), affording 4d as a white solid (54%): melting point 69°C–71°C; 1H NMR (399.93 MHz, CDCl3) (mixture of rotamers) δ 7.43–6.92 (m, 12H), 4.46 (m, 0.6H), 3.55 (m, 0.4H), 3.41 (m, 2H), 3.20–3.04 (m, 4H), 2.79 (m, 2H), 2.59 (m, 2H), 2.37 (s, 3H), 2.32 (m, 2H), 2.06 (brt, 2H,
5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-N-(7-(1-phenethylpiperidin-4-yl)propionamido) heptyl-1H-pyrazole-3-carboxamide (4e)

5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxamide (4f)

5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-N-(8-(1-phenethylpiperidin-4-yl)propionamido) octyl-1H-pyrazole-3-carboxamide (4g)

5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-N-(3-(1-phenethylpiperidin-4-yl)propionamido) phenyl-1H-pyrazole-3-carboxamide (4i)
(15 µL, 0.11 mmol) and N-(3-aminophenyl)-N-(1-phenethylpiperidin-4-yl)propionamide (3i) (35 mg, 0.09 mmol), affording 4i as a yellow solid (48%): melting point 105°C–108°C; 1H NMR (399.93 MHz, CDCl3) δ 7.53–6.73 (m, 16H), 4.58 (m, 1H), 3.91 (brs, 2H, J=10.5 Hz), 2.61 (m, 2H), 2.43 (m, 2H), 2.33 (s, 3H), 2.07 (brt, 2H, J=11.5 Hz), 1.91 (m, 2H), 1.75 (brt, 2H, J=14.0 Hz), 1.39 (m, 2H), 0.94 (t, 3H, J=7.5 Hz); 13C NMR (99.98 MHz, CDCl3) δ 173.4, 160.5, 144.5, 143.6, 139.5, 138.9, 136.2, 135.7, 135.1, 133.0, 132.9, 130.8, 130.5, 130.4, 129.6, 129.0, 128.6, 128.3, 127.9, 126.8, 126.0, 125.8, 121.4, 119.2, 118.3, 60.4, 53.1, 52.2, 33.8, 30.6, 28.5, 9.6, 9.5; MS (ES+) m/z (%): 716 (100) [M+H]+; HPLC 99% purity; anal. C39H38Cl3N5O2.3H2O (C, H, N, O).

5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-N-(3-(N-(1-phenethylpiperidin-4-yl)propionamido)benzyl)-1H-pyrazole-3-carboxamide (4j)
5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carbonyl chloride (2) (60 mg, 0.15 mmol), TEA (26 µL, 0.19 mmol) and N-(3-(aminomethyl)benzyl)-N-(1-phenethylpiperidin-4-yl)propionamide (3j) (60 mg, 0.15 mmol), affording 4j as a yellow solid (60%): melting point 83°C–85°C; 1H NMR (399.93 MHz, CDCl3) (mixture of rotamers) δ 7.40–7.06 (m, 16H), 3.84 (m, 1H), 3.47 (m, 1H), 3.05 (m, 2H), 2.57 (m, 2H), 2.53 (m, 2H), 2.31 (s, 3H), 2.27 (m, 2H), 2.02 (m, 2H), 1.86 (m, 2H), 1.63 (m, 2H), 1.99–1.31 (m, 20H), 1.19 (m, 3H); 13C NMR (99.98 MHz, CDCl3) δ 172.8, 161.9, 145.2, 142.9, 140.0, 136.0, 135.9, 134.8, 132.9, 130.8, 130.6, 130.3, 128.8, 128.6, 128.4, 127.8, 126.1, 117.7, 60.5, 56.0, 53.3, 48.5, 45.5, 37.5, 34.4, 33.1, 32.1, 32.0, 29.6, 29.3, 33.8, 28.6, 24.5, 9.6, 9.4; MS (ES+) m/z (%): 816 (100) [M+H]+; HPLC 99% purity; anal. C41H42Cl2N5O2.0.5H2O (C, H, N, O).