Thrombophilia and retinal vascular occlusion

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Purpose: The purpose of this research was to assess associations of thrombophilia with central retinal vein occlusion (CR VO), central retinal artery occlusion (CRAO), and amaurosis fugax (AF); to evaluate outcomes of normalizing high homocysteine; and to study CR VO, CRAO, and AF developing in estrogens/estrogen agonists in women subsequently shown to have thrombophilia.

Methods: Measures of thrombophilia–hypofibrinolysis were obtained in 132 CR VO cases, 15 CRAO cases, and 17 AF cases. Cases were compared to 105 healthy control subjects who did not differ by race or sex and were free of any ophthalmologic disorders. All cardiovascular disease (CVD) risk factors were compared to healthy general populations.

Main outcome measures: The main outcome measure of this study was thrombophilia.

Results: CR VO cases were more likely than controls to have high homocysteine (odds ratio [OR] 8.64, 95% confidence intervals [CI]: 1.96–38), high anticardiolipin immunoglobulin M (IgM; OR 6.26, 95% CI: 1.4–28.2), and high Factor VIII (OR 2.47, 95% CI: 1.31–7.9). CRAO-AF cases were more likely than controls to have high homocysteine (OR 14, 95% CI: 2.7–71.6) or the lupus anticoagulant (OR 4.1, 95% CI: 1.3–13.2). In four of 77 women with CRVO (two found to have high homocysteine, two with inherited high Factor XI), CRVO occurred after starting estrogen–progestins, estrogen–testosterone, or estrogen agonists. In one of eight women with CRAO found to have high anticardiolipin antibody IgG, CRAO occurred after starting conjugated estrogens, and AF occurred after starting conjugated estrogens in one of eleven women with AF (inherited protein S deficiency). Therapy for medians of 21 months (CR VO) and 6 months (CRAO-AF) was 5 mg folic acid, 100 mg B6, and 2000 mcg/day B12 normalized homocysteine in 13 of 16 (81%) CR VO cases and all five CRAO-AF cases with pretreatment hyperhomocysteinemia. The CRVO cases had an excess of hypertension; CRAO-AF cases had an excess of type 2 diabetes and hypertension.

Conclusion: Treatable thrombophilia, hyperhomocysteinemia in particular, is more common in RVO cases than in normal controls. RVO occurs after estrogens or estrogen agonists were administered in women subsequently shown to have thrombophilia.

Keywords: central retinal vein occlusion, central retinal artery occlusion, amaurosis fugax, retinal vascular occlusion, thrombophilia, estrogen, estrogen agonist

Introduction

Retinal vascular occlusion (RVO), which includes central retinal vein occlusion (CRVO), central retinal artery occlusion (CRAO), and amaurosis fugax (AF), can be caused by inherited and acquired thrombophilia, particularly homocysteinemia.1–5 Thrombophilia associated with exogenous estrogens,6 estrogen progestin oral contraceptives,7 clomiphene citrate,8 or selective estrogen receptor modulators (SERMS)9
may promote CRVO, AF, and CRAO. Most studies associating estrogen–progestin oral contraceptives with CRVO or CRAO are case reports of only one to three cases. Most reports of CRVO or CRAO associated with estrogens or estrogen agonists have not assessed interactions between pharmacologic thrombophilia conferred by estrogens or estrogen agonists and the inherited and acquired thrombophilia known to be causally associated with RVO.

Our specific aim was to assess associations of inherited and acquired thrombophilia-hypofibrinolysis with CRVO, CRAO, and AF; to evaluate outcomes of normalizing high homocysteine levels; and to study CRVO, CRAO, and AF development after the use of estrogens or estrogen agonists in women subsequently shown to have thrombophilia.

Materials and methods
Setting and study design
Cases
The study was carried out following a protocol approved by our Institutional Review Board, and with signed informed consent from both the cases and controls. In consecutive order of their referral by six vitreoretinal specialists (two academic, four community-based) in an outpatient clinical research center, we prospectively studied 164 RVO cases (68 men, 96 women). These 164 cases included 132 with CRVO (55 men, 77 women), 15 with CRAO (seven men, eight women), and 17 with AF (six men, eleven women). There were no known selection biases for referral. We excluded from this study any CRAO-AF cases who had hemodynamically significant ipsilateral carotid-vertebral atherosclerotic lesions by carotid-vertebral Doppler measures, or who had coronary left-to-right shunts determined by trans-esophageal echo studies.

One or more months after their RVO, serologic coagulation assays were done. No cases had taken warfarin or heparin within 3 months of blood sampling. Most cases with CRVO had bevacizumab intraocular injections. At each case’s initial visit, a detailed history and physical examination were carried out, with a focus on cardiovascular events, hypertension, diabetes, cigarette smoking, pulmonary embolus, deep venous thrombosis, reproductive history, estrogen-containing oral contraceptives, hormone replacement therapy, clomiphene citrate, SERMS, and therapy for hypertension, diabetes, and hyperlipidemia. Blood specimens for coagulation measures were obtained from seated cases and controls, as previously described, between 8 am and 10 am the morning after an overnight fast. CRVO was diagnosed by the referring vitreoretinal specialists based on the results of characteristic fundus features including retinal hemorrhages in all four quadrants of the fundus with a dilated, tortuous retinal venous system.

CRAO was diagnosed by the referring vitreoretinal specialists by the presence of acute painless loss of vision with central, dense visual loss. Funduscopic criteria included a whitened retina with a cherry red macula (the “cherry-red spot”), resulting from the obstruction of blood flow to the retina from the central retinal artery. A continued supply of blood to the choroid from the short ciliary arteries resulted in a bright red coloration at the thinnest part of the retina, the macula. We included only those CRAO cases without ipsilateral carotid atherosclerosis.

AF was diagnosed by transient monocular visual loss with normal funduscopic examination in cases without ipsilateral carotid atherosclerotic plaque and without evidence of temporal arteritis.

Cases with hyperhomocysteinemia at entry were treated with folic acid (5 mg/day), vitamin B6 (100 mg/day), and vitamin B12 (2000 mcg/day), with repeat measures of fasting serum homocysteine every 3–4 months and repeated funduscopic examinations every 3–6 months.

Controls
For comparison with the cases’ polymerase chain reaction (PCR) and serologic measures of thrombophilia-hypofibrinolysis, the 132 CRVO cases and the 32 CRAO-AF cases were compared to 105 healthy controls (45 men, 60 women) that had not sustained RVO and did not differ from the cases by ethnicity or sex. By selection, no male controls were taking testosterone and no female controls were taking estrogen–progestin oral contraceptives, hormone replacement therapy, clomiphene citrate, or SERMS. To assess the contributions of cardiovascular risk factors to CRVO and CRAO-AF, cases’ lipid levels were characterized by their percentile distributions within age- and gender-specific lipid distributions from healthy general populations documented in the Lipid Research Clinics Prevalence Study. Separately, the prevalence of type 2 diabetes, hypertension, and smoking in cases with CRVO and CRAO-AF were compared to US population estimates.

PCR assays
PCR measures of thrombophilia-hypofibrinolysis included G1691A Factor V Leiden, G20210A prothrombin, MTHFR C677T-A1298C, and 4G5G plasminogen activator inhibitor activity. These PCR measures were performed in cases with
CRVO and CRAO-AF and in healthy controls using previously published methods by laboratory staff blinded to the subjects’ status (case, control, and severity).15,24,25,28,29,39–44

Serologic measures of thrombophilia
Serologic measures of thrombophilia included anticardiolipin antibodies (IgG and IgM), antigenic protein C, total and free antigenic protein S, antithrombin III, resistance to activated protein C (RAPC), activated partial thromboplastin time, dilute Russell’s viper venom time (DRVVT), lupus anticoagulant, factors VIII and XI, and homocysteine. Established, previously published methods were used.39,44–46
To be considered abnormal, high anticardiolipin antibodies and the lupus anticoagulant had to be abnormal in a second test done 12 weeks after the first. Testing was not done for anti-beta 2 globulin.

Serologic measures of hypofibrinolysis
Serologic measures of hypofibrinolysis included lipoprotein a (Lp(a)) and plasminogen-activator inhibitor activity. These measures were performed using established methods.29,45,46

Statistical methods
All statistical comparisons were done using SAS software (SAS/STAT, Release 9.1; SAS Institution, Cary, NC). The proportions of RVO cases and healthy normal controls having abnormalities in coagulation measures were compared using odds ratios and 95% confidence intervals, and by $\chi^2$ analyses or the Fisher exact test when cell sizes were <5. Serum homocysteine levels pre- and on-treatment with folic acid, vitamin B6, and vitamin B12 were compared by paired Wilcoxon tests of difference.

Results
CRVO, CRAO, and AF in the total cohort
Of the 132 CRVO cases, 121 were white, six black, and five other; 55 were men and 77 women; the mean ± standard deviation (SD) age was 57 ± 14 years. Of the 32 CRAO-AF cases, 30 were white and two black; 13 were men; the mean ± SD age was 52 ± 16 years. CRVO and CRAO-AF cases did not differ from the 105 healthy controls by ethnicity (92 white, six black, seven other; $P = 0.54, P = 0.43$, respectively) or by sex (44 men, 61 women; $P = 0.97, P = 0.90$), but the controls were younger (44.2 ± 12 years, $P < 0.0001$, $P = 0.003$, respectively).
In all of the 132 CRVO and 32 CRAO-AF cases, CRVO and CRAO-AF were the cases’ first thrombotic event. By selection, CRAO-AF cases did not have carotid artery atherosclerosis or right-to-left heart shunts, which might have accounted for their RVO.

Case-control differences in thrombophilia
CRVO cases were more likely than normal controls to have high homocysteine, high anticardiolipin IgM, and high Factor VIII (Table 1). Low free-antigenic protein S was

<table>
<thead>
<tr>
<th>Coagulation measures (normal range)</th>
<th>Abnormal in CRVO</th>
<th>Abnormal in controls</th>
<th>$P$</th>
<th>OR, 95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Homocysteine (≤13.5 μmol/L)</td>
<td>15% (19/129)</td>
<td>2% (2/102)</td>
<td>$\chi^2 = 11.2, P = 0.0008$</td>
<td>8.64, 1.96–38.0</td>
</tr>
<tr>
<td>Free antigenic protein S (≥66%)</td>
<td>9% (11/120)</td>
<td>2% (2/92)</td>
<td>$\chi^2 = 4.4, P = 0.04$</td>
<td>4.54, 0.98–21.0</td>
</tr>
<tr>
<td>Anticardiolipin antibody IgM (&lt;10 MPL)</td>
<td>11% (14/128)</td>
<td>2% (2/104)</td>
<td>$\chi^2 = 7.3, P = 0.007$</td>
<td>6.26, 1.39–28.2</td>
</tr>
<tr>
<td>Factor VIII (≥150%)</td>
<td>20% (23/116)</td>
<td>7% (7/98)</td>
<td>$\chi^2 = 7.1, P = 0.008$</td>
<td>3.22, 1.31–7.86</td>
</tr>
<tr>
<td>Factor V Leiden mutation</td>
<td>5% CT (6/130)</td>
<td>2% CT (2/104)</td>
<td>Fisher’s $P = 0.3$</td>
<td></td>
</tr>
<tr>
<td>Resistance to activated protein C (detailed cutpoint)</td>
<td>8% (9/112)</td>
<td>5% (5/92)</td>
<td>$\chi^2 = 0.53, P = 0.5$</td>
<td></td>
</tr>
<tr>
<td>Prothrombin gene mutation</td>
<td>3% CT (4/127)</td>
<td>3% CT (3/105)</td>
<td>Fisher’s $P = 1.0$</td>
<td></td>
</tr>
<tr>
<td>Plasminogen activator inhibitor mutation</td>
<td>28% 4G4G (36/127)</td>
<td>26% 4G4G (26/100)</td>
<td>Mantel-Haenszel</td>
<td></td>
</tr>
<tr>
<td>46% 4G5G (59/127)</td>
<td>43% 4G5G (43/100)</td>
<td>Mantel-Haenszel</td>
<td>$\chi^2 = 0.67, P = 0.4$</td>
<td></td>
</tr>
<tr>
<td>MTHFR C677T mutation</td>
<td>22% CC (29/130)</td>
<td>31% CC (31/101)</td>
<td>Mantel-Haenszel</td>
<td></td>
</tr>
<tr>
<td>35% CT (45/130)</td>
<td>14% CT (14/101)</td>
<td>Fisher’s $P = 0.7$</td>
<td>$\chi^2 = 0.13, P = 0.7$</td>
<td></td>
</tr>
<tr>
<td>Antigenic protein C (≥73%)</td>
<td>6% (7/127)</td>
<td>7% (6/92)</td>
<td>$\chi^2 = 0.10, P = 0.8$</td>
<td></td>
</tr>
<tr>
<td>Antigenic protein S (≥63%)</td>
<td>2% (3/125)</td>
<td>4% (4/92)</td>
<td>Fisher’s $P = 0.5$</td>
<td></td>
</tr>
<tr>
<td>Antithrombin III (≥80%)</td>
<td>7% (8/121)</td>
<td>2% (2/92)</td>
<td>Fisher’s $P = 0.2$</td>
<td></td>
</tr>
<tr>
<td>Anticardiolipin antibody IgG (&lt;22 GPL)</td>
<td>10% (13/127)</td>
<td>7% (7/104)</td>
<td>$\chi^2 = 0.89, P = 0.4$</td>
<td></td>
</tr>
<tr>
<td>Plasminogen activator activity (=21.1 U/mL)</td>
<td>10% (11/107)</td>
<td>10% (10/97)</td>
<td>$\chi^2 = 0.0, P = 1.0$</td>
<td></td>
</tr>
<tr>
<td>Lp(a) (&lt;35 mg/dL)</td>
<td>20% (25/123)</td>
<td>20% (20/102)</td>
<td>$\chi^2 = 0.02, P = 0.9$</td>
<td></td>
</tr>
<tr>
<td>Resistance to activated protein C (detailed cutpoint)</td>
<td>8% (9/112)</td>
<td>5% (5/92)</td>
<td>$\chi^2 = 0.53, P = 0.5$</td>
<td></td>
</tr>
<tr>
<td>Factor XI (≥150%)</td>
<td>4% (5/114)</td>
<td>2% (2/96)</td>
<td>Fisher’s $P = 0.5$</td>
<td></td>
</tr>
<tr>
<td>Lupus anticoagulant</td>
<td>10% (9/87)</td>
<td>9% (7/81)</td>
<td>$\chi^2 = 0.14, P = 0.7$</td>
<td></td>
</tr>
</tbody>
</table>

Abbreviations: CI, confidence interval; C, mutant allele; OR, odds ratio; T, wild-type normal allele; Lp(a), lipoprotein a; CRVO, central retinal vein occlusion.
marginally more common (by Chi squared analysis) in cases than in controls (Table 1). There were no other case–control differences ($P > 0.05$) in any other CRVO case–control comparisons of coagulation measures (Table 1).

CRAO-AF cases were more likely than normal controls to have high homocysteine or the lupus anticoagulant (Table 2). There were no other case–control differences ($P > 0.05$) in any other case–control comparisons of coagulation measures (Table 2).

### Normalization of elevated serum homocysteine by folic acid, vitamin B6, and vitamin B12

Of the 19 CRVO cases with high pre-treatment homocysteine (Table 1), 16 had had treatment with folic acid–vitamin B6–vitamin B12 with a median follow-up of 21 months (interquartile range 3–60 months). Homocysteine levels fell in every case on treatment, from $19.1 \pm 8.7$ to $9.8 \pm 3.9 \mu mol/L, P < 0.0001$, and normalized ($\leq 13.5 \mu mol/L$) in 13 of 16 cases (81%). In these 16 cases, there were no new CRVO events during treatment.

Of the seven CRAO-AF cases with high pre-treatment homocysteine (Table 2), five were treated with folic acid–vitamin B6–vitamin B12 for a median of 6 months (interquartile range 5–8 months). Homocysteine levels normalized in all five, falling from $15.8 \pm 1.1$ to $9.3 \pm 3.1 \mu mol/L, P = 0.06$. In these five cases, there were no new CRAO-AF events during treatment. There were no adverse side effects associated with the folic acid–vitamin B6–vitamin B12 treatment.

CRVO, CRAO, and AF after starting estrogen–estrogen agonist therapy in six of the women subsequently shown to have underlying inherited or acquired thrombophilia

Of 96 women in the cohort, eleven (11%) sustained an RVO while using estrogens or estrogen agonists, and six were found to have a previously undiagnosed thrombophilia (Table 3). Five of the six were 55 years old or older (Table 3). RVO was the first clinical thrombotic event in all six women (Table 3).

Of 77 women with CRVO, four (5%)–aged 37 to 53 years–presented after taking estrogen–progesterin oral contraceptives ($n = 2$), estrogen–testosterone ($n = 1$), or tamoxifen ($n = 1$) (Table 3). Of these four, two were subsequently shown to have high homocysteine, two had inherited high factor XI, and one had the lupus anticoagulant (Table 3). CRVO occurred after as little as 11 months to as long as 5 years after starting estrogen–progestins, estrogen–testosterone, or tamoxifen for all four women (Table 3).

Of eight women with CRAO, one (13%) presented after taking conjugated estrogen tablets for 8 years, and was subsequently found to have high anticardiolipin antibody IgG (Table 3).

### Table 2 Comparisons of coagulation measures between 32 patients – 15 with CRAO, 17 with AF, and 104 healthy, normal controls

<table>
<thead>
<tr>
<th>Coagulation measures (normal range)</th>
<th>Abnormal in CRAO</th>
<th>Abnormal in controls</th>
<th>$P$</th>
<th>OR, 95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Homocysteine ($\leq 13.5 \mu mol/L$)</td>
<td>22% (7/32)</td>
<td>2% (2/102)</td>
<td>Fisher’s $P = 0.0006$</td>
<td>14.0, 2.7–71.6</td>
</tr>
<tr>
<td>Lupus anticoagulant</td>
<td>28% (7/25)</td>
<td>9% (7/81)</td>
<td>Fisher’s $P = 0.02$</td>
<td>4.1, 1.3–13.2</td>
</tr>
<tr>
<td>Factor V Leiden mutation</td>
<td>6% CT (2/32)</td>
<td>2% CT (2/104)</td>
<td>Fisher’s $P = 0.2$</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>Resistance to activated protein C (dated cutpoint)</td>
<td>7% (2/30)</td>
<td>5% (5/92)</td>
<td>Fisher’s $P = 1.0$</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>Prothrombin gene mutation</td>
<td>6% CT (2/32)</td>
<td>3% CT (3/105)</td>
<td>Fisher’s $P = 0.3$</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>Plasminogen activator inhibitor gene mutation</td>
<td>29% 4G4G (9/31)</td>
<td>26% 4G4G (26/100)</td>
<td>Mantel-Haenszel</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>MTHFR C677T mutation</td>
<td>42% CC (13/31)</td>
<td>31% CC (31/101)</td>
<td>Mantel-Haenszel</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>Antigenic protein C ($\geq 73%$)</td>
<td>13% (4/32)</td>
<td>7% (6/92)</td>
<td>Fisher’s $P = 0.3$</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>Antigenic protein S ($\geq 63%$)</td>
<td>0% (0/23)</td>
<td>4% (4/92)</td>
<td>Fisher’s $P = 0.3$</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>Antigenic-free protein S ($\geq 66%$)</td>
<td>3% (1/30)</td>
<td>2% (2/92)</td>
<td>Fisher’s $P = 1.0$</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>Antithrombin III ($\geq 80%$)</td>
<td>10% (3/31)</td>
<td>2% (2/92)</td>
<td>Fisher’s $P = 0.1$</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>Anticardiolipin antibody IgG ($&lt; 22$ GPL)</td>
<td>9% (3/32)</td>
<td>7% (7/104)</td>
<td>Fisher’s $P = 0.7$</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>Anticardiolipin antibody IgM ($&lt; 10$ MPL)</td>
<td>6% (2/32)</td>
<td>2% (2/104)</td>
<td>Fisher’s $P = 0.2$</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>Plasminogen activator inhibitor ($&lt; 21.1$ UI/mL)</td>
<td>13% (4/30)</td>
<td>10% (10/97)</td>
<td>Fisher’s $P = 0.7$</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>Lp(a) ($&lt; 35$ mg/dL)</td>
<td>13% (4/33)</td>
<td>20% (20/102)</td>
<td>$\chi^2 = 0.72$, $P = 0.4$</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>Factor VIII ($\leq 150%$)</td>
<td>19% (6/32)</td>
<td>7% (7/98)</td>
<td>Fisher’s $P = 1.0$</td>
<td>1.0, 0.5–2.0</td>
</tr>
<tr>
<td>Factor XI ($\leq 150%$)</td>
<td>9% (3/32)</td>
<td>2% (2/96)</td>
<td>Fisher’s $P = 0.1$</td>
<td>1.0, 0.5–2.0</td>
</tr>
</tbody>
</table>

**Abbreviations:** CI, confidence interval; C, mutant allele; OR, odds ratio; T, wild-type normal allele; Lp(a), lipoprotein a; CRVO, central retinal vein occlusion; Ig, immunoglobulin.
Of eleven women with AF, one (9%) presented after taking conjugated estrogen tablets for 2 years, and was subsequently found to have inherited protein S deficiency (Table 3).

Table 3 Central retinal vein, central retinal artery occlusion, and amaurosis fugax after the use of exogenous estrogens or estrogen agonists in six women subsequently shown to have inherited or acquired thrombophilia

<table>
<thead>
<tr>
<th>Case #</th>
<th>Age</th>
<th>Coagulation disorders</th>
<th>Exogenous estrogen or estrogen agonist</th>
<th>Duration of estrogen or estrogen agonist use prior to ocular thrombotic event</th>
</tr>
</thead>
<tbody>
<tr>
<td>Of 77 women with central retinal vein occlusion</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>51</td>
<td>High factor XI</td>
<td>Nolvadex</td>
<td>5 years</td>
</tr>
<tr>
<td>2</td>
<td>53</td>
<td>High serum homocysteine</td>
<td>Estrogen–testosterone</td>
<td>11 months</td>
</tr>
<tr>
<td>3</td>
<td>42</td>
<td>High serum homocysteine</td>
<td>Estrogen–progestin oral contraceptive</td>
<td>2 years</td>
</tr>
<tr>
<td>4</td>
<td>37</td>
<td>Lupus anticoagulant, high factor XI</td>
<td>Estrogen–progestin oral contraceptive</td>
<td>11 months</td>
</tr>
<tr>
<td>Of eight women with central retinal artery occlusion</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>55</td>
<td>High anticardiolipin antibody IgG</td>
<td>Conjugated estrogen tablets</td>
<td>8 years</td>
</tr>
<tr>
<td>Of eleven women with amaurosis fugax</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>79</td>
<td>Protein S deficiency</td>
<td>Conjugated estrogen tablets</td>
<td>2 years</td>
</tr>
</tbody>
</table>

Abbreviation: Ig, immunoglobulin.

Case-control differences in risk factors for atherosclerosis

Based on age–sex-specific lipid distributions from healthy general populations from the Lipid Research Clinics Prevalence Study, the mean percentiles in 132 CR VO cases were 34% for total cholesterol (TC), 52% for triglycerides (TGs), 49% for high-density lipoprotein cholesterol (HDLC), and 29% for low-density lipoprotein cholesterol (LDLC). In 32 CRAO-AF cases, the mean percentiles were 32% for TC, 47% for TG, 48% for HDLC, and 29% for LDLC. Thus, for TG and HDLC, CR VO and CRAO-AF cases were in the middle of the normal distribution, and both had TC and LDLC percentiles in the lower third of the distribution.

Of the 132 CR VO cases, 7% had type 2 diabetes, 43% had hypertension, and 17% smoked, compared to US population estimates of 8%, 24%, and 20%, respectively, with an excess of hypertension in CR VO cases. Of the 32 cases with CRAO-AF, 16% had type 2 diabetes, 34% had hypertension, and 16% smoked, with an excess of type 2 diabetes and hypertension in CRAO-AF cases.

Discussion

Our report, as well as earlier studies, has established thrombophilia as a common pathoetiologic cause of RVO. In this study, in agreement with previously published studies, we found significant enrichment in hyperhomocysteinemia in our 132 CVRO cases (15% versus 2% in controls; \( P = 0.0008 \)). Hansen et al reported a high prevalence of hyperhomocysteinemia in retinal venous thrombosis, even superseding the prevalence of venous thromboembolism in other compartments.

Sofi et al reported that low vitamin B6 levels, low folic acid levels, and elevated homocysteine levels were each independently associated with CRVO, offering therapeutic targets for normalization of serum vitamin B6 and B12 and folate levels to lower homocysteine, as was successfully done in our study.

In the current study of CR VO cases, and congruent with other reports, when compared to normal controls, we found that CR VO cases had high anticardiolipin antibodies, inherited low protein S, and inherited high factor VIII.

Congruent with previously published studies, we found that homocysteinemia was much more common in cases with CRAO-AF than in controls (22% versus 2%; \( P = 0.0006 \)). In agreement with previous reports, cases with CRAO-AF were more likely than normal controls to have the lupus anticoagulant.

A novel finding of the current study was that treatment with folic acid-B6-B12 normalized serum homocysteine in 81% of CR VO cases and in 100% of CRAO-AF cases with pre-treatment homocysteinemia. Normalizing high serum homocysteine levels in cases with CRVO and CRAO-AF may reduce the risk of subsequent ocular venous or arterial thrombosis, as well as reduce the risk of thrombi in other venous and arterial beds, especially the brain, since hyperhomocysteinemia is associated with both venous and arterial thrombosis.

However, in a placebo-controlled study, folic acid–vitamin B6–vitamin B12 therapy, which lowered homocysteine, did not reduce the risk for symptomatic venous thromboembolism. An optimal study of normalizing high homocysteine with folic acid–vitamin B6–vitamin B12 in patients with CRVO and CRAO-AF in an attempt to prevent recurrent RVO events or other thrombotic events would be blinded and placebo-controlled, and would run for 5 years.

In addition to thrombophilic risk factors for CRVO and CRAO-AF, atherosclerotic risk factors and cigarette smoking...
have been implicated as causative factors.\textsuperscript{2,6} In the current study, hypertension – but not hyperlipidemia – was common in CRVO and CRAO-AF cases, and type 2 diabetes was common in CRAO-AF cases.

Previous studies\textsuperscript{64} and case reports\textsuperscript{10-13,18,19} have emphasized that RVO can be triggered by estrogens, estrogen–progestins, clomiphene citrate,\textsuperscript{8} or SERMS, but these studies did not explore for underlying and/or previously undiagnosed inherited and/or acquired thrombophilia or hypofibrinolysis. A novel finding of the current study was that, of 96 women with RVO, six (6\%) first sustained RVO 3 years (on average) after beginning to take estrogens or estrogen agonists and were subsequently discovered to have inherited and/or acquired thrombophilia. Our report casts light on the pathophysiologic interaction\textsuperscript{65} leading to RVO between pharmacologic thrombophilia conferred by exogenous estrogens or estrogen agonists and inherited or acquired thrombophilia–hypofibrinolysis.\textsuperscript{46} When CRVO or CRAO-AF occurs in women receiving estrogens or estrogen agonists, particularly at 55 years old and older, we suggest evaluation for underlying inherited and acquired thrombophilia. Thrombophilia contributes to the risk of thrombosis in women using estrogen–progestin oral contraceptives or hormone replacement therapy.\textsuperscript{65} In postmenopausal women taking estrogen–progestin hormone replacement therapy, the presence of inherited thrombophilia (factor V Leiden, high factor VIII) increases the risk of deep venous thrombosis 17-fold compared to women without inherited thrombophilia who do not use hormone replacement therapy.\textsuperscript{46}

**Conclusion**

Treatable thrombophilia, particularly hyperhomocysteinemia, is more common in RVO cases than in normal controls. RVO occurring after estrogens or estrogen agonist use in women should stimulate an assessment for any underlying thrombophilia.

**Acknowledgments/disclosure**

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**References**


35. Scott JA, Arnold JJ, Currie JM, et al. No excess of factor V:Q506 genotype but high prevalence of anticardiolipin antibodies without antieno


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