

Knockout of the c-Jun N-terminal Kinase 2 aggravates the development of mild chronic dextran sulfate sodium colitis independently of expression of intestinal cytokines TNF α , TGFB1, and IL-6

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Introduction: The c-Jun N-terminal kinases (JNKs) are involved in signal transduction of inflammatory bowel diseases. The aim of this study was to examine the function of JNKs by using a low-dose dextran sulfate sodium (DSS) model in JNK1 knockout mice (Mapk8^{-/-}), JNK2 knockout mice (Mapk9^{-/-}), and wild-type controls (WT1, WT2).

Methods: The animals were evaluated daily using a disease activity index. After 30 days, the intestine was evaluated histologically with a crypt damage score. CD4⁺ and CD8⁺ cells were quantified using immunofluorescence. Analysis of tumor necrosis factor- α (TNF α), interleukin-6 (IL-6), and transforming growth factor β 1 (TGFB1) expression was carried out using LightCycler[®] real-time polymerase chain reaction.

Results: Cyclic administration of low-dose DSS (1%) was not able to induce features of chronic colitis in Mapk8^{-/-} WT2 mice. By contrast, DSS administration significantly increased the disease activity index in WT1 and Mapk9^{-/-} mice. In Mapk9^{-/-} mice, the crypt damage score and the number of CD4⁺ and CD8⁺ cells as features of chronic colitis/inflammation were also significantly elevated. Expression of TNF α , IL-6, and TGFB1 was not altered by the JNK knockout.

Conclusion: Administering DSS at a defined low concentration that is unable to induce colitis in WT animals leads to clinically and histologically detectable chronic colitis in Mapk9^{-/-} mice. The reason for this disease-inducing effect resulting from the loss of JNK2 remains to be elucidated. Expression of TNF α , IL-6, and TGFB1 does not appear to be involved; proapoptotic JNK2 may prolong the activity of proinflammatory immune cells, leading to perpetuation of the inflammation.

Keywords: inflammatory bowel diseases, proinflammatory cytokines, JNK knockout mice, T-cell immune response

Introduction

Inflammatory bowel diseases (IBDs) are disorders of unknown etiology, characterized by chronic, relapsing inflammation of the gastrointestinal tract. Inappropriate activation of the immune system leads to overproduction of immune cells, inflammatory cytokines, and tissue-destructive enzymes.¹⁻³ Mediators of IBDs activate a variety of intracellular signaling cascades, such as mitogen-activated protein kinases (MAPKs). MAPKs are components of inflammatory and immunological signaling cascades that integrate various extracellular stimuli and initiate inflammatory cellular responses.^{4,5} The MAPK signal transduction pathway appears to be fundamental in the initiation

of inflammatory responses.⁶ Several subgroups have been identified within the MAPK family, including p42/44 extracellular signal-related kinase, Jun N-terminal kinase (JNK), and p38 MAPK.⁷ The JNK class of enzymes comprises three main types: JNK1, JNK2, and JNK3. The first two are more ubiquitously distributed, whereas the third is confined to the central nervous system and cardiac myocytes.¹ JNK1 and JNK2 are involved in the signal transduction of IBDs, and they regulate the expression of proinflammatory cytokines.^{6,8,9} For this reason, JNKs are considered to be targets for IBD therapy.

Previous studies of the function of JNK in vitro and in vivo have been limited by the lack of a specific inhibitor. In a pilot clinical trial in patients with Crohn's disease, administration of CNI-1493, a synthetic guanylhydrazone that inhibits phosphorylation of both p38 MAPK and JNK, significantly decreased disease activity.⁶ Hollenbach et al¹⁰ and Assi et al¹¹ confirmed the beneficial effects of the p38 inhibitor SB203580 in both dextran sulfate sodium (DSS)-induced and 2,4,6-trinitrobenzenesulfonic acid-induced murine colitis, underscoring the importance of this pathway in gastrointestinal inflammation. SP600125 has been identified as a specific inhibitor of all three isoforms of JNK.^{12,13} The in vivo effects of SP600125 on inhibition of cytokine synthesis and protection against tissue injury have been evaluated in several animal models of inflammation.^{12,14,15} A cell-penetrating antiproteinase-resistant peptide inhibitor of JNK (D-JNKI-1), derived from the JNK binding domain of JNK-interacting protein-1, was developed to competitively inhibit JNK binding to its substrates.¹⁶ The peptide inhibitor D-JNKI-1 was more potent than the small-molecule inhibitor SP600125.¹⁶ D-JNKI-1 reduced the number of apoptotic cells, indicating suppression of the mitochondrial mechanism of apoptosis.¹⁷

The beneficial effects of JNK inhibitors have implicated JNK as a critical regulator in IBDs. However, the specificity of some of these inhibitors has been questioned.¹⁸ Parallel studies using JNK gene knockout approaches have therefore provided valuable insights into the roles of JNK. In this context, Kontoyiannis et al^{8,9} underlined the important role of JNK in IBDs by genetic deletion of JNK2 in the tumor necrosis factor- α (TNF α)-mediated TNF α Δ ARE model of experimental colitis. JNK gene knockout led to alleviation of the TNF α -mediated inflammatory reaction.

Several animal models for research on intestinal inflammation have been developed. In 1990, Okayasu et al¹⁹ described an experimental model of acute and chronic colitis in mice, induced by DSS. The pathological features resembled

those of IBDs. DSS administered in drinking water produced weight loss and bloody diarrhea. Histopathologically, the changes were characterized by colonic mucosal inflammation and ulceration. The inflammation was worse in the left colon than in the right colon and cecum. Due to its ease of administration and the induction of a well-characterized and reproducible mucosal injury, DSS has been widely used in animal models for acute and chronic colitis.

In the present study, a low-dose DSS model of chronic experimental colitis was used. Mild chronic colitis was induced in JNK1 knockout mice (Mapk8 $-/-$) and JNK2 knockout mice (Mapk9 $-/-$). The study investigated the effect of JNK knockout on intestinal inflammation, T-cell immune responses, and expression of JNK-dependent cytokines.

Materials and methods

Animals

The experiments were conducted in female JNK1 knockout mice (Mapk8 $-/-$), JNK2 knockout mice (Mapk9 $-/-$), and wild-type controls (WT1 and WT2). All of the mice were 10–12 weeks of age at the start of the experiment (weight 20–25 g). The mice were housed three per cage in standard laboratory conditions and supplied with drinking water and food ad libitum (ssniff M-Z, 10 mm; ssniff Spezialdiäten, Soest, Germany). They were kept at the Institute of Pharmacology at Schleswig-Holstein University Hospital, Campus Kiel, Germany, for 14 days before the study was initiated. At this time, the mice were mature and the body weights among the different strains were comparable.

Transgenic mice deficient in JNK1 and JNK2 (Mapk8 $-/-$ and Mapk9 $-/-$) were kindly provided by Roger J Davis (Program in Molecular Medicine, University of Massachusetts, Worcester, MA) and Richard A Flavell (Howard Hughes Medical Institute, Yale University School of Medicine, New Haven, CT), and bred at the Institute of Pharmacology at Schleswig-Holstein University Hospital. Genetic inactivation using targeted disruption of JNK1 and JNK2, resulting in Mapk8 $-/-$ and Mapk9 $-/-$ genotypes, has been described in detail elsewhere.^{20–22} In both genotypes, the deleted exons coded for the TPY motif of the activation loop and for the two alternatively spliced JNK1/2 α and β subdomains.^{20–22}

The Mapk8 $-/-$ and Mapk9 $-/-$ mice were back-crossed once with C57/Bl6 WT mice that had not been genetically altered, in order to obtain WT control animals for the investigations. These heterozygous F2 offspring were crossed to obtain F2 homozygous JNK knockout mice and homozygous WT controls. This F2 generation was used as parent

founders to breed the JNK knockout and WT control mice for the animal experiments (F3 generation). This breeding approach minimizes the genetic strain variability of knockout/control animals, provides a greater number of animals, and meets the criteria of the international guidelines (Banbury Conference).²³

Genomic DNA was isolated from mouse tail tips using the Roche DNA High Pure PCR Template Preparation Kit (Roche Diagnostics, Mannheim, Germany) in accordance with the manufacturer's instructions. To genotype the experimental animals, specific polymerase chain reaction (PCR) primer sets of three primers were used for each genotype to distinguish the endogenous locus from targeted alleles, as previously described.²⁴ For genomic PCR, 4 μ L genomic DNA, 2 μ L 10 \times PCR buffer, 1.6 μ L 2.5 mM dNTPs, 0.2 μ L Taq polymerase (Eppendorf, Hamburg, Germany; 5 U/ μ L), 2.7 μ L aqua dest, and 2.5 μ L of each primer (10 μ M) were used (total volume 20 μ L). Cycling conditions for PCR amplification were 95°C for 5 minutes and 32 cycles (for the JNK1 primer set) of 95°C for 60 seconds, 55°C for 60 seconds, and 72°C for 30 seconds, followed by a final extension at 72°C for 10 minutes. For PCR with the JNK2 primer set, the conditions were identical except for a reduction to 26 cycles. PCR products were blotted on a 1% agarose gel with electrophoresis.²⁵

Induction of mild chronic DSS colitis

The DSS concentration of 1% for induction of mild chronic colitis was determined in previous studies (data not shown) and is suitable for experimental protocols. The JNK1 knockout mice (Mapk8^{-/-}) and JNK2 knockout mice (Mapk9^{-/-}) and wild-type controls (WT1 and WT2) (n = 9 in each group) received three cycles of DSS treatment (molecular weight 36–44 kDa; MP Biomedicals, Aurora, OH, USA), each consisting of 1% (w/v) DSS for 5 days in distilled water. Each cycle of DSS treatment was followed by a 5-day interval with distilled water only. After completion of the third DSS cycle (after 30 days), the animals were killed by decapitation in deep anesthesia. The control groups for each strain (n = 9 in each group) received distilled water without DSS, and were also sacrificed after 30 days.

Assessment of disease activity

Disease activity was quantified in all of the animals daily using the disease activity index (DAI) described elsewhere.^{26–29} The mice were weighed daily and visually inspected for stool consistency, diarrhea, and rectal bleeding. Hemocult tests were performed daily to detect occult blood in the stool.

Tissue samples

After the mice had been sacrificed, the entire colon was dissected out. It was rinsed with normal saline and divided into four anatomical parts: cecum, proximal colon, mid-colon, and distal colon. All of the dissection steps were carried out in ice, and the tissue was snap-frozen in liquid nitrogen and stored at –80°C.

Histological evaluation of inflammation

For light microscopy, 7- μ m transverse cryostat sections (rotary cryostat microtome; Leica Microsystems, Nussloch, Germany) from the four anatomical parts (cecum, proximal colon, mid-colon, and distal colon) of each animal were stained with hematoxylin and eosin. The slides were examined under a Leica DM4000 B microscope and were evaluated independently by two investigators (EH and AMM). The degree of inflammation was graded semiquantitatively, with blinding to the respective group. The severity of mucosal injury was defined using the crypt damage score (CDS), as described previously.^{24,26,27,30} With regard to intra-observer agreement, differences were noted in only 5% of the observations. Any differences in grading were resolved by a joint examination that included a third blinded examiner (AMC).

Evaluation of colonic cytokine expression using real-time PCR

mRNA expression of TNF α , transforming growth factor β 1 (TGFB1), and interleukin-6 (IL-6) was evaluated as previously described.²⁶ Total RNA was prepared from specimens of the colon of JNK2 knockout mice (Mapk9^{-/-}) and WT controls using the RNeasy Mini Kit and Shredder Kit (Qiagen, Hilden, Germany). DNA digestion was carried out using the RNase-Free DNase Set (Qiagen). Thereafter, RNA was submitted to a second DNase digestion and transcribed using Promega M-MLV Reverse Transcriptase (Promega, Mannheim, Germany). Real-time PCR was performed using the LightCycler[®] System with Fast-Start DNA Master SYBR Green I (Roche Diagnostics). β -Actin was used as a reference gene. Calibrator-normalized relative quantification without efficiency correction was carried out using the LightCycler[®] Relative Quantification software, following the manufacturer's instructions. After normalization, a multiplication factor was used to adjust the final calibrator-normalized relative ratios to reasonable values.

The following primers were used:

- β -actin, 5'-ACGTTGACATCCGTAAAG-3' (upstream) and 5'-CAGTAACAGTCCGCCT-3' (downstream) (amplicon 286 bp);

- TNF α , 5'-CATCTTCTCAAATTCGAGTGACAA-3' (upstream) and 5'-TGGGAGTAGACAAGGTACAA CCC-3' (downstream) (amplicon 175 bp);
- TGFB1, 5'-TGACGTCCTGGAGTTGTACGG-3' (upstream) and 5'-GGTTCATGTCATGGATGGTGC-3' (downstream) (amplicon 170 bp);
- IL-6, 5'-GAGGATACCACTCCCAACAGACC-3' (upstream) and 5'-AAGTGCATCATCGTTGTT CATAA-3' (downstream) (amplicon 141 bp).

Immunofluorescence of activated lymphocytes

Immunofluorescence was carried out in 6- μ m transverse cryostat sections from the distal colon (rotary cryostat microtome; Leica Microsystems). The sections were mounted on poly-L-lysine slides, air-dried, and fixed in methanol at room temperature. They were then blocked with Aurion BSA-c (Aurion Immuno Gold Reagents and Accessories, Wageningen, The Netherlands). Sections were subsequently incubated with the primary antibodies for 2 hours at room temperature: monoclonal rat anti-CD4 antibody (BD Biosciences, Heidelberg, Germany) diluted 1:400 (0.2 mg/mL) in phosphate-buffered saline (PBS) and monoclonal rat anti-CD8 antibody (BD Biosciences) diluted 1:100 (0.2 mg/mL) in PBS. After washing in PBS, tissue-bound antibodies were detected using Cy3-conjugated donkey anti-rat IgG (Dianova, Hamburg, Germany) diluted 1:100 (1.5 mg/mL). Incubation with the secondary antibody was carried out for 2 hours at room temperature and then at 4°C overnight. Sections were then washed in PBS. Each section had its own control using the secondary antibody only. Finally, the sections were mounted and analyzed using a fluorescence microscope (Leica DM4000 B microscope). Quantification of CD4+ and CD8+ cells was performed by examining two randomly selected high-power fields (with a magnification of 40,000 \times).

Statistical evaluation

The results for the DAI and CDS were expressed as means \pm standard error of the mean (SEM). DAI values were analyzed using repeated-measures ANOVA over all time points. Comparison of CDS values and cytokine expression between the groups was carried out using one-way ANOVA; pair comparisons were conducted using Tukey's test. The labeling density of CD4+ and CD8+ cells was evaluated with the *t*-test. *P*-values of ≤ 0.05 were considered statistically significant. Survival rates and hazard ratios were analyzed using a log rank test and Cox regression analysis, with

P-values of ≤ 0.05 considered statistically significant; 95% confidence intervals were also indicated.

Statement of animal care

All of the animal experiments were performed in accordance with German legislation for the protection of animals and the US National Institutes of Health (NIH) guidelines for the use and care of laboratory animals. The experiments were all approved by the Ministry of Agriculture and the Environment (Ministerium fuer Landwirtschaft und Naturschutz), Kiel, Germany (V742-72241.121-22(10-2(05))).

Results

Genotyping of JNK knockout and wild-type controls

Genotyping of JNK-deficient mice and WT controls using PCR and gel electrophoresis confirmed the respective genotype in all of the experimental animals (data not shown).

Development of mild colitis in Mapk9 $^{-/-}$ as opposed to Mapk8 $^{-/-}$ mice

One of the nine animals (11.1%) died in the Mapk8 $^{-/-}$ group and three of the nine WT1 animals (33.3%) died after treatment with DSS. None of the animals in the H₂O control groups died. The differences in the mortality rates were not significant.

Differences in DAI values in the animals that survived up to the end point of the study (30 days) were statistically evaluated using repeated-measures ANOVA over all time points. As illustrated in Figure 1A, only WT1 animals developed significant chronic colitis, as assessed by the DAI, following cyclic administration of low-dose DSS (1.0%). The DAI was significantly higher in the WT1 DSS group, in comparison with the WT1 H₂O control group (4.2 ± 0.7 vs 1.2 ± 0.3 ; $P < 0.001$). By contrast, in Mapk8 $^{-/-}$ mice, cyclic administration of low-dose DSS (1.0%) was unable to induce chronic colitis with a significantly elevated DAI, in comparison with the H₂O control group (3.2 ± 0.2 vs 1.6 ± 0.5 ; $P = 0.056$) (Figure 1A). There were no significant differences in DAI values between the WT1 and Mapk8 $^{-/-}$ mice following DSS treatment.

None of the animals in the Mapk9 $^{-/-}$ and WT2 groups died (mortality 0%). Administration of 1.0% DSS in the WT2 animals did not induce chronic colitis, as reflected in a mean DAI of 1.1 (± 0.4), which was not significantly elevated, in comparison with the value of 0.6 (± 0.2) in the H₂O control group. By contrast, cyclic administration of DSS in the Mapk9 $^{-/-}$ animals resulted in mild triphasic colitis with a mean DAI of 2.3 (± 0.3), which was significantly higher, in comparison with the H₂O control group at 0.6 (± 0.2)

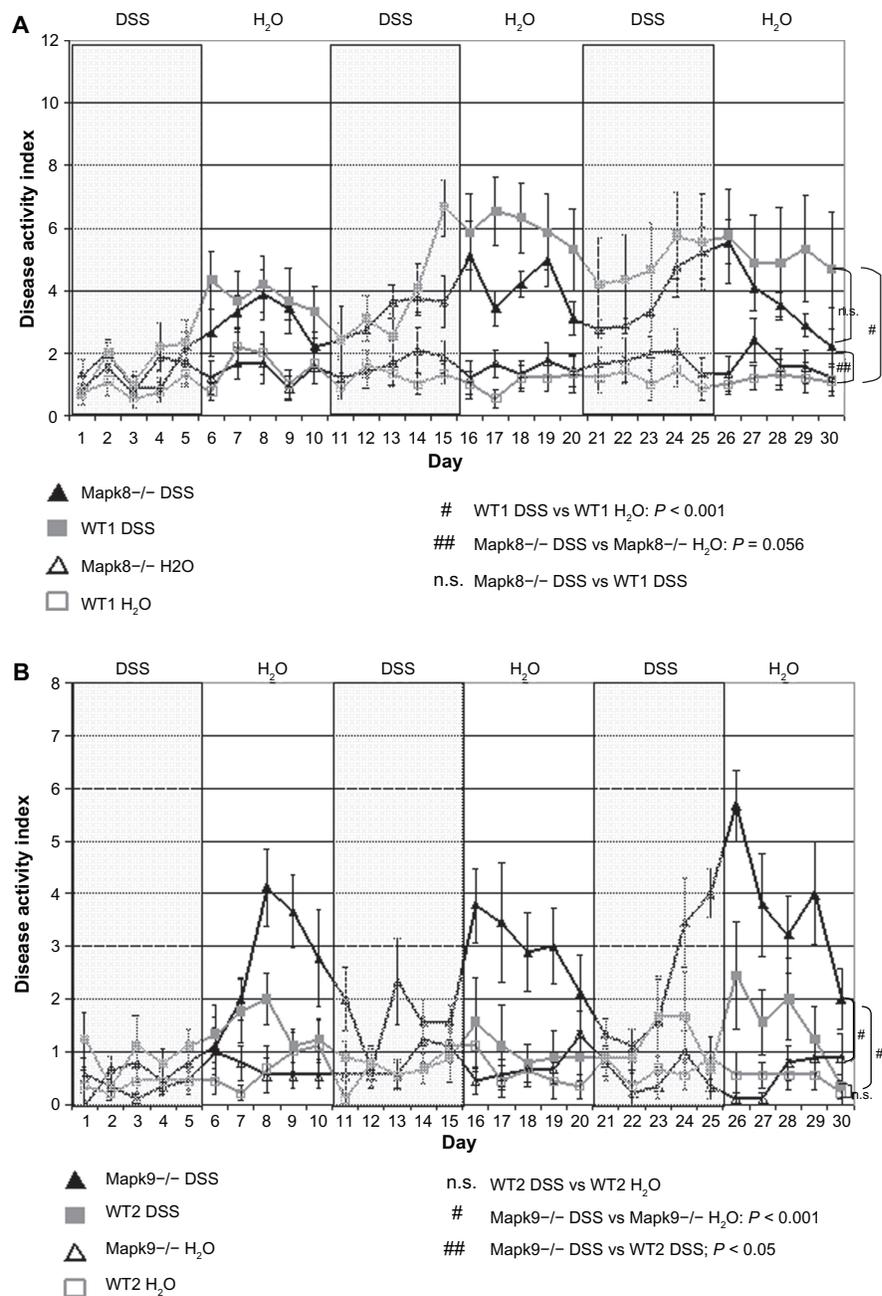


Figure 1 The disease activity index (DAI) after administration of three cycles of 1.0% dextran sulfate sodium (DSS) or H₂O as the control (means \pm SEM).

Notes: (A) After administration of 1.0% DSS, the DAI in WT1 mice was significantly elevated, in comparison with the H₂O control group ($P \leq 0.05$). Cyclic administration of DSS in Mapk8^{-/-} animals resulted in mild triphasic colitis, with no significant differences in comparison with the H₂O control group or the WT1 DSS group (statistical evaluation: ANOVA over all time points). (B) After administration of 1.0% DSS in the WT2 animals, the DAI was not significantly elevated, in comparison with the H₂O control group. Cyclic administration of DSS in Mapk9^{-/-} animals resulted in mild triphasic colitis, which was significantly greater, in comparison with the H₂O control group ($P \leq 0.001$) and in comparison with the WT2 DSS group ($P \leq 0.05$) (statistical evaluation: ANOVA over all time points).

($P \leq 0.001$), and in comparison with the WT2 DSS group ($P \leq 0.05$) (Figure 1B).

Histological scoring of the degree of inflammation

After DSS treatment, histological scoring using the CDS developed by Egger et al^{24,30} showed gradually elevated crypt

damage in the medial and distal colon in Mapk8^{-/-} mice (CDS in the mid-colon 2.6 ± 1.9 and in the distal colon 4.0 ± 2.4) and WT1 mice (CDS in the mid-colon 1.8 ± 1.2 and in the distal colon 5.1 ± 4.0). However, these differences in CDS did not reach statistical significance (Figure 2A).

In Mapk9^{-/-} mice, cyclic administration of 1% DSS also led to significant colitis in the mid-colon (CDS 4.0 ± 1.2)

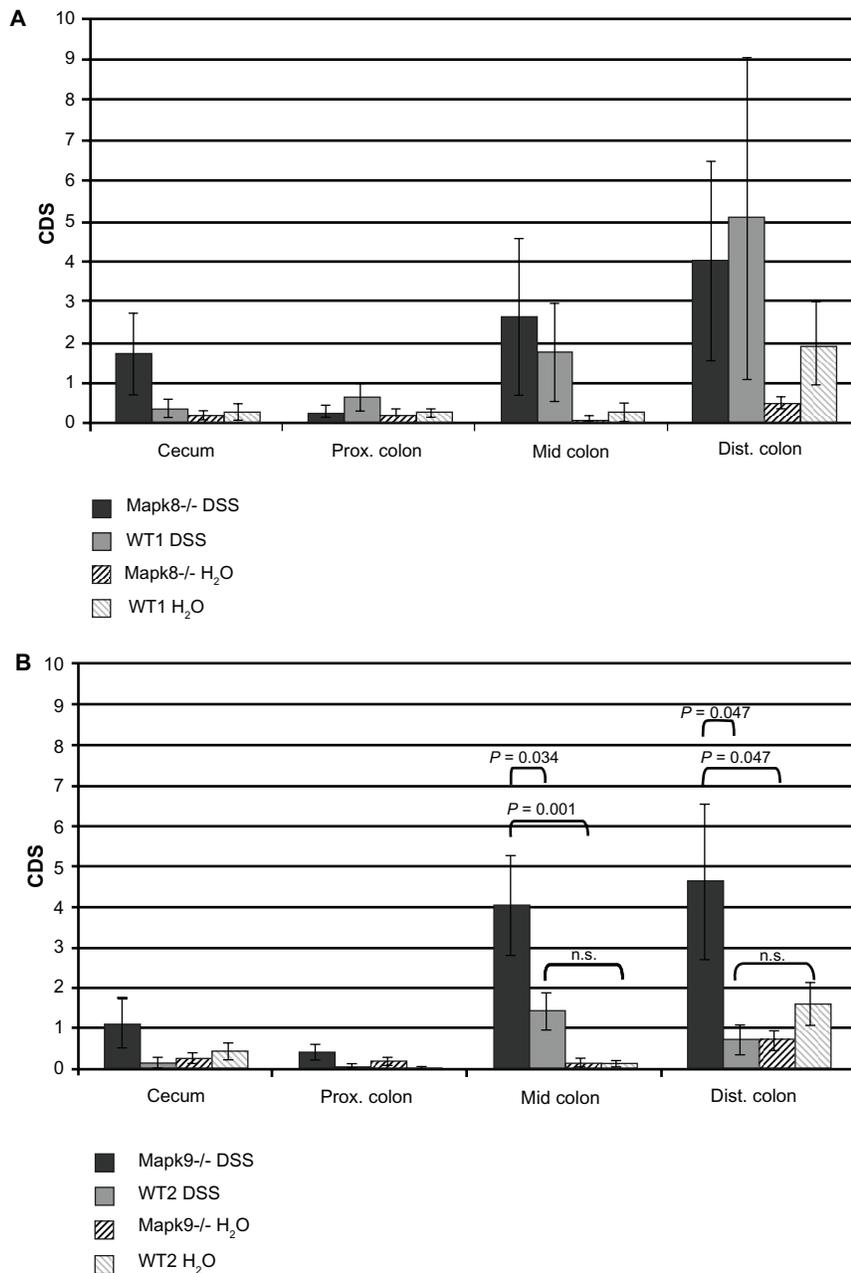


Figure 2 The crypt damage score (CDS) after administration of three cycles of 1.0% dextran sulfate sodium (DSS) or H₂O as the control (means \pm SEM).

Notes: The most affected regions were the mid-colon and distal colon. **(A)** Differences in the CDS in JNK1 knockout (Mapk8-/-) and wild-type controls (WT1) did not reach statistical significance (CDS value comparison: one-way ANOVA; pair comparisons: Tukey's test). **(B)** In the DSS-treated Mapk9-/- group, the CDS was significantly elevated, in comparison with the DSS-treated WT2 group (mid-colon, $P = 0.034$; distal colon, $P = 0.047$) and with the H₂O control group (mid-colon, $P = 0.001$; distal colon, $P = 0.047$) (CDS value comparison: one-way ANOVA; pair comparisons: Tukey's test).

and distal colon (CDS 4.6 ± 1.9). In the DSS-treated Mapk9-/- group, the CDS was significantly elevated, in comparison with the DSS-treated WT2 group (mid-colon, $P = 0.034$; distal colon, $P = 0.047$) and in comparison with the H₂O control groups (mid-colon, $P = 0.001$; distal colon, $P = 0.047$). DSS treatment of WT2 mice did not result in significant CDS changes, in comparison with H₂O treatment (Figures 2B and 3).

Inducing mild DSS colitis in JNK knockout mice did not alter the expression profile of cytokines TNF α , TGFB1, and IL-6

The expression profile of the cytokines TNF α , TGFB1, and IL-6 was examined in all of the animal groups (Table 1). Calibrator-normalized quantitative real-time PCR did not show any significant differences, either between the four Mapk8-/- WT1 groups or between the four Mapk9-/-

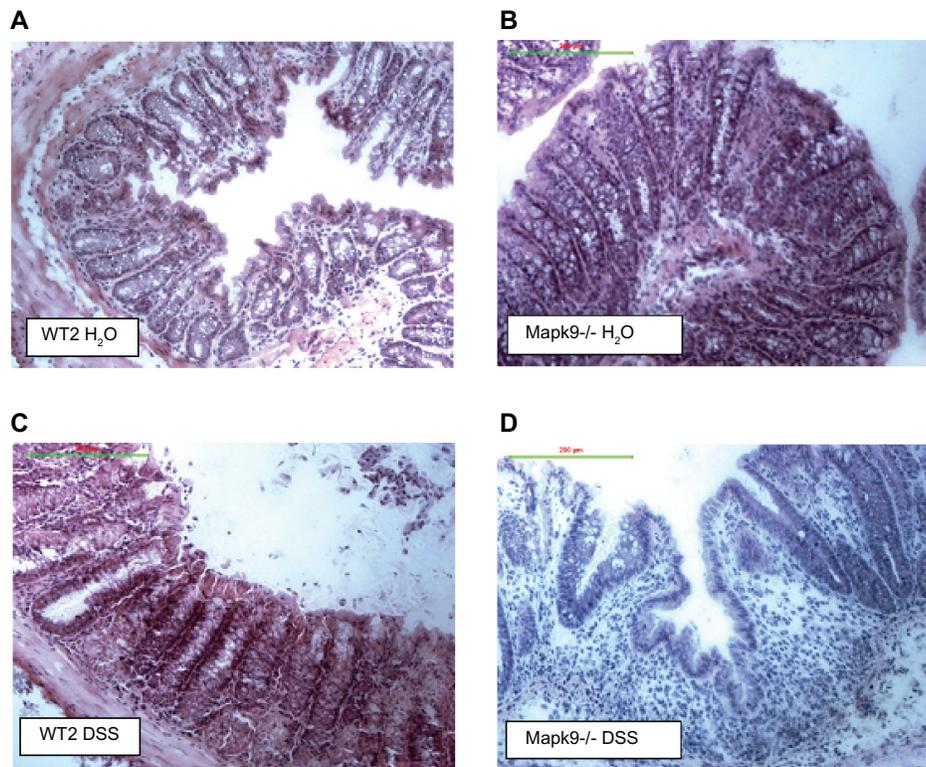


Figure 3 Hematoxylin–eosin staining of the distal colon.

Notes: (A) Wild-type controls (WT2) treated with H₂O: normal mucosa with a regular crypt architecture. (B) JNK2 knockout mice (Mapk9^{-/-}) treated with H₂O: normal mucosa with a regular crypt architecture. (C) Wild-type controls (WT2) treated with three cycles of 1.0% dextran sulfate sodium (DSS): mild changes in the crypt architecture. (D) JNK2 knockout mice (Mapk9^{-/-}) treated with three cycles of 1.0% DSS: severe changes in the crypt architecture, with ulceration and loss of crypts.

WT2 groups in the experiment, with regard to the expression of TNF α , TGF β 1, or IL-6.

Activated lymphocytes in the inflamed colon

In the Mapk8^{-/-} WT1 group, differences in the CDS did not reach statistical significance (Figure 2A). Analysis of the expression of CD8⁺ and CD4⁺ cells also did not show any notable differences.

Table 1 Relative mRNA expression (mean \pm SEM) for TNF α , TGF β 1, and IL-6

| | TNF α | TGF β 1 | IL-6 |
|---------------------------------------|-----------------|-----------------|------------------|
| Mapk8 ^{-/-} DSS | 744 \pm 238 | 574 \pm 111 | 1674 \pm 675 |
| WT1 DSS | 2048 \pm 1356 | 2319 \pm 850 | 10139 \pm 5838 |
| Mapk8 ^{-/-} H ₂ O | 739 \pm 268 | 1920 \pm 1197 | 975 \pm 528 |
| WT1 H ₂ O | 1244 \pm 610 | 2481 \pm 915 | 2171 \pm 622 |
| Mapk9 ^{-/-} DSS | 622 \pm 170 | 1402 \pm 310 | 1516 \pm 417 |
| WT2 DSS | 624 \pm 259 | 1237 \pm 572 | 1201 \pm 477 |
| Mapk9 ^{-/-} H ₂ O | 302 \pm 57 | 574 \pm 123 | 199 \pm 74 |
| WT2 H ₂ O | 662 \pm 293 | 1706 \pm 787 | 2336 \pm 1159 |

Abbreviations: DSS, dextran sulfate sodium; IL-6, interleukin-6; SEM, standard error of the mean; TGF β 1, transforming growth factor β 1; TNF α , tumor necrosis factor- α ; WT, wild-type.

In the Mapk9^{-/-} WT2 group, immunofluorescence analysis of CD4 and CD8 expression was carried out in tissue from the distal colon, as this part of the colon developed the highest grade of inflammation. After induction of chronic DSS colitis, CD8 staining was significantly higher in Mapk9^{-/-} animals, in comparison with WT2 mice ($P = 0.002$). CD8 staining was also significantly increased in Mapk9^{-/-} animals treated with DSS, in comparison with Mapk9^{-/-} animals treated with H₂O (Figures 4 and 5A). Although the number of CD4⁺ cells was lower than the total number of CD8⁺ cells, immunofluorescence analysis of CD4 also showed significantly greater staining in Mapk9^{-/-} animals treated with DSS, in comparison with Mapk9^{-/-} animals treated with H₂O. In DSS-treated Mapk9^{-/-} animals, CD4 staining was greater, but did not show any significant differences, in comparison with DSS-treated WT2 mice ($P = 0.071$) (Figure 5B).

Discussion

JNKs have a central position in the intracellular signaling that orchestrates inflammatory responses. Finding a way of inhibiting JNKs is therefore an attractive option in the treatment of inflammatory diseases, including IBDs.

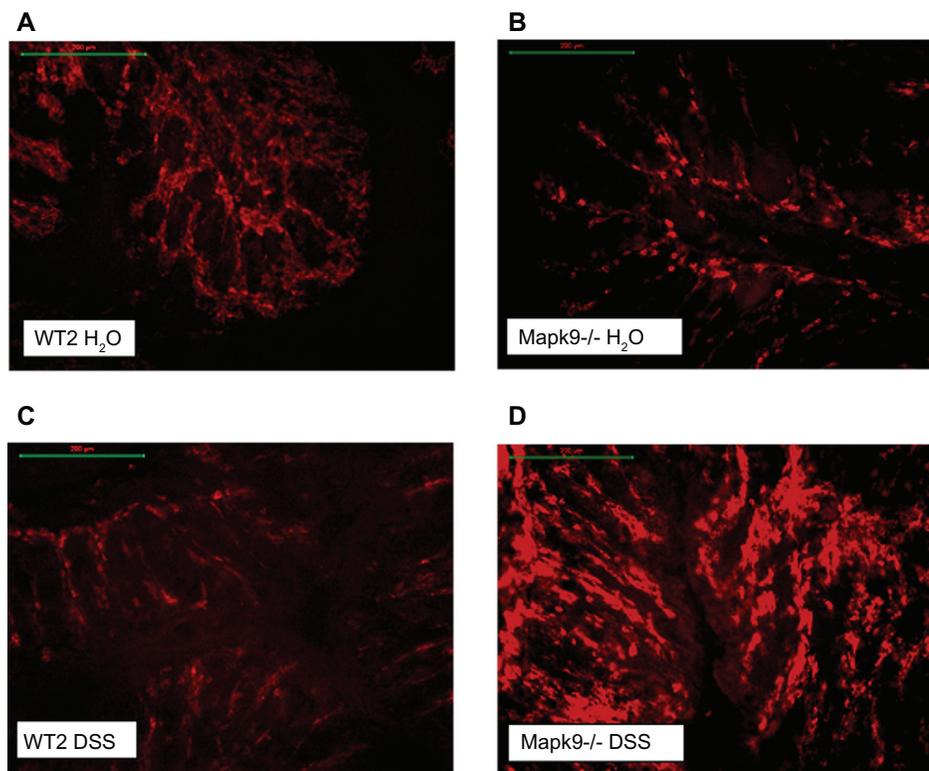


Figure 4 Immunofluorescence (Cy3-conjugated antibody) of CD8⁺ cells in the distal colon.

Notes: (A) Wild-type controls (WT2) treated with H₂O. (B) JNK2 knockout mice (Mapk9^{-/-}) treated with H₂O. (C) Wild-type controls (WT2) treated with three cycles of 1.0% dextran sulfate sodium (DSS). (D) JNK2 knockout mice (Mapk9^{-/-}) treated with three cycles of 1.0% DSS.

The present study investigated the function of the JNK isoforms JNK1 and JNK2 after induction of mild chronic colitis in JNK1 knockout mice (Mapk8^{-/-}) and JNK2 knockout mice (Mapk9^{-/-}). The clinical and histological features of inflammation were scored using the DAI and CDS, as described previously.^{24,30} As JNKs have multiple roles in T-cell immune responses and cytokine production,^{21,31} T-cell infiltration was evaluated using immunofluorescence, and the mRNA expression of the inflammatory cytokines TNF α , TGFB1, and IL-6 was quantified.

The observation that WT2 animals did not develop colitis after cyclic administration of low-dose DSS (1%), whereas WT1 animals developed chronic colitis, reflects a genetically determined differential susceptibility to DSS-induced colitis.³² The concentration of 1% DSS was apparently too low to induce chronic colitis in WT2 animals. A further increase in the DSS dosage to 1.7%, as described by our group in a previous study, resulted in chronic colitis even in WT2 animals.²⁷ Cyclic application of 1.7% DSS triggered severe colitis in Mapk8^{-/-} mice, Mapk9^{-/-} mice, and wild-type controls, with DAI values between 4 and 11 and CDS values between 5 and 13. Unfortunately, the development of severe chronic colitis following cyclic application of 1.7%

DSS was associated with high mortality rates of 75% in the Mapk9^{-/-} group and 33% in the WT2 group. This limited the interpretation of histological differences between Mapk9^{-/-} and WT2, due to the reduced number of surviving mice. For the same reasons, the further investigation of cytokines, CD4, and CD8 expression carried out in the present study could not be performed.²⁷

The intention in the present study was therefore to reduce the DSS dosage and achieve mild colitis with lower mortality rates. However, cyclic administration of 1% DSS in WT2 animals did not induce chronic colitis, reflected by a mean DAI of 1.1, which was not significantly elevated, in comparison with the H₂O control group. In the Mapk9^{-/-} animals, the same DSS dosage resulted in mild triphasic colitis with a mean DAI of 2.3, which was significantly higher, in comparison with the H₂O control group at 0.6 ($P \leq 0.001$) and in comparison with the WT2 DSS group ($P \leq 0.05$) (Figure 1B). The reason for this disease-inducing effect from the loss of JNK2 remains to be elucidated.

The fact that one of the nine animals in the Mapk8^{-/-} group died and three of the nine animals in the WT1 died after treatment with DSS, whereas none of the animals in the Mapk9^{-/-} WT2 groups died, may also reflect a genetically

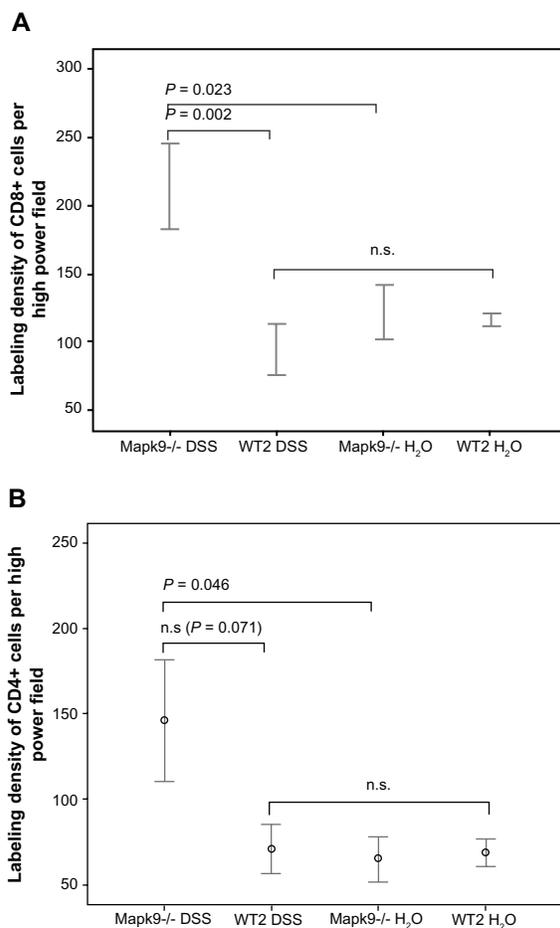


Figure 5 The labeling density of CD8 and CD4 (cells per high-power field) after administration of three cycles of 1.0% dextran sulfate sodium (DSS) or H₂O as the control.

Notes: (A) After induction of chronic DSS colitis, CD8 staining was significantly higher in Mapk9^{-/-} animals, in comparison with WT2 mice ($P = 0.002$). CD8 staining was also significantly elevated in Mapk9^{-/-} animals treated with DSS, in comparison with Mapk9^{-/-} animals treated with H₂O ($P = 0.023$) (statistical evaluation: *t*-test). (B) There was significantly greater staining in Mapk9^{-/-} animals treated with DSS, in comparison with Mapk9^{-/-} animals treated with H₂O ($P = 0.046$). In DSS-treated Mapk9^{-/-} animals, CD4 staining was greater, but did not show any significant differences in comparison with DSS-treated WT2 mice ($P = 0.071$) (statistical evaluation: *t*-test).

determined differential susceptibility to DSS-induced colitis in Mapk8^{-/-} WT1 animals, in comparison with Mapk9^{-/-} WT2 animals.³² Although the variability between mice was relatively high and the number of animals was limited ($n = 9$), inactivation of JNK2 resulted in a significant elevation of DAI and CDS, whereas there was no significant difference in Mapk8^{-/-} mice, in comparison with WT1 mice after three cycles of 1% DSS administration. Although the DAI and CDS values were higher in WT1 mice, no significant differences were observed, either in histologic inflammation or in cytokine expression. One possible reason for this might be the experimental protocol used, in which a very mild form of chronic colitis was induced.

The distribution pattern of CD4⁺ and CD8⁺ cells reflected the degree of inflammation. The incidence of CD8⁺ cells was significantly higher in highly inflamed colon sections from Mapk9^{-/-} mice, in comparison with WT2 mice and Mapk9^{-/-} animals treated with H₂O. These results are in accordance with the immune responses of JNK1-deficient and JNK2-deficient mice following infection with lymphocytic choriomeningitis virus.³³ In contrast to Mapk8^{-/-}, which reduced the virus load, lymphocytic choriomeningitis virus infection generated more virus-specific CD8⁺ cells in Mapk9^{-/-}, in comparison with the respective wild type. These results indicate that the JNK1 and JNK2 signal pathways have distinct roles in T-cell responses. JNK1 is involved in the survival of activated T cells during immune responses, and JNK2 plays a role in the control of CD8⁺ cell expansion. Previous studies showed that the absence of JNK2 causes increased proliferation of CD8⁺ cells, whereas JNK1-deficient CD8⁺ cells were unable to undergo antigen-stimulated expansion.³⁴ Experimental research on CD4⁺ cells revealed that the JNK signaling pathway is also required for differentiation of CD4⁺ cells into effector cells.^{21,35} The present data on CD4 expression do not support this conclusion, as increased CD4 labeling was found in the inflamed colon of Mapk9^{-/-} mice. CD4⁺ cell expression was lower than CD8⁺ cell expression, but the difference did not reach significance.

The mechanisms that abnormally and persistently activate inflammation, leading to ongoing tissue damage, are varied and complex. Cytokines also play a central regulatory role.^{2,3} JNK has been implicated as an important regulator of the coordinated release of cytokines by immunocompetent cells and of the response of neutrophils to inflammatory stimuli. It positively regulates a variety of genes involved in inflammation, such as the TNF α , IL-1, and IL-6 genes.^{36,37} Inhibition of JNK activation effectively modulates TNF α secretion from macrophages and mast cells.^{38,39} However, JNK-dependent expression of the inflammatory cytokines TNF α , IL-6, and TGF β 1 apparently did not play a major role in the present model of mild chronic DSS colitis. Calibrator-normalized quantitative real-time PCR did not show any significant differences, either between the four JNK1 groups or between the four JNK2 groups in the experiment, with regard to the expression of TNF α , TGF β 1, or IL-6. This finding is in line with observations by Alex et al,⁴⁰ who reported an effect of these cytokines in acute DSS colitis, but not in chronic DSS colitis. An important aspect of further investigations by our study group will be the expression of other cytokines, such as interferon- γ (IFN- γ), IL-13, and IL-17, in order to obtain

more information about the involvement of the Th1 and Th2 pathways in the exacerbated DSS colitis observed in JNK knockout mice.

The JNK branch of the MAPK signaling cascade has been implicated in the regulation of apoptosis in a variety of mammalian cell types.^{22,25,41,42} Apoptosis was not assessed in the present study. One possible hypothesis is that the proinflammatory effects caused by genetic deletion of JNK2 may be a consequence of inhibited JNK2-induced apoptosis. It might be speculated that functional deletion of proapoptotic JNK2 may prolong the activity of proinflammatory immune cells, leading to perpetuation of the inflammation. In addition, the increased inflammation in *Mapk9*^{-/-} mice may be induced by blockage of the anti-inflammatory potency of the JNK2 isoform in these animals.^{33,34} No differences were observed in *Mapk8*^{-/-} mice with regard to the degree of mucosal inflammation, in comparison with WT1 mice. A possible explanation for this might be that mice that are deficient in JNK1 are not able to develop the antiapoptotic activity of activated JNK1 that is achieved by transforming growth factor- β -associated kinase 1.⁴³ In view of the observation that the DSS colitis observed in JNK2 mice was not driven by the cytokines TNF α , TGF β 1, or IL-6 that were being investigated, further studies are necessary in order to investigate the role of apoptosis in this process of chronic inflammation.

Deletion of JNK2 resulted in a deterioration of colitis in *Mapk9*^{-/-} mice after DSS administration at a defined low concentration (1.0%). These results are in contrast to the expectations raised in studies that have reported a decrease in inflammation following administration of JNK inhibitors. It has been reported that the MAPK inhibitor CNI-1493 is able to alleviate human Crohn's disease, even in some cases in which anti-TNF α antibody has failed.⁶ It has also been demonstrated that p38 MAPK inhibition using SB203580 was effective in reducing disease in both DSS-induced and trinitrobenzenesulfonic acid-induced murine colitis. Both the clinical and the histological scores were significantly improved by treatment with SB203580.¹⁰ Assi et al¹¹ reported that the JNK inhibitor SP600125 reduced infiltration of inflammatory cells, macrophage infiltration, and epithelial cell apoptosis. The observation in the present study that JNK2 deficiency leads to more severe colitis, whereas the JNK inhibitors described in the current literature have an anti-inflammatory effect, may be explained by the diverse functions of JNK signaling pathways in cellular metabolism and immunological regulation. It is very likely that these functions of JNK are needed for immunocompetence in both normal and pathogenic conditions. Genetic deletion of JNK2 is not equivalent to inhibition eg,

by drugs. The former may result in compensatory (embryonic) changes whereas the pharmaceutical inhibition interferes with the acute situation only.

Conclusion

In contrast to the pharmacological inhibition of JNK, the present study shows that genetic deletion of JNK2 aggravates DSS colitis. It may be speculated that apoptosis of T cells is reduced, which is reflected in the high expression of CD4⁺ and CD8⁺ cells in the inflamed colon in DSS-treated *Mapk9*^{-/-} mice. This might result in perpetuation of chronic intestinal inflammation. Further research is needed to confirm the relevance of apoptosis in this context.

Disclosure

The authors report no conflicts of interest in this work.

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