

Epithelial cell-adhesion molecule-directed trifunctional antibody immunotherapy for symptom management of advanced ovarian cancer

Ramez N Eskander
Krishnansu S Tewari

Division of Gynecologic Oncology,
Department of Obstetrics and
Gynecology, University of California,
Irvine, CA, USA

Abstract: Despite advances in cytotoxic chemotherapy and surgical cytoreduction, disease recurrence continues to be a troubling problem in patients with advanced-stage epithelial ovarian cancer (EOC). Malignant ascites affects approximately 10% of patients with recurrent EOC and is associated with troublesome symptoms, including abdominal pressure, distension, dyspnea, pelvic pain, and bowel/bladder dysfunction. To date, no effective therapy has been identified for the treatment of malignant ascites in patients with recurrent, advanced-stage ovarian cancer. Recently, immune modulation has gained attention as a novel approach to anti-cancer therapy. This review explores the role of epithelial cell-adhesion molecule (EpCAM)-directed immunotherapy, with a specific focus on the mechanism of action of the trifunctional antibody catumaxomab (anti-EpCAM × anti-CD3). In addition, clinical trials exploring the use of catumaxomab in the treatment of malignant ascites in patients with ovarian cancer are reviewed.

Keywords: ovarian cancer, immunotherapy, catumaxomab, CD3, EpCAM

Introduction

Epithelial ovarian cancer (EOC) accounts for 25% of all malignancies affecting the female genital tract, and is the most lethal gynecologic malignancy. In 2013 there will be an estimated 22,240 new ovarian cancer cases in the USA, with 14,030 deaths.¹ Advanced-stage EOC is traditionally managed with cytoreductive surgery, followed by combination platinum- and taxane-based chemotherapy.² Despite aggressive treatment, the majority of these patients develop recurrent cancer, with selection of chemotherapy-resistant clones.³ The subset of patients who develop recurrent disease is a population that traditionally faces extended exposure to multiple cytotoxic chemotherapy regimens, dictated by their disease-free interval.⁴⁻⁷ Throughout this period, management of disease-associated morbidities becomes a priority in an effort to improve quality of life.

Malignant ascites, which affects approximately two-thirds of patients with EOC, primary peritoneal cancer (PPC), and fallopian tube cancer (FTC), is common, and to date few effective therapies have been identified.^{8,9} Importantly, ascites is associated with troublesome symptoms, including abdominal pressure and distension, dyspnea, pelvic pain, and bowel/bladder dysfunction.¹⁰ Historically, malignant ascites in patients with EOC was treated utilizing diuretics and salt restriction, as well as intraperitoneal administration of sclerosing agents and radioactive isotopes.^{11,12} Patients with malignant ascites secondary to EOC rarely suffer from fluid accumulation due to

Correspondence: Krishnansu S Tewari
Division of Gynecologic Oncology,
Department of Obstetrics and
Gynecology, University of California,
Irvine, 101 The City Drive South,
Building 56, Suite 260, Orange,
CA 92868, USA
Fax +1 714 456 6632
Email ktewari@uci.edu

intraparenchymal liver metastasis, and portal hypertension is uncommonly identified. Rather, these patients traditionally exhibit reduced intravascular volume, making diuretic use an unattractive option.¹³ With respect to the use of radioactive isotopes, poor tumor penetration and intestinal toxicity (necrosis and perforation) due to loculations and prolonged exposure have caused them to fall out of favor. Limited success rates, in combination with significant side effects, have resulted in the infrequent use of these modalities.

Mechanical drainage of accumulated ascitic fluid via therapeutic paracentesis results in relief in up to 90% of patients.¹⁴ However, recurrence/reaccumulation of ascites is common, and multiple paracentesis are required, with their associated risks of pain, visceral perforation, infection, and hematoma formation.^{15,16} Furthermore, these patients are likely to have intra-abdominal adhesions as a result of their extensive surgical cytoreduction, and the resultant fluid loculations limit the therapeutic benefit derived.¹⁷ Alternatively, placement of permanent intra-abdominal drains and peritoneovenous shunts has been explored. Experience with these modalities has been poor, as blockage of the shunt, infectious morbidity, as well as embolization and implantation of tumor cells in distant organs, were reported to be relatively common complications.^{12,18–21}

Unlike other solid malignancies, where ascites portends a universally poor prognosis, patients with EOC and ascites at the time of diagnosis can expect 5-year survival rates approaching 40%.^{15,22} This discrepancy is largely attributable to the biology of ovarian cancer, and the subsequent etiology of the abdominal fluid accumulation. Specifically, malignant ascites in patients with EOC is thought to be attributable to (1) lymphatic obstruction, (2) increased vascular permeability, (3) release of inflammatory cytokines, and (4) direct increase of fluid production by the cancer cells lining the peritoneal cavity.^{9,23–25}

Given the above, exploration into immune modulation as a novel anti-cancer approach for the treatment of malignant ascites in patients with advanced-stage EOC has been a clinical priority.⁹ Tumor immunology is complex, as activation of the immune system requires presentation of a foreign antigen via antigen-presenting cells (APCs) to T-cells.²⁶ In normal immune responses, the cytotoxic effects are driven by a combination of T-lymphocyte activity, antibody-dependent mechanisms, and natural killer (NK) cell activation.^{26–28}

Tumor cell destruction by tumor antigen-specific T lymphocytes has been demonstrated *in vitro* for a variety of both solid and hematologic malignancies. Certain cytokines, including interferon- γ and tumor necrosis factor (TNF), are

essential components of the above response. Conversely, antibody-mediated cytotoxicity relies on direct antibody binding to tumor cell surfaces, and the subsequent recruitment of granulocytes and macrophages, both of which contain surface receptors for the fragment crystallizable (Fc) portion of the antibody. Analogously, activated NK cells contain surface receptors for antibody Fc regions, and participate in antibody-mediated cytotoxicity. Furthermore, activated NK cells secrete TNF- α , inducing hemorrhage and tumor necrosis.

Unfortunately, tumor cells have adapted to escape immune destruction, utilizing tumor-related and host-related mechanisms. Cancer cells may fail to provide an antigenic target due to lack of an antigenic (foreign) epitope, lack of major histocompatibility complex-I molecule, or via antigenic modulation and tumor masking.^{29,30} Host-related mechanisms include immune suppression, regulatory T-cell (Treg) suppression of tumor immunity, deficient epitope expression by host APCs, and failure of host immune cells to reach the tumor due to stromal barriers.³¹

Ovarian cancer immunology

An increasingly robust body of literature supports the contribution of immune cells in both ovarian cancer therapeutics and pathogenesis.^{32–42} In 1988, immune studies performed on ovarian cancer specimens confirmed the presence of tumor-infiltrating lymphocytes (TILs).⁴³ Cluster of differentiation 3 (CD3)+ TILs have been found to independently predict tumor recurrence and prolonged survival.⁴² Conversely, lack of TILs has been associated with poor survival.^{34,36–40}

Certain subsets of T-cells exhibit a pro-cancer effect. Treg infiltration has been associated with higher cancer grade and advanced surgical stage.⁴⁴ Additionally, increasing numbers of circulatory Tregs identified in peripheral blood samples have been linked to disease progression.⁴⁵ These CD4+CD25+ Tregs are hypothesized to mitigate the immune response via two distinct mechanisms, inhibiting both cell-cell contact and interleukin-2 transcription.³¹

The impact of B-cell function on cancer immunology has been more difficult to discern as a result of conflicting data.^{46–49}

Epithelial cell-adhesion molecule (EpCAM)

Epithelial cell-adhesion molecule (EpCAM) is a calcium-independent transmembrane, glycoprotein cell-adhesion molecule, with a molecular weight of 39–42 kDa.⁵⁰ Traditionally, it is expressed in normal epithelium, with the

exception of squamous epithelium, epidermal keratinocytes, gastric parietal cells, myoepithelial cells, thymic cortical epithelium, and hepatocytes.^{51,52} EpCAM is abundantly expressed on human cancers, and was first described as a dominant antigen in patients with colon carcinoma.^{53,54} Since then, EpCAM overexpression has been associated with poor prognosis in patients with ovarian, breast, prostate, and gallbladder carcinoma, both functioning as an oncogene and suppressing CD4+ T-cell-dependent immune responses.^{55–58}

Initial interest in the utilization of EpCAM as a target for active immunotherapy emerged following a seminal publication reporting anti-tumor effects with the EpCAM-specific monoclonal antibody edrecolomab in patients with metastatic colorectal cancer.^{58,59} Despite initial promise, the therapeutic impact of this approach was found to be inferior to traditional cytotoxic chemotherapy regimens in colon cancer patients.

With respect to ovarian cancer, EpCAM expression has been demonstrated in the main histotypes, and is well documented in microarray studies.⁶⁰ In a study of 21 biomarkers in four distinct ovarian cancer subtypes (high-grade serous, clear cell, endometrioid, mucinous), only EpCAM exhibited consistent high expression.⁶⁰ Furthermore, data indicate that EpCAM appears to be a stable antigen in ovarian cancer patients, with preserved expression levels in primary, recurrent, and metastatic specimens, as well as malignant ascites and effusions.^{61,62}

The trifunctional antibody catumaxomab (anti-EpCAM × anti-CD3)

In an effort to circumvent the limitations initially encountered with monoclonal antibodies (mAb), bispecific antibodies,

which allow for simultaneous binding of both T-cell and accessory cells, were designed and tested. Ultimately, the design of trifunctional antibodies allowed for combination of two distinct anti-tumor functionalities (T-cell-mediated death and accessory cells).^{27,63,64}

Catumaxomab is a trifunctional mAb with two different antigen-binding sites and a functional Fc domain (Figure 1).^{65,66} It is composed of a mouse κ light chain, a rat λ light chain, a mouse immunoglobulin (Ig)-G2a heavy chain, and a rat IgG2a heavy chain. The two specific antigen-binding sites bind to epithelial tumor cells via the EpCAM and to T-cells via CD3. In addition, catumaxomab activates Fc γ -receptor I-, IIa-, and III-positive accessory cells (dendritic cells [DCs], macrophages, and NK cells) via its functional Fc domain, resulting in a comprehensive and complex immune reaction (Figure 1).^{65,66}

The functionality and selectivity of this novel antibody rely on the fact that tumor cells in malignant ovarian cancer-associated ascites have been shown to express EpCAM in 70%–100% of cases, while the mesothelial cells lining the peritoneal cavity lack expression.⁶⁷ Following EpCAM binding, catumaxomab results in recruitment and activation of immune effector cells, resulting in its antineoplastic activity. The two unique antibody–antigen binding sites of the trifunctional antibody enable recognition of both T-cells and tumor cells. The functional Fc domain can then activate neighboring Fc γ -receptor-positive macrophages, DCs, and NK cells, as described by Eissler et al in elegant animal model experiments.⁶⁸ Ultimately, tumor cell death results from cell lysis via perforin/granzyme-B, antibody-mediated cell death, and phagocytosis.^{66,69,70}

Catumaxomab has also been shown to induce anti-tumor immunity in animal models and in patients with peritoneal carcinomatosis. In a study conducted by Ströhlein and Heiss

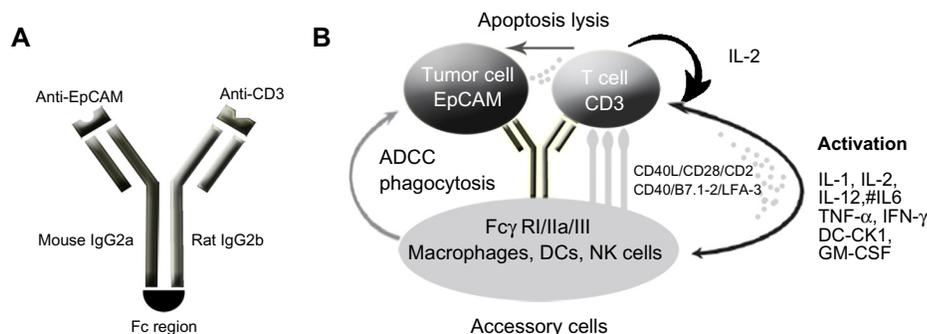


Figure 1 Schematic of the structure and mode of action of catumaxomab. **(A)** Catumaxomab is a trifunctional monoclonal antibody with two different antigen-binding sites and a functional Fc domain. **(B)** The two specific antigen-binding sites bind to epithelial tumor cells via the EpCAM and to T-cells via CD3, while activating Fc γ -receptor I-, IIa-, and III-positive accessory cells (dendritic cells, macrophages, and NK cells) via its functional Fc domain.

Abbreviations: CD3, cluster of differentiation 3; DCs, dendritic cells; EpCAM, epithelial cell-adhesion molecule; Fc, fragment crystallizable; IgG, immunoglobulin; NK, natural killer; IL, interleukin; INF, interferon; GM-CSF, granulocyte macrophage colony stimulating factor; ADCC, antibody dependent cell mediated cytotoxicity; DC-CK1, dendritic cell CK1.

, tumor-reactive CD4+/CD8+ T-cells were induced after an initial treatment course with escalating doses of trifunctional antibodies, followed by re-stimulation 4 weeks after completion of initial treatment.^{71–73}

Catumaxomab in the treatment of ovarian cancer ascites (anti-EpCAM × anti-CD3)

Malignant ascites affects approximately 10% of patients with recurrent EOC and is associated with troublesome symptoms. Intraperitoneal administration of catumaxomab was first studied in the treatment of eight patients (two of whom had ovarian cancer) with malignant ascites in 2005.⁷⁴ All patients had >2% EpCAM expression via flow cytometry, on nuclear ascites cells. Trifunctional antibodies were administered intraperitoneally over 6–8 hours, for at least four cycles. Seven of eight patients required no further paracentesis during follow-up or until death, with a mean paracentesis-free interval of 38 weeks (median 21.5, range 4–136). Clinical response, with disappearance of ascites accumulation, was seen in all patients, which was correlated with elimination of tumor cells ($P = 0.0014$).⁷⁴

Following this study, a multicenter phase I/II clinical trial was conducted evaluating the tolerability and efficacy of intraperitoneal catumaxomab in ovarian cancer patients with malignant ascites containing EpCAM-positive tumor cells.⁷⁵ Twenty-three women with recurrent ascites due to pretreated refractory ovarian cancer were treated with four to five intraperitoneal infusions of catumaxomab in doses of 5,200 µg within 913 days. Treatment with catumaxomab resulted in significant and sustained reduction of ascites. Of the 23 patients, 22 did not require paracentesis between the last infusion and the end of study at day 37.⁷⁵ The most commonly reported grade 2/3 adverse events in the study included fever, nausea, and vomiting.

Recently, a prospective, randomized phase II/III study was conducted comparing the efficacy of catumaxomab plus paracentesis with paracentesis alone in the treatment of malignant ascites.⁶⁷ Following paracentesis, catumaxomab was administered at doses of 10, 20, 50, and 150 µg on days 0, 3, 7, and 10, respectively, via an intraperitoneal catheter. The primary efficacy endpoint was puncture-free survival. Secondary efficacy parameters included time to next paracentesis, ascites signs and symptoms, and overall survival (OS). Puncture-free survival was significantly longer in the catumaxomab group (median 46 days) than in the control group (median 11 days) (hazard ratio [HR] 0.254; $P < 0.0001$), as was median time to next paracentesis

(77 versus 13 days; $P < 0.0001$). Within the ovarian cancer cohort, median puncture-free survival was 52 days in the catumaxomab arm versus 11 days in the placebo arm (HR 0.205; $P < 0.0001$). In addition, catumaxomab-treated patients had fewer signs and symptoms of ascites than control patients. The most common adverse events included fever, abdominal pain, nausea, and vomiting. One patient had a grade 3 gastric hemorrhage. Findings from the above trials ultimately resulted in the European Medicines Agency approval of catumaxomab for the treatment of malignant ascites in patients with EpCAM-positive tumors for whom no standard therapy is available.²³

Immunomonitoring studies performed as part of the clinical trial were notable for a significant decline in EpCAM-positive tumor cells from a median screening value of 6,510 EpCAM-positive cells (165 patients) to a median of 27 cells on day 3 (133 patients), and to 0 cells (115 patients) on day 11 in the catumaxomab-treated arm.⁷⁶ In the control group, the tumor cell number increased from 9,373 EpCAM-positive tumor cells at screening (85 patients) to 18,929 EpCAM-positive tumor cells (74 patients) at the puncture visit. Furthermore, catumaxomab treatment was associated with a significant decline (63%; $P < 0.001$) in ascites fluid levels of vascular endothelial growth factor (VEGF), inhibiting vascular permeability, translating into decreased ascites fluid production. Lastly, CD69+ (indicative of lymphocyte proliferation), CD4+, and CD8+ T-cell populations increased more than 2-fold in catumaxomab-treated subjects. The activation of peritoneal T-cells and concomitant decline in EpCAM-positive tumor cells establishes a cellular basis for the anti-tumor immunologic effects of the trifunctional antibody catumaxomab.⁷⁶

The palliative nature of the treatment of malignant ascites in patients with recurrent ovarian cancer necessitates prioritization of quality of life during treatment. Wimberger et al conducted a post-trial ad hoc analysis of the above-described phase II/III study to determine the impact of catumaxomab on health-related quality of life (HR-QOL).⁷⁷ Deterioration in QOL scores appeared more rapidly in the control than in the catumaxomab group (median 19–26 days versus 47–49 days). The difference in time to deterioration in QOL between the groups was statistically significant for all scores ($P < 0.01$).

The chronicity of disease in patients with recurrent malignant ascites related to ovarian carcinoma led to the exploration of intraperitoneal changes resulting from treatment with catumaxomab. In a small retrospective series, ten patients previously treated with intraperitoneal catumaxomab

underwent repeat surgical exploration for secondary cytoreduction, treatment of anastomotic leaks or ileus, or for colostomy reversal.⁷⁸ Catumaxomab treatment was associated with severe intra-abdominal adhesions, grade 3. Conversely, no patients had ascites volume >500 mL despite extensive carcinomatosis in eight of ten subjects.

Given the mouse–rat origin of catumaxomab, limitations in retreatment were anticipated due to formation of human anti-drug antibodies (HADA). However, in 2011, Pietzner et al described a case of successful re-treatment with catumaxomab for the management of malignant ascites.⁷⁹ A 74-year-old female patient with breast cancer and peritoneal carcinomatosis-associated ascites was treated with catumaxomab, with resolution of her symptoms. The patient remained puncture free for 45 days, and evaluation of HADA levels demonstrated increased levels after cycle 1, followed by a considerable decline and delayed increase in ascites HADA levels for each subsequent cycle. This experience suggested that a repeat cycle of catumaxomab might be feasible and effective in patients suffering from recurrent malignant effusions.

More recently, Ott et al⁸⁰ conducted a post-trial ad hoc analysis in order to determine the impact of human antimouse antibodies (HAMA) levels 8 days after the last infusion on clinical outcomes in patients treated on the European Union Drug Regulating Authorities Clinical Trials (EudraCT) clinical trial (NCT00836654).⁸¹ There was a strong correlation between humoral response and clinical outcome, with HAMA-positive patients showing improvement in median puncture-free survival, median time to next therapeutic puncture, as well as median OS.⁸⁰ Specifically, median OS in HAMA-positive ovarian cancer patients was 163 days, compared with 82 days in the HAMA-negative patients ($P = 0.0123$; HR 0.407). This finding remained significant in both the intention-to-treat and per-protocol analysis.

Conclusion

Despite aggressive surgical cytoreduction and adjuvant chemotherapy, disease recurrence continues to be problematic for patients with advanced-stage EOC. Immune modulation has gained significant attention in recent years as a novel anti-cancer approach. Catumaxomab is an innovative trifunctional antibody that relies on direct tumor cell and T-cell binding while simultaneously recruiting accessory immune cells to treat malignant ascites. This complex association results in tumor cell kill and simultaneously induces a humoral immune response. Currently, additional trials are being conducted to explore the safety of a 3-hour

infusion of catumaxomab in conjunction with steroid pre-medication (Catumaxomab Safety Phase IIIb Study With Intraperitoneal Infusion in Patients with Malignant Ascites Due to Epithelial Cancers [CASIMAS]),⁸² as well as the safety of re-treatment with intraperitoneal catumaxomab (Safety Study of Second Intraperitoneal Infusion Cycle of Catumaxomab in Patients With Malignant Ascites [SECI-MAS]).⁸³ Additionally, phase I/II clinical trials exploring combination treatment using catumaxomab and traditional cytotoxic chemotherapy are required, to determine if there is therapeutic efficacy to combined treatment. The ENGOT-ov8 study,⁸⁴ a multicenter prospective phase II clinical trial, is currently open, and is exploring the feasibility and clinical activity of intraperitoneal catumaxomab followed by systemic intravenous chemotherapy in patients with recurrent ovarian cancer.

Ultimately, as with all novel therapies, symptom relief and treatment goals must be weighed against patient discomfort and adverse events. Careful patient selection, and identification of risk factors, to help reduce significant side effects associated with treatment are required.

Disclosure

The authors have no conflicts of interest to disclose.

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