Biodistribution and pharmacokinetics of a telodendrimer micellar paclitaxel nanoformulation in a mouse xenograft model of ovarian cancer

Wenwu Xiao 1
Juntao Luo 2
Teesta Jain 3
John W Riggs 3
Harry P Tseng 1
Paul T Henderson 3
Simon R Cherry 4
Douglas Rowland 4
Kit S Lam 1,3

1Department of Biochemistry and Molecular Medicine, UC Davis Cancer Center, University of California Davis, Sacramento, CA; 2Department of Pharmacology, SUNY Upstate Cancer Research Institute, SUNY Upstate Medical University, Syracuse, NY; 3Department of Internal Medicine, Division of Hematology and Oncology, 4Department of Biomedical Engineering, UC Davis Cancer Center, University of California Davis, Davis, CA

Correspondence: Kit Lam
Department of Biochemistry and Molecular Medicine, UC Davis Cancer Center, University of California Davis, Sacramento, CA 95817, USA
Tel +1 916 734 0910
Fax +1 916 734 6415
Email kit.lam@ucdmc.ucdavis.edu

Correspondence: Juntao Luo
Department of Pharmacology, SUNY Upstate Cancer Research Institute, SUNY Upstate Medical University, 750 East Adams Street, Syracuse, NY 13210, USA
Tel +1 315 464 7965
Fax +1 315 464 5143
Email luoj@upstate.edu

Background: A multifunctional telodendrimer-based micelle system was characterized for delivery of imaging and chemotherapy agents to mouse tumor xenografts. Previous optical imaging studies demonstrated qualitatively that these classes of nanoparticles, called nanomicelles, preferentially accumulate at tumor sites in mice. The research reported herein describes the detailed quantitative imaging and biodistribution profiling of nanomicelles loaded with a cargo of paclitaxel.

Methods: The telodendrimer was covalently labeled with 125I and the nanomicelles were loaded with 14C-paclitaxel, which allowed measurement of pharmacokinetics and biodistribution in the mice using microSPECT/CT imaging and liquid scintillation counting, respectively.

Results: The radio imaging data showed preferential accumulation of nanomicelles at the tumor site along with a slower clearance rate than paclitaxel formulated in Cremophor EL (Taxol®). Liquid scintillation counting confirmed that 14C-labeled paclitaxel sequestered in nanomicelles had increased uptake by tumor tissue and slower pharmacokinetics than Taxol.

Conclusion: Overall, the results indicate that nanomicelle-formulated paclitaxel is a potentially superior formulation compared with Taxol in terms of water solubility, pharmacokinetics, and tumor accumulation, and may be clinically useful for both tumor imaging and improved chemotherapy applications.

Keywords: telodendrimer, nanomicelle, paclitaxel, microSPECT/CT, imaging guided drug delivery

Introduction
Cancer is primarily characterized by the uncontrolled proliferation of cells and their ability to metastasize. In spite of significant advances in the fight against cancer, it remains a challenging medical problem. Systemic chemotherapy is a common approach to the treatment of cancer. However, toxicity is a major side effect, which limits the utility and effectiveness of such nontargeted chemotherapeutics. Recent research in the development of drug delivery systems has focused on targeted delivery and controlled drug release at tumor sites in order to increase efficacy while reducing systemic toxicity. Imaging-guided drug delivery is considered a promising strategy to achieve this goal. Full implementation of imaging-guided drug delivery will require that drugs can be imaged or identified in the body as they enter the bloodstream. The goal of imaging-guided drug delivery is to optimize local delivery of the therapeutic pharmaceutical agent to the target tissue and provide microanatomical and functional imaging feedback of the therapeutic process, as well as longitudinal treatment and monitoring.
The explosive growth of biocompatible nanotechnologies has made the clinical utilization of molecular imaging and therapy with a host of novel agents a realistic near-term possibility. Various nanocarriers, such as superparamagnetic iron oxide nanoparticles, gold nanoparticles, liposomes, and polymeric nanoparticles loaded with large payloads of imaging agents, enable detection of cancer with standard imaging equipment via passive or active tumor-targeting effects. In order to fulfill the goal of imaging-guided drug delivery, nanocarriers need to be multifunctional, thus making them useful for both imaging and drug delivery. Our group has developed a well-defined and biocompatible amphiphilic telodendrimer system composed of polyethylene glycol (PEG), cholic acid, and a dendritic lysine core that can self-assemble into multifunctional water-soluble nanomicelles to enable efficient delivery of hydrophobic cargos such as paclitaxel and the hydrophobic dye, 3,3,3′,3′-dioctadecyl-3,3,3′,3′-tetramethylindodicarbocyanine perchlorate. Telodendrimers are easily labeled with fluorophores and radionuclides by covalent conjugation onto a lysine side chain at the junction site between the PEG and the cholic acid cluster, without compromising their propensity to form nanomicelles. For noncovalent labeling, telodendrimers can be mixed with radio-labeled drugs or fluorescent probes during nanomicelle formation. The tailored multifunctional features of the telodendrimer-based nanomicelle system are ideal for imaging-guided drug delivery applications.

Previous reports have demonstrated efficient tumor targeting and drug delivery by nanomicelles through optical imaging and efficacy studies using ovarian cancer xenograft models. Our nanomicelles were prepared via self-assembly of telodendrimers containing a linear 5 kDa PEG covalently attached to a dendritic lysine core with eight cholic acid groups (PEG8CA8). In order to characterize and quantitatively analyze the biodistribution and tumor targeting effect of nanomicelle nanocarriers further, the telodendrimer was radiolabeled with 125I using the Bolton-Hunter reagent, which was provided by Perkins Elmer through custom radiolabeling. Excess Bolton-Hunter reagent was removed by passage through an LH-20 size exclusion column using ethanol as the eluent, yielding 10 mCi of 125I-labeled telodendrimers with a radioactive purity of 99.1%.

Preparation of the paclitaxel-loaded 125I-labeled nanomicelles (PTX-125I-NM) was as follows: 3.2 mg of paclitaxel and 32 mg of PEG8CA8 telodendrimer were dissolved in 2 mL of chloroform. Immediately after mixing, 0.8 mCi of 125I-labeled PEG8CA8 solution in ethanol (0.32 mL) was added. The solution was dried via nitrogen blowing overnight in a radio safety fume hood. Phosphate-buffered saline 1.6 mL was added and the mixture was vortexed for 30 seconds and sonicated for 30 minutes to disperse the complex into micelles. Comparison of both the imaging and quantitative biodistribution profiles of paclitaxel loaded into nanomicelles will aid in understanding and optimizing the telodendrimer system for better targeting of systemic cancer therapies.

Materials and methods

Paclitaxel

14C-PTX was purchased from Moravek Biochemicals Inc, (Brea, CA). Unlabeled paclitaxel was purchased from AK Scientific (Union City, CA). A Cremophor EL formulation of paclitaxel (Taxol®) from AK Scientific (Mountain View, CA) was obtained from the UC Davis Cancer Center pharmacy.

Cell lines

Ovarian cancer (SKOV-3) cell line was obtained from the American Type Culture Collection (Manassas, VA). The cells were maintained in Macoy-5A media and split every 3–4 days.

Mouse ovarian cancer xenograft model

The animal studies were performed according to a protocol approved by the Institutional Animal Care and Use Committee of the University of California, Davis. Female athymic nude mice (nu/nu), obtained from Harlan Inc, (Indianapolis, IN) at 5–6 weeks of age, were injected subcutaneously in the right flank with 5 × 106 SKOV-3 cells suspended in 200 µL of phosphate-buffered saline. When the subcutaneous tumors reached 0.5–1.0 cm in diameter or 14–21 days after implantation, the tumor-bearing mice were subjected to microSPECT/CT imaging of the 125I-labeled nanomicelles or biodistribution studies of the 14C-PTX loaded nanomicelles.

Iodination of telodendrimers and micelles

Iodination of telodendrimers was accomplished via the 125I-Bolton-Hunter reagent, which was provided by Perkins Elmer through custom radiolabeling. Excess Bolton-Hunter reagent was removed by passage through an LH-20 size exclusive column using ethanol as the eluent, yielding 10 mCi of 125I-labeled telodendrimers with a radioactive purity of 99.1%.

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MicroSPECT/CT imaging

MicroSPECT/CT imaging of PTX-125I-NM in a human ovarian cancer xenograft model was performed at the Center for Molecular and Genomic Imaging, University of California, Davis. Nude mice bearing SKOV-3 xenografts were injected with 200 µL of 100 µCi paclitaxel-loaded PTX-125I-NM (400 µg of paclitaxel in 4 mg of PEG5kCA8) via the tail vein. Pinhole SPECT images were acquired at hours 1, 5, 18, 24, 48, 72, and 94 following injection. SPECT images were acquired using the InveonMM (Siemens Preclinical Solutions, Knoxville, TN). The collimator set used was the mouse medium energy collimator, which is a single pinhole with a 3 mm diameter. The scanner has two SPECT heads that rotate around the animal. The radius of rotation was set at 40 mm. This collimator and radius combination yields a spatial resolution of approximately 3.5 mm. A total of 20 projections were acquired over 180 degrees per SPECT head and the initial projection exposure time was 150 seconds. The exposure time at each projection is adjusted for nuclide decay to yield an equivalent number of counts per projection. List mode data are collected, including all photon events, and photon energy is recorded. The data were reconstructed with an energy cut of 28–42 keV. The images were reconstructed with OSEM using four subsets and 20 iterations. Low resolution CT images were acquired for anatomical registration with the SPECT images. The images are displayed as a percent of the maximum voxel in the image. All images were scaled to 100 as the maximum intensity value.

14C-PTX-NM preparation

The micelles were formulated in a manner similar to the protocol used above, except without 125I-labeling. Briefly, 4 µCi of 14C-PTX in ethyl acetate was added to 5 mL of chloroform solution containing 15 mg of unlabeled paclitaxel and 100 mg of PEG5kCA8 telodendrimer. The solution was dried by nitrogen blowing overnight, followed by addition of 5 mL of phosphate-buffered saline and vortex mixing for 30 seconds and sonication for 30 minutes. The ratio of radiocarbon-labeled and unlabeled paclitaxel in the resulting micelles was approximately 1:1100 by mass.

14C-PTX biodistribution

Nude mice bearing SKOV-3 xenografts were injected with 200 µL solutions containing of 100 µCi 14C-PTX-NM (400 µg total mass of paclitaxel that was mixed with 4 mg of PEG5kCA8 prior to nanomicelle synthesis) via the tail vein. The major organs and tissues collected were whole blood, brain, skin, heart, lung, liver, spleen, kidney, bladder, muscle, and tumor. The time points of data collection were at hours 0, 0.5, 1, 6, 16, 24, 48, and 72. At selected time points, blood was collected by heart puncture under temporary anesthesia with ether. After blood collection, the mouse was euthanized by cervical dislocation. All organs were collected and weighed, put into 1.5 mL Eppendorf centrifuge tubes, and kept on ice prior to homogenization. Organs and tissue were minced into chunks of approximately 1 mm in diameter. The minced tissue was then added to 1 mL of Biosol, agitated gently, and placed in an incubator shaker at 50°C and shaken for 6 hours at 650 rpm. When chunks of tissue were no longer visible in the suspension, all the solution in the tubes were transferred into 20 mL scintillation vials, and decolorized by addition of 0.2 mL of 30% hydrogen peroxide. The vials were capped loosely and incubated at 50°C for one hour. Finally, 18 mL of liquid scintillation counting cocktail was added into each scintillation vial, followed by running the samples in a Packard 1500 Tri-Carb liquid scintillation analyzer with standard protocols for radiocarbon detection. Data were processed on an Excel spreadsheet running Solutions pharmacokinetic software.

Data processing and statistics

For determination of the radiointensity of tumors and normal organs in the radioimaging, we calculated the average values of the tumor area and of the normal organ area using the three-dimensional regions of interest function of the Siemens microPET ASIPro WM 6.7.1.2 software program. All data are shown as the mean ± standard deviation of three independent measurements. The Student’s t-test was used for statistical analysis of the radioimaging intensity and pharmacokinetic data. Statistical significance was indicated by P < 0.05 and P < 0.001.

Results and discussion

MicroSPECT/CT imaging of nanomicelles

Our previous research has demonstrated excellent tumor targeting of telodendrimer-based nanomicelles using near-
infrared fluorescence imaging, whereby nanomicelles formed by PEG5kCA8 are loaded with the near-infrared dye, 1,1’-dioctadecyl-3,3,3′,3′-tetramethyl indodicarbocyanine perchlorate, and paclitaxel.7,8 Near-infrared fluorescence imaging offers unique advantages for diagnostic imaging of solid tumors with high sensitivity,9 given the low in vivo autofluorescence in the near-infrared range (700–900 nm) and the minimal near-infrared absorbance by biological components, such as hemoglobin, water, and lipids.10,11 However, near-infrared fluorescence imaging is not quantitative and has a limited penetration distance, as well as low spatial resolution.12 In contrast, radioimaging is superior for quantification due to good tissue penetration of gamma rays and the ability to measure count rates in tissue accurately, permitting whole body quantitative imaging not only in small animals, but also in humans.

For this report, radioimaging was performed using an established tumor xenograft model. The PEG5kCA8 telodendrimer was labeled with 125I via the Bolton-Hunter reagent as shown in Scheme 1A. The unbound labeling reagent was removed by passage through a size-exclusive column, with ethanol as the eluent to avoid formation of telodendrimer aggregates in which the iodination reagent could become trapped. The 125I-labeled telodendrimer was doped into a solution of unlabeled telodendrimers and loaded with paclitaxel during self-assembly according to a previously published protocol.7,8 Doses of PTX-125I-labeled nanomicelles were injected into nude mice bearing SKOV-3 xenografts. As shown in Figure 1, decay-corrected imaging indicated that the PTX-125I-NM accumulated specifically and continuously at the tumor site and reached a peak at around 18–24 hours post injection. The tumor signal then decreased gradually, with substantial retention of nanomicelles in the tumor with respect to background tissue for up to 94 hours post injection. Within the first 5 hours, the majority of the 125I-labeled material was observed in the blood circulation of the mouse and in the blood-rich organs of the chest and upper abdomen. The signal within normal organs, such as the heart, lung,
and liver, was dramatically higher than that in tumor tissue. For 18–24 hours post injection, the signal in the tumor tissue was significantly ($P < 0.05$) higher than in most of the rest of the body. Forty-eight hours post injection, significant retention of PTX-$^{125}$I-NM persisted at the tumor site. Whole body projection images (Figure 2) showed the tumor to be the brightest part of the whole body at 72 hours post injection. In general, the tumor-targeting profile demonstrated by radioimaging is consistent with previous results from optical imaging, in which the telodendrimer was fluorescently labeled with BODIPY, a hydrophobic near-infrared dye. This finding supports previous observations that nanomicelles are able to target the tumor site via the enhanced permeability and retention effect associated with porous tumor vasculature, which enables nanomicelles to release their cargo into localized tumor regions.

Quantitative analysis of the imaging data was performed in order to study the distribution profiles of PTX-$^{125}$I-NM further in normal organs and tumor tissue. The radio signal in the blood pool refers to the signal calculated from the heart cavity. Other normal organs, such as the liver, lung, and muscle, were analyzed in addition to tumor tissue. Dominant $^{125}$I accumulation was shown in the blood pool, liver, and lung within 24 hours. Distribution of $^{125}$I in the liver was much higher than that in tumor tissue for 5 hours post injection (Figure 3A), which is probably due to the high content of blood in the liver. Early accumulation in normal tissues, including muscle, had washed out quickly by one hour post injection. However, uptake in tumor tissue increased gradually and reached a plateau at 18–24 hours, then decreased slowly. This contrast in accumulation between tumor and normal tissue is consistent with enhanced permeation and retention-mediated accumulation of nanomicelles in tumor tissue. The $^{125}$I signal in normal organs and tumor over time was normalized by the signal in muscle to reflect the relative distribution and kinetic profiles. The signal change in muscle is usually in response to the basic clearance profile of the injected PTX-$^{125}$I-NM. Therefore, the signal in tumor tissue or organs normalized by the signal from muscle will subtract interference from the background to highlight accumulation.
and clearance trends of PTX-\(^{125}\)I-NM in tumor tissue or organs. In addition, it will facilitate comparison between tumor tissue and normal organs. The normalized results indicate that the blood pool/muscle ratio was higher than the ratios from other organs and tumor tissue within 24 hours, but decreased rapidly to the background level by 48 hours. The ratio of lung/muscle remained constant, reflecting the similar clearance profile of the \(^{125}\)I-labeled substance in lung and muscle. However, the ratio of tumor/muscle kept increasing over time, indicating gradual accumulation and much slower clearance of the nanoparticles from tumor tissue. The liver/muscle ratio slightly increased over time, probably owing to the reticuloendothelial system in the liver that continuously takes up the \(^{125}\)I-labeled materials remaining in the body (Figure 3B).

**Comparison of biodistribution of \(^{14}\)C-PTX-NM and \(^{14}\)C-PTX-Taxol**

An understanding of the fate and biological effects of nanoparticles in animals is critical to their medical applications.
in vivo. Pharmacokinetic and organ/tissue distribution properties of nanoparticles are of great interest from the clinical point of view because of their potential uses in cancer imaging and therapy. Systematic studies are required to evaluate the distribution of administered nanoparticles inside the body, especially at the tumor site, after injection.

The use of $^{14}$C-PTX enabled us to quantify the drug and its metabolites within the mouse tissue and organs accurately via liquid scintillation counting after intravenous administration. Because the predominant metabolites of paclitaxel include 3′-p-hydroxypaclitaxel, 6α-hydroxypaclitaxel, and 6α,3′-p-dihydroxypaclitaxel, without any metabolic loss of carbon atoms from the parent molecule, we assume that the radiocarbon measurements accounted for all of the parent paclitaxel and its metabolites present in each sample. In order to study the effects of formulation into nanomicelles on the distribution of paclitaxel, $^{14}$C-PTX was loaded into nanomicelles and injected into nude mice bearing SKOV-3 xenografts. As a benchmark for comparison, a small amount of $^{14}$C-PTX was added to Taxol, a Cremophor EL formulation of paclitaxel used in the clinic, which was also administered to the tumor-bearing mice. The tissues and organs were harvested at various time points after injection. The $^{14}$C concentration in blood and tissues was determined by liquid scintillation counting, which allowed calculation of total paclitaxel using the specific activity of each dose as a conversion factor. As shown in Figure 4, the biodistribution profiles of the $^{14}$C-PTX-NM in different organs and tumor tissue were similar to that of $^{14}$C-PTX-Taxol. Both formulations showed low brain uptake of paclitaxel, indicating a general lack of accessibility because of the blood-brain barrier. At the zero time point, blood was withdrawn by heart puncture within one minute of intravenous injection of the $^{14}$C-labeled drug formulation. Organs and tumor tissue were harvested immediately after euthanasia. Samples from all subsequent time points were harvested using the same procedure. Figure 4A shows that $^{14}$C uptake in most of the organs reach a maximum within minutes, followed by rapid washout of drug, as was observed in the $^{125}$I experiments. In contrast, the concentration of paclitaxel and its metabolites showed slower accumulation in tumor tissue, and reached a peak at around 30 minutes post injection, which then decreased slowly compared with normal tissues. The paclitaxel concentration in tumors of animals treated with the telodendrimer nanomicelle formulation was significantly ($P < 0.05$) higher than that in other organs, even in the liver, after 24 hours, which is consistent with the microSPECT/CT imaging results. Tumor uptake in animals treated with $^{14}$C-PTX-Taxol was also observed to increase within the first hour and to decrease gradually to a level similar to that in other organs, including skin, lung, and liver (Figure 4B). It is known that Taxol, which is a Cremophor EL (surfactant)
formulation of paclitaxel, can also form micelles of about 10 nm in size upon dilution in water or biological fluids. Therefore, Taxol also has a relatively long circulation time and is able to target tumor tissue via the enhanced permeation and retention effect. The biodistribution of Taxol in rodents has been described in tumor-bearing mice before. Liver and tumor have shown high paclitaxel concentrations at 24 hours after administration. Similar distribution behavior has also been observed with $^{111}$In-labeled paclitaxel. However, the stability of paclitaxel dissolved in Cremophor EL is poor in vitro upon dilution, as shown in our experiments. Paclitaxel has a tendency to leak out from the Cremophor micelles,
and crystals of paclitaxel were observed in the diluted Taxol solution (20× dilution by phosphate-buffered saline), whereas the telodendrimer nanomicelles loaded with paclitaxel at the same dilution remained very stable in size and solubility over months.

To demonstrate the relative trend of 14C-PTX uptake in different organs and tumor tissue, all values were normalized to muscle 14C levels for each time point, except in blood (Figure 5). Most organ ratios displayed continuously decreasing radiocarbon ratios, except for skin, elimination from which was slow compared with muscle, which is consistent with the high lipophilicity of paclitaxel. For 14C-PTX-NM, the tumor/muscle ratio of radiocarbon content kept increasing and reached a plateau at approximately 8.5-fold 24–48 hours post injection. However, the normalized tumor/muscle ratio of 14C-PTX-Taxol showed an increase of only 3.5-fold at the highest point (Figure 5B), which is much lower than that when 14C-PTX was loaded in nanomicelles. Furthermore, the ratio of tumor/muscle (14C-PTX-Taxol) was even lower than that of the skin/muscle ratio at earlier time points at least 24 hours post injection (Figure 5B). These observations indicated that telodendrimer nanomicelles enhanced the uptake of paclitaxel by tumor tissue.

The whole blood pharmacokinetic profile of paclitaxel after administration of 14C-PTX-NM or 14C-PTX-Taxol at dose levels of 15 mg/kg was calculated using a one-compartment model (Figure 6). A t = 0 time point was used

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**Figure 5B** Normalized biodistribution of 14C-labeled paclitaxel in Taxol® in nude mice with SKOV-3 xenografts (organs/muscle).

The ratio of liver/muscle showed higher levels for 6 hours post injection, but decreased rapidly at later time points. The ratio of skin/muscle kept increasing to a plateau at 6 hours post injection, and decreased after 24 hours. This was even higher than the ratio of tumor/muscle most of the time, and was only around 3.5-fold at its peak.

**Figure 6** Pharmacokinetic data for paclitaxel formulated in nanomicelles and Taxol®.

The terminal half-life (t_{1/2}) 70.3 hours) of paclitaxel loaded into PEG5kCA8 micelles was significantly longer than that for Taxol (t_{1/2} 46.6 hours, P < 0.05). The area under the curve for PTX-NM was slightly higher, indicating modestly longer systemic exposure to the drug as a result of the nanomicelle formulation.

**Abbreviations:** PTX, paclitaxel; NM, nanomicelles.
Abbreviations: AUC, area under the concentration-time curve; C\text{max}, peak plasma concentration; V\text{d}, volume of distribution; Cl, clearance; PTX, paclitaxel; NM, nanomicelles.

table 1 Pharmacokinetic parameters for paclitaxel in whole blood

<table>
<thead>
<tr>
<th>Dose level (mg/kg)</th>
<th>Formulation</th>
<th>C\text{max} (ng/mL)</th>
<th>AUC (ng-h/mL)</th>
<th>Elimination</th>
<th>V\text{d} (L/kg)</th>
<th>CI (L/h/kg)</th>
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<td>15</td>
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<td>70.3</td>
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<td>18306</td>
<td>0.153</td>
<td>46.6</td>
<td>54093</td>
</tr>
</tbody>
</table>

The longer half-life of PTX-NM compared with Taxol is consistent with slower hepatic elimination as a consequence of the 10–100 nm size of the nanomicelle nanocarrier. The slower blood terminal half-life observed for the nanomicelles is consistent with the slower hepatic filtration of the nanoparticle formulation, which has been previously reported for other nanoparticle-mediated drug delivery systems. The paclitaxel clearance of 805 L/hour/kg for the Taxol formulation was reduced to 687 L/hour/kg for the PTX-NM formulation. Despite the fact that higher blood levels are reached when paclitaxel is given in the Taxol formulation, the tissue levels were essentially similar for both of the tested drug preparations.

It has to be noted that two different radiolabeling strategies were used in this research, in which the radio isotope \textsuperscript{125}I was conjugated to telodendrimers for the SPECT imaging study and \textsuperscript{14}C-PTX was loaded inside nanomicelles for the pharmacokinetic experiments. There was a high radioactive signal found in the bloodstream during the first 24 hours in the microSPECT/CT images (Figures 1 and 3A). However, in the pharmacokinetic study, \textsuperscript{14}C-PTX-NM showed much less accumulation in the bloodstream than in the rest of the body, except for muscle, even within one minute post injection (Figure 4A). One explanation for this is that the current nanomicelle formulation is not stable in blood and that the nanomicelles disassemble partially and the loaded drug begins to release from the nanomicelles immediately after in vivo administration and diffusion into normal organs, resulting in a rapid decrease of \textsuperscript{14}C-PTX in the circulation.

In contrast, the \textsuperscript{125}I-telodendrimers, even if coming from dissociated nanomicelles, still remain in the blood circulation for a longer time, contributing to a much higher radioactive signal in the blood during the 24 hours post injection.

We recently reported on the development of disulfide or boronate-catechol cross-linked nanomicelles that are much more stable than the non-crosslinked micelles reported here. The main advantage of crosslinked micelles is that premature drug release in the circulation can be minimized. Furthermore, disulfide reduction can occur inside the tumor cells with high glutathione levels, or on demand with exogenously administered N-acetyl cysteine, a US Food and Drug Administration approved reducing agent. Similarly, the boronate catechol crosslinkages can be reversed with the acidic environment at the tumor site and inside the endosomes of the tumor cells, or with exogenously administered mannitol, an approved diuretic. As a result, such crosslinked nanomicelles may be therapeutically more efficacious than non-crosslinked micelles.

Work is underway in our laboratory using the techniques described in this study to determine the biodistribution and pharmacokinetics of such crosslinked nanomicelles.

Conclusion

The tumor-targeting properties of the telodendrimer PEG\textsuperscript{5kCA\textsubscript{8}} nanomicelle system have been previously demonstrated through near-infrared fluorescence optical imaging with improved preclinical outcomes compared with Taxol and Abraxane\textsuperscript{®}. MicroSPECT/CT imaging enabled by \textsuperscript{125}I-labeling of the nanomicelles further verified the highly efficient tumor targeting of the nanomicelles and provides a potential nanoplatform for cancer imaging and diagnosis. The biodistribution study of \textsuperscript{14}C-PTX indicates that sequestration...
of the drug in the PEG<sup>4</sup>CA<sub>4</sub> nanomicelle system markedly prolongs the paclitaxel circulation time in blood and improves the uptake of paclitaxel at the tumor site. The liver was the organ showing the highest uptake of paclitaxel at the beginning but the uptake had decreased rapidly to lower than tumor levels by 16 hours post injection. The results of radioimaging and biodistribution of the loaded drug support potential use of the PEG<sup>4</sup>CA<sub>4</sub> NM system as a promising nanocarrier for imaging-guided drug delivery.

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**Disclosures**

The authors report no conflicts of interest in this work. KL is the founding scientist of LannoTherapeutics Inc, which plans to develop the nanotherapeutics described in the manuscript.

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