

Porous microsphere and its applications

Yunpeng Cai^{1,2,*}

Yinghui Chen^{3,*}

Xiaoyun Hong²

Zhenguo Liu¹

Weien Yuan²

¹Department of Neurology,
Xinhua Hospital affiliated to
Shanghai Jiaotong University
School of Medicine, ²School
of Pharmacy, Shanghai JiaoTong
University, ³Department of Neurology
Jinshan Hospital, Fudan University,
Shanghai, People's Republic of China

*These authors contributed equally
to this work

Abstract: Porous microspheres have drawn great attention in the last two decades for their potential applications in many fields, such as carriers for drugs, absorption and desorption of substances, pulmonary drug delivery, and tissue regeneration. The application of porous microspheres has become a feasible way to address existing problems. In this essay, we give a brief introduction of the porous microsphere, its characteristics, preparation methods, applications, and a brief summary of existing problems and research tendencies.

Keywords: pore, porosity, porogen, suspension polymerization, seed swelling, pulmonary drug delivery, tissue regeneration

Introduction

Porous microspheres have external pores on the surface or internal pores in the core (usually interconnective), and active substances can be dissolved or dispersed on the surface or in the core of the microspheres. In the 1950s, cross-linked polystyrene with a macroporous structure in the dry state was prepared by a novel polymerization method.¹ Little progress was made until the 1990s when porous microspheres were identified as suitable materials for potential applications including as carriers for drugs, high speed chromatography, cell delivery, and tissue regeneration as a scaffold. Compared with traditional microspheres, they show many unique properties such as special behavior of drug absorption and drug release kinetics, large specific surface area, and low density.

What differentiates porous microspheres from traditional microspheres are the pores on the surface and inside. Porosity plays an important role in determining the capacity efficiency and release kinetics. The diameter, the amount, and the structure of the pores are the crucial factors affecting the properties of porous microspheres.² Most applications of porous microspheres are based on the porous structure (porosities, pore sizes, surface areas, and so on). To control the pore structure parameters is therefore of particular importance.

Characterization of porous microspheres

Size and morphology of porous microspheres

Porous microsphere morphology can be directly observed with microscopy or scanning electron microscopy (SEM). The diameter of microspheres can be determined by dynamic light scattering. Figure 1 shows typical porous microspheres.

Correspondence: Weien Yuan
School of Pharmacy, Shanghai Jiao Tong
University, 800 Dongchuan Road,
Shanghai 200240,
People's Republic of China
Tel +86 21 3420 5072
Email yuanweien@126.com

Zhenguo Liu
Department of Neurology, Xinhua
Hospital Affiliated To Shanghai Jiao Tong
University School of Medicine, 1665
Kongjiang Road, Shanghai 200092, People's
Republic of China
Tel/fax +86 21 6579 0000
Email zhenguoliu2004@yahoo.com.cn

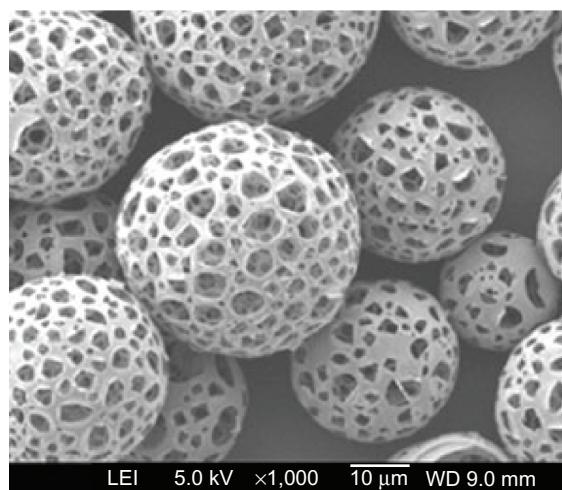


Figure 1 Porous microspheres.

Note: Springer, *Colloid and Polymer Science*, 291, 2013, [page 119]. Double emulsion-templated microspheres with flow-through pores at micrometer scale, Chen SF, Gao F, Wang QB, Su ZG, Ma GH, [Figure 3C], [License Number 3095270116326], with kind permission from Springer Science and Business Media.⁶⁸

Pore size of porous microspheres

Pore size is one of the most important features which influences the application of porous microspheres. Usually pore size can be determined by SEM, transmission electron microscopy (TEM), or confocal laser scanning microscopy (CLSM).

According to the International Union of Pure and Applied Chemistry classification,³ porous materials can be classified according to their pore sizes: microporous materials (less than 2 nm), mesoporous materials (2–50 nm), macroporous materials (50–200 nm), and gigaporous materials (more than 200 nm). The diversities in pore sizes meet requirements in many practical applications and are intensively studied for their promising virtues.

The pore size is an important factor in determining the adsorption and release of biomolecules. In the field of enzyme immobilization, many studies have proved that pore size of heterogeneous support is probably the most important parameter and have indicated that the ideal size of the pores is three to five times that of the protein size.⁴

As pore size plays an essential part in the porous microspheres, some efforts have been made to regulate the pore size, but there's still no universal method to deal with this problem. In many studies, pore size was controlled to some degree by choosing suitable porogens or by adjusting their concentrations. Only a few studies have tried to discover a universal technology. Cheng et al employed CO₂ bubbles as templates and precisely adjusted the system pressure.⁵ Their carbonated hydroxyapatite microspheres' pore sizes could be well regulated over a wide range from the microscale to the nanoscale. Their results

suggested that the gathering and growth of the CO₂ bubbles generated the formations of the porous structure, and with the increase of system pressure, the surface of the microsphere got denser and the flakes, which assembled the porous structure, became smaller.⁵ Li and Zhang developed a new method to produce a porous polymer film with size-tunable pores on its surface.⁶ The porous film was produced using polystyrene microspheres as a template and the surface pore size could be tuned without changing the size of the template microsphere. Polystyrene microspheres were self-assembled on a substrate precoated with a thin layer of polystyrene and heated at an appropriate temperature, prior to infusion of a polymer precursor solution into cavities of the template microspheres, solidification of the polymer, and removal of the microspheres. The surface pore sizes could be tuned by changing the temperature and time for the heat treatment.

Surface area and porosity of porous microspheres

The specific surface area of porous microspheres can be investigated by nitrogen adsorption/desorption analysis, and a Brunauer–Emmett–Teller absorption model is often adopted.⁷ The porosity equals the ratio of pore volume to total volume, which can be investigated by Barrett–Joyner–Halenda (BJH) measurement. In this method, N₂ adsorption–desorption is carried out at 77K. According to the BJH equation, both the pore size and pore volume can be calculated.⁸ Mercury-intrusion porosimetry is also used to investigate the porosity, in particular for relatively big pore size microspheres.⁹

The porosity and specific surface area have a direct relationship with the microsphere's density (for pulmonary delivery) and drug absorption amount, and a controllable porosity is preferable. Generally, in cases with porogens, porosity can be regulated by carefully adjusting the amount of porogen and the volume of porogen particulates.¹⁰

Materials for preparing porous microspheres

Framework material

Many materials employed in traditional microspheres can also be employed in preparing porous microspheres. The most commonly employed materials are polymers such as polylactic acid, poly(lactic-co-glycolic acid) (PLGA), chitosan, polycaprolactone (PCL), divinylbenzene, and other polymers or co-polymers.

Besides polymers, some other materials also have some superiority at fabricating porous microspheres, including both organic and inorganic materials such as calcium carbonate (CaCO₃), mesporous silica, hydroxyapatite, and biodegrad-

able porous starch foam.^{11–13} These materials have merits like biocompatibility or have a higher porosity or can be easily modified.

Porogen

Porogen is also called pore-forming agent. In most experiments, it's widely employed to form the pores. Porogens are often used in solvent methods and polymerization methods, which are the most common methods to prepare porous microspheres.

The mechanism of porogens can usually be described as solvent casting/particulate leaching, in which porogen is usually an unreactive agent added into the materials for porous microsphere preparation. Once the porogen particulates are removed, the spaces they leave become the pores both externally and internally. However, the process that requires the porogens to be extracted later causes other problems. The pore-forming strategy involving these porogens has inherent limitations: leaching with solvents facilitates diffusional mass exchange, which removes not only porogens but also encapsulated actives. Moreover, the high-shear inherent in

emulsification leads to microspheres with broad distributions in size and morphology and this can be further exacerbated by the inclusion of these porogens. Also, the porogens usually require a long time to leach out completely.^{14–16}

Though with some drawbacks, fabricating porous microspheres with a porogen is widely reported. The commonly used porogens are effervescent salts like ammonium bicarbonate, inorganic salts like sodium chloride, hydrocarbon waxes, linear polymers, carbohydrates, gelatin, ice, and sugar.^{17–19}

Figure 2 shows how the pores form with porogen.²⁰

Methods for preparing porous microspheres

Solvent evaporation method

The most widely used method to fabricate porous microspheres is the solvent evaporation method. Multi-emulsion solvent evaporation is commonly employed for the influx between the internal aqueous phase and external aqueous phase, and this influx forms pores easily. In this typical method, dissolved or dispersed polymer is added into a volatile organic phase, and it

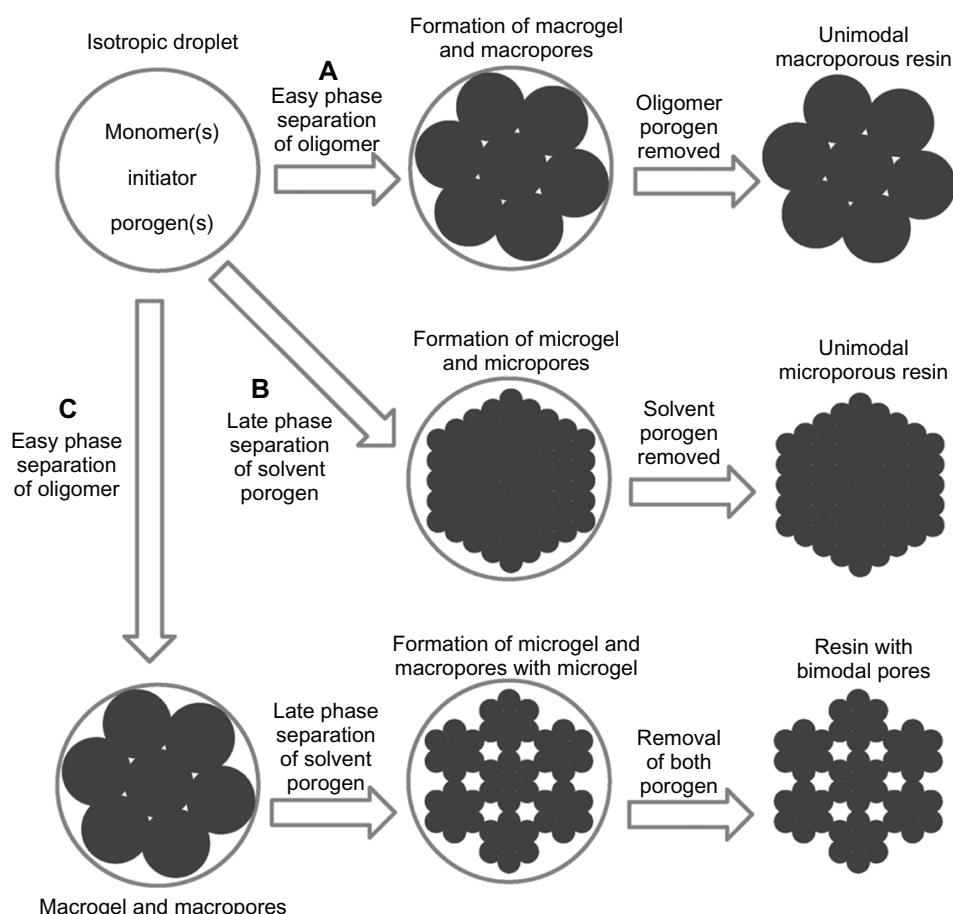


Figure 2 Schematic diagram of pore formation.

Note: Copyright © 2007, Taylor & Francis Ltd. Reproduced with permission from Liu QQ, Wang L, Xiao AG. Research progress in macroporous styrene-divinylbenzene co-polymer microspheres. *Designed Monomers and Polymers*. 2007;10(5):405–423.²⁰ <http://www.informaworld.com>.

is emulsified in an aqueous phase with a surfactant. The volatile solvent is removed as it diffuses into the aqueous medium and evaporates at the air/water interface under constant stirring, leaving only spherical polymer microspheres in the aqueous phase. Usually the drug is added in with the polymer, so that the drug is entrapped or imbedded in the polymer.

In this method, the diffusion of the inner phase can contribute to the formation of the pores. But to obtain an optimum porosity, porogens are often employed, which can adjust the porosity and the diameter of the pores more easily. The drawbacks of employing porogens have been discussed above. Another drawback of this method lies in the use of organic solvents which must be fully removed to avoid any possible damage concerning applications *in vivo*.

It has been reported that in a solid-in-oil-in-water emulsion system, the addition of the solid phase in the system is a key factor in forming the porous structure. The affinity between the solid and oil phases influences the speed of the oil phase diffusion. Results show that high affinity between solid-polymer and polymer-medium, and low affinity between solid-medium are the best combinations to obtain porous microspheres.²¹

A novel type of porous microsphere characterized by a dense core and a porous layer with an assembly of cone-like pores was reported.²² In the presence of NaCl, particles like hydroxyapatite can work as emulsifiers. Furthermore, hydrophilic particles such as metal oxides tend to stabilize oil-in-water emulsions while hydrophobic particles such as carbon tend to stabilize water-in-oil emulsions.²³

Large osmotic pressure differentials between the inner and outer aqueous phases can also be used to produce macroporous microspheres, because of the large influx of the inner liquid phase driven by the pressure gradient. Thus the polymer dissolved in the inner aqueous phase is a key to control porosity. The molecule weight and concentration can be controlled to fabricate microspheres with a desired porosity. Owen et al used high molecular weight poly(lactide-co-glycolide) at a concentration of 100 mg/mL in the internal aqueous phase and fabricated macroporous microspheres, with a particle size range of 5–20 µm and pore sizes of 1–5 µm.²⁴ Furthermore, by altering the inner aqueous volume fraction, extremely different porous and nonporous shell layer morphologies can be modulated.¹¹

Polymerization method

Suspension polymerization, precipitation polymerization, and emulsification polymerization methods are widely adopted to prepare porous microspheres. Sometimes they can be classified as phase separation methods.

In suspension polymerization, the reaction mixture consists of two phases, a liquid matrix and monomer droplets. The monomer and initiator (like crosslinker) are insoluble in the liquid phase, so they form drops within the liquid matrix. A suspension agent is usually added to stabilize the monomer droplets and hinder monomer drops from coming together. Micron-size monodispersed microspheres can be prepared with this method. Jayakrishnan and Thanoo prepared porous beads of poly(2-hydroxyethyl methacrylate) with the suspension polymerization method.²⁵

Precipitation polymerization is similar to the suspension polymerization method. The difference lies in the fact that precipitation polymerization gives larger and less regular particles, as a result of little or no stabilizer being present. In this method, the monomer and initiator are soluble, and upon initiation the formed polymer is insoluble and thus microspheres precipitate. Li and Stöver prepared porous poly(divinylbenzene) microspheres with precipitation polymerization, which have diameters between 4–7 µm, total pore volumes of up to 0.52 cm³/g, and surface areas of up to 800 m²/g.²⁶

In emulsification polymerization, an emulsion, monomer, and a surfactant are needed. The most common type of emulsification polymerization is an oil-in-water emulsion, in which droplets of monomer (the oil) are emulsified (with surfactants) in a continuous phase of water. Then the surfactant stabilizes the surface between microspheres and the continuous phase, and the charge of the surfactant stops the coagulation of microspheres. It has been reported that in some cases a surfactant is not required to realize emulsification polymerization.^{27,28} Sometimes an emulsifier is not a must in emulsification polymerization. In an experiment, Okubo et al fabricated a ‘golf ball-like’ microsphere with many dents on it by seeded emulsion polymerization without an emulsifier.²⁹

Seed swelling method

Monodispersed porous polymer microspheres can be prepared using the seed swelling method. The reaction system consists of seed, monomer, dispersed phase, initiator (crosslinker), and stabilizer.

A seed polymer is employed, which is often prepared by the suspension polymerization method. This seed acts as a template that retains its shape as it is swollen by the entrance of monomers and solvents. However, this procedure is time-consuming and also difficult to carry out compared with other methods, and the condition of the seed is vital. Some researchers have modified this seed swelling method into two

steps, which simplifies the process, although the process is still time consuming to prepare well monodispersed porous microspheres.^{30–32}

Generally, the seed swelling method can be classified as below:

1. One-step seed swelling method: In this method, seeds are prepared by suspension polymerization. Then in a polymer emulsion, in the presence of a crosslinker, monomers aggregate on the seeds as the seeds ‘swell’ into microspheres. A porous structure is formed by the separation of porogens (also called diluents), which are added in the suspension polymerization step. However, this method cannot prepare large porous microspheres (diameter above 10 µm).^{33,34}
2. Stepwise seed swelling method: The difference is that the swelling process is repeated many times. Though large microspheres can be prepared, it is rather time-consuming.³¹
3. Two-step seed swelling method: This method allows an enormous increase of absorption capacity in the swelling step. Thus, microspheres with large size can be prepared. By making the seeds absorb a water insoluble, relatively low molecular weight compound in the first step, the absorption capacity in the second swelling step can be increased.³⁵ A similar experiment used acetone as carriers to facilitate the diffusion of monomers into the seeds.³⁶
4. Dynamic seed swelling method: This method can also increase the absorption of monomers by simply changing the temperature of the system in the swelling process. Large monodispersed microspheres can be prepared.³⁷

Sinter method

In this method, porous microspheres are fabricated by the bonding of materials, and some materials are burned off under high temperature while the heat-resistant materials are left to form the matrix of microspheres. For example, hydroxyapatite is often used in orthopedic or bone surgery due to its biocompatibility, non-toxicity, and osteoconductivity. Hydroxyapatite is mixed with a chitosan solution, with a surfactant to stabilize the surface and glutaraldehyde to make the microspheres harden under constant stirring. When the microspheres form, they are dried at 60°C for 3 hours. These chitosan-bonded spheres are heated initially at 500°C for 1 hour to burn off the chitosan and are finally sintered at 1100°C for 1 hour, at which point the porous microspheres are completed. The pores consist of the burned-off chitosan in the hydroxyapatite’s structure.³⁸

Synthesis method

This method is often seen in preparing mesoporous silica microspheres. Silica and other metal oxides with uniform mesoscale pore channels (typically in the range of 2–10 nm) are usually employed. The uniform pore channels and narrow pore size distribution are ideal properties for porous microspheres.³⁹ Usually, inorganic pressure and organic templates are synthesized into mesoporous materials. For a typical synthesis, BmimBF₄ is added into a Na₂SiO₃ · 9H₂O solution, then hydrochloric acid is added, all while under shaking. The mixture is aged for 5 hours at room temperature under quiescent conditions and then transferred into an oven at 80°C for another 60 hours. The product is collected by centrifugation and washing with deionized water and ethanol several times. Finally, the solid product is calcined at 550°C for 4 hours to remove impurities.⁴⁰ Synthesis with an organic template method in an oil-in-water phase has also been reported.⁴¹

Phase separation method

In this method, the first step is usually to prepare a polymer solution with the substances to be encapsulated, and add in other substances or solvents that lower the polymer’s solubility, then the polymers which encapsulate the drugs separate in a new condensed phase. The pores form with the solvent diffusion. Mandal et al prepared PLGA porous microspheres with the phase separation method, and got a higher encapsulation rate (58%) than the water-in-oil-in-water solvent evaporation method (16%).⁴²

Spray drying method

In this method, polymers and the substances to be encapsulated are all dissolved or suspended in a liquid. When the liquid is sprayed out by a nozzle, heated air will make the liquid of the drops evaporate, leaving the polymer and substances to form microspheres. Straub et al prepared porous PLGA microspheres by spray drying, which had an average diameter of 2.3 µm.⁴³ Ammonium bicarbonate was employed as a porogen.

Other methods

Preparation methods combined with new technologies have also been reported in recent years. For example, with microfluidic techniques, the polydispersity in particle size can be significantly reduced.¹⁴ With molecular imprinting technology, molecularly imprinted polymers can afford specific recognition against an imprinting molecule and moderate recognition against the structurally-related compounds.^{44,45}

Baimark prepared porous MPEG-b-PCLDLL microspheres by a melt dispersion method.⁴⁶ Any organic solvents and surfactants can be neglected for this method.

Table 1 provides a summary of the porous microsphere preparation methods discussed here.

Applications of porous microspheres

Tissue regeneration scaffolds

Porous microspheres can form a higher structure for versatile applications by methods such as thermally induced phase separation, microsphere sintering, electrospinning, and hydrogelling. In this situation, a highly porous structure with well interconnected pores is required not only to achieve sufficient cell seeding density within the scaffold, but also to

facilitate in- and out-transport of nutrients and oxygen for subsequent cell proliferation and differentiation.⁴⁷

The use of three-dimensional porous microspheres as scaffolds in tissue engineering is widely reported. Usually the porous microspheres are heated just above their glass transition temperature to sinter into porous scaffolds. However, it's reported that heat sintering is not applicable across a broad spectrum of polymer types due to its dependence on specific physicochemical properties such as: glass transition temperature, specific heat, crystallinity, viscosity, and surface tension of the polymer to be sintered.⁴⁸

The porous scaffolds take on a porous structure and facilitate the adhesion and proliferation of cells and usually they carry some drugs like cell growth factors to assist the proliferation of cells during tissue regeneration. Both the

Table 1 Summary of porous microsphere preparation methods

	Particle size	Advantages	Disadvantages
Preparation method			
Solvent evaporation method	0.1–1000 µm	Easy to prepare	Diffusion of internal aqueous phase or oil phase affects pore size and porosity
Polymerization method			
Emulsification polymerization	100 nm to 1 mm	Fast reaction speed; low viscosity of medium; narrow size distribution; controllable reaction temperature	Surfactants are difficult to remove
Suspension polymerization	100 nm to 5 mm	Narrow size distribution; controllable reaction temperature	Suspension agents are difficult to remove
Precipitation polymerization	100 nm to 5 mm	No suspension agents; narrow size distribution; controllable reaction temperature	Relatively larger and less regular compared with suspension polymerization; sensitive to reaction condition
Seed swelling method			
One step seed swelling	0.1–10 µm	Monodispersity	Cannot prepare large porous microspheres over 10 µm; time consuming
Stepwise seed swelling method	1–100 µm	Monodispersity; larger size compared to one step seed swelling	Time consuming; complicated
Two-step seed swelling method	1–100 µm	Monodispersity; larger size compared to one step seed swelling	Complicated; time consuming
Dynamic seed swelling method	1–100 µm	Monodispersity; simpler than two-step seed swelling	Time consuming
Phase separation method	1–1000 µm	Easy	Sensitive to reaction conditions; poor monodispersity
Spray-drying method	10–1000 µm	Suitable for industry; easy	Not many ways to form a porous structure and control the pore size; not fit for heat-sensitive substances
Sinter method	0.1–1000 µm	Porous microspheres can be sintered into a three-dimensional scaffold with a controllable porosity and pore size	Not fit for heat-sensitive substances; microspheres' adhesion during sinter process
Synthesis method	0.1–1000 µm	Uniform pore channels and narrow pore size distribution	Here synthesis method only refers to mesoporous microspheres preparation; complicated

pores between the microspheres and the smaller pores on the microspheres' surface offer room for the vessels and cells ingrowth.^{49,50}

The sinter method is the most frequently used method to prepare a porous microsphere scaffold. However, the best pore parameters for tissue engineering are different depending on what the tissue is. For example, to enhance osteogenic signal expression and support new bone formation, a scaffold's best parameters are not fixed (see Table 2).⁵¹

Table 2 provides a comparison of the effects of porosity and pore size on osteogenic signal expression and differentiation.^{52–61}

Table 2 The effect of porosity and pore size on osteogenic signal expression and differentiation

Scaffold materials	Pore parameters' effects on tissue engineering
Collagen–GAG	Chondrocytes synthesized more GAG in a 20 µm pore than a 85 µm pore. Improved migration in pore sizes larger than 300 µm.
Calcium phosphate	Higher pore sizes (280 µm) resulted in greater osseointegration. Pore size also affected blood vessel formation. Bone growth was affected more by pore size than porosity. Larger pore size induced greater bone formation.
β-TCP	ALP expression was affected by size of pores and porosity of scaffolds. Higher mineralization with higher porosity.
PLGA	100 µm solid groups had significantly greater percentage of bone ingrowth compared with 500 µm hollow groups of new bone formation in a cranial defect.
Polycaprolactone	Pore sizes between 350 µm and 800 µm had limited roles in bone regeneration after 8 weeks of implantation subcutaneously in mice. The scaffold section with 380–405 µm pore size showed better cell growth for chondrocytes and osteoblasts, while the 186–200 µm pore size was better for fibroblasts growth. The 290–310 µm pore size showed faster new bone formation.
EH–PEG	The 100 µm group had a faster rate of ALP (alkaline phosphatase, an early marker for osteoblastic differentiation) expression compared with the 250 µm group.
HA	Higher ALP expression of human MSCs in smaller pore sizes (200 µm). Higher proliferation in larger pore sizes (500 µm).

Note: Data from.^{52–61}

Abbreviations: β-TCP, beta tricalcium phosphate; ALP, alkaline phosphatase; EH–PEG, 5-ethyl-5-(hydroxymethyl)-beta,beta-dimethyl-1,3-dioxane-2-ethanol-co-poly(ethylene glycol); GAG, glycosaminoglycan; HA, hydroxyapatite; MSC, mesenchymal stem cell; PLGA, poly(lactic-co-glycolic acid).

High-speed protein chromatography

Due to the huge specific surface area, porous microspheres are fit for adsorption and desorption substances. Thus a separation of substances can be realized. Many studies have indicated that porous microspheres could be used for high-performance protein chromatography with high column efficiency, high dynamic capacity, and low flow resistance due to their porous nature. During chromatography, the mobile phase can flow through the macropores or the superpores in the solid phases, thus leading to the enhancement of mass transport rate of solute in the materials. On the other hand, the micropores connecting the macropores or superpores provide a large surface area for solute binding.⁵² Employing novel porous microspheres, an efficient low-pressure preparative chromatography for purification of icariin from crude extract has been developed. The microspheres column even demonstrated a higher resolution and better selectivity than the C18 column.⁵³

The porous microspheres applied in chromatography have a size range of 5–10 µm, and a high porosity is preferred while keeping rigid. Generally, the macropores among the microspheres are 600–800 nm, and the micropores on the porous microspheres are 80–150 nm. Keeping the micropores' depth under 1000 nm can provide a high separation speed.⁵⁴

Carriers for biomacromolecules or other substances

Carriers loaded with biomacromolecules have potential applications in tissue engineering, drug delivery, and cell therapy. Biomacromolecules here often refer to proteins, peptides, and genes. These molecules can be absorbed by porous structures due to the large specific surface area, with which a high drug loading amount can be achieved.

The mechanism of the absorption process can be divided into physical and chemical mechanisms. The physical absorption is a weak interaction and usually reversibly driven by a hydrophobic interaction or hydrogen bond. In most cases, we have to take desorption into consideration, so that physical absorption is a better choice. The chemical absorption by contrast is a strong interaction which usually involves chemical reactions, and covalent bonds or ionic bonds form after the chemical reaction.

The pore size plays an essential role in the molecules' absorption and desorption as mentioned before. Li et al investigated how the pore size affects the lipase distribution, thermal stability, storage stability, and reusability.⁵⁵ Their results showed that the thermal stability, storage stability, and reusability were all improved significantly with increased

pore sizes; while Michaelis constant values were lower in large pore size microspheres.

To achieve better absorption and desorption results, modifying the materials like polymers is often adopted. Wang et al modified polymers with polyethylene glycol; their results showed excellent adsorption and desorption properties with bovine serum albumin and trypsin.⁵⁶

Porous microspheres can provide a high loading efficiency and a good release behavior. However, one of the biggest problems is how to keep the bioactivity of biomolecules. Some researchers have worked on this in recent years. One key factor is to protect the biomacromolecules from the changeable environment. A mild environment can be achieved by coating the biomacromolecules with friendly excipients or entrapping them with pH buffers or by other means. Furthermore, the use of porogens should also be carefully taken into consideration to avoid inactivation of proteins. An example is that the use of salts, such as calcium and sodium chloride, in the internal water phase has been reported to induce this process and enable the formation of highly porous particles. The use of this formulation strategy, however, can be dangerous due to the fact that salts can cause protein precipitation/aggregation/inactivation as a function of salt type and concentration used.⁵⁷

Kim et al fabricated a pore-closing porous PLGA microsphere, loaded with recombinant human growth hormone (rhGH).⁵⁸ For their controlled release, porous microspheres containing rhGH were treated with water-miscible solvents in an aqueous phase for the production of pore-closed microspheres, and this process was performed in an ethanol vapor phase using a fluidized bed reactor. The results exhibited a high protein loading amount.

As to carrying other substances, such as metals or compounds, porous microspheres can be employed as carriers for catalysts. Porous microspheres with active molecules or active groups can be an excellent catalyst carrier. The larger specific surface area means more active sites for catalyst. So a high porosity is usually preferred. Thus combining catalysts with porous microspheres is a promising method.

Lee et al prepared porous structured TiO₂ microspheres, which showed better photocatalytic degradation of phenol compared with P25 TiO₂.⁵⁹ Molvinger et al prepared porous chitosan-silica hybrid microspheres as a potential catalyst.⁶⁰

Pulmonary drug delivery

In recent years, the lung has gained much attention as a non-invasive alternative for protein delivery; biomacromolecules

can be absorbed efficiently in a lung surface area of approximately 100 m², crossing the thin pulmonary absorption barrier.⁶¹ Here porous microspheres can be adopted as the biomacromolecule vehicle due to their low density of porous structure. Making the porous microspheres into an inhalable aerosol, biomolecules like proteins can be free from first-pass effects and realize a sustained-release. As to pulmonary drug delivery, it is very important to control the diameter of the microspheres and a narrow diameter distribution is preferred. For high deposition of inhaled proteins throughout the lung, the porous microspheres' aerodynamic diameter should be about 5 μm.⁶² Briefly, microspheres fitted for pulmonary drug delivery should have a low mass density (<0.4 g/cm³, aerodynamic diameter of 1–5 μm) and a large size (>5 μm) which allows for a high accumulation of protein particles in the periphery of the lung, thus preventing phagocytic clearance and realizing a controlled release.^{63,64}

Other applications

New fields that microspheres can be applied to have been discovered. Jiang et al built a lotus-leaf-like superhydrophobic surface with a porous microsphere/nanofiber composite film prepared by electrohydrodynamics.⁶⁵ An electrically charged jet of polystyrene/dimethylformamide solution was pumped by a nozzle to a metal plate, and with the evaporation of the solvent, a porous structure formed. In other fields, porous microspheres can be employed for gastric delivery,⁶⁶ and high performance lithium ion battery anode materials.⁶⁷

Conclusion

Porous microspheres have been extensively studied in recent years for their applications in tissue regeneration, pulmonary drug delivery, and separation of substances. The key characteristics are the porous structure, large specific surface area, and low density. Porous microspheres have an excellent absorption capacity compared with traditional microspheres.

However, many problems still exist in the application of porous microspheres. The methods to prepare porous microspheres are not perfect. For example, the methods used with porogens: the removal of porogens not only leads to the diffusion of the encapsulated drugs, but can also be time consuming and incomplete. The porous structure makes a big difference in the absorption and desorption of substances. So the parameters of the pores should be controllable to perform the desired kinetic behavior. Furthermore, the irritation in pulmonary delivery is not easily addressed.

In future research, universal technologies to control the pore structure, pore size, and porosity should be studied. New porous materials with ideal pore parameters like mesoporous silica will be beneficial to large scale applications.

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Disclosure

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