Can the anti-inflammatory activities of β2-agonists be harnessed in the clinical setting?

Abstract: Beta2-adrenoreceptor agonists (β2-agonists) are primarily bronchodilators, targeting airway smooth muscle and providing critical symptomatic relief in conditions such as bronchial asthma and chronic obstructive pulmonary disease. These agents also possess broad-spectrum, secondary, anti-inflammatory properties. These are mediated largely, though not exclusively, via interactions with adenylyl cyclase-coupled β2-adrenoreceptors on a range of immune and inflammatory cells involved in the immunopathogenesis of acute and chronic inflammatory disorders of the airways. The clinical relevance of the anti-inflammatory actions of β2-agonists, although often effective in the experimental setting, remains contentious. The primary objectives of the current review are: firstly, to assess the mechanisms, both molecular and cell-associated, that may limit the anti-inflammatory efficacy of β2-agonists; secondly, to evaluate pharmacological strategies, several of which are recent and innovative, that may overcome these limitations. These are preceded by a consideration of the various types of β2-agonists, their clinical applications, and spectrum of anti-inflammatory activities, particularly those involving adenosine 3′,5′-cyclic adenosine monophosphate-activated protein kinase-mediated clearance of cytosolic calcium, and altered gene expression in immune and inflammatory cells.

Keywords: adenylyl cyclase, corticosteroids, cyclic AMP, muscarinic receptor antagonists, neutrophils, phosphodiesterase inhibitors

Introduction

Beta2-adrenergic agonists (β2-agonists) are widely used in clinical practice to treat patients with obstructive airway disorders, such as asthma, chronic obstructive pulmonary disease (COPD) and bronchiolitis obliterans. These agents relax airway smooth muscle, resulting in bronchodilatation, via interaction with G-protein-coupled β2-adrenoreceptors (β2ARs), linked to adenylyl cyclase. The consequence is elevation of intracellular cyclic adenosine monophosphate (cAMP) concentrations and activation of protein kinase A (PKA). In addition to their primary bronchodilatory effects, β2-agonists have been shown to attenuate the proinflammatory activities of a range of immune and inflammatory cells in vitro, such as neutrophils, monocytes, mast cells, eosinophils, basophils, and lymphocytes, all of which contribute to the pathogenesis of various acute and chronic respiratory diseases. In addition, these agents have demonstrated efficacy in animal models of experimental acute lung injury. Clearly, the combination of bronchodilatory and anti-inflammatory activities is of considerable potential value in the pharmacotherapy of acute and chronic diseases of the airways, of both infective and noninfective origin. Disappointingly, however, β2-agonists do not appear to possess significant anti-inflammatory activity in the clinical setting.
The current review is focused on the cellular targets and mechanisms of anti-inflammatory activity of β2-agonists, as well as on strategies, both current and future, that might enable these to be actualized in the clinical setting. This is preceded by a brief consideration of the current clinical applications and types of β2-agonists.

Types of β2-agonists

These agents are characterized according to their duration of action, the three categories being: short-acting beta-agonist (SABA), long-acting beta agonist (LABA) and ultra-LABA. Some commonly used examples of these are shown in Table 1.5–7,9–11 Together with their types of agonist activity, partition coefficients, and durations of action. The number of β2ARs per cell on various immune and inflammatory cells, together with their dissociation constants, is summarized in Table 2.12–19 In the case of LABAs, formoterol has a more rapid onset of action than salmeterol,5 while both agents provide sustained bronchodilatation for at least 12 hours.20 Although indacaterol is the only example shown of an ultra-LABA, several other such agents (abediterol, carmoterol, milveterol, olodaterol, vilanterol) are in the pipeline,5 while another, vilanterol, has recently received US Food and Drug Administration approval for therapy of COPD.

β2-adrenoceptor agonists and therapy of respiratory airway disorders

SABAs are commonly used as rescue bronchodilator therapy to provide symptomatic relief for patients with exacerbations of asthma or COPD. Longer-term control of airway inflammation in asthma is usually achieved using inhaled corticosteroids (ICS). Significantly, LABAs in combination with ICS, currently play an important role in the management of chronic persistent asthma.21 Both types of β2-agonists, as well as the more recently introduced ultra-LABAs, are generally considered to have good safety profiles, and these, as well as the cost-effectiveness of bronchodilator therapies, have been covered extensively in a recent review.1

There has been some concern that LABAs may mask ongoing airway inflammation in asthma and, accordingly, these agents should not be used as monotherapy in this condition.22 Beta-agonists are not recommended as monotherapy in asthma, as these agents may increase airway hyper-responsiveness, and increase the risk of death in patients with chronic asthma.23,24 In contradistinction to asthma, long-term use of LABAs as monotherapy in patients with COPD appears to be safe and effective.23 Importantly, however, the combination of low-dose ICS with a LABA has been shown to be more effective than high doses of ICS.25 This is supported by the findings of a study which demonstrated that the control of airway inflammation, determined by means of bronchial biopsies, was not compromised when asthmatic patients on high doses of ICS were switched to a combination of low-dose ICS and LABA.26 This is consistent with the well-documented anti-inflammatory interactions of combinations of LABAs and ICS, as discussed later in this review. Current guidelines for asthma therapy incorporate ICS/LABA combinations for patients not controlled on low doses of ICS.21,22

The role of LABAs in the management of COPD has been confirmed in a number of large randomized controlled trials, demonstrating improved pulmonary function tests, reduced frequency of exacerbations, and apparent slowing of the rate of decline of forced respiratory volume in 1 second.28 Patients with symptomatic COPD may be given long-acting anticholinergics or LABAs as first-line therapy. In addition, some studies have shown LABAs to attenuate airway inflammation in COPD, and to decrease systemic markers of inflammation.29

<table>
<thead>
<tr>
<th>Cell type</th>
<th>Number of receptors/ cell (Bmax)</th>
<th>Dissociation constant (Kd) (pM)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Neutrophils12</td>
<td>20878±2470 (females)</td>
<td>7331±3179 (males)</td>
</tr>
<tr>
<td>Macrophages12</td>
<td>5643±942</td>
<td>29±9</td>
</tr>
<tr>
<td>Monocytes14</td>
<td>1000–3000</td>
<td>38±29</td>
</tr>
<tr>
<td>Mast cells15</td>
<td>Variable btw preparations</td>
<td>40±10</td>
</tr>
<tr>
<td>Epithelial cells16</td>
<td>9908±1127 (Calu-3 cells)</td>
<td>6423±895 (16HBE14o(-) cells)</td>
</tr>
<tr>
<td>Eosinophils17</td>
<td>4333±318</td>
<td>25.3±1.4</td>
</tr>
<tr>
<td>T-cells18,19</td>
<td>776±183</td>
<td>19.7±1.6</td>
</tr>
</tbody>
</table>

Note: The results are expressed as the mean ± standard error of the mean or *standard deviation.

Abbreviation: btw, between.
Large clinical trials have indicated that LABAs are safe in COPD patients, as mentioned above. However, in patients with COPD, the use of ICS may be associated with an increased risk of pneumonia, which should be considered when using ICS/LABA combinations in this setting.

Bronchiolitis obliterans is a chronic fibrotic disorder of small airways and bronchioles which results in progressive and usually fixed airflow obstruction. Adjunctive therapy with a combination of a LABA and ICS has been shown to provide symptomatic relief and significant improvement in airflow obstruction, in patients with this condition.

Acute lung injury (ALI) is a common and serious disorder, characterized by alveolar flooding, neutrophil accumulation in the lungs, and markedly impaired gaseous exchange. Beta2-agonists have been reported to enhance fluid clearance from alveoli, to attenuate both neutrophil recruitment and proinflammatory activity, and to decrease endothelial permeability in experimental ALI. Consequently, β2-agonists were considered potentially useful in ALI, a contention which was supported by data from limited human trials. However, recent, larger, randomized controlled trials have not shown improved outcomes for patients with ALI, treated with aerosolized or intravenous albuterol/salbutamol.

Antibenflammatory activities of β2-agonists

Of the various types of immune and inflammatory cells involved in the immunopathogenesis of bronchial asthma and COPD, all have been reported to express β2ARs. Seemingly, neutrophils have the highest level of expression, albeit at lower density than airway smooth muscle cells (Table 2). Beta2-agonists have been reported to suppress the proinflammatory activities of all of these cell types (monocytes/macrophages, basophils, eosinophils, mast cells, neutrophils, T lymphocytes), as well as those of airway structural cells (epithelial cells, fibroblasts, smooth muscle cells) by several distinct mechanisms. These are broadly categorized as being either cAMP-dependent or cAMP-independent, with LABAs (especially formoterol) being most effective, while little is known about the anti-inflammatory potential of ultra-LABAs. The effects of β2-agonists on various immune and inflammatory cells and their mediators are summarized in Table 3.

| Table 3 The effects of β2-agonists on various immune and inflammatory cells |
|-----------------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| Neutrophils                 | ↓O2, ↑elastase release | ↓LTB4, ↑CR3, ↓IL-8, ↓chemotaxis | ↑Ca2+ clearance from cytosol | ↓Adhesion to bronchial epithelial cells | ↓Bacterial permeability-increasing protein | ↑Membrane-stabilizing effects |
| Mast cells                  | ↓Histamine release | ↓IL-5, TNF-α, GM-CSF, MIP-1α | ↓LTC4, LTD4, and PGD2 | | | |
| Epithelial cells            | ↓GM-CSF, VEGF | ↓IL-8, RANTES, eotaxin, IP-10 | ↓ICAM-1, VCAM-1 | | | |
| Eosinophils                 | ↓H2O2 | ↓Eosinophil cationic protein | ↓Eosinophil peroxidase release | ↓LTC4 | ↓Chemotaxis | ↓Adherence to fibronectin-coated plates | ↓Eosinophil-derived neurotoxin |
| Basophils                   | ↓Histamine release | ↓IL-4, IL-13 | | | | |
| T-cells                     | ↓Proliferation in response to anti-CD3 Ab | ↓IL-2, IL-3, IL-4, IL-5, IL-13 | ↓IFN-γ, GM-CSF, TNF-α | ↑IL-10 | | |

**Abbreviations:** O2, superoxide anion; iNOS, inducible nitric oxide synthase; ERK, extracellular-regulated kinase; LTB4, leukotriene B4; PGE2, prostaglandin E2; TXB2, thromboxane B2; GM-CSF, granulocyte-macrophage colony-stimulating factor; TNF-α, tumor necrosis factor alpha; IL-1β, interleukin-1 beta; IL-6, interleukin-6; MIP-1α, macrophage inflammatory protein 1 alpha; LTD4, leukotriene D4; PGD2, prostaglandin D2; VEGF, vascular endothelial growth factor; RANTES, regulated upon activation normal T cell expressed and secreted; ICAM-1, intercellular adhesion molecule 1; VCAM-1, vascular cell adhesion protein 1; anti-CD3 Ab, anti-CD3 monoclonal antibody; IFN-γ, interferon gamma; CR3, complement receptor 3; LTC4, leukotriene C4; IP-10, interferon γ inducible protein.

**cAMP-dependent anti-inflammatory mechanisms**

Intracellular cAMP has two major targets, PKA and guanine nucleotide exchange protein directly activated by cAMP-1 (Epac1), both of which regulate the proinflammatory
activities of various cell types, with the former mechanism being the best characterized. The major cellular targets of PKA are Ca\(^{2+}\) mobilization and clearance mechanisms,\(^{40-50}\) and regulation of the expression of genes encoding anti-inflammatory/proinflammatory proteins at both the transcriptional and translational levels.\(^{51-74}\) Epac1 also appears to modulate gene expression.\(^{75-77}\)

### Regulation of Cytosolic Ca\(^{2+}\) Fluxes by cAMP/PKA

Several distinct, albeit interactive, mechanisms have been described, by which cAMP/PKA downregulates the Ca\(^{2+}\)-dependent proinflammatory activities of immune and inflammatory cells, as well as structural cells. These mechanisms, which are best characterized in activated human neutrophils, are regulated by various types of cAMP-elevating agents, including β2-agonists, agonists of subtype A2A adenosine receptors, and inhibitors of cAMP phosphodiesterases (PDEs), especially PDE4.\(^{40-43}\)

In the case of neutrophils, receptor-mediated stimulation of these cells is accompanied by activation of phospholipase Cβ3 (PLC), which, in turn, cleaves membrane phosphatidylinositol to generate inositol triphosphate (IP3). This second messenger interacts with Ca\(^{2+}\)-mobilizing IP3 receptors on intracellular Ca\(^{2+}\) stores (calciosomes, endoplasmic reticulum), resulting in an abrupt, transient increase in cytosolic Ca\(^{2+}\). This is a critical event, which precedes, and is a prerequisite for, activation of most of the proinflammatory activities of these cells, including: i) generation of both reactive oxygen species and the eicosanoid, leukotriene B4 (LTB\(_4\)); ii) mobilization of cytosolic granules; and iii) β2-integrin-mediated firm adhesion to vascular endothelium.\(^{42}\)

To counter neutrophil hyper-reactivity, these events are stringently regulated by several cAMP/PKA-dependent mechanisms, which interact to restore Ca\(^{2+}\) homeostasis. These are: i) phosphorylative inactivation of PLC by PKA;\(^{44}\) ii) upregulation of the activity of the Ca\(^{2+}\)-resequestering endomembrane Ca\(^{2+}\) ATPase (adenosine triphosphatase; probably via phosphorylation of serine (Ser)16 on the phospholamban regulatory subunit);\(^{41}\) iii) inactivation of the IP3 receptor;\(^{45}\) iv) inhibition of influx of extracellular Ca\(^{2+}\), via regulation of store-operated Ca\(^{2+}\) channels;\(^{46}\) and v) phosphorylative inactivation of p38 mitogen-activated protein kinase (MAPK), resulting in failure of activation of 5′-lipoxygenase (5-LO), with resultant attenuation of an autocrine, LTB\(_4\)-mediated secondary wave of Ca\(^{2+}\) influx;\(^{47}\) in addition, PKA-mediated phosphorylation of 5-LO inhibits the nuclear import of this enzyme.\(^{48}\)

In this setting of restoration of Ca\(^{2+}\) homeostasis to activated neutrophils, the autocrine interaction of adenosine with G-protein/adenyl cyclase-coupled A2A receptors appears to be the primary mechanism for generation of cAMP,\(^{49,50}\) which is complemented by β2-agonists and PDE4 inhibitors.\(^{41}\)

### PKA Modulation of Inflammatory/Anti-Inflammatory Gene Expression

Cyclic AMP/PKA-dependent, gene-targeted, anti-inflammatory mechanisms are also operative at several levels, including: i) antagonism of transcription factors;\(^{51,52}\) ii) induction of synthesis of the anti-inflammatory cytokine, interleukin (IL)-10;\(^{53-57}\) and iii) potentiation of glucocorticoid (GC)/glucocorticoid receptor (GR)-activated gene transcription, which contributes to the beneficial therapeutic interactions of LABAs with ICS.\(^{58-74}\)

### Antagonism of Transcription Factors by PKA

Activation of PKA results in phosphorylation of the transcription factor, cAMP response binding protein (CREB), which, in turn, competes with proinflammatory transcription factors, such as nuclear factor kappa B (NF-κB), and activator protein-1, for limited binding sites on the transcriptional coactivator, with intrinsic histone acetyl transferase activity and CREB-binding protein. This mechanism, which has been described in epithelial cells and macrophages, results in decreased expression of genes encoding a range of proinflammatory proteins.\(^{51,52}\)

### Modulation of IL-10 Gene Expression by PKA

The promoter region of the IL-10 gene contains a cAMP response element. Gene transcription results, in turn, from PKA-mediated phosphorylation of CREB, a known transcription factor for IL-10 promoter activation. This anti-inflammatory mechanism is operative in several cell types, such as dendritic cells and macrophages,\(^{53-56}\) and may be particularly important in controlling neutrophilic inflammation, as these cells respond to, but do not produce, IL-10.\(^{57}\)

### Potentiation of Glucocorticoid Receptor-Activated Gene Transcription by cAMP/PKA

Cyclic AMP/PKA-activating agents, particularly LABAs, have well documented enhancing actions, both direct and indirect, on the GR/glucocorticoid response element
(GRE) dependent transactivation of genes encoding anti-inflammatory proteins. These effects are complex, and are achieved primarily via positive, site-specific phosphorylation of serine residues situated in the regulatory N-terminal region of the GR, either by PKA directly, or indirectly, via PKA-mediated phosphorylation of other kinases, such as cyclin-dependent kinases.59,60 These events have been described in a variety of cell types, including airway epithelial cells, smooth muscle cells, fibroblasts, and monocytes/macrophages, as well as hippocampal progenitor cells. They result in the following: i) increased stability and nuclear translocation of the GR, both ligand-dependent and ligand-independent; and ii) increased transcriptional efficiency, via enhanced GRE binding of the activated GR, apparently via recruitment of transcription-promoting coregulatory proteins.58-69 The consequence is activation of genes encoding a series of anti-inflammatory proteins. One of these, MAPK phosphatase-1, also promotes stabilization of the GR via dephosphorylative inactivation of p38 MAPK and c-jun-N-terminal kinase (JNK), both of which phosphorylate the GR at position Ser226, inhibiting translocation and GRE binding.69 Inactivation of these kinases by MAPK phosphatase-1 also attenuates their ability to activate several proinflammatory transcription factors. These GR-targeted activities of LABAs are complemented by several other cAMP/PKA-dependent mechanisms, including: i) increased expression of functional GRs by stabilization of GR messenger (m)RNA; ii) phosphorylative inhibition of phosphatidylinositol-3 kinase (PI3 K)/protein kinase B (Akt) signaling, which prevents inactivation of histone deacetylase 2, increasing the efficiency of GR-mediated repression of genes encoding proinflammatory proteins;70,71 and iii) interference with LTB4 production, thereby antagonizing the antiapoptotic/proinflammatory effects of GR activation on neutrophils and monocytes, which are mediated via upregulation of expression of the LTB4 receptor 1, BTL1.72

As well as underpinning the improved efficacy of ICS in patients receiving combination therapy with LABAs, these various cAMP/PKA-mediated mechanisms may also attenuate the development of corticosteroid (CS) resistance.26,58,69,71,73,74

**Epac1**

Cyclic AMP has also been reported to suppress the proinflammatory activities of several cell types, including macrophages, microglia, and epithelial cells, by a PKA-independent mechanism. In this setting, the alternative target of cAMP is Epac1, the substrate of which is the Ras-like, small guanosine triphosphatase, Rap1.73 Notwithstanding effects on the cytoskeleton,74 Epac1, acting via Rap1, modulates the proinflammatory activities of NF-κB and glycogen synthase kinase-3 β, resulting in decreased synthesis of tumor necrosis factor by target cells.77

These various mechanisms of cAMP-mediated anti-inflammatory activity are summarized in Table 4.

**Cyclic AMP/PKA-independent mechanisms of β2-agonist-mediated anti-inflammatory activity**

Although the anti-inflammatory mechanisms of β2-agonists are mediated predominantly by cAMP/PKA-dependent mechanisms, the existence of alternative mechanisms has been described in various cell types, including bronchial epithelial cells and macrophages.3,76-80 Perhaps the most significant of these was described in a recent study, in which formoterol was found to cause β2AR-independent activation of the serine/threonine phosphatase, PP2A.80 Activation of PP2A, in turn, reversed JNK-mediated phosphorylation of the GR at Ser226, thereby increasing CS sensitivity.

**Table 4 cAMP-mediated mechanisms of anti-inflammatory activity**

<table>
<thead>
<tr>
<th>Target</th>
<th>Effect</th>
</tr>
</thead>
<tbody>
<tr>
<td>Protein kinase A-mediated</td>
<td>Downregulation of Ca2+-dependent proinflammatory activities60-68</td>
</tr>
<tr>
<td>• Regulation of Ca2+-mobilization and cytosolic clearance in activated immune and inflammatory cells</td>
<td>Decreased expression of genes encoding proinflammatory proteins51,52</td>
</tr>
<tr>
<td>• Antagonism of proinflammatory transcription factors (NFκB, AP-1, others)</td>
<td>Activation of synthesis of IL-1053-54</td>
</tr>
<tr>
<td>• Interaction of CREB with the promoter region of the IL-10 gene</td>
<td>Activation of synthesis of other anti-inflammatory proteins55-57</td>
</tr>
<tr>
<td>• Potentiation of GR/GRE-mediated gene transcription (trans-activation)</td>
<td>Decreased production of proinflammatory leukotrienes, and antagonism of glucocorticoid-mediated anti-apoptotic effects on neutrophils40,48,72</td>
</tr>
<tr>
<td>• Inhibition of S′-lipoxygenase</td>
<td>Decreased production of TNF75,77</td>
</tr>
</tbody>
</table>

**Epac1-mediated**

- Antagonism of NFκB and glycogen synthase kinase

Abbreviations: cAMP, cyclic adenosine monophosphate; NFκB, nuclear factor kappa B; AP-1, activator protein-1; CREB, cAMP response element-binding protein; IL-10, interleukin 10; GR, glucocorticoid receptor; GRE, glucocorticoid response element; TNF, tumor necrosis factor; Epac1, guanine nucleotide exchange protein directly activated by cAMP-1.
Mechanisms which restrict the anti-inflammatory efficacy of β2-agonists

Beta2-agonists, especially LABAs, are ineffective, possibly harmful, when used as monotherapy in bronchial asthma, administered intravenously, or used in aerosols in patients with ALI.\(^{26,73,81}\) This seems surprising, given the broad range of cellular targets and inflammatory mediators which are negatively regulated by cAMP.

Notwithstanding differences in molecular structure, agonist activity, and duration of action, the following mechanisms, operative in airway smooth cells, are also likely to undermine the anti-inflammatory efficacy of β2-agonists: i) homologous receptor desensitization;\(^{26,73,81,82}\) and ii) induction of cAMP-specific PDE4.\(^{82,73,83}\) These mechanisms are compounded by the lower numbers of β2ARs on immune and inflammatory cells, relative to those present on airway smooth muscle.\(^{38}\)

Receptor desensitization

Homologous receptor desensitization and downregulation of receptor expression, following extended use of β2-agonists, are considered to be major limitations of these agents, with respect to anti-inflammatory activity.\(^{2,26,73,81}\) In this setting, prolonged activation of the β2AR results in sequential uncoupling of the receptor from adenyl cyclase, internalization and phosphorylation by βAR kinases, PKA and G-protein-coupled receptor kinases, with resultant desensitization.\(^{73}\) Extended exposure to β2-agonists also results in decreased β2AR gene transcription and receptor expression.\(^{73}\) These mechanisms, compounded by the relatively low numbers of β2ARs on immune and inflammatory cells,\(^{38}\) are likely to restrict not only the anti-inflammatory potential of β2-agonists, but also that of endogenous catecholamines.

In addition, prolonged exposure to β2-agonists has also been reported to cause heterologous receptor desensitization (cross-desensitization) in airway smooth muscle cells, via the aforementioned mechanisms.\(^{82,84}\) Although it is speculative, heterologous receptor desensitization, if operative in vivo, may attenuate other types of endogenous, receptor-mediated, cAMP-dependent, anti-inflammatory mechanisms, such as those involving activation of adenosine A2A receptors.\(^{85}\)

Induction of cAMP-specific PDE4

Prolonged exposure of airway smooth muscle cells has also been reported to result in increased transcription of the gene encoding PDE4D5, the predominant isoform present in these cells, by mechanisms involving PKA and extracellular-regulated kinase 1/2.\(^{82,84,86}\) The consequence is markedly decreased levels of intracellular cAMP and loss of responsiveness to β2-agonists, which can be prevented by pretreatment of the cells with selective and nonselective inhibitors of PDE4.\(^{82,84,86}\)

β2-adrenoreceptors on immune and inflammatory cells

Because they express lower numbers of β2ARs, immune and inflammatory cells are likely to be particularly prone to loss of sensitivity to β2-agonists by the aforementioned mechanisms. The probable consequence is attenuation of the cAMP-dependent anti-inflammatory mechanisms described above.

Pharmacological strategies which potentiate the anti-inflammatory activities of β2-agonists

Several pharmacological strategies, some well-established and others currently in the preclinical and clinical phases of evaluation, have been described that may counter β2AR desensitization and increased activity of PDEs.

Combining LABAs with CS

As described above, combining LABAs with CS into a single inhaler device has proven to be a major innovation in the therapy of bronchial asthma.\(^{26,73}\) The magnitude of improvement in asthma control resulting from a LABA/ICS combination is superior to that achieved with high doses of ICS alone.\(^{87}\) Similarly, for patients with COPD, the clinical efficacy of a LABA/ICS combination is significantly greater than with either agent used as monotherapy.\(^{88}\) Commonly used LABA/ICS combinations, together with their times of onset of action, efficacies, and adverse effects are shown in Table 5.

It is generally assumed that the primary role of the LABA is to augment the anti-inflammatory actions of the CS, by the mechanisms described in the preceding sections.\(^{83}\) However, CS can also augment the anti-inflammatory actions of LABAs, consistent with synergistic interactions between the two components. These mechanisms include CS-mediated increases in: i) the numbers of β2ARs, by potentiating receptor gene transcription; and ii) the efficacy of coupling between the β2AR and its Gs-protein subunit. The consequence is enhanced and sustained cellular responses to β2-agonists.\(^{73}\)
Table 5 Inhaled bronchodilator therapies that may be used alone or in combination for the management of chronic stable COPD

<table>
<thead>
<tr>
<th>Class of agent</th>
<th>Onset of action</th>
<th>Therapeutic efficacy</th>
<th>Adverse effects</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>LABA</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Formoterol</td>
<td>10 minutes</td>
<td>↑ Lung functions</td>
<td>Tremor</td>
</tr>
<tr>
<td>Salmeterol</td>
<td>3 hours</td>
<td>↑ Quality of life</td>
<td>Headache</td>
</tr>
<tr>
<td></td>
<td></td>
<td>↓ Symptoms</td>
<td>Cough</td>
</tr>
<tr>
<td><strong>LABA + ICS</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Formoterol/budesonide</td>
<td>As above for LABAs</td>
<td>↑ Lung functions</td>
<td>Tremor</td>
</tr>
<tr>
<td>Salmeterol/fluticasone</td>
<td>As above for LABAs</td>
<td>↑ Quality of life</td>
<td>Headache</td>
</tr>
<tr>
<td></td>
<td></td>
<td>↓ Symptoms</td>
<td>Cough</td>
</tr>
<tr>
<td><strong>LAMA</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tiotropium</td>
<td>3–4 hours</td>
<td>↑ Lung functions</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>↑ Quality of life</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>↓ Symptoms</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>↓ Exacerbations</td>
<td></td>
</tr>
<tr>
<td><strong>LABA + LAMA</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Formoterol + tiotropium</td>
<td>As above</td>
<td>As above</td>
<td>As above</td>
</tr>
<tr>
<td>Salmeterol + tiotropium</td>
<td>As above</td>
<td>As above</td>
<td>As above</td>
</tr>
<tr>
<td><strong>LABA + LAMA + ICS</strong></td>
<td>As above</td>
<td>As above</td>
<td>As above</td>
</tr>
<tr>
<td><strong>Ultra-LABA</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Indacaterol</td>
<td>5 minutes</td>
<td>↑ Lung functions</td>
<td>Cough</td>
</tr>
<tr>
<td></td>
<td></td>
<td>↑ Quality of life</td>
<td>Nasopharyngitis</td>
</tr>
<tr>
<td></td>
<td></td>
<td>↓ Symptoms</td>
<td>Worsening</td>
</tr>
<tr>
<td></td>
<td></td>
<td>↓ Exacerbations</td>
<td>COPD</td>
</tr>
<tr>
<td><strong>Experimental agents</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Various Ultra-LABAs</td>
<td>To be established</td>
<td>See Ultra-LABA</td>
<td>See Ultra-LABA</td>
</tr>
<tr>
<td>New LAMAs</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Glycopyrronium</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GSK-573719</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>BEA-21808R</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aclidinium bromide</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Darotropium bromide</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Notes: *Randomized placebo-controlled trials of duration >24 weeks have been selected, as this duration allows the best assessment of treatment effects. Data from Decramer et al.111  
Abbreviations: COPD, chronic obstructive pulmonary disease; LABA, long-acting beta agonist; ICS, inhaled corticosteroid; LAMA, long-acting muscarinic agent.

Combining LABAs with inhibitors of PDE4

Clearly, pharmacological strategies that target PDEs (the enzymes that inhibit the hydrolysis of cyclic nucleotides) are attractive for potentiating the anti-inflammatory actions of cAMP-elevating agents, including β2-agonists. One such agent, roflumilast, is a selective inhibitor of type 4 PDE, the predominant type in human neutrophils, and which is also present in other types of immune and inflammatory cells. This agent has been approved for clinical application in COPD, while several other such agents are in the pipeline.83,89,90 Interestingly, roflumilast has been reported to enhance the potentiating action of formoterol on CS-mediated gene transcription in human airway epithelial cells in vitro.91 However, drug intolerance due to gastrointestinal and central nervous system side-effects may be limiting, while therapeutic efficacy has also been questioned by some.92 To counter these problems, inhaled formulations of roflumilast, for use in combination with ICS/LABAs, are currently under development, as are other novel PDE4 inhibitors, for both oral and inhaled administration, with apparently improved windows of therapeutic efficacy.89 Moreover, given that PDE4 is not the only isoform present in immune and inflammatory cells (with one or both of PDE3 and PDE7 also being present, both of which hydrolyze cAMP), there is an increasing realization that simultaneous targeting of several PDE isoforms may be a more effective strategy.90

In this respect, it is noteworthy that montelukast, an antagonist of type 1 cysteinyl leukotriene receptors (CysLT1Rs), which is used primarily in the therapy of exercise-induced asthma (and as add-on therapy to ICS/LABAs in severe asthma, as well as in some cases of...
moderate-to-severe COPD), has also been reported to possess secondary, nonspecific PDE-inhibitory activity. Exposure of human neutrophils to montelukast alone, but especially in combination with formoterol, was found to suppress the proinflammatory activities of these cells by a CysLT1R-independent, apparently cAMP-mediated, mechanism. Interestingly, montelukast has recently been reported to prevent salbutamol-mediated β2AR desensitization in airway smooth muscle cells, via direct inhibition of PDE4D5.

Other agents which possess this combination of CysLT1R-antagonism and nonspecific PDE inhibitory activity include ibudilast (also known as KC-404, AV-411 and MN-166) and CR3465. Ibudilast is currently undergoing clinical evaluation as a treatment for chronic neuropathic pain and multiple sclerosis, while CR3465 is under early assessment as a potential therapy for bronchial asthma and interstitial cystitis. Clearly, all three of these agents have the potential to augment the anti-inflammatory activities of LABAs in the clinical setting, with supporting evidence, albeit limited, in the case of montelukast.

Combinations of anticholinergics with β2-agonists

Anticholinergic agents induce bronchodilation by antagonism of muscarinic receptors on smooth muscle cells. The short-acting anti-muscarinic agent, ipratropium bromide provides short-term symptomatic relief for patients with airflow obstruction, while longer-acting muscarinic agents (LAMAs), such as tiotropium, maintain bronchodilation for a sustained period.

A number of new-generation LAMAs are currently being developed, as shown in Table 5. Because LAMAs and LABAs act via separate mechanisms, the combination of these two agents may induce greater bronchodilation than either agent alone. Clinical trials support the safety and efficacy of LABA/LAMA combinations for the management of COPD.

Triple therapy with a LABA/LAMA/ICS combination has been used for patients with severe COPD who do not respond adequately to other therapeutic regimens. There does appear to be some additional benefit to be gained by following this strategy of triple therapy for selected patients.

In addition to its bronchodilating actions, tiotropium also possesses anti-inflammatory activities, albeit by mechanisms that remain to be conclusively established. Given their potential complementary bronchodilatory and anti-inflammatory actions, several recent studies have addressed the therapeutic potential of combinations of tiotropium with either LABAs or ultra-LABAs only, or with ICS/LABAs, in the clinical setting of COPD. Improvements in several clinical and inflammatory indices were observed. Likewise, combining tiotropium as add-on therapy to ICS/LABAs in patients with severe, uncontrolled asthma was found to improve lung function over 24 hours. Although promising, firm recommendations on the clinical utility of combining tiotropium with either LABAs/ultra-LABAs, or as a single bifunctional molecule, with or without CS, in the therapy of COPD or asthma cannot be made on the basis of the limited data currently available. This needs to be reinforced, not only by additional clinical trial data, but also by the acquisition of compelling mechanistic data on the possible additive/synergistic anti-inflammatory interactions of tiotropium with β2-agonists and CS.

Conclusion

LABAs are used in the management of obstructive airways diseases, primarily for their very effective bronchodilator activity. They are an established therapy for COPD, often as single agents, and are considered to be safe and effective in this condition. However, since their introduction, controversy has surrounded the use of LABAs in asthma with concerns that their use as monotherapy is associated with adverse outcomes, including an increase in asthma exacerbations and mortality. Accordingly, they are always used in combination with a conventional anti-inflammatory agent, most commonly ICS. However, the improved control of asthma, together with reduced airway responsiveness to allergen challenge, achieved by combining LABAs with an ICS (as opposed to simply increasing the dose of the ICS) has been interpreted by some as evidence for anti-inflammatory activity of β2-agonists. A better understanding of the mechanisms which both underpin and counteract the anti-inflammatory potential of β2-agonists may facilitate the design of pharmacological strategies to optimize the clinical benefit of these agents.

Disclosure

CF has acted on the Advisory Board and/or received honoraria for lectures and/or received assistance for congress travel from Aspen-GSK, Boehringer Ingelheim, Cipla Medpro, and Novartis. The other authors have no conflicts of interest to declare.

References


87. Pauwels RA, Löfdahl CG, Postma DS, et al. Effect of inhaled for-
meterol and budesonide on exacerbations of asthma. Formoterol and 
Corticosteroids Establishing Therapy (FACET) International Study 
88. Donohue JF. Combination therapy for chronic obstructive pulmo-
275–281.
89. Loukides S, Bartziotakis K, Vebstho J, Singh D. Novel Anti-inflammatory 
agents in COPD: Targeting lung and systemic inflammation. Curr Drug 
90. Page CP, Spina D. Selective PDE inhibitors as novel treatments for 
augment the ability of formoterol to enhance glucocorticoid-dependent 
gene transcription in human airway epithelial cells: A novel mechanism 
for the clinical efficacy of roflumilast in severe COPD. Mol Pharmacol. 
92. Anonymous. Roflumilast: doubtful efficacy but clear harms in COPD. 
newer inhaled bronchodilators: A meta-analysis of randomized con-
94. Anderson R, Theron AJ, Gravett CM, Steel HC, Tintinger GR, 
Feldman C. Montelukast inhibits neutrophil pro-inflammatory 
with inhaled corticosteroids alone or inhaled corticosteroids and 
long-acting beta-2-agonists in the management of patients diagnosed 
with asthma and concurrent allergic rhinitis (the RADAR trial). Can 
Respir J. 2009;16 Suppl A:17A–31A.
Efficacy of add-on montelukast in patients with non-controlled asthma: 
97. Bjerg A, Lundbæk B, Løttvall J. The future of combining inhaled drugs 
98. Calverley PM, Boonsawat W, Cseke Z, Zhong N, Peterson S, 
Olsson H. Maintenance therapy with budesonide and formoterol in 
chronic obstructive pulmonary disease. Eur Respir J. 2003;22(6): 
912–919.
twice-daily salmeterol for COPD: a placebo-controlled comparison. 
100. Szafranski W, Cukier A, Ramirez A, et al. Efficacy and safety of 
budesonide/formoterol in the management of chronic obstructive 
and-as-needed treatment of chronic obstructive pulmonary disease. 
102. Taskhin DP, Rennard SI, Martin P, et al. Efficacy and safety of 
budesonide and formoterol in one pressurized metered-dose inhaler 
in patients with moderate to very severe chronic obstructive pulmonary 
disease: results of a 6-month randomized clinical trial. Drugs. 2008; 
103. Taskhin DP, Doherty DE, Kerwin E, et al. Efficacy and safety of a fixed-
dose combination of mometasone furoate and formoterol fumarate in 
subjects with moderate to very severe COPD: results from a 52-week 
propionate and salmeterol combination delivered via the Diskus device 
in the treatment of chronic obstructive pulmonary disease. Am J Respir 
salmeterol 50 microg bid to anticholinergic treatment in patients with 
COPD: a randomized, placebo controlled trial. Chronic obstructive 
fluticasone in the treatment of chronic obstructive pulmonary disease: 
Health outcomes following treatment for six months with once daily 
tiotropium compared with twice daily salmeterol in patients with 
108. Hanania NA, Darken P, Horstman D, et al. The efficacy and safety of 
fluticasone propionate (250 microg)/salmeterol (50 microg) combined 
834–843.
109. Stockley RA, Chopra N, Rice L. Addition of salmeterol to existing 
treatment in patients with COPD: a 12 month study. Thorax. 2006; 
61(2):122–128.
110. Donohue JF, van Noord JA, Bateman ED, et al. A 6-month, placebo-
controlled study comparing lung function and health status changes 
in COPD patients treated with tiotropium or salmeterol. Chest. 
111. Decramer ML, Hanania NA, Lötvall JO, Yawn BP. The safety of 
long-acting beta2-agonists in the treatment of stable chronic obstructive 
pulmonary disease. Int J Chron Obstruct Pulmon Dis. 2013;8: 
53–64.
112. Vogelmeier C, Hederer B, Glaab T et al. Tiotropium versus salme-
113. Wedzicha JA, Calverley PM, Seemungal TA, et al. The prevention of 
chronic obstructive pulmonary disease exacerbations by salmeterol/ 
fluticasone propionate or tiotropium bromide. Am J Respir Crit Care 
114. Larsson K, Janson C, Lisspers K, et al. Combination of budesonide/
formoterol more effective than fluticasone/salmeterol in preventing 
exacerbations in chronic obstructive pulmonary disease: the PATHOS 
of chronic obstructive pulmonary disease with tiotropium, a once-daily 
inhaled anticholinergic bronchodilator: a randomized trial. Ann Intern 
116. Dusser D, Bravo ML, Iacono P. The effect of tiotropium on exacerba-
tions and airflow in patients with COPD. Eur Respir J. 2006;27(3): 
547–555.
on sputum and serum inflammatory markers and exacerbations in 
118. Chan CK, Maltais F, Sigouin C, Haddon JM, Ford GT; SAFE Study 
Group. A randomized controlled trial to assess the efficacy of tiotro-
pium in Canadian patients with chronic obstructive pulmonary disease. 
119. Casaburi R, Mahler DA, Jones PW. A long-term evaluation of 
one-daily inhaled tiotropium in chronic obstructive pulmonary disease. 
120. Cazzola M, Taskhin DP. Combination of formoterol and tiotro-
pium in the treatment of COPD: effects on lung function. COPD. 
121. van Noord JA, Aumann JL, Janssens E, et al. Comparison of tiotropium 
one day, formoterol twice daily and both combined once daily in 
with and without formoterol on airflow obstruction and resting hyper-
123. Taskhin DP, Pearle J, Iazzoni D, Varghese ST. Formoterol and tiotro-
pium compared with tiotropium alone for COPD. COPD. 2009;6(1): 
17–25.
124. Taskhin DP, Littner M, Andrews CP, Tomlinson L, Rinehart M, 
Denis-Mize K. Concomitant treatment with nebulized formoterol and 
tiotropium in subjects with COPD: a placebo-controlled trial. Respir 


