

Hippuric acid nanocomposite enhances doxorubicin and oxaliplatin-induced cytotoxicity in MDA-MB231, MCF-7 and Caco2 cell lines

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Background: The aim of the current study is to design a new nanocomposite for inducing cytotoxicity of doxorubicin and oxaliplatin toward MDA-MB231, MCF-7, and Caco2 cell lines. A hippuric acid (HA) zinc layered hydroxide (ZLH) nanocomposite was synthesized under an aqueous environment using HA and zinc oxide (ZnO) as the precursors.

Methods: The hippuric acid nanocomposite (HAN) was prepared by the direct reaction of a HA solution with an aqueous suspension of ZnO.

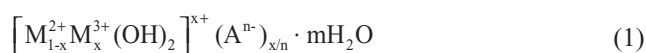
Results: The basal spacing of the nanocomposite was 21.3 Å, which is average of four harmonics at $2\theta = 8.32^\circ$, 12.50° , 16.68° , and 20.84° . This result indicates that the hippurate anion was successfully intercalated into the interlayer space of ZLH. The combinations of HAN with chemotherapy (drugs) has inhibited the cell growth of the MDA-MB231, MCF-7, and Caco2 cancer cells when compared to drugs alone. An IC_{50} value for the combination of HAN with doxorubicin toward MCF-7 is $0.19 \pm 0.15 \mu\text{g/mL}$ and toward MDA-MB231 is $0.13 \pm 0.10 \mu\text{g/mL}$. Similarly, the IC_{50} for the combination of HAN with oxaliplatin toward Caco2 is $0.24 \pm 0.11 \mu\text{g/mL}$. In the antiproliferative results, the equal combination of HAN ($0.5 \mu\text{g/mL}$) with doxorubicin ($0.5 \mu\text{g/mL}$) has reduced the cell proliferation in MCF-7 and MDA-MB-231 cells into 37.3% and 17.6%, respectively after 24 hours. Similarly, the antiproliferation percentage for equal combination HAN with oxaliplatin ($5.00 \mu\text{g/mL}$) toward Caco2 is 72.7% after 24 hours.

Conclusion: The resulting combination HAN with drugs has exhibited higher inhibition in cells growth in all cancer cell lines.

Keywords: hippuric acid nanocomposite, zinc-layered hydroxide, doxorubicin, oxaliplatin, MDA-MB231, MCF-7, Caco2 cell lines

Introduction

The structure of layered double hydroxides (LDHs) and zinc layered hydroxide (ZLH) are derived from the structure of brucite, $\text{Mg}(\text{OH})_2$. Brucite is the natural mineral form of magnesium hydroxide. The magnesium ions with a (2+) charge are centered to six octahedrally hydroxides with a (1-) charge. Each hydroxide is bonded to three magnesium atoms, resulting in neutral layers that interact with each other through weak hydrogen bonds.¹ The chemical structure of LDHs can be represented by the general formula:²



and the layered hydroxide salts can be represented as follows:



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where M^{2+} is a divalent cation, M^{3+} is a trivalent cation, and A^{n-} is an exchangeable anion with a charge (n^-).³

The flexibility in composition of LDH and ZLH, and easily replaced the anions located in the interlayer regions led to an increased level of interest in these materials. As a result of the relative ease to synthesize, LDHs and LHSs represent inexpensive and versatile compounds that can be used in different advanced applications, such as in a controlled release system for drugs.⁴

When applying two drugs in a system, the interaction may yield an additive effect, where the total response is the sum of the two drugs individually. In addition, the combination of these two drugs may result in a greater response, which is called a synergistic response. On the other hand, the interaction between the two drugs may also lead to a smaller response, whereby one drug blocks the effects of the other. It is important to note that the synergistic effect of the combination of two drugs against one target has recently been of particular interest.^{5,6}

Hippuric acid (benzoylaminoethanoic acid) can be obtained from the urine of horses and other herbivores. A biological investigation of hippuric acid shows that it has limited antimicrobial activity at acidic pH values,⁷ and has synergistic potentiating effects on the selective toxicity of a mixture of 13 substances within the circulatory system.⁸ From our previous work, hippurate nanocomposite HAN has shown synergistic activity with tamoxifen against the HepG2 cell line.⁹ Therefore the aim of this present work is to study the synergistic properties of HAN when combined with doxorubicin and oxaliplatin against different cell lines (MDA-MB231, MCF-7, and Caco2). It should be known that oxaliplatin has been commonly used for chemotherapy in colon cancer,¹⁰ while doxorubicin is the most common treatment for breast cancer.^{11,12}

Materials and methods

Materials

Hippuric acid (98% purity) was purchased from Merck (Darmstadt, Germany) and was used as received. Doxorubicin (98% purity) and oxaliplatin was purchased from Sigma-Aldrich (St Louis, MO). Zinc oxide (ZnO) of American Chemical Society reagent grade was purchased from Fisher Scientific (Waltham, MA), and dimethyl sulfoxide (DMSO) was purchased from Ajax Finechem (Thermo Fisher Scientific, Waltham, MA) and used without further purification. The cell lines MCF-7, MDA-MB231, and Caco2 were purchased from the American Type Culture Collection. Deionized water was used in all the experiments.

Preparation of hippuric acid nanocomposite, HAN

The hippuric acid nanocomposite (HAN) was synthesized by the direct method using ZnO as the starting material as reported previously,^{9,13,14} with minor modifications. The hippuric acid solution (0.01 Mole) was prepared using 1.8 g of hippuric acid in 20 mL of DMSO, and was adjusted to 50 mL by the addition of deionized water. The ZnO powder (0.2 g) was suspended in 50 mL of water. Hippurate solution was added slowly dropwise to the suspended ZnO, with vigorous stirring until the addition was complete and the solution became clear. The pH was adjusted to 7.9 using an aqueous solution of NaOH (0.5 Mol/L) to get the white precipitate; the resulting precipitate was magnetically stirred for 18 hours at 70°C. The resulting product was centrifuged, thoroughly washed with deionized water, dried in an oven at 60°C overnight, and kept in a sample bottle for further use.

MTT cytotoxicity assay

Breast cancer cell lines (MCF-7 and MDA-MB231) and heterogeneous human epithelial colorectal adenocarcinoma cells (Caco2) were seeded into 96-well plates and kept at 5% CO₂ at 37°C for 24 hours, at a cell density of 40–50% confluence. The cells were then treated with doxorubicin and oxaliplatin alone, and with an equivalent cytostatic mixture of either hippuric acid or HAN. After 72 hours of incubation, 20 µL of tetrazolium salts (3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) MTT solution (5 mg/mL) was added to each well and incubated for 4 hours. The microplates were turned swiftly to discard the medium, and the formazan precipitate was dissolved in 10% sodium dodecyl sulfate in DMSO containing 0.6% acetic acid. The microplates were then gently shaken in the dark for 30 minutes, and absorbance at 570 nm and 630 nm (background) was measured with a microtiter plate reader. All experiments were carried out in triplicate. The IC₅₀ was generated from the dose-response curves for the cell line.

Antiproliferation assay

MCF-7, MDA-MB231, and Caco2 cells were first seeded in six-well plates. After incubation for 24 hours for cell attachment, exponentially growing cells were exposed to doxorubicin (0.5 µg/mL) alone, or to oxaliplatin (5 µg/mL) and its combination with hippuric acid or HAN. The plates were incubated at 37°C at 5% CO₂ for 24, 48, and 72 hours. After incubation, the media was aspirated off, washed with cold phosphate buffered saline to get rid of the dead cells, and replaced with 1 mL of 0.05% (2 mg/mL) trypsin-EDTA.

The plates were incubated at 37°C for 10–15 minutes, until the majority of the cells had lifted off. The cells were then harvested. The cell suspension was centrifuged at 1000 rpm for 10 minutes, and the supernatant was discarded. Twenty μL of cell suspension was mixed with 20 μL of 0.4% trypan blue solution. Cells were resuspended and dye-excluding viable cells were microscopically counted using a hemocytometer.

Statistical analysis

All data were expressed as the means \pm standard deviation of the values obtained from three replicates. Using analysis of variance, statistical significance was determined. Mean values with probability values of $P < 0.05$ were taken as statistically significant.

Characterization

Powder X-ray diffraction patterns were recorded with a Shimadzu XRD-6000 instrument (Shimadzu Corporation, Tokyo, Japan) using CuK_α radiation ($\lambda = 1.5418 \text{ \AA}$) and a dwell time of 4 degrees per minute. Cell count was carried out with a Neubauer hemocytometer (Weber Scientific International Ltd, Middlesex, UK) by way of clear field microscopy (Nikon Corporation, Tokyo, Japan).

Results and discussion

Powder X-ray diffraction

Figure 1 shows the powder X-ray diffraction patterns for hippuric acid, ZnO, and HAN, respectively, which was reported previously in our work.⁹ The mechanism of formation of hippurate nanocomposite synthesized by direct reaction of ZnO in an aqueous environment can be described as follows.¹⁵ Hydrolyzed ZnO in an aqueous environment leads to the formation of thin layers of zinc hydroxide (Zn(OH)_2) on the surface of the solid particles. Zn^{2+} species form of dissociation of Zn(OH)_2 in a solution-solid interface. In addition, Zn^{2+} ions react with hydroxyls, hippurate anions, and water in the solution to form the layered HAN nanocomposite. Figure 1A shows that ZnO exhibited three intense peaks between 30° and 40° , corresponding to diffractions due to the 100, 002, and 101 planes. Complete disappearance of the intense peaks of the ZnO phase and the appearance of a new intense peak at low 2θ with a d value of 21.3 \AA indicated that the sample of HAN is pure phase, and that the ZnO was completely converted to ZLH. In addition, Figure 1B shows another four harmonics at $2\theta = 8.32^\circ$, 12.50° , 16.68° , and 20.84° , with d values of 10.70 \AA , 7.10 \AA , 5.30 \AA , and 4.30 \AA , respectively, resulting in an average d value of 21.30 \AA .⁹

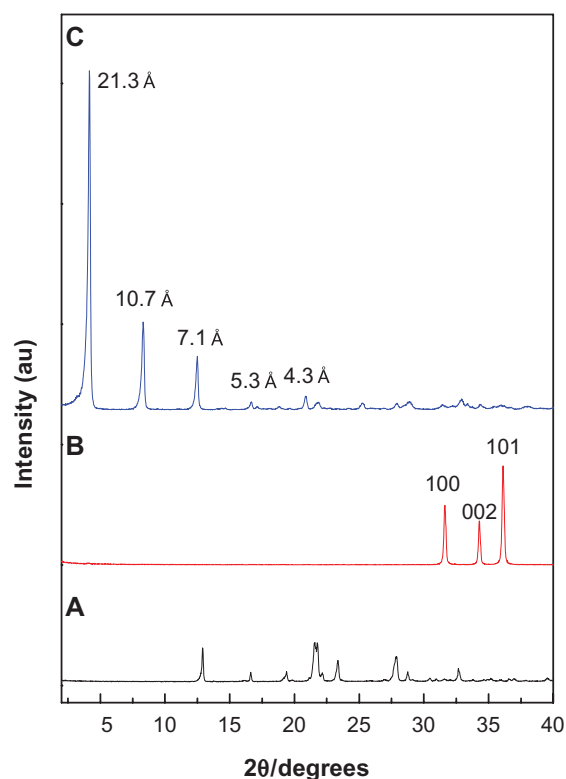


Figure 1 Powder X-ray diffraction patterns of (A) hippuric acid, (B) ZnO, and (C) HAN.

Abbreviations: ZnO, zinc oxide; HAN, hippuric acid nanocomposite.

Cytotoxicity of the combination of HAN with doxorubicin and oxaliplatin

In this work we used breast cancer cells MCF-7 and MDA-MB231 because they retained several ideal characteristics particular to the mammary epithelium.^{16,17} Caco2 was used as an in vitro model for colon cancer, which is the second most common cause of cancer in women, and the third most common in men.¹⁸ We found that HAN could potentiate the killing of breast and colon cancer cells induced by chemotherapeutic agents. In the absence of hippuric acid or HAN, the doxorubicin suppresses the cells growth of MCF-7 and MDA-MB231 cell lines with IC_{50} values $0.29 \pm 0.09 \mu\text{g/mL}$ and $0.20 \pm 0.06 \mu\text{g/mL}$, respectively. In Figure 2 and Table 1, the combination of hippuric acid with doxorubicin dose not shown significant reduction in the cell viability ($P > 0.05$). When closely examining each part in Figure 2, it is apparent that the combination of doxorubicin with HAN has a higher tumor inhibition efficiency compared to doxorubicin alone, and the IC_{50} values were $0.19 \mu\text{g/mL} \pm 0.15 \mu\text{g/mL}$ and $0.13 \mu\text{g/mL} \pm 0.10$ for the MCF-7 and MDA-MB231 cell lines, respectively. There was a significant decrease of MDA-MB231 and Caco2 viability at the combination concentration as low as $0.31 \mu\text{g/mL}$ till $20 \mu\text{g/mL}$ ($P < 0.05$, $P = 0.01$,

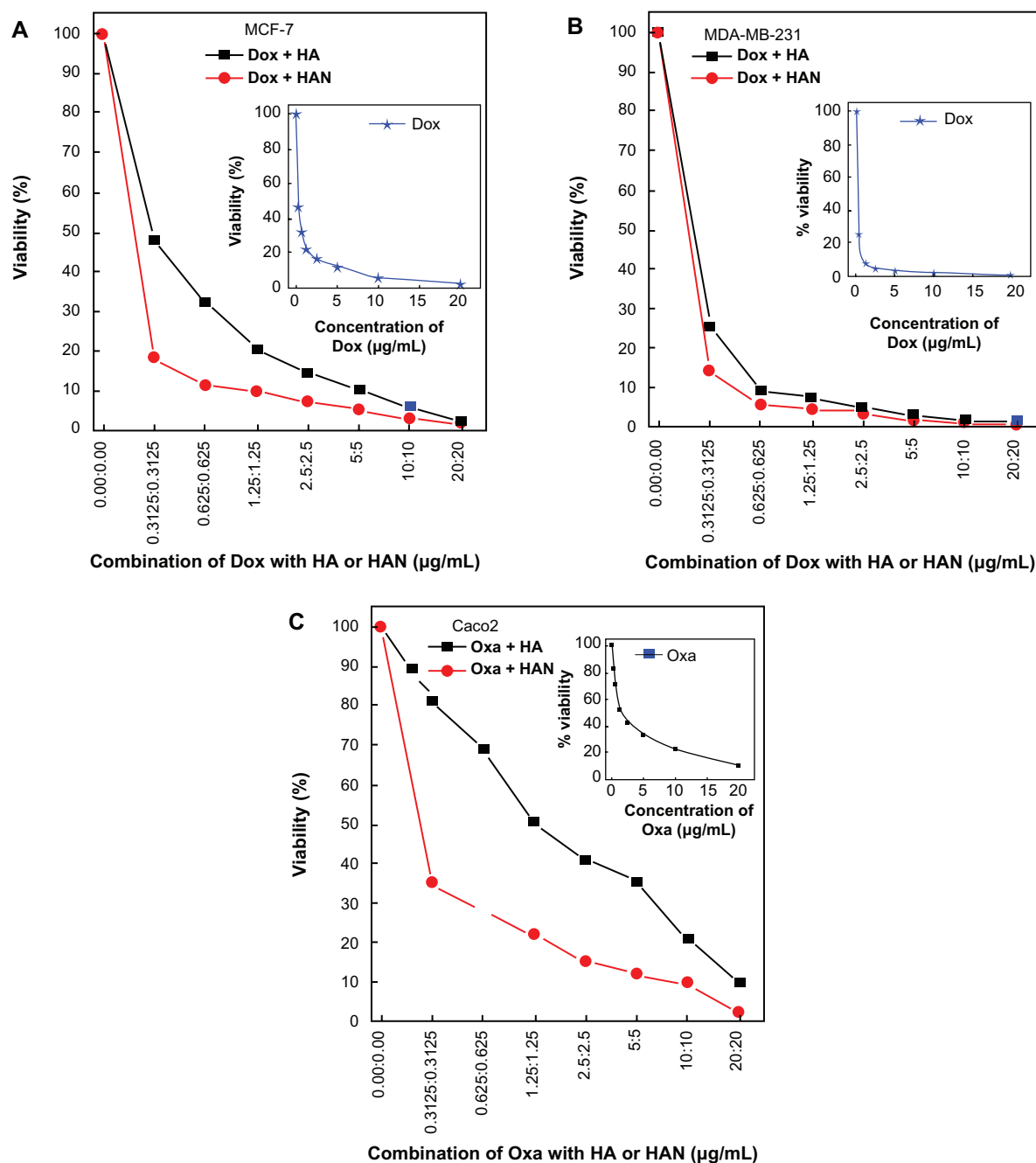


Figure 2 MTT assays of (A) MCF-7, (B) MDA-MB231, and (C) Caco2 cell lines after 72 hours of treatment.

Abbreviations: Dox, doxorubicin; HA, hippuric acid; HAN, hippuric acid nanocomposite; Oxa, oxaliplatin.

respectively). This result indicates that the suppression percentage in cells MCF-7 and MDA-MB231 are 34.4% and 35.0%, respectively.

Similarly, the combination of oxaliplatin with HAN against the Caco2 cells shows an suppression efficiency with IC_{50} of $0.24 \mu\text{g/mL} \pm 0.11$ compared with $1.48 \mu\text{g/mL} \pm 0.10$ for oxaliplatin alone. Our findings demonstrated that the co-treatment of Caco2 cell by oxaliplatin and HAN mixture shows maximal efficacy, this may be due to effect of the ZLH

nanolayer, which facilitates the entry of the oxaliplatin drug, as compared to doxorubicine.

Antiproliferative effect of free doxorubicin or oxaliplatin and a combination of drugs with hippuric acid and HAN

Figure 3 shows the effect of free doxorubicin and oxaliplatin drugs, and the combination of hippuric acid or HAN with that drugs on the proliferation of MCF-7, MDA-MB231,

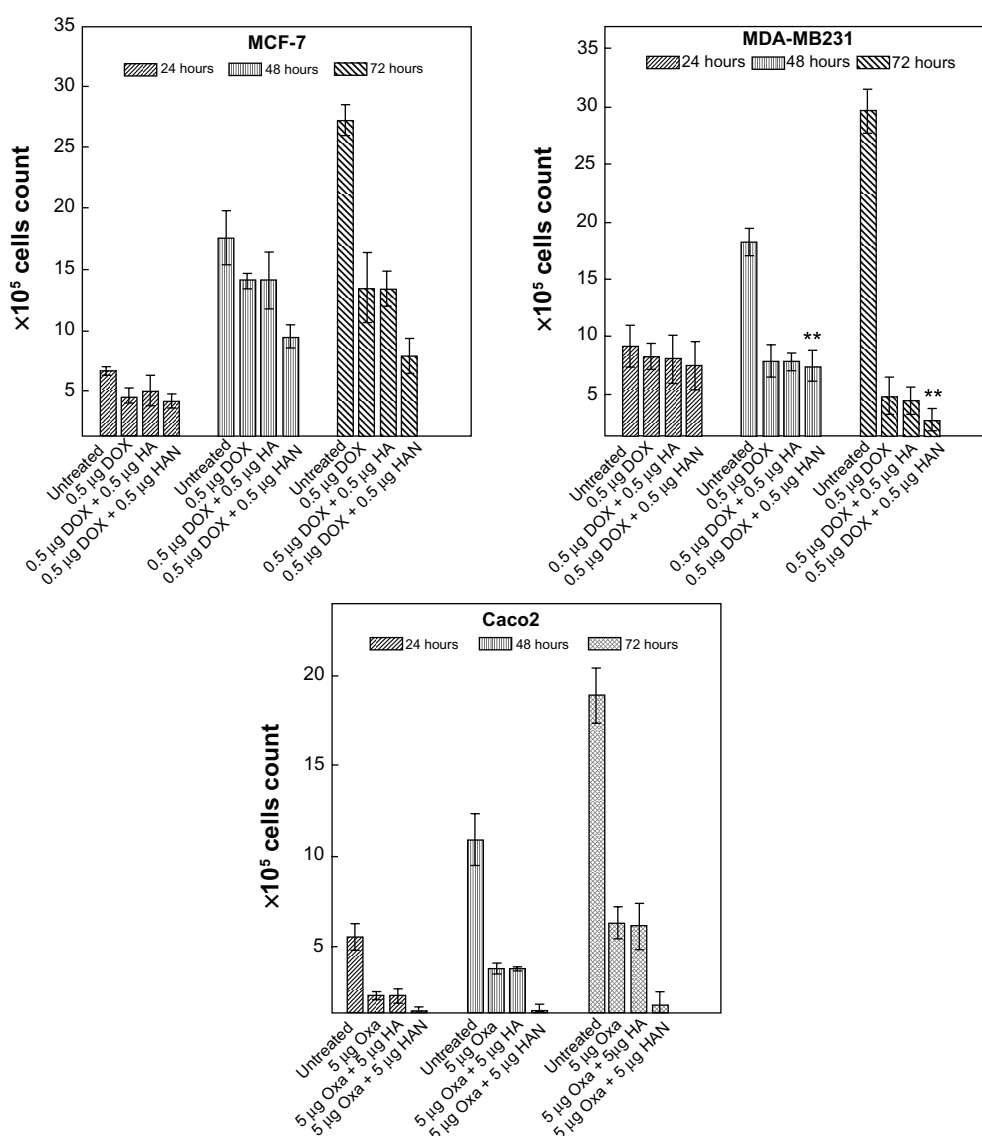
Table 1 IC₅₀ value of different cell lines treated with free doxorubicin or oxaliplatin, and a combination of free drugs with hippuric acid and HAN

	MCF-7			MDA-MB231			Caco2		
	Dox	Dox + HA	Dox + HAN	Dox	Dox + HA	Dox + HAN	Oxa	Oxa + HA	Oxa + HAN
IC ₅₀	0.29 ± 0.09	0.30 ± 0.06	0.19 ± 0.15	0.20 ± 0.06	0.21 ± 0.07	0.13 ± 0.10	1.48 ± 0.10	1.29 ± 0.03	0.24 ± 0.11

Abbreviations: HAN, hippuric acid nanocomposite; Dox, doxorubicin; HA, hippuric acid; Oxa, oxaliplatin.

and Caco2 cell lines at incubation times (24, 48, and 72 hours). However, doxorubicin and oxaliplatin drugs as well as combination of these drugs with HAN suppress the proliferation of MCF-7, MDA-MB231, and Caco2 tumor cells. The combination of these drugs with HAN are more efficient than the drugs alone against the growth of tumor cells. This result indicates that drug delivery to

the tumor cell is noticeably enhanced by nanocomposites containing ZLH; however, in the nanocomposites system, drugs can reach the tumor cell membrane without early decomposition, since the drugs are stabilized and protected in the interlayer space of the ZLH layers. These results clearly confirm that the intercalation reaction not only prevents drug denaturation, but it also enhances the

**Figure 3** Antiproliferative assays of (A) MCF-7, (B) MDA-MB231, and (C) Caco2 cell lines after, 24, 48, and 72 hours of treatment with free doxorubicin or oxaliplatin, and with a combination of doxorubicin or oxaliplatin with hippuric acid and HAN.

Abbreviations: Dox, doxorubicin; HA, hippuric acid; HAN, hippuric acid nanocomposite; Oxa, oxaliplatin.

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