Uptake and transport of a novel anticancer drug-delivery system: lactosyl-norcantharidin-associated N-trimethyl chitosan nanoparticles across intestinal Caco-2 cell monolayers

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Abstract: In this paper, novel liver-targeting nanoparticles (NPs), lactosyl-norcantharidin (Lac-NCTD)-associated N-trimethyl chitosan (TMC) NPs (Lac-NCTD-TMC-NPs), were prepared using ionic cross-linkage. The physical properties, particle size, and encapsulation efficiency of the nanoparticles were then investigated. The continuous line of heterogeneous human epithelial colorectal adenocarcinoma cells (Caco-2) cell monolayer model was used to study the transport mechanism of Lac-NCTD, and the effects of factors such as time, temperature, pH level, drug concentration, enhancers, and inhibitors. This model was also used to indicate the differences among Lac-NCTD, Lac-NCTD-associated chitosan NPs (Lac-NCTD-CS-NPs), and Lac-NCTD-TMC-NPs in the absorption and transportation of membranes. Drug concentration levels were measured using high-performance liquid chromatography. Active transport and paracellular transport were suggested to be both the primary and secondary mechanisms for Lac-NCTD absorption, respectively. Lac-NCTD uptake and absorption were not controlled by pH levels, but were positively correlated to uptake time, and negatively correlated to temperature. The basolateral to apical apparent permeability coefficients (Papp) were higher than those of the apical to basolateral values. The inhibitor of P-glycoprotein and the multidrug resistance-associated protein 2 significantly enhanced the uptake amount of Lac-NCTD. Compared with Lac-NCTD, Lac-NCTD-CS-NPs and Lac-NCTD-TMC-NPs significantly enhanced drug absorption. Additionally, the latter exhibited stronger action. Lac-NCTD-NPs could penetrate the plasma membrane of Caco-2 cells and translocate into the cytoplasm and even into the nucleus. Nanoparticles were uptaken into Caco-2 cells through the endocytosis pathway.

Keywords: Lac-NCTD, Lac-NCTD-TMC-NPs, Caco-2 cell, transport

Introduction

Lactosyl-norcantharidin (Lac-NCTD) is synthesized by modifying NCTD with a lactobiose-bearing galactose group to achieve active targeting. The galactose group can combine with the asialoglycoprotein receptor situated on the hepatocyte membrane.¹,² A previous study by the present researchers demonstrated that, compared with NCTD, Lac-NCTD has significantly higher therapy efficiency and lower toxicity.³

Chitosan (CS) is a biocompatible, biodegradable, nontoxic polymer. It has been extensively investigated as a formula carrier in delivery systems for therapeutic molecules.⁴,⁵ However, the application of CS has been hampered largely by its solubility only in acidic environments, with pH values below 6.0. On the other hand, N-trimethyl chitosan (TMC), a partially quaternized chitosan (CS) derivative, can be soluble in the entire pH range and
encompasses all the advantages of CS. It can also form nano-
complexes with anionic compounds such as drugs, proteins, and
DNA through ionotropic gelation. Hence, TMC has potential
applications in targeted drug-delivery applications.6

To improve the therapeutic efficacy of Lac-NCTD and
decrease its side effects, Lac-NCTD-loaded liver-targeting
chitosan nanoparticles (Lac-NCTD-CS-NPs) were successfully
developed in previous studies.3 Novel liver-targeting
nanoparticles Lac-NCTD-associated N-trimethyl chitosan
nanoparticles (Lac-NCTD-TMC-NPs) were then investigated
and discussed in the present study.

For most therapeutic agents, the oral route is still the most
convenient for drug administration. Based on this
consideration, the present study investigated the absorption mech-
nisms of Lac-NCTD, and two nanoparticles. The human
Caco-2 cell line has been widely used as a potent in-vitro
model to predict drug absorption in humans, to explore
the mechanisms of drug absorption, and to identify substrates
or inhibitors of transporters.7

The present study aims to use Caco-2 cells to develop a
theoretical basis for the understanding of the oral absorption
of Lac-NCTD and its nanoparticles, as well as to further char-
acterize both its uptake and transepithelial transport across
Caco-2 cells. The present study could provide a theoretical basis
for the formulation and development of a rational design of new
agents, as well as for the selection of its administration route.

Methods and materials

Materials

Lac-NCTD and TMC were synthesized in our laboratory.
CS (molecular weight [MW] of 8 kDa to 10 kDa,
deacetylation degree [DD] 93.1%) was purchased from
Xingcheng Biochemical Co, Ltd (Nantong, China). Sodium
tripolyphosphate (TPP) was obtained from Sinopharm Group
Co, Ltd (Shanghai, China).

Caco-2 cells were obtained from Fu Dan University
(Shanghai, China). Cell culture reagents were purchased from
Gibco (Grand Island, New York, NY). All other reagents
and solvents were of analytical grade and commercially
available.

High-performance liquid chromatography (HPLC) was
carried out using a chromatograph consisting of two pumps
(LC-10AT VP, Shimadzu, Japan) and a UV-vis detector
(SPD-M10A VP, Shimadzu, Japan) set at 220 nm. The
reversed-phase column used was a Hypersil ODS-C18 (5 μm
in 4.6 × 250 mm, Elite, Dalian, China) set to 25°C. The
mobile phase was a mixture of acetonitrile and H2O (at a ratio
of 10:90, adjusted to pH 3.2 by adding phosphoric acid) and
the flow rate was 0.8 mL/minute.

Preparation of Lac-NCTD-TMC-NPs

Lac-NCTD-TMC-NPs were prepared according to the
procedure first reported by Calvo and colleagues,8 which was
based on the ionic gel of TMC with TPP anions. Briefly,
0.15 g TMC was dissolved in 50 mL distilled water contain-
ing 0.03 g Lac-NCTD. Then, 25 mL of the TPP (1.2 mg/mL)
aqueous solution was drip-fed into the TMC solution under
magnetic stirring (500 rpm) at 30°C, resulting in crosslink-
age. Finally, the opalescent suspension was filtered through
a 0.45 µm filter to remove any insoluble aggregate residue,
and to obtain the Lac-NCTD-TMC-NPs suspension.

Evaluation of Lac-NCTD-TMC-NPs

The particle distribution and zeta potential of Lac-NCTD-
TMC-NPs’ suspension were measured using a Zetasizer
nanoparticle analyzer (model HPP-5001; Malvern Instru-
mements, Malvern, UK), which works on the principle of laser
diffraction analysis. The shape of the Lac-NCTD-TMC-NPs
particles was determined through transmission electron
microscopy (TEM) analysis, which was performed using an
H-600 transmission electron microscope (Hitachi, Tokyo,
Japan).

The drug entrapment efficiency (EE) and drug-loading
(DL) amount of Lac-NCTD-TMC-NPs were determined as
described previously.9 Briefly, the Lac-NCTD-TMC-NPs’
suspension was ultracentrifuged (250,000 × g for 30 minutes
at 10°C) in an Optima MAX Centrifuge (Beckman Instru-
mements, Palo Alto, CA). The supernate was sampled, and the
Lac-NCTD concentration in the supernate was tested using
a reversed-phase HPLC method.

Cell culture

Caco-2 from a passage between 30 and 50 were used in the
experiments. The cells were grown in conditions similar
to those reported earlier. The cells were maintained in
Dulbecco’s modified Eagle’s medium (DMEM) with high
glucose (4.5 g/L), which was supplemented with 15% fetal
bovine serum, 1% nonessential amino acids, 1% L-glutamine,
and 0.5% penicillin/streptomycin. All cell cultures were main-
tained in a humidified incubator at 37°C in an atmosphere
of 5% CO2 and 95% relative humidity. The culture medium
was changed every two days, and the cells were passaged at
a ratio of 1:3, every 3 to 5 days (at 70% to 80% confluence),
using 0.25% trypsin and 0.02% EDTA at 37°C.
Uptake studies
Caco-2 cells were plated on 12-well tissue culture plates (Corning, Corning, NY) at a density of $1 \times 10^5$ cells/cm². The cell monolayers were used on the 15th day for uptake experiments.

Hank’s balanced salt solution (HBSS; pH 7.4) was used as the incubation medium. The Caco-2 cell monolayers were washed twice with the incubation medium, and then preincubated with 1 mL of the incubation medium for 15 minutes at 37°C. The medium was then removed after the preincubation period and uptake was initiated by adding 1 mL of the preincubated drug solution (Lac-NCTD, Lac-NCTD-CS-NPs, or Lac-NCTD-TMC-NPs). At various time points, the cells were quickly washed three times with cold HBSS, and then scraped off using a cell scraper (Corning). Next, the cells were homogenated, vortexed, and ultracentrifuged. The supernate was sampled, and the accumulated amount of Lac-NCTD was assayed by HPLC. The protein content was measured according to Bradford’s method.10

Transport studies
Caco-2 cells were seeded at a density of $1 \times 10^5$ cells/cm² onto a permeable polycarbonate insert (1.1 cm², 1 µm pore size) (Millipore, Billerica, MA) in 12-well tissue culture plates (Corning). On the 21st day, the cell monolayers were used for transepithelial transport experiments. The integrity of the monolayers grown on the permeable membrane was assessed using the transepithelial electrical resistance (TEER) of the monolayers, through a Millicell-ERS apparatus (Millipore) and phenol red transport. Caco-2 monolayers could only be used in the experiments when TEER values reached 700 Ω·cm² and phenol red permeability was <0.5% per hour.

The culture medium of both sides was removed by aspiration, and the Caco-2 cell monolayers were washed twice with the incubation medium. After washing, the monolayers were preincubated for 15 minutes at 37°C with 0.5 and 1.5 mL of the incubation medium in the apical and basolateral sides, respectively. After the preincubation, the medium was removed immediately, and the incubation medium containing Lac-NCTD and its nanoparticles (Lac-NCTD-CS-NPs and Lac-NCTD-TMC-NPs) were added to either the apical side (0.5 mL) or the basolateral side (1.5 mL). The unmodified incubation medium was then added to the opposite side (basolateral, 1.5 mL; apical, 0.5 mL). Next, 50 µL samples were taken from the receiver compartment at intervals of 5, 15, 30, 45, and 60 minutes, and replaced with a fresh buffer. Concentrations in the transepithelial transport samples were determined using the HPLC method as described above.

Subcellular localization of NPs
To detect cellular uptake using a laser scanning confocal microscope, Lac-NCTD-CS-NPs and Lac-NCTD-TMC-NPs were labeled with fluorescein isothiocyanate (FITC). The FITC-labeled CS and TMC were prepared by the reaction between the isothiocyanate group of FITC and the primary amino group of CS and TMC, as described by Huang and colleagues.11

The Caco-2 cells were seeded on 6-well dishes at a density of $1 \times 10^5$ cells/cm². FITC-Lac-NCTD-TMC-NPs and FITC-Lac-NCTD-TMC-NPs were added to the cells and incubated for 15, 30, and 60 minutes. The cells were washed three times with PBS, fixed with 4% paraformaldehyde, and then stained with 10 µg/mL Hoechst 33258 (Sigma-Aldrich, St Louis, MO). Micrographs were taken on a Nikon laser scanning confocal microscope (Nikon Eclipse TE2000-E; Nikon, Tokyo, Japan).

Uptake process of NPs
The Caco-2 cells were seeded on 6-well dishes at a density of $1 \times 10^5$ cells/cm². Lac-NCTD-TMC-NPs were added to the cells and incubated for 4 hours. The cells were then washed three times with PBS, twice with distilled water, and then fixed with 4% paraformaldehyde. Micrographs were taken on an atomic force microscope (AFM; MultiMode V; Veeco Instruments, Woodbury, NY).

Data and statistical analysis
The Lac-NCTD EE and DL amounts were calculated as follows:

$$EE = \frac{T - F}{T} \times 100\% \quad (1)$$

$$DL = \frac{T}{W} \times 100\% \quad (2)$$

where $T$ is the total Lac-NCTD in the colloid, $F$ is the free Lac-NCTD in the supernate, and $W$ is the weight of Lac-NCTD-TMC-NPs.

The $P_{app}$ was calculated according to the equation

$$P_{app} = \frac{dQ/dt}{AC_0} \quad (3)$$

where $dQ/dt$ is the slope of the cumulative amount transported during the time course of the period studied, $A$ is the area of the inserts, and $C_0$ is the starting concentration.
Permeability direction ratios (PDR) were calculated according to the following equation:

$$PDR = \frac{P_{app}(BL-AP)}{P_{app}(AP-BL)}$$

where $AP-BL$ is the apical to basolateral transport and $BL-AP$ is the basolateral to apical transport.

All data were reported as mean ± standard deviation for at least three independent samples. Data were analyzed by one-way ANOVA with the post hoc Tukey’s test applied for paired comparisons using SPSS software (version 16.0; SPSS, Chicago, IL). The criteria for statistical significance were taken as $*P < 0.05$, $**P < 0.01$, $\sigma P < 0.05$, and $\#P < 0.01$ (Figures 5–7; Tables 1,3).

## Results

### Preparation and evaluation of Lac-NCTD-TMC-NPs

Lac-NCTD-TMC-NPs were prepared through an ionic cross-linking process and evaluated in terms of the NPs EE, DL, particle size, and polydispersity index (PDI). The particle size of Lac-NCTD-TMC-NPs was around 120 nm (PDI < 0.3) (Figure 1). The zeta potential of Lac-NCTD-CS-NPs was 28.08 ± 3.95 mV, whereas that of Lac-NCTD-TMC-NPs was 37.37 ± 3.62 mV, mainly because of the high degree of TMC quaternization (65%). The EE and DL of Lac-NCTD-TMC-NPs were 69.29% ± 0.76% and 9.1% ± 0.07%, respectively. A TEM image of the Lac-NCTD-TMC-NPs is shown in Figure 2. Under TEM, most of the NPs appeared to be uniform and round, with the majority of particles separated from one another.

### Linearity, precision, and accuracy of Lac-NCTD determination

Under the conditions described above, the Lac-NCTD retention was at 5.1 minutes. The active component test was uncontaminated, and also had high specificity. In a 0.1 µg/mL to 52 µg/mL concentration range, Lac-NCTD in PBS/blank cells homogenate showed a good linear relationship. Recovery was 98.99% to 102.46% within the linear range, and relative standard deviation was below 5%, both of which fitted the requirements for analyzing the biological specimen.

### Uptake of Lac-NCTD into Caco-2 monolayers

Effect of time on the uptake of Lac-NCTD

The uptake of Lac-NCTD (250 µg/mL) into Caco-2 monolayers was shown to be linear in relation to time, from 0 minutes to 30 minutes at 37°C (Figure 3). To ensure the optimal absorption rate, the uptake time of the drug should be determined...
within its linear absorption range. Considering the nature of the drug, cell characteristics, and operational feasibility, 15 minutes was set as the uptake time for the subsequent experiments.

**Effect of concentration on the uptake of Lac-NCTD**
Concentration dependence or saturation was observed for the uptake of Lac-NCTD in the concentration range of 100 µg/mL to 400 µg/mL (Figure 4).

**Effect of pH and temperature**
The uptake amount of Lac-NCTD (250 µg/mL) into Caco-2 monolayers in pH 6.0 and 7.4 were (0.82 ± 0.23) and (0.80 ± 0.15) mg/g protein, respectively. There were no significant differences between the uptake amounts in the two pH values (P > 0.05). Hence, the subsequent experiments were performed using the medium at pH 7.4.

The uptake amount of Lac-NCTD (250 µg/mL) into Caco-2 monolayers in 4°C and 37°C were (1.35 ± 0.13) and (0.80 ± 0.15) mg/g protein, respectively. The uptake of Lac-NCTD across Caco-2 cell monolayers showed temperature dependence, and was enhanced at low temperatures.

**Effect of protein inhibitor**
The saturation possibility of the P-glycoprotein (P-gp) and the multidrug resistance-associated protein 2 (MRP2) function was investigated in the present study. Hence, the uptake of Lac-NCTD across Caco-2 in solutions with added P-gp or MRP2 inhibitors 20 µmol/L cyclosporine (CyA) or 50 µmol/L MK-571 was investigated. Results are shown in Figure 5, which indicate that CyA and MK-571 can facilitate the uptake procedure. However, the latter had an even stronger action.

**Uptake of Lac-NCTD-NPs into Caco-2 monolayers**
The uptake of Lac-NCTD-NPs (Lac-NCTD-CS-NPs, Lac-NCTD-TMC-NPs) and Lac-NCTD solution at a dose of 250 µg/mL at 37°C was investigated. The results are shown in Table 1. Among the three formulations, the order of uptake amount was Lac-NCTD-TMC-NPs > Lac-NCTD-CS-NPs > Lac-NCTD solution.

**Transport of Lac-NCTD across Caco-2 monolayers**
**Effect of concentration on the transport of Lac-NCTD**
The influence of concentration on the transport of Lac-NCTD across the Caco-2 cell monolayers was measured (Table 2). The bidirectional transport data for Lac-NCTD showed that the secretory Papp (BL-AP) of Lac-NCTD was higher than its absorptive Papp (AP-BL) at every concentration point, indicating the existence of efflux proteins.12–13

**Table 1** Uptake of Lac-NCTD on Caco-2 cells with different preparations (n = 3)

<table>
<thead>
<tr>
<th>Preparations</th>
<th>Lac-NCTD</th>
<th>Lac-NCTD-CS-NPs</th>
<th>Lac-NCTD-TMC-NPs</th>
</tr>
</thead>
<tbody>
<tr>
<td>Uptake (mg/g protein)</td>
<td>0.80 ± 0.15</td>
<td>1.54 ± 0.32**</td>
<td>1.73 ± 0.17**</td>
</tr>
</tbody>
</table>

Notes: **P < 0.01 versus Lac-NCTD; *P < 0.05 versus Lac-NCTD-CS-NPs.**

**Abbreviations:** Lac-NCTD, lactosyl-norcantharitin; Caco-2, continuous line of heterogeneous human epithelial colorectal adenocarcinoma cells.
Effects of protein inhibitor on bi-directional transport of Lac-NCTD

The efflux transport of Lac-NCTD (250 µg/mL) was investigated in the presence of CyA (20 µmol/L) and MK-571 (50 µmol/L). The results are shown in Figure 6. When CyA was added, the \( P_{\text{app}} \) of the apical to basolateral direction increased, and the \( P_{\text{app}} \) of the basolateral to apical direction decreased. When MK-571 was added, the \( P_{\text{app}} \) of the apical to basolateral direction significantly increased, and the \( P_{\text{app}} \) of the basolateral to apical direction lightly decreased.

Effect of endocytosis inhibitors and bypass transport enhancers

The transport of Lac-NCTD (250 µg/mL) across Caco-2 in solutions with added endocytosis inhibitor oxophenylarsine 25 mmol/L or bypass transport enhancer sodium deoxycholate (SDCh) 100 mmol/L was also investigated. As shown in Figure 7, the \( P_{\text{app}} \) significantly increased after SDCh \((P < 0.05)\) was added. However, no statistical difference in \( P_{\text{app}} \) existed between the oxophenylarsine group and the control group.

**Table 2** \( P_{\text{app}} \) of Lac-NCTD at different concentrations in Caco-2 cells \((n = 3)\)

<table>
<thead>
<tr>
<th>Lac-NCTD (µg/mL)</th>
<th>( P_{\text{app}} \times 10^{4})</th>
<th>Ratio</th>
<th>AP-BL BL-AP ( P_{\text{app}} )</th>
<th>( P_{\text{app}} )</th>
<th>( P_{\text{app}} )</th>
</tr>
</thead>
<tbody>
<tr>
<td>100</td>
<td>3.29 ± 0.34</td>
<td>6.29 ± 0.43</td>
<td>1.91</td>
<td></td>
<td></td>
</tr>
<tr>
<td>250</td>
<td>3.67 ± 0.17</td>
<td>6.76 ± 0.28</td>
<td>1.845</td>
<td></td>
<td></td>
</tr>
<tr>
<td>400</td>
<td>3.34 ± 0.22</td>
<td>4.58 ± 0.15</td>
<td>1.37</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Abbreviations:** Lac-NCTD, lactosyl-norcantharitin-associated; Caco-2, continuous line of heterogeneous human epithelial colorectal adenocarcinoma cells.

**Table 3** \( P_{\text{app}} \) of Lac-NCTD on Caco-2 cells with different preparations \((n = 3)\)

<table>
<thead>
<tr>
<th>Preparations</th>
<th>( P_{\text{app}} \times 10^{4})</th>
<th>Ratio</th>
<th>AP-BL BL-AP ( P_{\text{app}} )</th>
<th>( P_{\text{app}} )</th>
<th>( P_{\text{app}} )</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lac-NCTD solution</td>
<td>3.67 ± 0.68</td>
<td>6.76 ± 1.12</td>
<td>1.85</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lac-NCTD-CS-NPs</td>
<td>5.38 ± 0.23**</td>
<td>6.01 ± 0.45*</td>
<td>1.11**</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lac-NCTD-TMC-NPs</td>
<td>5.73 ± 0.44***</td>
<td>6.12 ± 0.67**</td>
<td>1.07**</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Notes:** 
* \( P < 0.05 \) versus Lac-NCTD; ** \( P < 0.01 \) versus Lac-NCTD; *** \( P < 0.001 \) versus Lac-NCTD-CS-NPs. 

**Abbreviations:** Lac-NCTD, lactosyl-norcantharitin; Caco-2, continuous line of heterogeneous human epithelial colorectal adenocarcinoma cells.

Abbreviations: FITC-labeled, fluorescein isothiocyanate-labeled; Lac-NCTD-CS-NPs, lactosyl-norcantharitin (Lac-NCTD)-associated N-Trimethyl chitosan nanoparticles; Lac-NCTD-TMC-NPs, lactosyl-norcantharitin (Lac-NCTD)-associated N-Trimethyl chitosan nanoparticles.

Uptake process of NPs
The uptake of Lac-NCTD-TMC-NPs by human Caco-2 cells was observed using atomic force microscopy (Figure 9).

Figure 9A shows the surface of Caco-2 cells without any treatment. Figure 9B shows the cells’ surface morphology when treated with nanoparticles for 4 hours. The surface is relatively flat in Figure 9A, whereas multiple depressions are present in Figure 9B. These changes indicated the occurrence of endocytosis.

A statistical analysis of the height of the protruding particles on the cell surface (Figure 10) then followed. Figure 10A shows the Caco-2 cell surface without any treatment. The height of the protruding particles was mostly located from 100 nm to 150 nm. Figure 10B shows the cell surface morphology when treated with nanoparticles for 4 hours. The height of protruding particles mostly gathered at 280 nm. The height difference between 10A and B was consistent with the particle size of the nanoparticles.

Discussion
In the present study, a novel liver-targeting drug-delivery system (Lac-NCTD-TMC-NPs) with stable EE and high DL efficacy was prepared. The particle size of Lac-NCTD-TMC-NPs was 120.6 ± 1.7 nm. The majority of fenestrated liver sinusoids were, usually, smaller than 200 nm in diameter. Additionally, several abnormalities existed between tumor blood vessels and physiologically normal vessels, including a relatively high proportion of proliferation in vivo, increased tortuosity, aberrant basement membrane formation, and enhanced permeability. Thus, the prepared Lac-NCTD-TMC-NPs, with a mean particle size of around 120 nm, can pass through the fenestrated sinusoids and accumulate in the tumor sites of the liver. The small particle size reduced the adverse reaction of Lac-NCTD-TMC-NPs on normal hepatic cells, by reducing the passive targeting in the liver. The TEM image suggested that these nanoparticles were probably ionically stabilized against particle agglomeration, due to the positive charges on their surface.

The intestinal transport mechanism of Lac-NCTD and its NPs was also investigated in the present study using Caco-2 cells. In recent years, the Caco-2 cell model has become a popular model of the intestinal mucosa due to the fact that these cells have a colonic origin, and express transporters which are normally found in the intestinal mucosa. However, others (P-gp, MRP2, and MRP1) are known to restrict drug absorption. In the uptake study, concentration dependence or saturation was observed for the uptake of Lac-NCTD in the concentration range of 100 µg/mL to 400 µg/mL. This finding suggests that active transport occurred during the uptake process. The effects of pH level and temperature on the uptake of Lac-NCTD in Caco-2 monolayers were also studied. The uptake amount of Lac-NCTD in Caco-2 cells was observed to be dependent on temperature, but not on pH values. Generally, in low temperatures, the generation of adenosine triphosphate (ATP) is blocked, and so active...
transport is inhibited; therefore the uptake amount of Lac-NCTD in Caco-2 cells should be reduced. However, the experimental result was contrary to this finding. According to the literature,$^{21,22}$ Caco-2 cells highly express drug efflux transporters, which are energy-dependent drug efflux pumps. The uptake amount was enhanced at a low temperature, suggesting the presence of efflux proteins.$^{22–24}$

Previous studies have demonstrated that a class of proteins is related to drug uptake and intracellular transport.$^{25,26}$ In addition to P-gp, MRP2 also plays a targeted transport pump role, affecting the intestinal absorption of compounds. These two types of protein are both expressed in Caco-2 cells; thus, specific inhibitors were applied in the present study to observe the interaction between Lac-NCTD and inhibitors. Results showed that P-gp and MRP2 each play an important role in the uptake and transport process of Lac-NCTD. When either CyA (20 $\mu$mol/L) or MK-571 (50 $\mu$mol/L) was added to the apical side of Caco-2 cell monolayers, the $P_{\text{app}}$ of the apical to basolateral direction significantly increased by 1.41 and 2.29 times, respectively.

Both P-gp and MRP2 efflux pumps are located predominantly in the apical membranes of the epithelia (eg, small intestine, colon), making the secretion rate faster than the absorption rate.$^{27}$ Efflux proteins show a strong efflux effect in low drug concentrations, and become saturated in high drug concentrations. When the Lac-NCTD concentration increased from 100 $\mu$g/mL to 400 $\mu$g/mL, the ratios of $P_{\text{app}}$ BL-AP to $P_{\text{app}}$ AP-BL decreased from 1.91 to 1.37, respectively. Between the two types of efflux proteins, MRP2 showed a stronger action. This conclusion needs to be verified further, using a MDCK-MRP2 cell model.

Figure 7 shows the effects of oxophenylarsine and SDCh, which were added to the apical side of Caco-2 cell monolayers on the transport of Lac-NCTD. The paracellular transport enhancer SDCh promoted the absorption of Lac-NCTD, while the endocytosis inhibitor oxophenylarsine had no effect. This indicates that only a small proportion of Lac-NCTD was transported by the paracellular mode, which speculation is consistent with the physical and chemical properties of Lac-NCTD. Water-soluble small molecule compounds and paracellular transport are common modes of transport for these types of drugs.

The uptake and transport studies of NPs have shown that in the same concentrations, absorption of Lac-NCTD-CS-NPs and Lac-NCTD-TMC-NPs was greater than that of the Lac-NCTD solution. Additionally, Lac-NCTD-TMC-NPs have a stronger action; thus, NPs can improve the cell uptake and transportation of drugs. Adsorptive endocytosis has been suggested to exist as a nonspecific interaction of the cell
membrane with the NPs. Meschini and colleagues reported that higher intracellular content of a drug via endocytosis of a carrier can remarkably increase its therapeutic effect against the target cells.\(^{16}\) On the other hand, CS and TMC can improve uptake. The mechanism is that CS and TMC can open tight junctions.\(^{17}\) TMC’s high degree of quaternization (65%) enables high absorption-enhancing properties. Moreover, TMC-containing formulations reportedly exceed the potency of CS in mucoadhesion.\(^{28}\) In particular, as shown in Table 3, the \(P_{\text{app}}\) \(\text{BL-AP}\) of NPs was greater than the \(P_{\text{app}}\) \(\text{AP-BL}\), and the ratios of \(P_{\text{app}}\) \(\text{BL-AP}\) to \(P_{\text{app}}\) \(\text{AP-BL}\) were largely reduced compared with the Lac-NCTD solution.

These results suggested that after being encapsulated in CS or TMC, a small portion of Lac-NCTD was directly transported into the cells in nanoparticle form, effectively bypassing the need for efflux proteins. Therefore, for multidrug-resistant cells with a decreased drug uptake, CS-NPs or TMC-NPs can act as a transmembrane delivery vehicle, to help transport the drug molecules into the cells. Thus, CS-NPs or TMC-NPs can decrease drug efflux as well as increase cell internalization and intracellular accumulation of the drugs.\(^{29}\)

Efficient cellular internalization of nanoparticles is necessary, for both intracellular drug delivery, and efficient therapy. The results in Figure 8 proved that Lac-NCTD-NPs can penetrate the plasma membrane of Caco-2 cells, and, from there, translocate into the cytoplasm, and even into the nucleus. CS was regarded as a versatile polymer.\(^{30}\) Nanoparticles made of polymer materials may pass through the nuclear pore, and then enter the nucleus during the cell division process.\(^{31}\) Additionally, the positive charge of CS-NPs and TMC-NPs enables an electrostatic interaction with the phosphate groups of nucleic acids.\(^{32}\) Isolated spots of green fluorescence showed higher fluorescence intensity in some cells, demonstrating the cluster of the nanoparticles. The figures obtained from atomic force microscopy (Figure 9) indicate that nanoparticle uptake into cells occurred through an endocytosis pathway. The combination of the nanoparticles and the cells is relatively secure.

**Conclusion**

Active transport and paracellular transport are suggested as both main and secondary mechanisms for the absorption of Lac-NCTD, respectively. Lac-NCTD uptake and absorption are not controlled by pH levels; however, they are positively correlated to uptake time, and negatively correlated to temperature. The \(P_{\text{app}}\) of the basolateral to apical direction is higher than that of the apical to basolateral direction. The P-gp and MRP2 inhibitor significantly enhances the uptake amount of Lac-NCTD. Compared with Lac-NCTD, Lac-NCTD-CS-NPs and Lac-NCTD-TMC-NPs also significantly enhance drug absorption. Lac-NCTD-NPs can penetrate the plasma membrane of Caco-2 cells and translocate into the cytoplasm, and even into the nucleus.

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