Toll-like receptor 8: augmentation of innate immunity in platinum resistant ovarian carcinoma

Taylor J Brueseke
Krishnansu S Tewari
Department of Obstetrics and Gynecology, Division of Gynecologic Oncology, University of California, Irvine, Orange, CA, USA

Abstract: Ovarian cancer is the most deadly gynecologic cancer, with 15,000 anticipated deaths within the United States alone in 2012, and new treatment strategies are needed. Ovarian cancer tumors are known to host an immunosuppressive microenvironment. This suppression may be reversible via activation of the innate immune response. Toll-like receptor 8 activates innate immunity while simultaneously inhibiting the effects of regulatory T cells within the ovarian cancer tumors. VTX-2337 is a novel small molecule ligand of Toll-like receptor 8 and is currently the subject of a Phase II randomized, double-blind, placebo-controlled trial Gynecologic Oncology Group (GOG)-3003 for patients with recurrent platinum-resistant ovarian cancer. We look forward to the results of this trial as support for the paradigm of process therapy in the treatment of ovarian cancer.

Keywords: immunotherapy, ovarian cancer, Gynecology Oncology Group partners, VTX 2237

Introduction
Ovarian cancer is the deadliest gynecological cancer, with 22,000 new cases and 15,000 deaths anticipated within the United States in 2012.1 Despite years of intense research, the etiology of this disease remains unknown. There is currently no consistent early symptom or screening test, and consequently, most patients present with advanced-stage disease. Traditional therapy for ovarian cancer has included maximal cytoreductive surgery followed by cytotoxic chemotherapy with a platinum/taxane-based regimen. While most ovarian cancer is initially chemosensitive, recurrence of the disease is common and may be categorized as either platinum-sensitive or refractory. Current treatment regimens for platinum resistant recurrence include single agent paclitaxel, liposomal doxorubicin, or topotecan. Outcomes with these regimens are poor, with significant potential toxicity, thus, new treatment modalities are needed. The Gynecologic Oncology Group (GOG) is actively pursuing alternative treatment regimens including intraperitoneal chemotherapy, dose-dense paclitaxel, and anti-angiogenesis therapy. To date, there have been four positive Phase III clinical trials demonstrating improved progression-free survival with the anti-angiogenesis monoclonal antibody bevacizumab, in patients with ovarian cancer.2–5

Additional research has focused on immunotherapy and includes:6 administration of tumor-directed antibodies,7,8 administration of immune-stimulatory cytokines,9,10 peptide cancer vaccines, adoptive cell transfers,11 depletion of regulatory T cells, and dysfunctional immune cosignaling blockade. Each of these has met with
modest results. Further insights were gained with the mapping of the ovarian cancer genome atlas, which elucidated multiple aberrant cellular pathways within ovarian tumor cells. These discoveries have generated interest in specific pathway inhibition including: poly (adenosine diphosphate [ADP]-ribose) polymerase (PARP) inhibitors, anti-folic acid receptor inhibitors, heat shock protein 90 inhibition, gamma secretase inhibitors, and aurora kinase inhibitors. However, tumors often possess multiple aberrant pathways with a high degree of cross talk between signaling cascades, and thus, therapeutics directed at pathway inhibition may not have optimal success if the complexity of the pathway is not fully recognized or if a given patient does not possess the targeted aberrant pathway.

Reversing the process of tumor-induced immunosuppression is a promising alternative in immunotherapy. Ovarian cancer tumors are known to contain tumor-infiltrating lymphocytes (including T cells and dendritic cells [DCs]). These lymphocytes, however, are quiescent and do not readily attack tumor cells. The reason for this is multifactorial; however, regulatory T cells and inert DCs are postulated to play a role in the creation of this immunosuppression. Activation of Toll-like receptors (TLRs) holds potential for the reversal of this immunosuppressive microenvironment.

As mentioned in the awarding of the 2011 Nobel Prize in Medicine or Physiology, TLRs and DCs are the link between innate and adaptive immunity, thus, triggering the innate immune response in ovarian cancer tumors may result in activation of cytotoxic T cells and natural killer cells and in the elimination of ovarian cancer cells.

**Innate immunity**

Ralph Steinmann, Bruce Beutler, and Jules Hoffmann were awarded the 2011 Nobel Prize in Medicine or Physiology for discovering the roles that DCs and TLRs play as the gatekeepers of innate immunity. The innate immune system is the first line of defense against foreign organisms and includes natural killer cells, mast cells, eosinophils, basophils, physical barriers, and phagocytic cells, including DCs, macrophages, and neutrophils. DCs possess TLRs, which were the first pathogen-associated pattern-recognition receptors to be discovered. Activation of these receptors by exposure to foreign molecules results in the activation of a signal cascade, with multiple downstream effects. Upon activation, DCs increase their production of major histocompatibility complex (MHC) class II molecules and migrate to draining lymph nodes, where they present antigens to naïve T cells. The presentation of antigens via MHC class II molecules to T helper cells type 1 and 2 results in the activation of the adaptive immune response, with clonal expansion of T cells and the activation of B cell-mediated antibody secretion.

**Tumor microenvironment**

Tumor-infiltrating lymphocytes were described in the microenvironment of ovarian cancer as early as 1988. The types of lymphocytes present include CD8+ T cells, macrophages, a relatively low concentration of natural killer cells, B cells, polymorphonuclear cells, and rare mast cells. Significantly, the presence of tumor-infiltrating lymphocytes is associated with improved overall survival. However, these lymphocytes do not actively target ovarian cancer cells. Rather, an immunosuppressive microenvironment is present within the tumor. Active evasion of the immune response involves at least two cell types: (1) CD4+ CD25+ T cells (T regulatory cells [Tregs]), and (2) CD11c+ MCH-II myeloid DCs (mDCs), which are the most abundant subset of leukocytes in the solid ovarian cancer microenvironment.

Recent research has exploded regarding the role and function of Tregs and suggests that Tregs actively induce cytotoxic T cell anergy in at least two ways: (1) direct cell-to-cell contact inhibition, and (2) humoral inhibition, including the downregulation of interleukin-2. The presence of Tregs has been associated with poor prognosis. In one study, individuals with the highest Treg counts per high-powered field of ovarian cancer tumor tissue had a 25-fold higher death hazard ratio (95% CI 6.8–92.1) compared with those with the lowest Treg cell counts, even after controlling for the stage of disease and surgical debulking. Additionally, mDCs have recently been shown to actively inhibit T cell antitumor activity by expressing functional levels of the immunosuppressive proteins PD-L1.

**Toll-like receptors**

We believe that TLRs may hold the key to reversing this immunosuppressive microenvironment. TLRs constitute a family of highly conserved pattern-recognition receptors (see Table 1). Ten unique TLRs have been characterized in humans. Of these, TLRs 1, 2, 4, 5, and 6 are expressed on the cell membrane, while TLRs 3, 7, 8, and 9 are expressed within the endolysosomal compartmental pathway. TLRs detect a number of different foreign molecules including: double stranded deoxyribonucleic acid (DNA) (TLR3), lipopolysaccharides (TLR4), flagellin (TLR5), single-stranded viral ribonucleic acid (RNA) (TLR7/8), and unmethylated CpG sites of DNA of bacteria and viruses.
Table 1 Breakdown of Toll-like receptors: their cellular expression, intracellular location, natural ligands.

<table>
<thead>
<tr>
<th>TLR</th>
<th>Cellular expression</th>
<th>Location within cell</th>
<th>Natural ligand</th>
</tr>
</thead>
<tbody>
<tr>
<td>TLR1</td>
<td>Monocyte, mDC, B cell, NK, neutrophil, basophil</td>
<td>Cell membrane</td>
<td>Triacyl lipopeptide</td>
</tr>
<tr>
<td>TLR2</td>
<td>Monocyte, mastocytes, mDC</td>
<td>Cell membrane</td>
<td>Triacyl lipopeptide, lipoteichoic acid, zymosan, porins, macrophage-activating lipopeptide, bacterial peptidoglycan, lipoarabinomannan</td>
</tr>
<tr>
<td>TLR3</td>
<td>mDC</td>
<td>Endosomal membrane</td>
<td>dsDNA</td>
</tr>
<tr>
<td>TLR4</td>
<td>Monocytes, macrophages, mDC, mastocytes, basophil</td>
<td>Cell membrane</td>
<td>LPS, mannan, phospholipids, envelope proteins</td>
</tr>
<tr>
<td>TLR5</td>
<td>mDC, monocyte, NK, T cell</td>
<td>Cell membrane</td>
<td>Flagellin</td>
</tr>
<tr>
<td>TLR6</td>
<td>Monocyte, mastocytes, mDC</td>
<td>Cell membrane</td>
<td>Triacyl lipopeptide, lipoteichoic acid, zymosan, porins, macrophage-activating lipopeptide, bacterial peptidoglycan, lipoarabinomannan</td>
</tr>
<tr>
<td>TLR 7</td>
<td>pDC, eosinophil</td>
<td>Endosomal membrane</td>
<td>ssRNA (viral)</td>
</tr>
<tr>
<td>TLR8</td>
<td>mDC, T and B cells, eosinophils, monocytes</td>
<td>Endosomal membrane</td>
<td>ssRNA (viral)</td>
</tr>
<tr>
<td>TLR9</td>
<td>pDC, B cells, basophil, eosinophil</td>
<td>Endosomal membrane</td>
<td>DNA (bacterial/viral)</td>
</tr>
<tr>
<td>TLR10</td>
<td>pDC, neutrophil, B cell, basophil</td>
<td>Cell membrane</td>
<td>Unknown</td>
</tr>
</tbody>
</table>

Abbreviations: DNA, deoxyribonucleic acid; dsDNA, double-stranded DNA; LPS, lipopolysaccharide; mDC, myeloid dendritic cells; NK, natural killer cells; pDC, plasmatic dendritic cells; ssRNA, single stranded ribonucleic acid; TLR, Toll-like receptors.

(TLR9). TLRs are expressed broadly on hematopoietic cells, including DCs, plasma cells, monocytes, and B cells.

TLR activation induces the expression of selectins, chemokines, and chemokine receptors, the combination of which regulates migration of inflammatory cells. As reviewed by Smits et al, the administration of TLR7/8 agonists results in the secretion of multiple proinflammatory Th1-type cytokines, including tumor necrosis factor (TNF)-alpha, interleukins 1, 6, and 8, interferon-inducible protein 10, monocyte chemotactic protein 1, interleukin 12p40, interleukin-12p70, and interleukin 18. The molecular pathways in TLR signaling have previously been reviewed.

TLR7 is the therapeutic target of imiquimod, which has been used successfully in the treatment of skin cancers. Given that TLR9 is expressed predominantly in B cells, it is currently being explored in clinical trials for cutaneous lymphoma and leukemia.

TLR8 is unique in the TLR family: the activation of TLR8 has a direct inhibitory effect on Tregs. The exact mechanism for this is unknown; however, Tregs express TLR8 and the ability of TLR8 to reverse Treg inhibition is dendritic cell independent. TLR8 also likely has direct and indirect activating effects on natural killer cells, causing them to produce interferon-gamma. The overall result of TLR8 activation is robust enhancement of the innate immune response. Additionally, TLR8 receptors have been demonstrated to have variable expression within epithelial ovarian cancer tumors.

**VTX-2337**

The unique ability of TLR8 to reverse Treg immunosuppression and augment immunity in the tumor microenvironment makes it an ideal target for ovarian cancer therapy. VTX-2337 is a synthetic small-molecule agonist that is specific to TLR8, with a molecular weight of 458.6 Daltons and a molecular structure based on a 2-aminobenzazepine core. A recently published in vitro study using TLR-transfected human embryonic kidney cells showed that VTX-2337 selectively activates TLR8. The same study showed that VTX-2337 has the ability to augment the immune response on multiple levels (Figure 1): it induces mDCs and monocytes to produce high levels of interleukin-12 and TNF-alpha. It directly stimulates natural killer cells to produce interferon-gamma and increases their lytic activity. Furthermore, VTX-2337 augments antibody-dependent cell-mediated cytotoxicity. This process works synergistically with standard chemotherapy. Contrary to the traditionally immunosuppressive effects of most systemic cytotoxic chemotherapy, doxorubicin has been reported to increase the tumor cell surface exposure of calreticulin and the internalization of tumor cell antigens by phagocytic DCs. Furthermore, dying cancer cells have been shown to release high-mobility group box 1 protein, a ligand for TLR4 that has been reported to synergize with TLR8 to activate DCs and prime T-cell response.
VTX-2337 has the potential to induce robust activation of the innate and adaptive immune response. A Phase I clinical trial (NTC00688415), sponsored by VentiRx Pharmaceuticals (Seattle, WA, USA), found VTX-2337 to be well tolerated and to have dose-dependent pharmacologic activity. As presented by Cohen et al at the American Society of Clinical Oncology (ASCO) 2011 meeting, the inclusion criteria for this study were: (1) locally advanced or metastatic solid tumors or lymphoma, and (2) Eastern Cooperative Oncology Group (ECOG) performance status of 0–1. The exclusion criteria were: (1) anticancer therapy within 2 weeks, (2) immunosuppressive therapy within 2 weeks, and (3) active autoimmune disease. In this study, VTX-2337 was administered to 33 patients with advanced solid tumors (the most common histologies were colorectal cancer, pancreatic cancer, and melanoma), using a modified Fibonacci dose escalation scheme. Eight cohorts of 3–8 patients received doses from 0.1 mg/m² to 3.8 mg/m². The median age of the patients was 65 years. There was an average of 3.1 years between the time of diagnosis and study entry. The most common drug-related adverse events were grade 1–2 injection site reaction (85%), chills (58%), fever (42%), and flu-like symptoms (24%). Grade 3 hypotension occurred in 1/6 patients and was the only dose-limiting toxicity. The primary outcome measures were safety and the identification of dose-limiting toxicities and pharmacokinetics. The secondary outcome measures were the identification of pharmacodynamics and mean toxic dose. The results showed that the plasma levels of multiple immune mediators (including granulocyte colony-stimulating factor [G-CSF], monocyte chemotactic protein [MCP]-1, macrophage inflammatory protein [MIP]-1-beta, and TNF-alpha) were increased. As measured by the Response Evaluation Criteria in Solid Tumors (RECIST) criteria, 25% of patients demonstrated disease stabilization at 8 weeks.

**VTX-2337 and ovarian cancer**

Two clinical trials are currently investigating VTX-2337 in patients with ovarian cancer. GOG-9925 (study chair BJ Monk) is a Phase I trial undergone in collaboration with the National Cancer Institute, NCT01294293, evaluating the side-effect profile and best dose of VTX-2337 and liposomal doxorubicin, in patients with persistent or recurrent ovarian epithelial, fallopian tube, or peritoneal cancer. The primary outcomes are first-cycle dose-limiting toxicity and the frequency/severity of toxicities. The secondary outcome measures are immune activation and identification of the pharmacokinetics of VTX-2337 and pegylated liposomal doxorubicin. Enrollment criteria include measurable disease by RECIST 1.1, or detectable disease by CA-125 measurement, or other solid abnormalities on radiographic imaging. Enrolled patients have one prior platinum-based chemotherapeutic regimen and recurrence of disease within a 12-month platinum-free interval and do not have evidence of central nervous system (CNS) metastasis. This multicenter, dose-escalating study enrolled patients from April 2011 to June 2012.17 No results have yet been published from this trial.

A Phase II randomized, double-blind, placebo-controlled study is also now underway. Known as GOG Partners-3003 (study chair BJ Monk), NCT01666444 this trial is sponsored by VentiRx Pharmaceuticals in collaboration with the GOG partners and enrolls patients with recurrent or persistent platinum-resistant epithelial ovarian, fallopian tube, or primary peritoneal carcinoma, with measurable disease by RECIST 1.1 criteria and at least one target lesion. Figure 2 shows a schema of the trial structure: the investigational arm is VTX-2337 plus standard regimen pegylated liposomal doxorubicin (PLD). The reference arm is placebo plus standard regimen PLD. Patients who have previously been treated with VTX-2337, doxorubicin, PLD, or other anthracycline, patients receiving immunosuppressive therapy for any reason, and patients with active autoimmune disease are excluded from this study. 3.0 mg/m² of VTX-2337 or placebo will be administered as a subcutaneous injection on days 3, 10, and 17 of the first four 28-day treatment cycles and thereafter, on cycle day 3 only. The primary outcome will be overall survival, with a planned analysis to occur approximately 15 months after the last patient is enrolled. Secondary outcomes
will be the frequency and nature of side-effects of VTX-2337 and progression-free survival using Immune Related Response Evaluation Criteria in Solid Tumors (RECIST) 1.1. Exploratory objectives include the assessment of the effect of TLR8 polymorphisms and immune cell subsets (as measured by immunohistochemistry in primary tumor tissue) on clinical outcomes, such as overall survival, progression-free survival, and overall response rate.58

Conclusion

In conclusion, ovarian cancer remains a deadly disease, and new therapeutic approaches are needed. The immunosuppressive microenvironment present in ovarian tumors represents a pathologic process that is a prime target for novel therapies. Indeed, the 2011 Nobel Prize in Medicine or Physiology highlights that TLRs are the gateway to activating the innate immune response, and it appears that TLR8 ligands are in a unique position to augment the current treatment of patients with ovarian cancer. Novel immunotherapeutic strategies and manipulation of the microenvironment may hold the key to future success in ovarian cancer therapy.

Disclosure

The authors report no conflicts of interest in this work.

References


