Expression of transient receptor potential channel 6 in cervical cancer

Background: Recent studies have reported that aberrant expression of transient receptor potential channel C6 (TRPC6) in a variety of human cancers is associated with aggressive behavior. However, the functional significance of TRPC6 in human cervical cancer is not known. This study was planned to detect whether TRPC6 is expressed in cervical cancer tissue and to evaluate the association between TRPC6 expression and clinicopathologic features.

Methods: Tissue samples were collected from the West China Second UNIV Hospital of Sichuan University. TRPC6 expression was detected by quantitative real-time reverse transcription polymerase chain reaction and Western blotting. TRPC6 expression was evaluated by immunohistochemistry analysis of 40 cervical cancer specimens, and correlations were sought between elevated expression of TRPC6 and clinicopathologic features.

Results: Increased expression of TRPC6 was detected in 25 of the 40 cervical cancer samples. Positive cells found in cervical carcinomas were significantly increased in numbers compared with specimens without lymphovascular space invasion. Elevated expression of TRPC6 was neither related to International Federation of Gynecology and Obstetrics stage nor pelvic lymph metastases. Indeed, the clinicopathologic analysis indicated that overexpression of TRPC6 was significantly associated with lymphovascular space invasion.

Conclusion: These results indicate that elevated expression of TRPC6 might be associated with an aggressive cervical cancer phenotype.

Keywords: cervical cancer, transient receptor potential channel C6, overexpression, invasion
cation channel, which is expressed in a variety of tissues and is involved in the proliferation of cancer cells.\textsuperscript{3,5}

The transient receptor potential (TRP) superfamily is one of the largest ion channel families and consists of diverse groups of proteins. About 28 genes encode the TRP ion channel subunits in mammals. It comprises six subfamilies, including TRPC, which consists of seven proteins named TRPC1 to 7, and can be divided into three subgroups by sequence homology and functional similarities, ie, C1/C4/C5, C3/C6/C7, and C2.\textsuperscript{6} TRPC6 has been reported to be expressed in brain, lung, kidney, muscle and human platelets.\textsuperscript{7-11}

Overexpression of this channel has also been found in Ca\textsuperscript{2+} signal generation involving many malignant tissues and cells, but the role and mechanism for this remain elusive. There is much evidence suggesting that TRPC6 is involved in the oncogenic process, and is selectively expressed in aggressive cancers and intimately involved in metastases.\textsuperscript{3,5,12,13} Recently, it has been reported that TRPC6 is also expressed in esophageal, gastric, breast, and prostate tissue, where elevated channel expression has been implicated in cancer of these organs. The role of TRPC6 in cell proliferation has been demonstrated in several types of cancer cells, using specific inhibitors, ie, SKF96365 or short interfering RNA blockade of TRPC6 channels, which suppress cancer cell proliferation.\textsuperscript{3,5,11,14} However, it is not clear how it works or even whether it works in cervical cancer.

It is clinically important to understand the role of ion channels in the development of cancer. Indeed, TRPC6 is strongly expressed in prostate and breast cancers. Moreover, its expression correlates with Gleason score and aggressiveness. On the basis of these observations, we hypothesized that this protein might be expressed in cervical carcinoma and an increased level of human TRPC6 protein might be associated with tumor invasion. To evaluate this hypothesis, we investigated the expression of TRPC6 protein in benign and malignant human cervical tissue, and correlations were sought between TRPC6 protein immunostaining and stage, L VSI, and pelvic lymph node metastases of the tumors.

### Materials and methods

#### Samples

Forty paraffin-embedded specimens and 12 paired fresh cervical samples were collected from the West China Second UNIV Hospital of Sichuan University (Sichuan, China) with ethics committee approval and consent from the patients. Tumor specimens were obtained from 40 cervical cancer patients, staged following International Federation of Gynecology and Obstetrics (FIGO) criteria, ie, 18 stage I tumors, 15 stage II tumors and 7 stage III tumors. All of the samples were squamous cell carcinomas. Normal tissue samples were collected from six healthy women who had undergone elective total hysterectomies due to benign uterine disease (leiomyoma). These samples did not contain any histologic evidence of cervical intraepithelial neoplasia or invasive carcinoma. Clinicopathologic parameters, including age at time of surgery, tumor grade, lymph node status, and presence of L VSI, were extracted from the histopathologic examination results.

#### Total RNA extraction

All the samples used were analyzed by two experienced pathologists to ensure that they were correctly identified as either benign cervical tissue or cervical cancer. Total tissue RNA was extracted using Trizol reagent (Invitrogen, Carlsbad, CA), and cDNA was synthesized from total RNA using oligodT primers (Invitrogen). RNA integrity was confirmed by electrophoresis on 1% agarose gels stained with ethidium bromide.

#### Quantitative reverse-transcription PCR

Quantitative reverse-transcription polymerase chain reaction (PCR) was utilized to compare the relative amounts of TRPC6 in the fresh cervical tissue biopsy samples and was carried out on a Bio-Rad System (CFX96, Hercules, CA). For each reaction, cDNA was added to 20 \( \mu \)L of reaction mixture containing 10 \( \mu \)L of SYBR Green PCR Master Mix (Toyobo, Osaka, Japan) and 300 nM primers (Invitrogen). For quantification, the target sequence was normalized to GAPDH mRNA levels. PCR was performed under the following conditions: 95°C for 30 seconds, 57°C for 15 seconds, and 72°C for 30 seconds for 40 cycles. Heating for 2 minutes at 95°C preceded the cycles. We quantified the results using the comparative CT method. PCR products were analyzed by electrophoresis with 1.5% agarose gel and visualized by ethidium bromide staining. Primer sequences used for amplification were as follows: TRPC6 (product size 152 bp) upstream primer, 5’-GCCAATGAGCATCTGGAAAT-3’; downstream primer, 5’-AACCTCTTGGCTTACGAAA-3’; GAPDH (product size 200 bp) upstream primer, 5’-AACTGCTTAGCACCCCTGGC-3’; downstream primer, 5’-ATGACCTTGCCCACAGCCTT-3’.

#### Western blotting

Cervical tissue proteins were extracted using ice-cold lysis buffer containing a protease inhibitor cocktail (Roche, Meylan, France), and the proteins in the supernatant were...
quantified using the bicinechonic acid method (Pierce, Rockford, IL). Fifty micrograms of protein were separated per lane by 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis. After electrophoretic transfer of the proteins to a polyvinylidene difluoride membrane (Millipore, Bedfordshire, UK) using a MiniProtein III system (Bio-Rad), immunoblotting was performed using rabbit polyclonal primary anti-TRPC6 antibody (diluted 1:2000, Abcam, Cambridge, UK) and primary anti-β-actin (diluted 1:4000, Santa Cruz Biotechnologies, Santa Cruz, CA), and developed in an enhanced chemiluminescence system (Pierce) using specific peroxidase-conjugated anti-IgG secondary antibodies (1:4000 dilution, Santa Cruz). For quantification purposes, densitometric measurements were performed using the Quantity One image analysis software for Windows (Bio-Rad). All TRPC6 values were normalized to β-actin levels.

### Immunohistochemistry

Paraffin sections were cut and mounted on glass slides, and 5 μm sections from formalin-fixed and paraffin-embedded specimens were deparaffinized using xylene and rehydrated in graded ethanol. Samples were then preincubated with 3% H2O2 to eliminate endogenous peroxidase activity. Antigen retrieval was achieved by heating the sections (for 2 minutes to 100°C) in citric acid buffer (0.01 mol/L, pH 6.0). Sections were incubated at 4°C overnight in a 1:800 dilution of the primary anti-TRPC6 antibody (Abcam). After three washes, goat anti-rabbit horseradish peroxidase-conjugated antibody (Envision detection kit, Gene-Tech, Shanghai, China) was applied for one hour. Sections were developed using a peroxidase substrate DAB kit (Gene-Tech) and counterstained using hematoxylin. Sections were subsequently dehydrated before they were mounted on coverslips. Images were collected using an Eclipse TE2000-U microscope system (Nikon, Tokyo, Japan). The primary antibody was replaced with phosphate-buffered saline in the control experiments. All of the sections diagnosed as cervical squamous cell carcinoma were evaluated using conventional hematoxylin and eosin staining.

TRPC6 staining in immunohistochemistry was evaluated by the percentage of positive cells: 0%–10% of positive tumor cells; 10%–25% of positive tumor cells; 25%–50% of positive tumor cells; and >50% of positive tumor cells. Tumors were evaluated by random observation of five fields on every section by two investigators. The mean score served as the tumor TRPC6 expression score. Overexpression was defined as >25% of positive cell staining in the study without consideration of the intensity of staining.

### Statistical analysis

All the experiments were performed at least three times. Contingency table, χ2 analyses, and group means were compared by one-way analysis of variance using the Statistical Package for Social Sciences software version 17.0 (SPSS Inc, Chicago, IL). Differences resulting in P < 0.05 were considered to be statistically significant.

### Results

#### Increased TRPC6 expression in human epithelial cervical cancer

To determine whether TRPC6 mRNA expression differs in benign and malignant cervical tissue, tissue mRNA extracted from different patients was analyzed using quantitative PCR. The results presented are the means of the measurements for 12 individual specimens. From our study, it can be seen that both benign and diseased tissue expressed TRPC6 at the genetic level. TRPC6 expression was significantly higher in malignant tissue than in benign cervical tissue (P < 0.05, Figure 1).

#### Expression of TRPC6 in benign and malignant human cervical tissues

To our knowledge, no data are available on expression of TRPC6 in human cervical carcinoma tissue. We verified TRPC6 expression in tissues freshly collected after surgery. Transcripts for TRPC6 were expressed in these tissues, and expression was clearly higher in cancer tissue than in benign tissue by semiquantitative Western blotting (P < 0.05, Figure 2). Analysis of TRPC6 protein expression showed results consistent with quantitative PCR.
Association of TRPC6 protein expression and LVI in cervical carcinoma

In the present study, 40 cervical carcinoma specimens in paraffin sections were analyzed using immunohistochemistry in order to determine expression of TRPC6. TRPC6 protein was localized to the plasma membrane, and only the channels expressed in the zones affected by carcinoma were included. Consequently, overexpression of the protein was detected in 10 of 18 stage I tumors (55.6%), eight of 15 stage II tumors (53.3%), and seven of seven stage III tumors (100%). A Chi-square test showed that TRPC6 expression did not significantly correlate with clinicopathologic factors, including age, FIGO stage, and lymph node metastatic status \((P > 0.05)\). TRPC6 expression was significantly associated with LVI of cervical carcinoma \((P < 0.05, \text{Table } 1)\). Staining was less intense when the primary antibody was substituted with phosphate-buffered saline (Figure 4). Although more than 50% of tumor cells expressing TRPC6 protein was noted in 11.1% of stage I, 26.6% of stage II, and 42.9% of stage III tumors, no significant difference was found in FIGO stage \((P > 0.05)\).

Discussion

In our study, we investigated the association between expression of TRPC6 in squamous cell carcinoma and LVI. We initially assessed the expression of TRPC6 channels in benign and cervical cancer tissue samples removed at surgery. Increased expression of TRPC6 in patients with malignant cervical tumors was found. Second, results from the Western blotting and quantitative PCR were consistent. Expression of TRPC6 was shown in all of the tissues, while overexpression was only found in malignant tissues. Third, on the basis of results from histologic sections of cervical cancer tissue, it can be concluded that LVI is associated with increased numbers of TRPC6 positive cells. About 25%–50% of positive cells were found in 11 of 16 (68.75%) LVI samples, and more than 50% were shown in six of nine (66.7%) LVI samples. There was a significant association between LVI and TRPC6 positive cells \((P < 0.05)\).

LVI, including lymphatic vessel invasion and blood vessel invasion, is an important factor in the metastatic potential of cervical carcinoma. Therefore, it plays a predictive role in lymph node metastases and ovarian metastases.\(^{15-18}\) Patients with LVI may have completely different clinical outcomes. This study shows that overexpression of TRPC6 is associated with malignant tissue and LVI, and therefore it may be a useful biomarker for metastases in cervical cancer. Determination of levels of this protein may help to guide adjuvant therapies, such as radiotherapy and/or chemotherapy, for an individual patient with cervical cancer postoperatively. Otherwise, blockers targeted toward these channels may provide promising new therapeutic solutions to protect against development of tumors.

Indeed, evidence from existing research has shown that blocking TRPC6 using short interfering RNA inhibition of calcium ion influx induces cell cycle arrest in the G2/M phase and growth retardation in AGS and MKN45 gastric cancer

Table I Comparison of TRPC6 expression to tumor characteristics on 40 patients using \(\chi^2\) analysis

<table>
<thead>
<tr>
<th>Age</th>
<th>n</th>
<th>TRPC6 expression (positive cells ratio)</th>
<th>(\chi^2)</th>
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<tbody>
<tr>
<td>&gt;40</td>
<td>27</td>
<td>0%–10% 10%–25% 25%–50% &gt;50%</td>
<td>2.197</td>
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<tr>
<td>&lt;40</td>
<td>13</td>
<td>0%–10% 10%–25% 25%–50% &gt;50%</td>
<td>7.068</td>
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<tr>
<td>Pathological stage</td>
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<td></td>
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<tr>
<td>I</td>
<td>18</td>
<td>0%–10% 10%–25% 25%–50% &gt;50%</td>
<td>7.068</td>
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<tr>
<td>II</td>
<td>15</td>
<td>0%–10% 10%–25% 25%–50% &gt;50%</td>
<td>0.716</td>
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<tr>
<td>Pelvic lymph metastasis</td>
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<td></td>
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<tr>
<td>+</td>
<td>17</td>
<td>0%–10% 10%–25% 25%–50% &gt;50%</td>
<td>9.85</td>
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<tr>
<td>Lymphovascular space invasion</td>
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<tr>
<td>+</td>
<td>20</td>
<td>0%–10% 10%–25% 25%–50% &gt;50%</td>
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<td>0%–10% 10%–25% 25%–50% &gt;50%</td>
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Abbreviations: TRPC6, transient receptor potential channel C6.
out that tumor cells in the G2/M phase are most sensitive for the small sample size in our study. It should be pointed samples at an advanced clinical stage, and this is the reason Ι
ommended instead of surgery for patients with stage FIG
25 squamous cell carcinoma tissue slices, did not indicate histochemistry to detect overexpression TRPC6 protein in breast and prostate cancer, our study, which used immuno TRPC6 expression predicts a poor clinical prognosis in the growth of tumor cells. Although previous studies have shown that positive TRPC6 expression predicts a poor clinical prognosis in breast and prostate cancer, our study, which used immuno cell lines. Similar results were also observed in esophageal and ovarian cancer cell lines. Moreover, inhibition ofTRPC6 or TRPC3 can suppress tumor formation in nude mice. It is thought to regulate by capacitative Ca⁺⁺ entry. Ca⁺⁺ signaling is important for proliferation and survival of cancer cells, and TRPC6-mediated Ca⁺⁺ influx also increases RhoA activity, which may regulate cell migration. Therefore, we have reason to speculate that TRPC6, which is mainly permeable to Ca⁺⁺, may play some role in the development of cancer. TRPC6 and TRPC3 are nonselective cation channels, and are directly activated by diacylglycerol through a mechanism independent of protein kinase C. Increases in Ca⁺⁺ influx not only control gene expression, cell cycle progression, and apoptosis, but also modify cell growth and differentiation. However, overexpression of TRPC6 was not observed in esophageal and HEK293 cell proliferation or an alteration in G2/M phase transition. Based on these results, it is suggested that Ca⁺⁺ regulated by TRPC6 channels might be involved in tumorigenesis, but it does not control the growth of tumor cells.

Although previous studies have shown that positive TRPC6 expression predicts a poor clinical prognosis in breast and prostate cancer, our study, which used immunohistochemistry to detect overexpression TRPC6 protein in 25 squamous cell carcinoma tissue slices, did not indicate that TRPC6 expression was significantly correlated with FIGO stage in cervical cancer (P > 0.05).

Radiotherapy and chemotherapy is conventionally recommended instead of surgery for patients with stage III and IV cervical cancer. Therefore, it is difficult to find tissue samples at an advanced clinical stage, and this is the reason for the small sample size in our study. It should be pointed out that tumor cells in the G2/M phase are most sensitive to radiation, with other studies reporting that downregulation of TRPC6 contributes to G2/M cell cycle transition of cancer cells. We propose that FIGO III stage cervical cancer with upregulated expression of TRPC6 is likely to be more sensitive to radiotherapy.

In our experiment, we only detected expression of TRPC6 and the relationship between its expression and the prognosis of cervical cancer. The mechanism of increased TRPC6 channel expression in cervical cancer remains unknown. We postulate that the development of cervical carcinoma via increased TRPC6 channel expression might be associated with exchange of calcium ions. Further research is required to clarify the role of TRPC6 in this process.

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**Disclosure**

The authors report no conflicts of interest in this work.

**References**