

Rainfall and temperature variation does not explain arid species diversity in outback Australia

Lochran W Traill¹
Thomas C Wanger^{2,3,4}
Siobhan C de Little¹
Barry W Brook¹

¹Environment Institute and School of Earth and Environmental Sciences, University of Adelaide, Adelaide, SA, Australia; ²Center for Conservation Biology, Department of Biology, Stanford University, CA, USA; ³Ecosystem Functions, Leuphana University Lüneburg, Lüneburg, Germany; ⁴Agroecology, Department of Crop Sciences, University of Göttingen, Göttingen, Germany

Abstract: Steps toward conserving biodiversity should start at understanding the components across spatial scales and a determination of the drivers of these. Here we determine additive species diversity for arid South Australia, based on over 50 years of survey data. Elevation and soil data were sourced through the Australian Government, and climate data from the WorldClim database. Alternative hypotheses relating the effect of climatic and environmental parameters to diversity were tested using generalized linear models and ranked according to information-theoretic statistics. Total species richness for the region was 1824, similar to all arid regions. α -diversity values were low, relative to the contributions made by β -diversity toward total γ -diversity, similar to additive diversity indices for nonarid biomes. There was a lack of statistical support for our hypothesis that regional spatial variation in arid region diversity can be explained by climate topography. Arid South Australian species diversity appears to be largely driven by environmental parameters at the localized scale – beyond the resolution of available survey data. Heterogeneity in habitat, provided by mountainous regions, likely contributes toward the high β -diversity values. Our research is the first application of the additive (not multiplicative) approach toward understanding diversity within arid Australia.

Keywords: additive diversity, arid biome, South Australia, generalized linear models, global climate models, species richness

Introduction

The maintenance of biodiversity is central to the practice of conservation biology.¹ Biodiversity confers system resilience to change and disturbance² – given that different species respond in different ways to environmental pressures – thereby providing a more robust aggregate community.^{3,4} Such ecosystem resilience is pertinent to present-day conservation management as human populations continue to grow⁵ and exert impacts (eg, habitat alteration, introduction of invasive species) that may act in synergy with climatic shifts to alter community composition and structure.^{6,7} In this context, biodiversity-related research will likely move from a focus on understanding spatial-temporal changes in diversity toward improved explanation of the underlying mechanisms of change and the capacity of ecological communities to adapt.

The term “biodiversity” is synonymous with species diversity, or the temporal and spatial heterogeneity of communities,⁸ and has been in common usage since the Rio 1992 International Convention on Biological Diversity. Numerous biotic and abiotic factors can determine such heterogeneity, including rainfall, topography, temperature, aspect, evolutionary history, and anthropogenic activity.^{1,9,10} Naturally, species diversity will vary across broad biomes, as a consequence of both localized

Correspondence: LW Traill
Imperial College London, Silwood Park,
Berkshire SL5 7PY, UK
Tel +44 207 594 2470
Email lochran.traill@gmail.com

and environmental effects,^{1,11} but as a generalization, data indicate a relationship between diversity and productivity, such as soil or climate effects,³ with an asymptotic increase in richness with increased rainfall.¹² This most basic definition of diversity is based on the number of species within a region, but another estimate of diversity is the difference in the number of species between regions, with quite different implications. To elaborate, the most common measure of diversity is species richness, or simply the number of genetically distinct species that occupy a predefined region.¹³ α -diversity accounts for species' relative abundance within a plot or community, and is thereby a measure of average diversity. The magnitude of species turnover or spatial variation between communities is quantified through β -diversity,¹³ and together these contribute toward total, or γ -diversity, within a region.

The factors that drive α -diversity within a region may be different from those factors that drive β -diversity. For example, within central arid Australia, β -diversity tends to be high relative to the more tropical north, but α -diversity is lower.¹⁴ Localized factors such as habitat heterogeneity or human activity can lead to relatively high levels of β -diversity within a region,^{14,15} but mean α -diversity tends to be associated with mean annual rainfall.¹⁴ It is for these reasons that γ -diversity can be useful in determining "total" diversity for a particular region, or as a comparative index of change through time.

Most commonly, a multiplicative approach is used to determine γ -diversity; thus, $\gamma = \alpha \times \beta$, as per Whittaker.¹⁶ The practical disadvantage of this approach is that the diversity components are not weighted equally when partitioned across more than one spatial scale.¹⁷ As an alternative to the multiplicative approach, measures of diversity can be partitioned additively: eg, Veech et al.¹⁸ Thus, the observed γ -diversity (γ_{obs}) can be partitioned as $\gamma_{\text{obs}} = \alpha + \beta_{\text{S}} + \beta_{\text{R}}$, where β_{S} is the between-site β -diversity, and β_{R} is the between-region β -diversity.¹⁹ This provides for estimation of the relative contributions of α - and β -diversity, thereby allowing for comparative spatial studies that are more pertinent to conservation decision making.¹⁹ Further, the additive approach is more straightforward than the multiplicative approach, and as the diversity components are expressed in the same units, the additive approach allows for partitioning on an unlimited number of scales.¹⁹

To date, analyses that have used an additive partition approach for species diversity have been applied within temperate,²⁰ tropical, and subtropical regions,^{21,22} as well as marine systems.²³ However, there is an apparent lack

of additive diversity research within arid and semiarid regions (a search using the ISI Web of Science [<http://www.isiknowledge.com>] showed just five peer-reviewed articles since 1973). In fact, relatively little diversity-related research has been conducted in arid and semiarid regions at all, despite many threats to arid species persistence being identified.^{24,25} What is more, the world's arid regions cover up to 41% of the global ground surface, and support up to 38% of the global human population.²⁶

Moreover, although it is accepted that climate and topography influence species diversity,⁹ and that projected shifts in climatic parameters will shape future distributions in diversity,²⁷ we know of few studies that have used additive diversity indices to test for correlative climatic effects, and no studies for arid regions. Bowman¹⁴ and, more recently, James and Shine⁹ did explore the influence of climate on arid species diversity, but these were based on the multiplicative approach.

Here we apply the additive diversity approach to an extensive arid region likely to experience shifts in temperature and rainfall regimes in the coming decades – namely, outback South Australia (SA). We determine diversity indices based on SA survey data, develop a competing hypothesis framework to tease apart the regional drivers of diversity, including climatic factors, and discuss the conservation implications for arid region diversity under global change.

Material and methods

Data acquisition and processing

Data were restricted to the Alinytjara Wilurara Natural Resource Management (AWNRM) region, SA (Figure 1). This extensive (~250,000 km²) yet sparsely populated region is set aside for conservation and indigenous land use (see <http://www.awnrm.sa.gov.au>).

The study area falls within the arid climatic zone,⁹ or the desert and xeric shrubland ecoregion. Temperatures for our (entire) study region ranged from 3.6°C to 35.4°C (mean 20.8°C), and mean annual rainfall was 18 mm.²⁸

Major vegetation communities, as classified by the National Vegetation Information System (NVIS),²⁹ include Acacia woodlands, Mallee woodlands and shrublands, Eucalypt woodlands, and hummock grasslands. Human communities are widely dispersed, with an estimated total population of ~4000 people (<http://www.awnrm.sa.gov.au>).

All species data were sourced by permission of the South Australian Department of Environment, Water and Natural Resources (<http://www.environment.sa.gov.au>). These were part of a state-wide, ongoing survey of threatened and

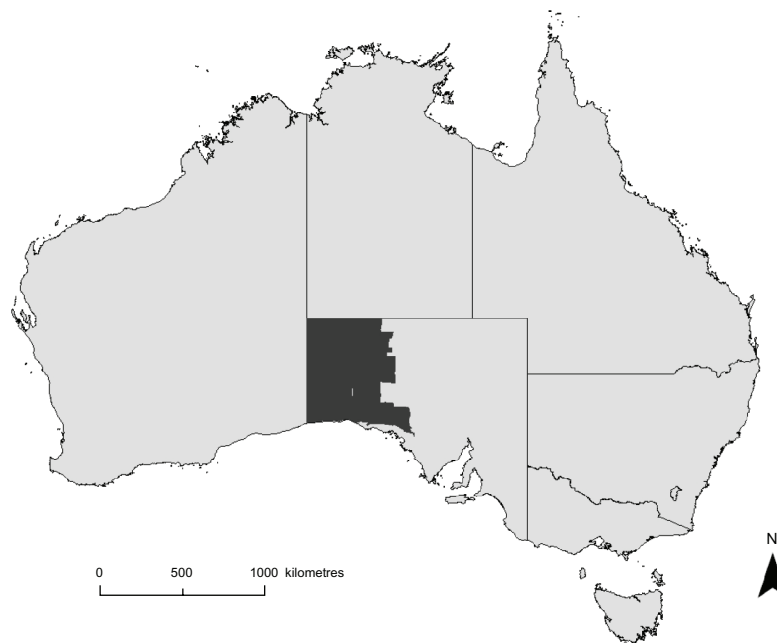


Figure 1 Outline of the Alinytjara Wilurara Natural Resource Management region (black) of South Australia, with the continent of Australia outlined in grey and lines demarcating each state and territory.

nonthreatened species (plants and vertebrates only). Each year, the South Australian Department of Environment, Water and Natural Resources conducts transect- and quadrat-based biological surveys to catalogue information on plant and terrestrial vertebrate species distribution.

Data were first segregated into general taxonomy (class for vertebrates [amphibians, birds, mammals, and reptiles] and then plants), and duplicate or erroneous data were omitted. Abundance data were not available for most survey counts, so only presence data were used. Species location data allowed spatial representation within the Geographic Information System (GIS) software ArcGIS (ArcMap software, v.9.3, ESRI, Redlands, CA, USA).

Before we could derive diversity indices we needed to delineate ecological boundaries. Biological survey data were not collected in a stratified random manner; rather, they were widely distributed (apparently following roads). To account for the broad spatial scale of these data and uneven spatial sampling, we chose major vegetation groups (MVGs) as the ecological communities to analyze, following the NVIS.²⁹ GIS spatial data (shapefiles) for these groups were sourced from the NVIS web link (<http://www.environment.gov.au/erin/nvis>) and processed using ArcGIS software.

Diversity assessment

Species data were first resampled (bootstrapped random samples, with replacement, within each MVG) to mitigate

against the confounding effects of spatial autocorrelation.³⁰ Diversity measures were then calculated following the formulae provided by Clough et al¹⁹ – first α -diversity, then β_s (between-sites, within MVGs) and β_R (between region, or MVG), and from these γ -diversity. Coding for these was implemented in the R v2.12 language (<http://www.r-project.org>) and is available from the authors on request.

Climate and environmental data

Given the global climate model-based forecasts of climate change in the region over the next few decades, including a general warming and drying trend,³¹ we were interested in the effects of temperature and rainfall on arid region diversity. Thus, we sourced climate data (as GIS raster files) through the WorldClim project,²⁸ and collated mean annual temperature and mean annual rainfall for the region. In order to calculate the mean values for each MVG, we used the extract function in ArcMap to allocate temperature and rainfall values to each datapoint (species location data), and averaged these across MVGs.

WorldClim's current climate data account for variance in rainfall and temperature over the last 50 years, so we excluded all species location data that predated 1960. Thus, diversity indices were derived from pooled survey data, ranging over the past five decades.

In addition to the influence of climate on additive diversity, we sourced data for those environmental parameters likely

to determine diversity. These were rainfall (mean annual), temperature (mean annual), elevation, geology or dominant soil type, fire history, and disturbance (in particular, the presence of large ungulates and other ferals).

We did not have spatial data available for fire history, given the vastness of the area and the lack of records for Aboriginal burning programmes (practiced in an informal, traditional manner). Further, we considered the impact of feral vertebrates to be homogenous, given that their ubiquitous distribution³² and abundance data were not available for cattle (*Bos taurus*).

Past research has indicated the importance of elevation and soil type in driving diversity patterns.^{14,33} Therefore, we sourced the Advanced Spaceborne Thermal Emission and Reflection Radiometer Global Digital Elevation Model, with a posting interval of 1 arc-second (<http://asterweb.jpl.nasa.gov/data.asp>). Elevation data were processed in ArcGIS, and we clipped an area appropriate to our study area. We then used species presence data to extract elevation at occupied points in the landscape, and averaged these across each community to derive mean elevation by MVG.

Soil data were extracted as a GIS shapefile from the Australian Natural Resources Data Library (<http://www.daff.gov.au/abares/data>), based on the Northcote³⁴ principal profile classification scheme. We required a soil value across each vegetation group, and so determined dominant soil type (majority of spatial distribution) for each community and allocated data (to MVGs) accordingly.

Correlative models

We used generalized linear models (γ -distribution with log link) using maximum likelihood estimation to fit the models, and information-theoretic, multi-model inference metrics³⁵

were then used to identify the suite of climate and environmental parameters that best explained variation in additive species diversity. Analyses were implemented within the R language, and model weights were assessed using Akaike's Information Criterion, adjusted for finite sample size (AIC_c).

Of note, we were limited to fitting one- or two-term models only, because initial analyses showed our saturated model to be overparameterized (too few data points per estimated parameter, given that we used averaged values for each MVG and therefore had a limited number of independent data points available). Thus, our competing models (hypotheses) consisted of single-term models for each predictor, and the possible additive combinations between climatic and environmental parameters.

Results

Summed species occurrence across NVIS communities showed high values within Mallee woodlands and shrublands, Acacia open woodlands, and Acacia and Chenopod shrublands (Table 1). Proportionally, the NVIS groups Acacia shrublands and Mallee woodlands and shrublands represent the greater part of land within the AWNRM region, at ~46% of land cover. Acacia open woodlands represent just ~9% of total land cover, however, despite having relatively high species representation (Table 1). The summed total of unique species across the entire region was 1824. Note that this value is different from the summed total for values in Table 1 (~7989), as many species occur across multiple communities.

α -diversity was relatively low in Acacia forests and woodlands and high for Acacia open woodlands, Acacia shrublands, Chenopod shrublands, and Samphire shrublands and forblands (Table 1). Summed β -diversity values were

Table 1 Major vegetation groups (MVGs) and species richness for the Alinytjara Wilurara Natural Resource Management region, South Australia, based on the National Vegetation Information System (NVIS; DEWR 2007)

Vegetation type, based on the NVIS	MVG code	Area km ²	Total species	α -diversity
Acacia forests and woodlands	6	5575	444	2.97
Acacia open woodlands	13	26225	928	10.66
Acacia shrublands	16	62300	838	10.39
Casuarina forests and woodlands	8	7050	473	9.26
Chenopod shrub, samphire shrub, and forblands	22	44300	839	10.79
Eucalypt open woodlands	11	32125	726	9.82
Hummock grasslands	20	7275	757	9.67
Inland aquatic	24	50	85	6.31
Mallee woodlands and shrublands	14	69175	1279	9.62
Other shrublands	17	10300	719	6.88
Tussock grasslands	19	23100	772	6.98

Note: Data include NVIS coding (see <http://www.environment.gov.au/erin/nvis>), spatial area (km²), sum of species within each respective community (Alinytjara Wilurara Natural Resource Management region only), and α -diversity values derived at the community scale.

substantially higher than α -diversity (Figures 2 and 3), and contributed greatly to γ -diversity (mean of 97% across all MVGs). Percentage α -diversity contribution toward γ -diversity was low across most of the major NVIS communities (Figure 3), except for Casuarina forests and woodlands (NVIS MVG8) and inland aquatic (MVG24).

Models testing statistically for the influence of climate, elevation, and soil on arid-zone diversity were not supported by the data, with the null expectation (ie, the same average diversity across all MVG types) being selected for all three diversity compartments (Table 2). This may be partly a problem with statistical power, because we had so few data points (climate data were averaged across MVGs, leaving eleven independent points, or 3.5 data points per parameter). Mean annual rainfall did capture some of the variance (% deviance explained [DE]), however, for all diversity indices (Table 2), as did elevation, showing that the structural goodness of fit of the model was adequate.

Mean annual rainfall and mean elevation by MVG were strongly correlated ($R = 0.903$). Of note, Fawcett et al³⁶ showed that there was an increase in annual rainfall (over the last century) in the northern part of SA, where elevation is higher (by as much as 550 m), and this was captured by the WorldClim rainfall data.

Further, we derived evidence ratios (ER; as a measure of comparative support) for the mean annual rainfall and mean annual temperature models. These were calculated by summing the weighted AIC_c values (Table 2) and dividing the sum of these for all rain and temperature models (thus, the ratio for

evidence between models that contained these parameters). Rainfall models showed twice as much support from the data for all three diversity components (rainfall versus temperature ER, α -diversity = 2.2; β -diversity = 2.2; γ -diversity = 2.3).

Discussion

The total species richness for plants and vertebrates across arid and semiarid SA, at 1824 species, is similar to analogous arid regions. For example, Stohlgren et al³³ estimated total species occurrence (plants only) at ~700 in Utah, US, and Waide et al³⁷ list total species richness, again in the arid US, at 535 and 814 (excluding plants), and 1341 and 2261 (including plants). Total species count for plants in arid SA was 1387, and total vertebrates was 437.

Of interest, and to provide a global context, the International Union for Conservation of Nature and Natural Resources (<http://www.iucnredlist.org>) lists the total number of described vertebrate species as 63,161 and total described plants as 307,674. The greater part of these species occur in the tropics, with a general biogeographic trend showing a peak in the tropics, apparently correlated with annual rainfall.³⁸ Thus, Bowman¹⁴ recorded relatively high α -diversity in tropical northern Australia and a decline along the southwards latitudinal gradient toward arid central Australia (but a noted increase in β -diversity). Further, anuran diversity has been found to be relatively low in arid Australia compared with other regions across the continent,³⁹ and songbird species are notably scarce in the outback, compared with more tropical parts of the country.⁴⁰ The latitudinal generalization does not hold for all taxa, however. For example, James and Shine⁹ found that α -diversity values for Australian *Ctenotus* skinks peaked in the arid center, suggesting that the relative homogeneity of arid climatic conditions allowed for greater sympatric speciation.^{9,41}

Our data exclude tropical northern Australia, but we do note that our diversity indices were correlated with annual rainfall (up to 16% of DE, Table 2) and elevation (9% of DE), with a trend of increased β_s -diversity following the latitudinal gradient from south to north (elevation in arid outback Australia increases toward the center). Although we recognize the correlation between rainfall and elevation in our data, ongoing research has indicated the importance of elevation in driving species diversity, particularly β -diversity. For example, Bowman¹⁴ noted a strong correlation with β -diversity and elevation, as did Simmons and Cowling⁴² in South Africa. These authors suggest that the heterogeneity in habitat provided by areas of greater elevation, such as the increased influence of aspect (due to rocky or broken terrain)

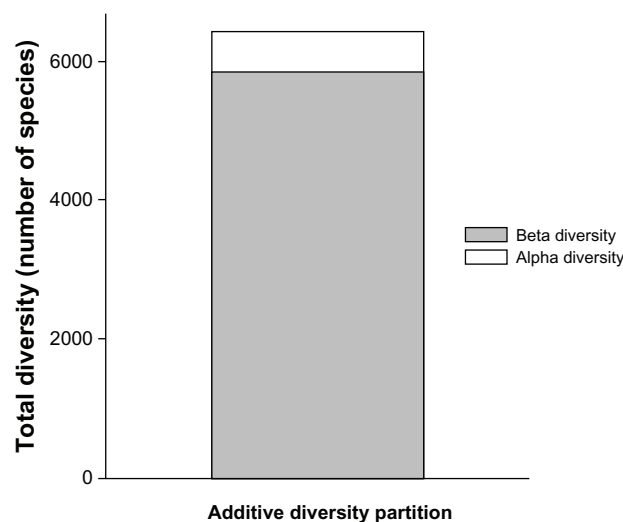


Figure 2 Additive species diversity for the entire Alinytjara Wilurara Natural Resource Management region (all vegetation groups), showing both α - and β -diversity ($\beta_s + \beta_r$) and respective contribution toward γ -diversity, where α is the mean α -diversity per site, β_s is the between-site β -diversity and β_r the mean between-region β -diversity, and the respective contribution of these to total regional diversity, or γ -diversity.

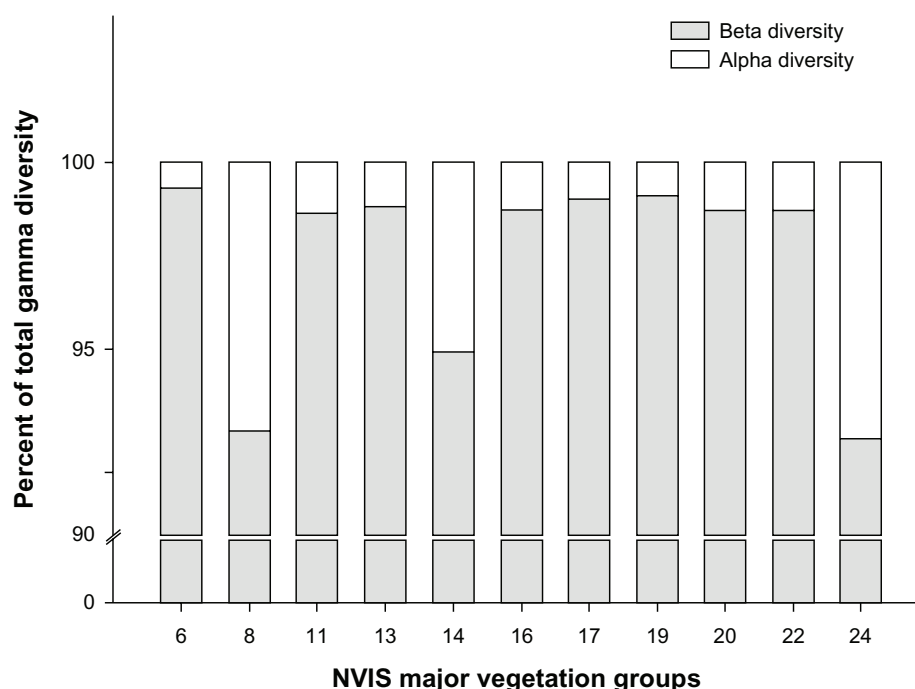


Figure 3 Additive site diversity across vegetation communities in the Alinytjara Wilurara Natural Resource Management region, South Australia. Shown are the percentage contributions made by α - and β -diversity toward γ -diversity. Note the break from 10% to 80% on the Y-axis, done to provide a clearer illustration of additive differences between communities: where α is the mean α -diversity per site, β_s is the between-site β -diversity and β_R the mean between-region β -diversity, and the respective contribution of these to total regional diversity, or γ -diversity.

Abbreviation: NVIS, National Vegetation Information System.

and altitude, provide for resource partitioning and thus greater diversity of species between sites.

Such findings are particularly relevant to biodiversity conservation under shifting climates. For example, recent research in the Swiss Alps⁴³ has shown that variation in micro-habitat temperature (driven by micro-topography and aspect) allows for greater diversity in (plant) species richness, and that a warmer world will affect only the species confined to the coldest micro-habitats, with the majority of plants finding suitable micro-habitats within just a few meters. We speculate that topographic variability in arid central Australia might provide biodiversity refuge to climate change, relative to more low-lying, flat arid areas.⁴³

That said, we did not predict diversity shifts based on climate change projections, as our null (mean-field) models were consistently ranked as the best model. Projections from global climate models forecast an increase in temperatures for the region ($\sim 5^\circ\text{C}$) and a decrease in annual rainfall by as much as 25%³¹ by 2070. This implies a possible decrease in diversity, given the positive (albeit weak) correlation with annual rainfall.

This leads to a further interesting finding of our study: ie, the relatively high contribution of β -diversity (both β_s and β_R) toward γ -diversity for the region (Figures 2 and 3), and

thus the usefulness of the additive approach. This pattern appears to be typical of arid regions, both within Australia and globally. For example, a meta-analysis of (mammal) diversity across global desert regions found a marked pattern of both low α - and high β -diversity,⁴⁴ and Bowman¹⁴ found the same disparity for Australian woody plant species (using the multiplicative approach). Similar findings have also been shown for dissimilar biomes – among Caribbean reef fishes,²³ temperate plants from northern Germany,^{19,20} and subtropical Chinese bird communities.²²

High β -diversity within arid Australia could be attributed to micro-habitat variation (as determined by elevation). Other factors could include spatial heterogeneity in soil type, although our data indicate otherwise (4% of DE, Table 2), or human activity.

Recent human impacts in arid Australia, notably pastoral activity, introduced species, and shifts in fire regimes, may have contributed toward habitat patchiness and thus high β -diversity. Pastoral activity can, through disturbance, lead to enhanced β_s -diversity.⁴⁵ Conversely, some studies have specifically tested the impacts of grazing on β -diversity in arid regions (including Australia) and have found no or little effect.^{10,46} Although pastoral activity did occur on some properties in the southern parts of the AWNRM region, cattle

Table 2 Outcome of generalized linear models (γ -distribution with log link) and information-theoretic model inference metrics, where species diversity is the response, and possible predictor variables are mean annual rainfall, mean annual temperature (averaged across vegetation groups), mean elevation, and dominant soil types (within each vegetation group)

	k	AIC _c	Δ AIC _c	w _i	%DE
(A) α-diversity as response and competing predictive variables					
Null	1	55.14	0.00	0.405	0.0
Mean annual rainfall	2	56.75	1.61	0.181	12.0
Elevation	2	57.53	2.39	0.123	5.8
Mean annual temperature	2	58.13	3.00	0.091	0.5
Soil type	2	58.19	3.06	0.088	0.0
Mean rainfall and soil	3	59.16	4.02	0.054	23.0
Mean rainfall and elevation	3	60.43	5.30	0.029	14.0
Mean temperature	3	61.46	6.32	0.017	5.8
and elevation					
Mean temperature and soil	3	62.04	6.90	0.013	0.7
(B) β_s-diversity as response and competing predictive variables					
Null	1	161.43	0.00	0.359	0.0
Mean annual rainfall	2	162.49	1.06	0.211	15.0
Elevation	2	163.36	1.93	0.137	9.1
Soil type	2	163.95	2.52	0.102	4.4
Mean annual temperature	2	164.17	2.74	0.091	2.6
Mean rainfall and elevation	3	166.15	4.71	0.034	17.0
Mean rainfall and soil	3	166.31	4.88	0.031	16.0
Mean temperature	3	167.24	5.81	0.020	9.4
and elevation					
Mean temperature and soil	3	167.82	6.39	0.015	4.9
(C) γ-diversity as response and competing predictive variables					
Null	1	161.45	0.00	0.355	0.0
Mean annual rainfall	2	162.45	1.00	0.215	16.0
Elevation	2	163.34	1.90	0.137	9.3
Soil type	2	163.96	2.51	0.101	4.5
Mean annual temperature	2	164.18	2.73	0.090	2.7
Mean rainfall and elevation	3	166.09	4.64	0.035	18.0
Mean rainfall and soil	3	166.26	4.81	0.032	17.0
Mean temperature	3	167.22	5.77	0.020	9.7
and elevation					
Mean temperature and soil	3	167.83	6.38	0.015	5.0

Notes: Data are for (A) α -diversity, (B) β_s -diversity, and (C) γ -diversity. Shown are the number of fitted model parameters (k; includes intercept), Akaike's corrected information criterion (AIC_c), difference from best model (Δ AIC_c), Akaike weight scaled relative to a total sum of 1 (w_i), and percentage deviance explained (%DE). %DE is a measure of the structural goodness of fit of the model.

are now mostly restricted to parts of the north of the AWNRM region, particularly the Anangu Pitjantjatjara Yankunytjatjara lands (see <http://www.awnrm.sa.gov.au>). We did not have the cattle abundance data required to test for a density effect on species diversity.

Recent experimental plot-based studies suggest that fire has no substantial effect on β -diversity, particularly where underlying drivers (of diversity) such as gradient remained unchanged.⁴⁷ Conversely, other research has shown that fire

can drive plant species turnover, but typically where the species are either fire tolerant or fire dependent.⁴⁸ We lack information on fire history for the study region, but speculate that the relatively low fuel loads in arid regions do not support frequent hot fires.

Although our findings provide insight into the relative contributions made by diversity indices to communities, the correlative models failed to provide adequate explanation of the underlying mechanisms of diversity distribution in arid SA. We concede that the spatial scale of available data (broad and nonrandom, thus requiring determination of diversity across MVGs) may have contributed toward a lack of evidence for our models (power to resolve effects is low because we could fit only eleven data points, see Burnham and Anderson³⁵), and thus advocate the site-specific collation of parameters likely to determine diversity, including local rainfall and temperature patterns, fire history, aspect, and cattle densities. Indeed, the biological surveys run through the relevant Government authorities require reconsideration, and new transects should be developed using a stratified random approach. Systematic collation of data, along south-north transects, for example, will provide the data needed to better address concerns regarding outcomes under global change.

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Disclosure

The authors report no conflicts of interest in this work.

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