VCAM-1-targeted core/shell nanoparticles for selective adhesion and delivery to endothelial cells with lipopolysaccharide-induced inflammation under shear flow and cellular magnetic resonance imaging in vitro

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Abstract: Multifunctional nanomaterials with unique magnetic and luminescent properties have broad potential in biological applications. Because of the overexpression of vascular cell adhesion molecule-1 (VCAM-1) receptors in inflammatory endothelial cells as compared with normal endothelial cells, an anti-VCAM-1 monoclonal antibody can be used as a targeting ligand. Herein we describe the development of multifunctional core-shell Fe₃O₄@SiO₂ nanoparticles with the ability to target inflammatory endothelial cells via VCAM-1, magnetism, and fluorescence imaging, with efficient magnetic resonance imaging contrast characteristics. Superparamagnetic iron oxide and fluorescein isothiocyanate (FITC) were loaded successfully inside the nanoparticle core and the silica shell, respectively, creating VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles that were characterized by scanning electron microscopy, transmission electron microscopy, fluorescence spectrometry, zeta potential assay, and fluorescence microscopy. The VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles typically had a diameter of 355 ± 37 nm, showed superparamagnetic behavior at room temperature, and cumulative and targeted adhesion to an inflammatory subline of human umbilical vein endothelial cells (HUVEC-CS) activated by lipopolysaccharide. Further, our data show that adhesion of VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles to inflammatory HUVEC-CS depended on both shear stress and duration of exposure to stress. Analysis of internalization into HUVEC-CS showed that the efficiency of delivery of VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles was also significantly greater than that of nontargeted Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticles. Magnetic resonance images showed that the superparamagnetic iron oxide cores of the VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles could also act as a contrast agent for magnetic resonance imaging. Taken together, the cumulative adhesion and uptake potential of these VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles targeted to inflammatory endothelial cells could be used in the transfer of therapeutic drugs/genes into these cells or for diagnosis of vascular disease at the molecular and cellular levels in the future.

Keywords: silica nanoparticles, vascular cell adhesion molecule-1, endothelial cells, adhesion, magnetic resonance imaging

Introduction

In recent years, there has been increasing interest in fabricating carriers in the nanometer and micrometer range with important applications in biomedicine. Of these, magnetic nanoparticles, eg, Fe₃O₄, are a promising candidate because of their wide range of...
sophisticated biomedical applications, including targeted drug delivery, magnetic hyperthermia, magnetic resonance imaging, biosensing, and magnetic separation of biological materials.\(^1\)\(^-\)\(^3\) Although magnetic nanoparticles have many advantages as magnetic carriers, they have at least two limitations. One is poor dispersion with easy aggregation and the other is the limited number of functional groups on their surfaces. To overcome these limitations, modifications have been made by adding surface coatings to the magnetic nanoparticles. In addition to its well-developed surface chemistry, silica is considered to be an exceptional material for encapsulating magnetic nanoparticles because of its good biocompatibility, excellent physicochemical stability, and ease of multifunctionalization.\(^2\)\(^,\)\(^4\)

In general, coating of Fe\(_3\)O\(_4\) nanoparticles with silica can be achieved using the Stöber process and the reverse (water-in-oil) microemulsion method.\(^5\)\(^,\)\(^6\) Using the reverse microemulsion method, it has been shown that aqueous Fe\(_3\)O\(_4\) nanoparticles can be incorporated within silica to form core/shell silica particles with single Fe\(_3\)O\(_4\) cores\(^7\) which can efficiently label human mesenchymal stem cells. However, the superparamagnetic behavior and saturation of magnetization decreases after coating with silica. One possible strategy is to encapsulate more Fe\(_3\)O\(_4\) cores into the silica shell,\(^8\) which would create hybrid silica nanoparticles with a large magnetic component. Consequently, the superparamagnetic behavior, saturation of magnetization, and magnetic resonance signal intensity is significantly increased. Other studies have shown that the magnetic core can be replaced by semiconductor quantum dots or fluorescence molecules doped into the silica shell to form fluorescent core/shell silica nanoparticles.\(^9\)\(^-\)\(^11\) These silica-coated fluorescent magnetic nanoprobes enabled development of biomedical platforms for simultaneous imaging, diagnosis, and therapy.\(^12\)\(^-\)\(^14\) However, it is still a challenge to fabricate fluorescent magnetic nanoprobes with high photo-stability, high payloads of dye, and desirable outer surfaces for further modification with functional or target moieties.

Avoiding nonspecific delivery is another important concern in the design of nanoparticles. An attractive means of targeted delivery is use of cell-specific targeting ligands linked to nanoparticle surfaces, because these enhance adhesion and cellular uptake (internalization) of nanoparticles via use of unique molecular markers that are preferentially overexpressed in diseased tissue. This strategy is particularly effective for receptor-mediated endocytosis using specific ligands to recognize their receptors on cell membranes.

Inflammation and endothelial dysfunction are major threshold developments in the progression of atherosclerosis. Expression of endothelial cell adhesion molecules, eg, vascular cell adhesion molecule-1 (VCAM-1), plays an important role in recruitment of leukocytes, and is often increased at sites of pathological inflammation. Persistent expression of VCAM-1 in dysfunctional endothelial cells mediates adhesion, rolling, and tethering of mononuclear leukocytes and facilitates their transmigration to developing atherosclerotic plaques. Previous studies, including those of our group, have reported overexpression of VCAM-1 and/or other cell adhesion molecules associated with endothelial cell dysfunction and progression of atherosclerosis.\(^15\)\(^,\)\(^16\) Therefore, VCAM-1 is a potential marker and ideal target for drug/gene delivery and early detection by imaging.

Here we report a new class of multifunctional VCAM-1-targeted Fe\(_3\)O\(_4\)@SiO\(_2\)(FITC) nanoparticles. We used a parallel flow chamber system to investigate adhesion of these nanoparticles on a monolayer of endothelial cells under controlled shear conditions. Using confocal laser microscopy and flow cytometry, we confirmed that the VCAM-1-targeted Fe\(_3\)O\(_4\)@SiO\(_2\)(FITC) nanoparticles accumulated on the surfaces of endothelial cells with high efficiency. Moreover, we demonstrated that these biofunctionalized nanoparticles could be used as both fluorescent and magnetic imaging probes at the cellular level. These nanoparticles are expected to be an important candidate for enhancing contrast in diagnostic imaging for atherosclerosis in the future.

**Materials and methods**

**Chemicals and materials**

Polyoxyethylene(5) nonylphenylether (Igepal CO-520), tetraethyloxysilicate, 3-aminopropyltrimethoxysilane (APS), fluorescein isothiocyanate (FITC), N-(3-dimethylaminopropyl)-N′-ethylcarbodiimide hydrochloride, N-hydroxysuccinimide, sodium silicate solution (Na\(_2\)O(SiO\(_2\))\(_3\), 27 wt% SiO\(_2\)), and lipopolysaccharide were obtained from Sigma-Aldrich (St Louis, MO, USA). Iron oxide nanoparticles (10 nm) were obtained from Nanjing Emperor Nano Material Co, Ltd (Nanjing, People’s Republic of China). RPMI-1640 cell culture medium, newborn calf serum, and trypsin were purchased from Gibco (Grand Island, NY, USA). A phycoerythrin-labeled anti-VCAM-1 monoclonal antibody was obtained from BioLegend (San Diego, CA, USA). All chemicals were used as received without further purification.

**Synthesis of VCAM-1-targeted Fe\(_3\)O\(_4\)@SiO\(_2\)(FITC) nanoparticles**

Fe\(_3\)O\(_4\)@SiO\(_2\)(FITC) nanoparticles were synthesized by hydrolysis and condensation of tetraethyloxysilicate in

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an ammonia ethanol/water solution in the presence of iron oxide particles and FITC using a modification of a previously reported sol-gel method. Briefly, 2 mg of FITC was reacted with 125 µL of APS in 1 mL of anhydrous ethanol in the dark at room temperature for 24 hours. The reaction solution of FITC-APS conjugates was then stored at 4°C. Next, 1 mL of aqueous solution containing magnetic particles was diluted with 1 mL of water and 7.5 mL of anhydrous ethanol; 300 µL of tetraethylorthosilicate and ammonia solution (28–30 wt% by NH₃) were added to this reaction mixture while stirring at 300 rpm for 24 hours at room temperature, followed by addition of an as-synthesized APS-FITC solution with continuous stirring for 48 hours. The reaction was terminated by adding methanol. The Fe₃O₄@SiO₂(FITC) particles were separated and washed several times with Milli-Q water (Millipore, Billerica, MA, USA).

The Fe₃O₄@SiO₂(FITC) nanoparticles obtained were resuspended in 5 mL of anhydrous acetic acid (1 mM), and reacted further with 200 µL of APS in a sealed vessel. The reaction was allowed to proceed at room temperature for 24 hours with gentle stirring at 300 rpm. The Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticles were then separated by centrifugation at 12,000 rpm for five minutes and washed five times with deionized water to remove excess reactants.

To attach anti-VCAM-1 monoclonal antibodies to the surfaces of the Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticles, 20 µL of anti-VCAM-1 monoclonal antibody were mixed with 20 mL of N-(3-dimethylaminopropyl)-N’-ethylcarbodiimide hydrochloride and 10 mL of N-hydroxysuccinimide solution, with gentle shaking at 130 rpm for 30 minutes in ice-cold water. The mixture was then added to a flask containing 1 mL of the Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticle suspension and stirred at 300 rpm for 12 hours at room temperature. The VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles were then centrifuged and washed with deionized water, finally dispersed in phosphate-buffered saline at 4°C, and then stored. The procedure used for synthesis of the VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles is shown in Figure 1.

**Characterization**

Particle morphology and size were investigated by scanning electron microscopy (JSM-6490 LV, JEOL, Tokyo, Japan) and transmission electron microscopy (JEM-100CX, JEOL). The size distribution and surface charge on the nanoparticles were

![Figure 1](https://www.dovepress.com/1899-vcam-1-core-shell-nanoparticles-targeted-to-inflammatory-endothelial-cells)

**Figure 1** Schematic illustration of the synthesis procedure for VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles. (A) Fe₃O₄@SiO₂(FITC) nanoparticles were synthesized by reverse microemulsion. (B) The coupling agent APS was used to continue the reaction to form Fe₃O₄@SiO₂(FITC)-NH₂ in acetic acid; surface modification of the phycocerythrin-labeled anti-VCAM-1 monoclonal antibody resulted in VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles.

**Abbreviations:** Fe₃O₄@SiO₂(FITC), fluorescein isothiocyanate-loaded silica-coated superparamagnetic iron oxide nanoparticles; VCAM-1, vascular cell adhesion molecule-1; TEOS, tetraethylorthosilicate; NPs, nanoparticles; EDC, N-(3-dimethylaminopropyl)-N’-ethylcarbodiimide; NHS, N-hydroxysuccinimide.
analyzed using a Mastersizer 2000 (Malvern Instruments, Worcestershire, UK) in accordance with the manufacturer’s operating manual. The presence of FITC doped into the silica shell was confirmed using an inverted fluorescence microscope (TE-2000U, Nikon, Tokyo, Japan). Magnetic measurement of the samples was performed on a superconducting quantum interference magnetometer (MPMS55, Quantum Design Inc, San Diego, CA, USA). For measurement, about 10 mg of powder samples was inserted into a gelatin capsule. The magnetic hysteresis loop was measured at room temperature (300 K).

Cell culture
An inflammatory subline of human umbilical vein endothelial cells (HUVEC-CS) was obtained from the American Type Culture Collection (Manassas, VA, USA) and cultured in RPMI-1640 cell culture medium supplemented with 10% newborn calf serum and 1% penicillin-streptomycin.15 The cells were maintained in an incubator at 37°C with 95% humidity and 5% CO₂, subcultured after trypsinization (0.025% trypsin, 0.5 mM EDTA, 1 mM sodium pyruvate, 10 mM HEPES), and grown to approximately 90% confluence. The HUVEC-CS were then subcultured in 12-well plates at a density of 4×10⁵/mL, and maintained in an incubator overnight. The cultured HUVEC-CS were stimulated with 1 µg/mL lipopolysaccharide for five hours to induce inflammation.19

Shear flow simulation and nanoparticle adhesion
The interaction (adhesion) between VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles and a monolayer of HUVEC-CS was measured in a parallel plate flow chamber at shear stresses of 1.1, 5.15, and 9.94 dynes/cm², as previously described by our laboratory.15,20 In brief, a flow channel in the chamber was constructed using a silicon gasket. The cell-seeded glass slide and the gasket were fastened between a polycarbonate base plate and a stainless plate. The chamber was connected to a perfusion loop system, kept in a constantly temperature-controlled enclosure, and maintained at pH 7.4 by continuous gassing with a humidified mixture of 5% CO₂ in air. Flow of perfusate in a flow channel is laminar with a parabolic velocity profile. The fluid shear stress (τ) generated on the cells seeded on the glass slide can be estimated as τ = 6µQ/wh², where Q is the flow rate, µ is the dynamic viscosity of the perfusate, w is the width of the flow field, and h is the height.21 The shear stress can be regulated through the flow rate, Q. Before perfusion, the VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticle suspension was mixed with the perfusion medium. Adhesion was recorded using an inverted fluorescence microscope. Six random fields (objective magnification 10×) were chosen to quantify the fluorescence intensity on the endothelial cell monolayer.

Specific accumulation on cell surfaces and confocal microscopy
The HUVEC-CS (0.5×10⁵ cells/mL) were cultured on coverslips, which were kept in a six-well plate for 12 hours before treatment. After treatment with Fe₃O₄@SiO₂(FITC)-NH₂ or VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles with or without lipopolysaccharide stimulation, the cells were washed three times with phosphate-buffered saline (pH 7.4), and then fixed with 4% paraformaldehyde solution in phosphate-buffered saline for 15 minutes at 37°C. The coverslips were then washed three times with phosphate-buffered saline to remove nonspecific binding. Finally, the samples were examined under a confocal laser scanning microscope (FluoView™ FV1000, Olympus, Japan).

Flow cytometry assay
To quantify specific accumulation of the nanoparticles, HUVEC-CS were seeded in 12-well plates at a density of 0.5×10⁵ cells/well and allowed to grow for 12 hours. The cells were then activated with lipopolysaccharide 1 µg/mL for five hours. The cells were washed three times with sterilized phosphate-buffered saline, and 2 mL of RPMI-1640 culture medium was then added. The HUVEC-CS were exposed to nanoparticles at a concentration of 850 µg/mL for 12 hours, then washed repeatedly with sterilized phosphate-buffered saline three times, trypsinized, and suspended in phosphate-buffered saline for flow cytometry analysis (FACSCanto II, Becton Dickinson, Franklin Lakes, NJ, USA).

Magnetic resonance imaging in vitro
HUVEC-CS were seeded onto 12-well plates at a density of 0.5×10⁵ cells/mL, grown overnight, and then activated with lipopolysaccharide 1 µg/mL for five hours. The cells were washed three times with sterilized phosphate-buffered saline, and 2 mL of RPMI-1640 was then added for 10 hours of culture in the incubator. Next, VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles at various concentrations (0, 100, 200, 300, and 400 µg/mL) were added, and a saline solution without any nanoparticles was used as the control. After 12 hours of incubation, the cells were washed three times in phosphate-buffered saline to remove the free nanoparticles, collected, and suspended in 1.5 mL of phosphate-buffered saline containing 1% agarose. Magnetic resonance imaging
experiments were performed in a clinical 3.0 Tesla Clinical Siemens Trio scanner (Discovery MR750, GE Healthcare, Boulder, CO, USA). T₂-weighted images were acquired using spin-echo imaging sequences.

**Statistical analysis**

The quantitative data are presented here as the mean ± standard deviation. Statistical analyses between different groups were performed using one-way analysis of variance followed by Bonferroni tests, and a P value of <0.05 was considered to be statistically significant.

**Results and discussion**

Magnetic nanoparticles have been attracting interest as a result of their unique properties and potential applications in biomedicine and bioengineering. However, most of these applications require magnetic nanoparticles to be embedded in a nonmagnetic matrix to avoid aggregation and sedimentation, as well as to endow them with particular surface properties for specific applications. This has been attempted by several groups. Although few research groups have concentrated on how nanoparticles adhere to targeted cell surfaces, attempts have been directed towards interactions between targeted nanoparticles and endothelial cells under hemodynamic shear flow conditions.

In this study, synthesis of Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticles was based on sol-gel growth of silica in a limited domain of water in water-in-oil microemulsion. Using this method, it is possible to grow an iron oxide in a silica core/shell structure of high uniformity. This synthesis process enables coencapsulation of Fe₃O₄ nanoparticles and a large number of FITC dye molecules together within a silica shell. The diameter of the Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticles increased from 331 ± 49 nm (Figure 2A) to 355 ± 37 nm after modification with anti-VCAM-1 monoclonal antibody (Figure 2B). Surface modification of Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticles with phycoerythrin-labeled anti-VCAM-1 monoclonal antibody was likely responsible for the slight increase in particle size. Moreover, these as-synthesized nanoparticles are uniform, and can be well dispersed in aqueous solution, thus making it feasible to measure relaxivity and to run cell experiments. In our earlier report, we also showed that Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticles have low cytotoxicity.

FITC-APS conjugates were prepared in advance via an addition reaction between the isothiocyanate group of FITC dye and the primary amine group of APS. This synthetic process enables the coencapsulation of Fe₃O₄ nanoparticles and a large number of FITC dye molecules together within the silica shell, and grafting of primary amines into the silica surface. Using fluorescence microscopy, we confirmed that both Fe₃O₄@SiO₂(FITC)-NH₂ and VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles showed well dispersed and distinct fluorescence (Figure 3). We also found that anti-VCAM-1 monoclonal antibody modification had no obvious influence on fluorescence intensity (Figure 3C). The final surface layer incorporating primary amines was formed in situ by hydrolysis of APS. It was confirmed that the zeta potential changed from negative (~20.1 ± 4.1 mV) to positive (8.4 ± 2.9 mV, Figure 4). The zeta potential also did not change significantly, even after modification with anti-VCAM-1 monoclonal antibody. Further, fluorescence spectroscopy was carried out to determine whether anti-VCAM-1 monoclonal antibody molecules conjugated with the Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticle surfaces. The fluorescence emission spectra showed an emission peak at a wavelength of approximately 575 nm for the VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticle suspension (Figure 5), which is the characteristic

![Figure 2](image-url) Scanning electron microscopic images of Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticles with a diameter of 331 ± 49 nm (A) and VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles with a diameter of 355 ± 37 nm (B).

**Notes:** Inset shows a higher magnification transmission electron microscopic image indicating the core/shell structure of Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticles. The Fe₃O₄ nanoparticle core is marked by arrows.

**Abbreviations:** Fe₃O₄@SiO₂(FITC), fluorescein isothiocyanate-loaded silica-coated superparamagnetic iron oxide nanoparticles; VCAM-1, vascular cell adhesion molecule-1.

![Figure 3](image-url) (A) Light microscopic images for aqueous suspension of Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticles in solution. (B and C) Fluorescence microscopy images of aqueous suspension of Fe₃O₄@SiO₂(FITC)-NH₂ and VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles in solution, respectively.

**Abbreviations:** Fe₃O₄@SiO₂(FITC), fluorescein isothiocyanate-loaded silica-coated superparamagnetic iron oxide nanoparticles; VCAM-1, vascular cell adhesion molecule-1.
fluorescence emission peak of phycoerythrin. This suggests that anti-VCAM-1 monoclonal antibody molecules were successfully conjugated with the Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticle surfaces. For assessment of the magnetic properties and sensitivity of the formulated Fe₃O₄@SiO₂(FITC)-NH₂ and VCAM-1-targeted Fe₃O₄@SiO₂(FITC), magnetic hysteresis loops were recorded using a magnetometer, and both types of nanoparticles showed superparamagnetic behavior without magnetic hysteresis (Figure 6) at room temperature (about 300 K). They were found to have no coercive fields, thereby confirming their superparamagnetic nature. Their saturation of magnetization was relative low at 3.0 T because of the presence of the thick outer silica shell. It is interesting that there was no significant difference in their magnetic hysteresis loops and magnetization saturation values even when the Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticles were modified with the anti-VCAM-1 monoclonal antibody.

The vascular endothelium is a potential target for therapeutic intervention in diverse pathological processes, including inflammation, atherosclerosis, and thrombosis. When targeted nanoparticles enter the bloodstream after injection, they will inevitably interact with endothelial cells, and be retained in the inflamed endothelium via specific endothelial cell adhesion molecules (eg, VCAM-1), expression of which is upregulated on endothelial cell surfaces. This interaction provides the basis for targeted drug/gene delivery or imaging of the abovementioned diseases. Therefore, it was necessary to profile VCAM-1-targeted Fe₃O₄@SiO₂(FITC) adhesion under flow conditions in vitro. Firstly, we compared adhesion with HUVEC-CS between the Fe₃O₄@SiO₂(FITC)-NH₂ and VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles under static conditions. We found no significant difference for nontargeted Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticles regardless of activation by lipopolysaccharide (Figure 7). However, the degree of adhesion between VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles and HUVEC-CS was markedly higher when the cells were activated by lipopolysaccharide or under a magnetic field, and the degree of adhesion decreased after blocking of the VCAM-1 receptors on the HUVEC-CS surfaces by the anti-VCAM-1 monoclonal

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**Figure 4** Zeta potential measurements of Fe₃O₄@SiO₂, Fe₃O₄@SiO₂(FITC), Fe₃O₄@SiO₂(FITC)-NH₂, and VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles in aqueous suspension.

**Abbreviations:** Fe₃O₄@SiO₂(FITC), fluorescein isothiocyanate-loaded silica-coated superparamagnetic iron oxide nanoparticles; VCAM-1, vascular cell adhesion molecule-1.

**Figure 5** Fluorescence emission spectra of Fe₃O₄@SiO₂(FITC)-NH₂ and VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles.

**Notes:** The fluorescence excitation peak at a wavelength of approximately 575 nm indicates successful coupling of the phycoerythrin-labeled anti-VCAM-1 monoclonal antibody to the particle surfaces.

**Abbreviations:** Fe₃O₄@SiO₂(FITC), fluorescein isothiocyanate-loaded silica-coated superparamagnetic iron oxide nanoparticles; VCAM-1, vascular cell adhesion molecule-1.

**Figure 6** Magnetization curves of Fe₃O₄@SiO₂(FITC)-NH₂ and VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles as a function of the applied magnetic field at room temperature (300 K).

**Abbreviations:** Fe₃O₄@SiO₂(FITC), fluorescein isothiocyanate-loaded silica-coated superparamagnetic iron oxide nanoparticles; VCAM-1, vascular cell adhesion molecule-1.
antibody (Figure 7). It was shown that VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles could adhere cumulatively and aggregate onto the inflammatory HUVEC-CS, and have the potential for targeted delivery. Secondly, the adhesion of nontargeted Fe₃O₄@SiO₂(FITC)-NH₂ and VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles was compared further under several sets of flow conditions. For both nontargeted Fe₃O₄@SiO₂(FITC)-NH₂ and VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles, their degree of adhesion decreased with increasing of shear stress (Figure 8) and duration of exposure to stress (Figure 9). Comparing Figures 8 and 9, the adhesion for each group was also found to show a shear-dependent relationship. The degree of nanoparticle adhesion decreased significantly when the exposed shear stresses increased from 1.1 to 9.94 dynes, indicating that interaction between the nanoparticles and HUVEC-CS depended on both shear stress and duration of exposure to stress. However, the degree of adhesion of VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles to endothelial cells was significantly greater than that of nontargeted Fe₃O₄@SiO₂(FITC)-NH₂ nanoparticles in the same experimental group. This specific adhesion could be used in clinical applications such as targeted magnetic resonance imaging or drug delivery to localized disease sites in the future.

To explore the biological applications of VCAM-1-targeted Fe₃O₄@SiO₂(FITC) nanoparticles further, their
accumulation was investigated by confocal microscopy and flow cytometry under in vitro conditions. We first compared the cellular distribution and accumulation of nontargeted Fe₃O₄@SiO₂ (FITC)-NH₂ and VCAM-1-targeted Fe₃O₄@SiO₂ (FITC) nanoparticles without stimulation by lipopolysaccharide. It was found that there was no marked difference, as shown in Figure 10A. A low level of VCAM-1 expression was possible without applying lipopolysaccharide stimulation to HUVEC-CS. However, the accumulation of VCAM-1-targeted Fe₃O₄@SiO₂ (FITC) nanoparticles on the cell surfaces was considerably higher compared with nontargeted Fe₃O₄@SiO₂ (FITC) nanoparticles (top two panels in Figure 10B). This was because of upregulation of VCAM-1 expression on lipopolysaccharide-activated endothelial cell surfaces causing more adhesion of VCAM-1-targeted Fe₃O₄@SiO₂ (FITC) nanoparticles on HUVEC-CS. This phenomenon also existed in a magnetic field (third panel in Figure 10B). To verify whether adhesion is mediated via VCAM-1, lipopolysaccharide-activated HUVEC-CS were pretreated with anti-VCAM-1 monoclonal antibody to block the VCAM-1 receptors. Accumulation of VCAM-1-targeted Fe₃O₄@SiO₂ (FITC) nanoparticles on the cell surfaces slightly decreased after pretreatment with the anti-VCAM-1 monoclonal antibody (the lowest panel in Figure 10B). It seems reasonable that some VCAM-1 receptors were occupied by anti-VCAM-1 monoclonal antibody, decreasing accumulation of VCAM-1-targeted Fe₃O₄@SiO₂ (FITC) nanoparticles on HUVEC-CS. These data also suggest that VCAM-1-mediated adhesion is an important avenue for delivery of modified nanoparticles, and some other accumulation or cellular uptake mechanisms in addition to VCAM-1-mediated endocytosis may also be operating. The quantitative accumulation of nanoparticles was further determined using detection of mean fluorescence intensity by flow cytometry, as shown in Figure 10C. The results agreed well with the data obtained by confocal microscopy. Our data showed that the VCAM-1-targeted Fe₃O₄@SiO₂ (FITC) nanoparticles were effectively delivered to the HUVEC-CS (ie, a dual-targeted delivery system).

To assess the magnetic sensitivity and imaging feasibility of VCAM-1-targeted Fe₃O₄@SiO₂ (FITC) nanoparticles as biomedical probes, HUVEC-CS were treated with various concentrations of VCAM-1-targeted Fe₃O₄@SiO₂ (FITC) nanoparticles. Magnetic measurement of the VCAM-1-targeted Fe₃O₄@SiO₂ (FITC) nanoparticles at 300 K showed no hysteresis (Figure 6), indicating that they were superparamagnetic and favorable as T₂ magnetic resonance contrast agents. Figure 11A shows the image-darkening effect with increasing concentration of VCAM-1-targeted Fe₃O₄@SiO₂ (FITC) nanoparticles. Because of their relatively low magnetization saturation, the contrast was more obvious at a high particle concentration of 400 µg/mL, indicating that the functionalized nanoparticles could be used as a probe in simultaneous T₂ magnetic resonance imaging and
optical fluorescence imaging (FITC tracking). We also determined the $T_2$ relaxivity ($R_2$, efficiency of a contrast agent) of the VCAM-1-targeted $\text{Fe}_3\text{O}_4@\text{SiO}_2$(FITC) nanoparticles, and it was found that they retained good magnetism (Figure 11B).

**Conclusion**

We have developed a strategy for synthesis of novel multifunctional VCAM-1-targeted $\text{Fe}_3\text{O}_4@\text{SiO}_2$(FITC) nanoparticles with a diameter of $355 \pm 37$ nm, which are stable and well dispersed in aqueous solution. The interaction between VCAM-1-targeted $\text{Fe}_3\text{O}_4@\text{SiO}_2$(FITC) nanoparticles and HUVEC-CS under various flow conditions was evaluated further using a parallel plate flow chamber system. The VCAM-1-targeted $\text{Fe}_3\text{O}_4@\text{SiO}_2$(FITC) nanoparticles could adhere specifically to inflammatory endothelial cells activated by lipopolysaccharide, and their adherence depended on the shear stress and duration of exposure to stress. We also demonstrated that the VCAM-1-targeted $\text{Fe}_3\text{O}_4@\text{SiO}_2$(FITC) nanoparticles were able to accumulate specifically on cell surfaces and enter endothelial cells with high efficiency via VCAM-1-mediated binding. Moreover, we have demonstrated that VCAM-1-targeted $\text{Fe}_3\text{O}_4@\text{SiO}_2$(FITC) nanoparticles have potential as probes in magnetic resonance imaging. The possibility of selective delivery of VCAM-1-targeted $\text{Fe}_3\text{O}_4@\text{SiO}_2$(FITC) nanoparticles to sites of inflammation and their accumulation or uptake by targeted cells give them high potential in vascular magnetic resonance imaging for clinical diagnosis of cardiovascular disease, eg, atherosclerosis and thrombosis.

**Disclosure**

The authors report no conflicts of interest in this work.

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