Endoglin: a critical mediator of cardiovascular health

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Abstract: Endoglin (CD105) is a type III auxiliary receptor for the transforming growth factor beta (TGFβ) superfamily. Several lines of evidence suggest that endoglin plays a critical role in maintaining cardiovascular homeostasis. Seemingly disparate disease conditions, including hereditary hemorrhagic telangiectasia, pre-eclampsia, and cardiac fibrosis, have now been associated with endoglin. Given the central role of the TGFβ superfamily in multiple disease conditions, this review provides a detailed update on endoglin as an evolving therapeutic target in the management of cardiovascular disease.

Keywords: endoglin, transforming growth factor beta, vascular, cardiac remodeling

Introduction

Endoglin (CD105) is a type III auxiliary receptor for the transforming growth factor beta (TGFβ) superfamily. Over the past three decades, several lines of evidence suggest that endoglin plays a critical role in maintaining cardiovascular homeostasis. First, mutations in human endoglin result in the autosomal dominant vascular dysplastic syndrome, hereditary hemorrhagic telangiectasia (HTT).1 Second, Endoglin null mice (Eng−/−) die at embryologic day 10–11.5 because of impaired cardiovascular development.2 Third, endoglin expression is increased in human atherosclerotic plaques3 and in models of balloon angioplasty-mediated vascular injury.4 Fourth, endoglin is abundantly expressed by neovascularized tumors in prostate, breast, and colon cancer.5 Finally, endoglin has recently been shown to regulate cardiac remodeling in heart failure6 and myocardial infarction.7 Given the central role of the TGFβ superfamily in multiple disease conditions, endoglin represents a potentially unique target of therapy for several debilitating conditions involving the cardiovascular system.

Endoglin structure, function, and distribution

Endoglin is a 180 kDa homodimeric integral membrane glycoprotein composed of disulfide-linked subunits. The receptor includes a large 561-amino acid extracellular domain with a single transmembrane domain and a serine/threonine-rich cytoplasmic region, with 47 amino acids in the long isoform (L-endoglin) and 14 residues in the short isoform (S-endoglin).8 The cytoplasmic domain of the predominantly expressed L-endoglin constitutes the region of the protein with the highest degree of conservation in mammalian species. A low-resolution structural analysis of the extracellular domain revealed a dome comprised of antiparallel-oriented monomers with a cavity at one end. Each endoglin monomer is comprised of three well
defined domains. A large N-terminus domain extending from Glu26 to Ile359 forms a large flat area and does not show any degree of identity with known protein domains and is therefore referred to as the orphan domain. This domain mediates interactions with ligands of the TGFβ superfamily. Initial studies demonstrated that endoglin can be found in the receptor complex for the TGFβ1 and TGFβ3 isoforms in association with the type II receptor (TBR2).9 Subsequently, it was shown that endoglin also interacts with activin as well as bone morphogenetic proteins (BMP)-2 and BMP7 in association with the ligand binding receptor, which can be either a type I or type II receptor depending on the ligand involved.10 More recent studies have shown that endoglin may bind directly to BMP9 and BMP10.11,12

The orphan domain is followed by two zona pellucida subdomains, organized into an open U-shaped monomer and containing eight conserved cysteine residues localized to Lys362–Asp561.12,14 This zona pellucida domain is highly conserved among endoglin proteins from human, mouse, rat, pig, and dog models, and constitutes more than 30% of the extracellular region, suggesting an important functional role.11,13 The extracellular domain of endoglin also contains consensus motifs for several O-linked and N-linked oligosaccharide chains, with glycosylation accounting for about 30% of the molecular weight. The sequence of human endoglin originally revealed an arginine-glycine-aspartic acid (RGD) tripeptide at Arg399–Asp401, suggesting that endoglin may play a role in cell adhesion.14 However, that sequence was not conserved in other species. A recent study demonstrated that endoglin can bind to the α5β1 integrin via its RGD motif or a functionally equivalent murine TDD peptide that may facilitate leukocyte transmigration.26

The extracellular domain of endoglin can be proteolytically cleaved by matrix metalloproteinase 14 and circulate as soluble endoglin, which may serve as a naturally occurring antagonist for TGFβ signaling and therefore play an important role in cardiovascular disease.6,8,17 The cytoplasmic domain of endoglin is phosphorylated by serine-threonine kinase TBRI and TBR2 TGFβ domain of endoglin is phosphorylated by serine-threonine kinase TBRI and TBR2 TGFβ-

zyxin and zyxin-related protein-1.18,19

Endoglin is present at high levels on the vascular endothelium in adults and is expressed early during development on vascular endothelium and on mesenchymal tissue derived from the endocardium.20 Stromal cells of mesenchymal origin,21 smooth muscle cells,22 the placental synctiotrophoblast, mesenchymal and hematopoietic stem cells, pre-erythroblasts, leukemic cells of lymphoid and myeloid lineage, and activated blood monocytes also express endoglin. In cardiac tissue, endoglin is expressed by the endocardium and fibroblasts, with minimal expression by cardiomyocytes.8,21

**Endoglin maintains vascular homeostasis**

In quiescent nonproliferative endothelium, TGFβ signal transduction is predominantly mediated by the type I TGFβ receptor, activin receptor-like kinase-5,23 and phosphorylation of signaling mediators known as Smads-2/3. Alternatively, TGFβ-mediated activation of the activin receptor-like kinase-1 pathway phosphorylates Smads-1/5/8, which promotes endothelial proliferation and migration24 (Figure 1). Endoglin negatively regulates TGFβ-mediated activin receptor-like kinase-5 signaling in quiescent endothelium, while promoting activin receptor-like kinase-1 signaling in activated endothelium.24,25 Conversely, in the absence of endoglin, activin receptor-like kinase-1 signaling is attenuated, while activin receptor-like kinase-5-mediated growth arrest of endothelial cells is stimulated25 (Figure 1). Li et al determined that suppressed endoglin expression enhances the inhibitor effects and further that endoglin overexpression promotes endothelial proliferation.26,27 These pathways have important implications for the development of arteriovenous malformations associated with HHT, as described below.

Further substantiating the role of endoglin in maintaining vascular homeostasis, cells from Endoglin heterozygous (Eng+/−) mice exhibit lower levels of endothelial nitric oxide synthase (eNOS), which results in less nitric oxide generation and more eNOS-derived superoxide production due to an uncoupling of eNOS activity.28 Endoglin may modulate vascular homeostasis via its colocalization to endothelial membrane-associated caveolae.26 Consistent with this observation, resistance arteries from Eng+/− mice demonstrate impaired myogenic responses and enhanced eNOS-dependent vasodilatation despite reduced eNOS levels. These findings may reflect increased stabilization of eNOS protein expression and enhanced calcium-induced activation of eNOS by endoglin.28,29 Subsequent data have also shown that TGFβ1 directly induces eNOS-dependent vasorelaxation, which is blocked by high circulating levels of soluble endoglin18 and may contribute to elevated vascular resistance.17 In addition to regulating eNOS activity, endoglin expression may correlate with myeloperoxidase activity and expression of vascular cell
adhesion molecule 1, inducible nitric oxide synthase, and CD68 during ischemia-reperfusion injury.10

Endoglin modulates cardiac development

The importance of endoglin in hematopoiesis, angiogenesis, cardiovascular development, and vascular remodeling has been well established11 and is supported by the observation that Endoglin null mice (Eng⁻/⁻) die at embryonic day 10–11.5 as a result of cardiovascular abnormalities.2,20,32,33 The critical role of endoglin in cardiovascular development is further supported by high levels of endoglin expression in the endocardial cushion during valve formation and heart septation.34 Originating from the lateral plate mesoderm, both cardiac myocytes and cardiac endothelial progenitor cells ultimately constitute the primitive spongy heart tube. A subpopulation of endothelial cells in the region of the future cardiac valve and septal formation transforms into mesenchymal cells, which migrate into the cardiac jelly and begin to express α-actin.35 A critical aspect of this process is the ability of the developing myocardium to induce endothelial transformation via endocardial-myocardial signaling pathways, which is mediated in part by signaling through TGFβ1, TGFβ3, and endoglin.35–37 The role of endoglin in valve formation was recently highlighted by a systems biology approach which identified a haplotype within the endoglin (ENG) gene that is strongly associated with the presence of a bicuspid aortic valve in patients.38

Regulators of endoglin expression

Endoglin is localized to human chromosome 9 and contains 15 exons of which 13 code for the extracellular domain. The 5′ flanking region of the gene lacks consensus TATA and CAAT boxes, but contains two regions rich in G-C and consensus motifs for SP1, ETS, GATA, AP-2, NFκB, MAD, and response elements for TGFβ, glucocorticoids, vitamin D, and estrogen.39 Importantly, both endoglin and type I collagen promoters contain Smad-binding elements in close proximity to Sp1 binding sites, suggesting a possible mechanism by which TGFβ1 may coactivate endoglin and type I collagen expression.40 Factors known to induce endoglin expression in various cell types include angiotensin II,41 TGFβ,42 hypoxia-inducible factor-1,43 hypercholesterolemia,44 and balloon-mediated vascular injury via the Kruppel-like zinc finger transcription factor, KLF6.4 The inflammatory cytokine, tumor necrosis factor-alpha (TNFα), attenuates endoglin expression by endothelial cells in vitro.26 Several pharmacologic agents also influence endoglin expression. Tranexamic acid is a competitive inhibitor of plasminogen activation to plasmin, and at much higher concentrations, a noncompetitive inhibitor of plasmin. In endothelial cells, tranexamic acid promotes endoglin gene and protein expression and TGFβ1 signaling via activin receptor-like kinase-1.45 Conversely, inhibitors of angiotensin-converting enzyme, such as trandolapril,46 and angiotensin type I receptor inhibitors, such as valsartan, losartan, and candesartan, attenuate angiotensin II-induced endoglin expression.42 The 3-hydroxy-3-methyl-glutaryl-CoA reductase inhibitor, atorvastatin, also decreases endoglin expression in mouse models of hyperlipidemia, which appears to correlate with the hypolipidemic effects of statin therapy.44,47

Acting as a biosensor for humoral and mechanical stimuli, the vascular endothelium plays a primary role in
maintaining local and systemic homeostasis under conditions of mechanical stress. As a central marker of endothelial cell integrity and function, the effect of hemodynamic forces and mechanical deformation on endoglin activity remains largely uncharacterized. Several lines of evidence suggest that endoglin expression is regulated by mechanical forces such as stretch and strain. First, the evolutionarily conserved zona pellucida domain in the extracellular region of endoglin is mechanically responsive in fruit flies.15 Second, in vivo models of placental hypertension and balloon-induced arterial injury are characterized by increased endoglin expression in vascular tissue.4,17 Third, endoglin expression is increased in hemorrhoidal tissue and correlates with an increased propensity for thrombosis, suggesting that increases in venular wall tension may modulate endoglin expression and vascular thromboreistance.48 Recent data showed that mechanically stretching myoblasts increases endoglin expression; a process which appears to be regulated by microRNA-208A.49

As a central regulator of cardiovascular homeostasis, a functional role for endoglin has been identified in several important clinical disease conditions, including HHT, pre-eclampsia, cardiac remodeling, cancer, pulmonary hypertension, and other fibroproliferative disorders. Several of these conditions are discussed in detail below.

**Hereditary hemorrhagic telangiectasia**

HHT, known historically as Osler-Weber-Rendu syndrome, is a disorder of vascular homeostasis affecting small and large vessels. The prevalence of HHT varies depending on the population studied, from as low as about 1:39,000 in northern England to as high as about 1:1330 in the Afro-Caribbean population of Curacao and Bonaire, with an average prevalence estimated at 1:5000–1:8000.50–53 The hallmark of HHT is the formation of arteriovenous malformations, which may occur in small vessels, leading to cutaneous telangiectases, nosebleeds, and gastrointestinal bleeds, or in large vessels of the lungs, brain, and liver. Important clinical sequelae of large-vessel arteriovenous malformations include hemorrhage, stroke, and brain abscess for pulmonary arteriovenous malformations, and pulmonary hypertension and high output heart failure arteriovenous malformations in the liver. Rupture of arteriovenous malformations, either spontaneous or associated with pregnancy or other physiological stressors, represents a significant source of morbidity in patients with HHT.54–57 The diagnosis of HHT is based on clinical observations that are summarized in consensus criteria known as the “Curaçao criteria.”56,58

Mutations in endoglin (ENG, OMIM#187300) and activin receptor-like kinase-1 (also known as ACVRL1, OMIM#600376) genes lead to HHT1 and HHT2, respectively, and account for >80% of cases.59 Mutations in Smad-4 (MADH4, OMIM #175050) are also associated with 2%–3% of cases with a combined juvenile polyposis-HHT syndrome.59 Two other loci have been shown to be in linkage disequilibrium with HHT; one on chromosome 5, defining HHT3, and the other on chromosome 7, defining HHT4.60,61 The precise genes involved remain to be identified and would give clues about the pathways deficient in HHT.

Transgenic mouse models of deficient expression of endoglin or ALK-1 recapitulate clinical features of HHT.62,63 In EngΔ/Δ mice, complete loss of endoglin is embryonically lethal due to impaired angiogenesis, while heterozygosity leads to the clinical syndrome of HHT1 with variable penetrance.62–64 In humans with HHT1, endoglin expression in peripheral blood monocytes and newborn umbilical vein endothelial cells is half of control levels and mutant proteins are rarely detected.65 These and other data support the role of haploinsufficiency in the pathogenesis of HHT1 as opposed to reduced local expression of endoglin through a “second hit” process or interference by the mutated allele with the function of the normal allele.59,62

Mutations in ALK-1 and to a lesser degree ENG are associated with pulmonary arterial hypertension, which can occur in the absence of HHT.64–66 Both endoglin and ALK-1 can associate with the BMP type II receptor, which is primarily responsible for most familial forms of pulmonary arterial hypertension.67 The mechanistic link between pulmonary arterial hypertension and HHT was recently studied in adult ALK-1+/− and Eng−/− mice, which develop signs of pulmonary arterial hypertension that can be attenuated by treatment with the antioxidant, tempol.67,68 These findings highlight the importance of eNOS-derived superoxide in these mice due to uncoupling of eNOS, which contributes to impaired vascular tone in either HHT or pulmonary arterial hypertension. Furthermore, endoglin and ALK-1 are coexpressed in the terminal-most segments of the pulmonary vasculature, which is most affected in mouse models and where most HHT-associated arteriovenous malformations tend to form.69 However, physiologically, pulmonary arterial hypertension is associated with increased pulmonary vascular resistance, while arteriovenous malformations are associated with abnormal vasodilatation.70

An important treatment option for patients with HHT is embolization of visceral arteriovenous malformations, which can prevent major adverse clinical events, including stroke,
high-output heart failure, pulmonary hypertension, and hemorrhage. Hormonal and antiangiogenic agents have also been explored as potential therapy for HHT. Clinical trials of estrogen preparations including the estrogen receptor antagonist, tamoxifen, and the selective estrogen receptor modulator, raloxifene, can reduce episodes of epistaxis and transfusion requirements.71,73,74 Inhibition of angiogenesis through the use of agents such as thalidomide, lenalidomide, and bevacizumab reduces the incidence of nasal and gastrointestinal bleeding in some patients.75–80 However, the lack of randomized controlled trial data for bevacizumab and the absence of long-term safety or efficacy data for antiangiogenesis therapy limits enthusiasm for these HHT treatment approaches, which remain experimental and limited to highly selected patients. The therapeutic action of thalidomide in HHT derives in part from increased platelet-derived growth factor expression at low doses and a direct antiangiogenic effect at high doses.77 Raloxifene and tranexamic acid increase in vitro mRNA and protein expression of endoglin and ALK-1 in endothelial cells.53,71 Whether future therapies such as gene transfer methods can target the underlying genetic defect associated with HHT remains unknown.51,82 Further insight into the role of endoglin and the TGFβ signaling pathway in the pathogenesis of HHT, together with evolving molecular approaches, may lead to novel therapeutic approaches for this debilitating condition.

**Pre-eclampsia**

Pre-eclampsia is a systemic disorder occurring in 3% of pregnant women after 20 weeks of gestation with symptoms of new-onset hypertension and proteinuria defined as a systolic blood pressure > 140 mmHg or diastolic blood pressure > 90 mmHg in a previously normotensive patient and at least 0.3 g of protein in a 24-hour urine specimen. Eclampsia is defined as pre-eclampsia with grand mal seizures without an identifiable cause.93,84 The incidence of maternal death associated with pre-eclampsia has been estimated at one in 100,000 live births.85 Maternal complications include central nervous system involvement, acute renal or liver failure, and hematological dysfunction.86 Incident pre-eclampsia is associated with increased risk of subsequent cardiovascular morbidity, including hypertension and ischemic heart disease, although it is not clear if pre-eclampsia is causal or reflects an underlying predisposition to cardiovascular disease.87 Fetal effects of pre-eclampsia include premature, fetal growth restriction, oligohydramnios, and placental abruption.88,89 Notable risk factors for the development of pre-eclampsia include a prior personal or family history of pre-eclampsia, advanced maternal age, obesity, and pregestational diabetes.90

The pathophysiology of pre-eclampsia remains poorly understood. Early changes in the placenta prior to the appearance of clinical manifestations of pre-eclampsia include failure of migrating placental cells to express endothelial surface adhesion markers, resulting in incomplete invasion of maternal arteries by the developing trophoblast.91–94 Pathological evidence of placental ischemia has been observed in some cases of pre-eclampsia,95 which is thought to trigger release of antiangiogenic factors, including soluble endoglin and soluble fms-like tyrosine kinase (sFlt1) into the maternal circulation.96–97 sFlt1 antagonizes the angiogenic factors, vascular endothelial growth (VEGF) factor and placental growth factor, while soluble endoglin antagonizes TGFβ1 and TGFβ3 activity.9,17,19,97 Furthermore, systemic disruption of vascular hemostasis caused by elaboration of these antiangiogenic factors promotes pre-eclampsia.95,98

The VEGF and TGFβ axes are the major signaling pathways implicated in the pathogenesis of pre-eclampsia (Figure 2). sFlt1, or the type 1 VEGF receptor, can be generated by either alternative splicing of the Flt1 gene or by cleavage of the Flt1 extracellular domain in response to hypoxia or thrombin.99,100 Elevated levels of sFlt1 in the maternal circulation correspond to a fall in VEGF and placental growth factor levels.97,101–103 A putative role for sFlt1 in pre-eclampsia is supported by studies showing that administration of exogenous sFlt1 to pregnant rats induces severe pre-eclampsia, while removal of sFlt1 by immunoprecipitation normalizes the angiogenic response of cells derived from placental villous explants.17,96 Similarly, pre-eclamptic placentae express increased levels of membrane-bound and soluble endoglin.17 A recent study demonstrated that oxysterol

![Figure 2 Circulating angiogenic peptides in pre-eclampsia.](https://www.dovepress.com/)

**Figure 2** Circulating angiogenic peptides in pre-eclampsia.

**Notes:** Soluble endoglin is released by ectodomain cleavage of membrane-associated endoglin, thereby disrupting TGFβ1 signaling. Soluble Flt1 is generated by either ectodomain cleavage of the VEGF receptor type I or alternative splicing of Flt-1 pre-mRNA and interferes with VEGF signaling.

**Abbreviations:** TGFβ1, transforming growth factor beta-1; PIGF, placental growth factor; VEGF, vascular endothelial growth factor; sEng, soluble endoglin; mRNA, messenger RNA.
activation promotes matrix metalloproteinase 14-mediated cleavage of soluble endoglin in cells of trophoblast origin,\textsuperscript{104} which suggests that hypoxia and oxidative stress are important triggers for release of soluble endoglin. Once in the circulation, soluble endoglin antagonizes TGF\(\beta\)-induced vasodilation, thereby promoting hypertension.\textsuperscript{28,105,106} A rise in circulating maternal soluble endoglin heralds the onset of symptoms in patients with pre-eclampsia.\textsuperscript{102,107,108} Individually and in concert with sFlt1, soluble endoglin induces a pre-eclamptic state in pregnant rats.\textsuperscript{17} Investigation into the regulation of sFlt and soluble endoglin in pre-eclampsia is ongoing.\textsuperscript{109}

At present, the primary mode of therapy for pre-eclampsia is accelerated delivery of the placenta. The available evidence strongly implicates an imbalance of the antiangiogenic factors, sFlt1 and soluble endoglin, and the proangiogenic factors, placental growth factor, TGF\(\beta\), and VEGF, in the pathogenesis of pre-eclampsia. Strategies targeting maternal depletion of soluble endoglin or sFlt1 would plausibly de-escalate the disease process. Given the growing appreciation for the role of sFlt1 and soluble endoglin in the pathophysiology of pre-eclampsia, novel therapeutic approaches include the use of antibodies and small molecules to sequester or limit synthesis of antiangiogenic molecules. Alternatively, modulating the balance of angiogenic factor levels has been recently explored in a study that showed improvement in blood pressure and renal function after administration of exogenous VEGF in a preclinical model of pre-eclampsia.\textsuperscript{110–112} More recently, directly removing sFlt1 from the maternal circulation by extracorporeal apheresis using a dextran sulfate column has been introduced as a potential therapeutic approach for pre-eclampsia.\textsuperscript{113} Other approaches include induction of hemoxygenase-1 with cobalt protoporphyrin in a rat model of pre-eclampsia\textsuperscript{114} and direct inhibition of matrix metalloproteinase 14 to prevent the release of soluble endoglin.\textsuperscript{115,116} Limiting fetal consequences of any treatment approach is mandatory and remains a challenging limitation to the development of new treatments.

**Cardiac remodeling**

Heart failure is a major cause of global morbidity and mortality that affects nearly 23 million individuals worldwide.\textsuperscript{117,118} Heart failure commonly occurs secondary to hypertensive heart disease, myocardial infarction, infection, and inherited cardiomyocyte dysfunction. A decline in cardiac function activates several signaling cascades that promote cardiomyocyte hypertrophy and cardiac fibrosis, a process known as cardiac remodeling.\textsuperscript{119} At each phase of cardiac remodeling from acute load to compensatory hypertrophy, various signaling cascades are implicated. Among these, TGF\(\beta\) promotes the cardiac hypertrophy and fibrosis associated with heart failure. In response to angiotensin II, TGF\(\beta\) is increased, converts fibroblasts into myofibroblasts, and generates extracellular matrix proteins, such as type I collagen.\textsuperscript{120} Excess collagen deposition exaggerates mechanical stiffness of the left ventricle, impairs myocyte contractility, disrupts electrical coupling, and worsens tissue hypoxia,\textsuperscript{119} thereby promoting heart failure.

TGF\(\beta\)-induced fibrosis involves both canonical and noncanonical signaling (Figure 3). Among the canonical pathways, Smad-3 plays a central role.\textsuperscript{120} First, Smad-3 mediates TGF\(\beta\)-induced activation of a TGF\(\beta\)-response element located in the promoter region of type I collagen.\textsuperscript{121} Second, Smads-2/3 and Smad-4 form a complex with activated transcription factor-2 to stimulate collagen synthesis. Activated transcription factor-2 phosphorylation requires activation of TGF\(\beta\)-activated kinase.\textsuperscript{122} Third, Smad-3 mediates

![Figure 3 Canonical and noncanonical transforming growth factor beta-1/endoglin signaling in fibrosis.](https://www.dovepress.com/)

**Figure 3** Canonical and noncanonical transforming growth factor beta-1/endoglin signaling in fibrosis.

**Notes:** On binding of transforming growth factor beta-1 to a heteromeric complex of receptors including type I, type II, and endoglin, downstream signaling via Smad-dependent or Smad-independent pathways occurs. Smad-3 promotes collagen synthesis by activating transforming growth factor beta-1 activated kinase or CTGF. Smad-3 can also directly activate collagen synthesis by binding a Smad-binding element within the collagen promoter. Noncanonical signaling can stimulate collagen synthesis via several mitogen-activated protein kinases including ERK, JNK, or p38 kinase.

**Abbreviations:** CTGF, connective tissue growth factor; ERK, extracellular regulated kinase; JNK, jun-n-terminal kinase; Eng, endoglin; MAPK, mitogen-activated protein kinase.
TGFβ1-induced expression of connective tissue growth factor via a TGFβ1 response element located in the connective tissue growth factor promoter. Connective tissue growth factor is a critical downstream mediator of tissue fibrosis. Noncanonical pathways contributing to TGFβ1-induced collagen synthesis involve activation of mitogen-activated protein kinases including: extracellular regulated kinases, jun-n-terminal kinases, and p38 kinase. While all three cascades are activated in advanced heart failure, the extracellular regulated kinase pathway is required for TGFβ1-induced connective tissue growth factor expression and collagen synthesis. In cardiac fibroblasts, connective tissue growth factor is an important downstream mediator of TGFβ1-induced fibrosis because it promotes binding of TGFβ1 to TBR2 and blocks inhibition of the pathway by Smad-7. Given its central role in stimulating fibrosis, TGFβ1 has been nonselectively targeted in heart failure models using multiple approaches, none of which have had clearly beneficial therapeutic effects.

Recently, endoglin expression was found to be increased in left ventricular tissue from patients with end-stage heart failure and in a murine model of thoracic aortic constriction-induced heart failure. Further analysis showed that endoglin is highly expressed by cardiac fibroblasts and endothelium, but poorly on cardiomyocytes. Compared with Eng+/+ wild-type mice, Eng+/- mice demonstrated improved survival, limited cardiac fibrosis, and enhanced myocardial capillarity after thoracic aortic constriction (Figure 4). Loss-of-function studies in vitro confirmed the dependence of TGFβ1 activity on endoglin expression in human cardiac fibroblasts. Paradoxically, adenoviral-mediated overexpression of full-length endoglin also blocked TGFβ1-induced collagen synthesis. Further study showed that levels of soluble endoglin were elevated in the conditioned medium after treatment with the adenovirus, thereby implicating soluble endoglin as a negative regulator of TGFβ1 activity. This observation was confirmed by adenoviral-mediated overexpression of human soluble endoglin or treatment with recombinant human soluble endoglin in vitro. Treatment with adenoviral-mediated overexpression of human soluble endoglin in wild-type mice after thoracic aortic constriction. These findings suggest that endoglin is required for TGFβ1 signaling in cardiac fibroblasts and that selectively inhibiting TGFβ1 signaling by reducing endoglin activity attenuates cardiac fibrosis and improves survival in a mouse model of heart failure. In contrast with the functional role of endoglin in promoting TGFβ1 signaling, soluble endoglin limits TGFβ1 signaling, type I collagen synthesis, and ultimately cardiac fibrosis.

**Figure 4** Functional role of endoglin in cardiac fibrosis.

**Notes:** In heart failure, increased ventricular pressure overload stimulates TGFβ1 expression. In mice with normal endoglin expression (Eng+/+), endoglin promotes TGFβ1-induced type I collagen synthesis and cardiac fibrosis. Reduced endoglin expression in Eng−/− mice attenuates TGFβ1-induced pSmad-2/3, type I collagen expression, and cardiac fibrosis.

**Abbreviation:** TGFβ1, transforming growth factor beta-1.
In a separate study, circulating levels of soluble endoglin were significantly increased in patients with suspected left ventricular dysfunction and were strongly correlated with predictors of mortality in heart failure, such as elevated left ventricular end-diastolic pressure, reduced left ventricular ejection fraction, and worsening New York Heart Association class symptoms. Soluble endoglin levels demonstrated superior sensitivity, specificity, accuracy, and predictive value compared with atrial and brain natriuretic peptides for identifying subjects with increased left ventricular end-diastolic pressure. Furthermore, levels were reduced in patients receiving medical therapy for congestive heart failure and correlated with reduced cardiac filling pressures. These findings identify elevated soluble endoglin levels as a biomarker of increased cardiac filling pressure and perhaps a potential biomarker of endoglin activity in heart failure. These findings also represent an important paradox in endoglin biology. Expression of membrane-associated endoglin is increased in heart failure and correlates with a direct increase in circulating soluble endoglin. However, membrane-associated endoglin promotes adverse cardiac remodeling by driving cardiac fibrosis, while soluble endoglin may serve as a negative feedback loop to limit TGFβ1 signaling. The exact functional role of soluble endoglin remains to be determined, but appears to serve as a potentially important diagnostic marker of heart failure as well as a participant in the cardiac remodeling associated with heart failure.

The role of soluble endoglin in heart failure remains poorly understood. First, the mechanism underlying increased expression of soluble endoglin in heart failure is unknown. Proteolytic cleavage of soluble endoglin from membrane endoglin may occur both locally in cardiac tissue or systemically because levels of matrix metalloproteinase 14 are known to be elevated in heart failure. Second, the mechanism by which higher soluble endoglin levels interrupt TGFβ1 signaling remains poorly characterized. First, endoglin may modulate signaling via several TGFβ-family ligands and soluble endoglin could serve as a ligand trap for ligands of the TGFβ superfamily. However, recent studies indicate that BMP9 and BMP10 may be the only ligands that bind to soluble endoglin with high affinity. Second, soluble endoglin may promote alternate signaling pathways that indirectly inhibit TGFβ1 signaling, such as BMP7. Finally, release of soluble endoglin by ectodomain shedding may render the receptor nonfunctional by removing the primary active component of endoglin, the extracellular domain, and leaving behind a nonfunctional cytoplasmic tail that cannot interact with TBR2.

Further supporting the role of endoglin in cardiac remodeling, studies of acute myocardial infarction have shown that Eng−/− mice exhibit showed neoangiogenesis in the infarct zone, progressive left ventricular dilatation, worsening cardiac function, and elevated cardiac filling pressures compared with wild-type controls. In this model, adverse cardiac remodeling in Eng−/− mice was partially rescued by intravenous injection of mononuclear cells from healthy human donors with normal endoglin expression but not by those from patients with HHT1 harboring mutant endoglin.

In contrast with heart failure, circulating levels of soluble endoglin in patients presenting with acute myocardial infarction were significantly lower than in healthy controls, and decreased further in the first 48 hours of admission. Reduced levels of soluble endoglin were an independent predictor of short-term and long-term cardiovascular mortality. Unlike other known circulating angiogenic proteins, including sFlt1 and VEGF, levels of soluble endoglin are not altered by treatment with the anticoagulants commonly used to manage patients with myocardial infarction. Whether alterations in soluble endoglin levels correlate with atherosclerotic burden remains under investigation. Collectively, these data suggest that both membrane-associated and soluble endoglin serve important roles in cardiac remodeling due to heart failure or acute myocardial infarction, and may serve as potential targets of therapy for these debilitating conditions.

In summary, the role of endoglin as a critical mediator of cardiovascular homeostasis has been well established over the past two decades. Studies have now begun to explore the potential utility of targeting endoglin and the signaling program it regulates as a novel therapeutic approach for patients suffering from cardiovascular diseases, including HHT1, pre-eclampsia, pulmonary hypertension, heart failure, myocardial infarction, atrial fibrillation, pulmonary hypertension, and atherosclerosis.

**Disclosure**

The authors report no conflicts of interest in this work.

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