Dear Editor

Concerning the recent article published in your journal on antibiofilm surface functionalization of catheters. This is an admirable approach to inhibit biofilm formation on the surfaces of various implants. Currently, a number of biomedical devices and implants are commonly used in hospitals and clinics. Over the past few decades, a number of knee and hip implants have been introduced to save lives and restore quality of life. Moreover, a significant increase in the use of stents, heart valves, vascular grafts, catheters, and other implantable devices are being introduced worldwide. However, regrettably, these surfaces are prone to microbial infections and hence device-related infections have become a major source of infection which may ultimately lead to a high mortality rate in the hospital setting.

Another major problem is the worldwide spread of multidrug resistance, especially of the newly discovered NDM-1 superbug. The latest metallo beta-lactamase, named NDM-1 (New Delhi metallo beta-lactamase) has been identified as a novel class of carbapenemases in enterobacteriaceae. Due to its spread, only a limited number of antibiotics can control infections with this agent. In this situation, there is an urgent need for research on biomaterials and design of surfaces that are resistant to infection and biofilms so that microbial infection and biofilm formation can be controlled. A number of promising approaches have been developed to control these infections on the surfaces of various devices. However, nanotechnology-based approaches are anticipated to provide new breakthroughs for prevention of biofilm-mediated infections via the broadened use of atomic-scaled nanomaterials.

Lellouche et al coated catheters with MgF2 nanoparticles using a sonochemical synthesis protocol. This preparation and coating procedure resulted in a uniform MgF2 nanoparticle layer on both sides of the catheter. These nanoparticle-coated catheters were investigated for their ability to control infections with this agent. In this situation, there is an urgent need for research on biomaterials and design of surfaces that are resistant to infection and biofilms so that microbial infection and biofilm formation can be controlled.

Recently, we explored gold nanoparticles (21 ± 2.5 nm and 0.2 mg/mL) and methylene blue (20 µg/mL) conjugation as a potential treatment for Candida biofilm. Type I phototoxicity against biofilm was demonstrated and confirmed by
fluorescence spectroscopy. Therefore, gold nanoparticle conjugate-mediated photodynamic therapy may also be used against common nosocomially acquired refractory *Candida albicans* biofilm.\(^5\)

In one of the relevant studies, Chifiriuc et al\(^6\) explored functionalized magnetite nanoparticles with oleic acid as a surfactant. Core-shell nanoparticles (Fe\(_3\)O\(_4\)/oleic acid:CHCl\(_3\)) were used to coat pieces of catheter by applying a magnetic field on nanofluid, while the CHCl\(_3\) diluted essential oil was pertained by adsorption in a secondary covering treatment. Fungal adherence and development of biofilm were strongly inhibited in the presence of these new core-shell/coated shell-based essential oil of *Rosmarinus officinalis*. Hence, it can be used to inhibit fungal adherence and could be of a great interest in the biomedical field, opening up new vistas for the design of film-coated surfaces with antibiofilm properties.\(^6\)

Silver nanoparticles are recognized as promising candidates due to their significant antimicrobial activity; however, there are two major problems with these nanoparticles, ie, they are highly toxic to healthy cells and cannot eradicate bacterial biofilms. Therefore, new technologies are being developed to overcome these limitations, consisting of multimodal nanoparticles having a magnetic core and a silver ring with a ligand gap, which have been used to eradicate bacterial biofilms successfully with no detrimental effect on healthy cells.\(^7\) This will require special consideration in future work, in which alternative nonmaterial compounds that can inhibit bacterial and fungal colonization of catheter surfaces should be developed.

**Disclosure**

The author reports no conflicts of interest in this letter.

**References**