Histologically proven epithelial ingrowth in failed Descemet stripping automated endothelial keratoplasty (DSAEK) managed by repeat DSAEK

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Purpose: To report a case of corneal graft failure due to epithelial ingrowth after an uneventful combined Descemet stripping automated endothelial keratoplasty (DSAEK) and phacoemulsification cataract surgery with intraocular lens implant treated successfully with a repeat DSAEK.

Methods: A 77-year-old male patient underwent combined DSAEK and phacoemulsification with intraocular lens implantation for Fuchs’ endothelial dystrophy plus cataract in the right eye. The donor cornea was cut on the Moria ALTK system and introduced using a suture pull-through technique. After an episode of endothelial rejection, the graft failed, with signs suggesting epithelial ingrowth. It was stripped from the host cornea using a Descemet’s membrane stripper, and a Simesce irrigation-aspiration cannula was used to remove all traces of interface material. The excised lenticule was examined histologically using a hematoxylin and eosin stain.

Result: The patient regained and maintained excellent visual acuity with no sign of recurrence of epithelial ingrowth. Histopathological evaluation of the donor tissue of the first graft showed epithelial ingrowth on the stromal surface of the graft and very few endothelial cells, in keeping with the diagnosis of graft failure.

Conclusion: Epithelial ingrowth is a possible cause of endothelial graft failure, but histologically proven cases are rare. Surgical intervention can achieve successful clearance, with the potential for cure and an excellent outcome.

Keywords: epithelial ingrowth, Descemet stripping automated endothelial keratoplasty, graft failure

Introduction

Endothelial transplantation has overtaken penetrating keratoplasty (PK) in popularity for the treatment of endothelial dysfunction.¹ The technique was originally described by Melles et al,² who named it “posterior lamellar keratoplasty,” and later modified by Terry and Ousley³ and renamed “deep lamellar endothelial keratoplasty.” The technique of stripping Descemet’s membrane (descemetorhexis) was again described by Melles et al⁴ and termed “Descemet’s stripping endothelial keratoplasty.” Further modification of the procedure used an automated blade microkeratome to create a lamellar dissection of the donor cornea, as described by Gorovoy,⁵ and was termed “Descemet stripping automated endothelial keratoplasty” (DSAEK). This technique is now widely used by corneal surgeons for treatment of a variety of corneal disorders characterized by compromised endothelial function.

Epithelial ingrowth is a rare but well-documented complication of anterior segment surgery or ocular trauma, and has been documented after corneal graft surgery.⁶–¹⁰
Corneal graft failure attributed to histologically proven epithelial ingrowth or downgrowth after DSAEK has very rarely been reported in the literature.\textsuperscript{11–18} We report a case of histologically proven epithelial ingrowth at the interface of the graft and host, leading to graft failure after uneventful DSAEK, treated successfully with stripping and careful aspiration of interface material, followed by repeat DSAEK.

**Case report**

A 77-year-old male with Fuchs’ endothelial dystrophy presented with decreased vision in the right eye. Best-corrected visual acuity (BCVA) was 20/30. He had a past history of phacoemulsification cataract surgery with intraocular lens (IOL) implantation in the left eye. Following this he had developed corneal edema for which he had undergone DSAEK with an excellent outcome (BCVA 20/25). Combined DSAEK with phacoemulsification and IOL insertion was therefore planned for the right eye.

After routine phacoemulsification and implantation of a posterior chamber IOL under a cohesive viscoelastic (Microvisc Plus\textsuperscript{TM} [sodium hyaluronate 1.4%], Bohus BioTech, Strömstad, Sweden), a descemetorhexis was performed. The donor lenticule was cut using the Moria ALTK system (Moria, Antony, France) and an 8.5 mm Barron corneal punch (Katena Products, Inc, NJ, USA). A slight irregularity and lip at one side of the donor lenticule was noted. The donor lenticule was introduced through an incision enlarged to 5 mm, using a 10-O-polypropylene double-armed suture and a pull-through technique. The wound was closed with interrupted 10/0 nylon sutures, all viscoelastic was carefully removed, and the donor lenticule tamponaded in position with an air bubble, which was reduced before leaving theater, to avoid pupil block. No venting incisions were made. The immediate postoperative period was uneventful. His BCVA improved to 20/30 at 5 months after the operation. Ten months after the procedure he had an episode of endothelial rejection, which was treated with intensive topical corticosteroid eye drops. Though this settled, a white linear interface opacity was noted just within the superior margin of the DSAEK lenticule immediately after the rejection episode (Figure 1A–C). Slit-lamp examination failed to find any relation to the region of irregular lenticule, as noted preimplantation, and the area of white opacity. This gradually enlarged over the ensuing 12 months with increasing upper corneal edema, and BCVA dropped to 20/200. Intraocular pressure remained normal. Unfortunately, an endothelial cell count attempt after the rejection episode was unsuccessful.

A diagnosis of graft failure secondary to possible epithelial ingrowth was made, and the patient underwent a repeat DSAEK procedure, using a tissue-matched donor lenticule. The original surgical wound was carefully re-entered, the failed donor lenticule was stripped using a Descemet’s membrane stripper, and the interface meticulously cleared of any foreign material using the Descemet’s membrane stripper followed by thorough aspiration using a Simcoe manual irrigation/aspiration cannula. The replacement lenticule was introduced using the suture pull-through technique and the operation concluded as previously described.

Postoperatively, BCVA improved to 20/25 and there was no further rejection. At 18 months postoperatively there was no sign of recurrence (Figure 1D). BCVA remained at 20/25 and was possibly limited by mild dry age-related macular degeneration.

Histolopathological examination of the explanted failed donor lenticule showed a thin layer of epithelium overlying the stroma. The anterior portion of the stroma in this area also showed a portion of thickened, warty Descemet’s membrane. The appearances suggested that there had been entrapment of host Descemet membrane at the graft host interface and ingrowth of epithelium at this point. Lack of endothelial cells on the donor lenticule was consistent with graft failure (Figure 2A–D).

**Discussion**

Epithelial proliferation in the anterior segment of the eye is a rare but serious complication of any ocular trauma or surgery.
Although many authors use the term “downgrowth” to describe this phenomenon, others, including the authors, prefer the term “ingrowth”. Whichever term is used, in cases involving corneal transplantation the origin of the ectopic epithelium may be conjectured to be from either the host or the donor. Epithelial migration into the anterior chamber post DSAEK has rarely been reported. Our results of a literature search on epithelial downgrowth, epithelial interface implantation, or epithelial ingrowth after DSAEK are given in Table 1. Not all such cases lead to graft failure. To the best of our knowledge, there are only four reported cases of histologically proven epithelial ingrowth at the interface after DSAEK leading to graft failure, only two of which were successfully treated by a repeat DSAEK, with the other two cases treated by PK and posterior mushroom keratoplasty, respectively. These cases were different from ours as they were complicated by graft dislocation or attempted suturing and rebubble, which increase the risk of epithelial cells being dislodged into the anterior chamber or at the interface of the graft and host. In our case, the postoperative period was entirely uneventful, but suspicion falls on the slightly irregular cutting of the donor lenticule as the source of epithelial implantation, which contributed to the eventual failure of the graft.

Reviewing the other reported cases, Suh et al, in a series of 118 DSAEK eyes, found one case of presumed epithelial implantation in the interface documented clinically and by anterior segment optical coherence tomography. The graft was clear and no histological evidence was documented. In another case series, the same authors described five further cases of epithelial ingrowth after DSAEK, but none of their cases developed graft failure. Culbertson documented a case that the author thought was of epithelial downgrowth, but no histological evidence was provided. Walker et al described a case as epithelial downgrowth after DSAEK with multiple white opacities at the interface. Confocal microscopy showed the cells to be epithelial and at the interface. Though the patient did not progress to graft failure, that case was treated with a PK.

Prasher et al reported two cases of epithelial ingrowth after DSAEK. In the first, the ingrowth was only on the endothelial surface of the donor cornea and was treated with a repeat DSAEK. In the second, interface epithelial ingrowth was histologically confirmed as causing graft failure in a patient treated with PK. Phillips et al reported a case of two failed DSAEKs where histology of the removed failed graft showed conjunctival epithelial cells over the surgical margin and on to the posterior surface. It was not clear whether the downgrowth of epithelium was the cause or the result of the repeat graft failure. Gorovoy and Ratanasit documented one case of epithelial ingrowth that was not at the interface in a patient who had DSAEK. They treated the patient with a repeat DSAEK. Saelens et al documented epithelial lined cysts at the interface after DSAEK, which was of donor origin, as revealed by X-Y karyotyping, which they treated with a penetrating posterior mushroom keratoplasty. Lee et al, in a retrospective histopathologic study of eight corneas in seven patients who had DSAEK graft failure, found one case of epithelial ingrowth at the interface. They documented that this case had donor graft dislocation during the first DSAEK procedure, which failed to reattach despite repeat rebubble and attempted transcorneal suturing of the graft. Bansal et al reported a case of intracorneal epithelial ingrowth after Descemet stripping endothelial keratoplasty with stromal puncture for phakic bullous keratopathy, which they treated conservatively.

We used tissue-matched graft only for the repeat DSAEK procedure and not for the original graft. The benefit of human leukocyte antigen (HLA) matching in keratoplasty is controversial and repeatedly questioned. There are certain studies that support the role of HLA matching by suggesting that it has a role in extending the graft survival, although...
there are other studies that question its benefit, especially as there can be a considerable delay in obtaining such a matched graft. Although the Collaborative Corneal Transplantation Studies (CCTS) Research Group and the Corneal Transplant Follow-up Study (CTFS) failed to show a beneficial effect of HLA matching, there have been questions raised as to the result of these studies, due to the aggressive immunosuppressant regimen used, which might alter the beneficial effect of HLA matching. Others, like Reinhard et al and Ross et al, have shown, respectively, that in patients with pseudophakic bullous keratopathy and in normal-risk patients there is a beneficial effect of an HLA-matched graft.

Given the controversy around HLA matching, and because of the fact that we were not planning to use aggressive systemic immunosuppressant therapy postoperatively, we opted for a tissue-matched graft in the hope of a better chance of graft survival in this case of repeat DSAEK.

Our case was unique because graft failure occurred after histologically proven epithelial ingrowth in an otherwise uneventful DSAEK procedure. One may argue that the combination of cataract extraction and IOL implantation along with the DSAEK increases the amount of surgical manipulation and provides a portal of entry for host epithelial cells to enter the anterior chamber. In our suture pull-through method we pass the straight needle of the polypropylene suture from the stroma/Descemet side and come out of the endothelium of the donor lenticule. If there is an eccentric punch of the donor graft, there can be a possibility that the remaining epithelial cells on the donor lenticule may be embedded in suture track in the graft during the process of suture passage. This may increase the risk of epithelial ingrowth. However, if such a case of eccentric trephination occurs, we do not pass the needle through that area of the donor lenticule, in order to avoid this possibility.

In our case, both the donor and the recipient were male patients, and hence X-Y karyotyping of the implanted epithelium was not possible to determine whether it was of donor or host origin as documented in certain cases. However, the lack of continuity between the host corneal DSAEK wound and the epithelial interface possibly argues against invasion by the host epithelium. The DSAEK wound was closed with interrupted 10-0 nylon sutures, and there was no evidence of wound leak or tissue incarceration, both of which are considered risk factors for epithelial ingrowth.

We did not use any venting incision, which can sometimes lead to epithelial cells of the host origin being implanted in the interface. We made a side port incision at 11 o’clock,

### Table 1 Literature review of epithelial migration into anterior chamber after DSAEK

<table>
<thead>
<tr>
<th>Study</th>
<th>Number of eye(s)</th>
<th>Description</th>
<th>Diagnosis</th>
<th>Graft failure</th>
<th>Treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Suh et al*</td>
<td>1</td>
<td>Epithelial growth at the interface</td>
<td>Clinically anterior segment optical coherence tomography</td>
<td>No</td>
<td>Nil</td>
</tr>
<tr>
<td>Culbertson*</td>
<td>1</td>
<td>Epithelial downgrowth</td>
<td>Confocal microscopy</td>
<td>No</td>
<td>PK</td>
</tr>
<tr>
<td>Koenig and Covert*</td>
<td>1</td>
<td>Epithelial ingrowth-interface</td>
<td>Histology</td>
<td>Yes</td>
<td>Repeat DSAEK</td>
</tr>
<tr>
<td>Walker et al</td>
<td>1</td>
<td>Epithelial downgrowth-at the interface</td>
<td>Confocal microscopy, histology</td>
<td>No</td>
<td>PK</td>
</tr>
<tr>
<td>Prasher et al*</td>
<td>2</td>
<td>Case 1-epithelial downgrowth-interface</td>
<td>Histology</td>
<td>Yes</td>
<td>Case 1 had PK</td>
</tr>
<tr>
<td>Phillips et al</td>
<td>1</td>
<td>Conjunctival epithelial downgrowth-over donor endothelium</td>
<td>Histology</td>
<td>Yes</td>
<td>Repeat DSAEK</td>
</tr>
<tr>
<td>Gorovoy and Ratanasit</td>
<td>1</td>
<td>Epithelial downgrowth-not at the interface</td>
<td>Histology</td>
<td>Yes</td>
<td>Repeat DSAEK</td>
</tr>
<tr>
<td>Saelens et al</td>
<td>1</td>
<td>Epithelial ingrowth in the flap-graft interface</td>
<td>Histology</td>
<td>Yes</td>
<td>Posterior mushroom keratoplasty</td>
</tr>
<tr>
<td>Lee et al</td>
<td>1</td>
<td>Epithelial ingrowth at the interface</td>
<td>Histology</td>
<td>Yes</td>
<td>Repeat DSAEK</td>
</tr>
<tr>
<td>Suh et al</td>
<td>5</td>
<td>Epithelial ingrowth interface-1 Interface + retrocorneal-4</td>
<td>AS-OCT-1, spectral domain</td>
<td>None documented</td>
<td>Observation in 4 cases</td>
</tr>
<tr>
<td>Bansal et al</td>
<td>1</td>
<td>Epithelial ingrowth after stromal puncture</td>
<td>Ultrahigh resolution OCT-3, histology-1</td>
<td>Clinical</td>
<td>Block excision and corneoscleral graft in 1 case</td>
</tr>
</tbody>
</table>

**Abbreviations:** DSAEK, Descemet stripping automated endothelial keratoplasty; PK, Penetrating Keratoplasty; AS-OCT, Anterior segment optical coherence tomography; OCT, optical coherence tomography.
which was hydrated at the end of the operation. We did not suture the side port. The original site of the epithelial ingrowth was noted to be from the 2 o’clock position and spread superiorly, which was not continuous with the side port incision.

We suspect that irregular trephination at the point of punching the lenticule to size may have resulted in the implantation of a small amount of donor epithelium, but we acknowledge that in the circumstances we have no proof of this. Our case therefore also serves as a reminder of the extreme importance of meticulous technique at all stages of the DSAEK procedure. It is possible that the epithelial implantation occurred as a result of irregular trephination and was therefore potentially avoidable.

Our case indicates that repeat DSAEK may be a successful surgical solution for this very rare but serious complication. We emphasize, however, the importance of very careful stripping and cleaning of the interface, as any epithelial cell rests remaining would risk a recurrence of the problem. We found the combination of manual scraping with a Descemet’s membrane stripper plus the Simcoe manual irrigation/aspiration cannula to be highly effective in this regard. However, not every suspected epithelial implantation leads to graft failure, and we have at least one patient (unpublished observation) in whom apparent interface epithelial inclusions have remained static or even regressed over time. Our literature review suggests that other surgeons have shared this experience.9,10,12,18,19

In summary, we have demonstrated that graft failure due to epithelial ingrowth can occur after an apparently uneventful DSAEK. Early recognition of the condition, careful removal of the implanted epithelium, and repeat DSAEK may help to achieve a successful outcome without the need for more invasive treatments, such as PK.

**Disclosure**

The co-authors have been equally involved in the management of the cases. None of the authors has any proprietary interest. Previously presented in part as a poster at the UKISCRS annual meeting, 2011.

**References**
