Optimal management of equine keratomycosis

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Abstract: Keratomycosis in the horse exists in several unique clinical forms. This paper discusses the diagnosis and clinical management of keratomycosis in the horse associated with tear film instability, epithelial keratopathy, subepithelial infiltrates, superficial and deep ulcers, plaques, melting ulcers, descemetoceles, iris prolapse, and stromal abscesses. Prompt diagnosis and aggressive treatment of equine keratomycosis can make a major difference in the maintenance of a cosmetic and visual eye.

Keywords: fungal keratitis, keratomycosis, horse, cornea, melting, keratoplasty

Introduction

Keratomycosis in the horse is associated with significant ocular morbidity, is considered a common vision-threatening disease of the horse, and remains a diagnostic and therapeutic challenge to the ophthalmologist, equine veterinarian, and horse owner. Once considered mainly a posttraumatic eye problem, keratomycosis in horses is now also being diagnosed more frequently in temperate geographical regions, although warm, humid, and subtropical environments are still important risk factors for the various forms of equine keratomycosis.

Fungi are normal inhabitants of the equine environment and corneal/conjunctival microflora, and exist in symbiosis with ocular surface bacteria on the horse. Some of these fungal species may be innately pathogenic, while others can become pathogenic following corneal injury and/or alterations in the microenvironment of the ocular surface. A lack of integrity and stability of the precorneal tear film, and corneal epithelial cell injury predispose and encourage fungal adhesion, invasion, and infection of the horse cornea. It may be that unique characteristics of the horse tear film, low resting temperature of the horse cornea, a large corneal surface, and/or interspecies alterations in corneal immunoprotection make the horse more susceptible than other animal species to keratomycosis.

The most often proposed pathogenesis of ulcerative fungal keratitis in horses begins with slight to severe corneal trauma, resulting in an epithelial defect. Corneal epithelial cell loss allows the ocular surface or environmental fungal organisms to adhere, invade, and infect the cornea. Seeding of fungi from a foreign body of plant origin is also possible. A concurrent viral or bacterial corneal infection may be present to weaken the corneal defense mechanisms of the ocular surface. Fungal infection may be suspected when there is a history of corneal trauma, or when a corneal ulcer has been treated with antibiotic therapy for a long
period, or if corticosteroids were previously used topically to treat ocular surface disease.\textsuperscript{2,14,17,18} We suspect other factors may also play a role in the development of equine keratomycosis.

**General management of equine keratomycosis**

Keratomycosis of the horse is found as a continuum of lesions, and in ulcerative and nonulcerative forms. Fungal keratitis in the horse can be grouped into three basic categories, ie, superficial keratomycoses (including tear film alterations and microerosions, superficial ulceration, and plaque formation), stromal ulcerative keratomycosis (including deep ulcers with corneal furrowing, melting ulcers, and corneal perforation with iris prolapse), and stromal abscesses.\textsuperscript{3}

Slight to severe corneal haze, varying amounts and depths of white to yellow cellular infiltration, no to intense corneal vascularization, and varying degrees of painful iridocyclitis can be present in the several forms of equine keratomycosis. *Aspergillus*, *Fusarium*, *Cylindrocarpon*, *Curvularia*, *Penicillium*, *Cystodendron*, yeasts, and molds are known causes of ulcerative and nonulcerative keratomycosis in horses,\textsuperscript{16} and *Mortierella wolfii* was also recently described.\textsuperscript{19} Clinical signs like microerosion, nonulcerative or ulcerative lesions with whitish to yellow opacification, corneal vascularization, plaque formation, melting, and perforation, as well as secondary uveitis and signs of ocular pain are common in equine keratomycosis.\textsuperscript{3}

Diagnostic tests in horses with corneal diseases should include fluorescein and rose Bengal staining,\textsuperscript{8} tear film breakup time, superficial and deep corneal cytology, superficial and deep corneal cultures with attempted growth on both fungal and aerobic plates, and corneal biopsy if surgery is performed.\textsuperscript{16,20} Rose Bengal retention indicates instability of the tear film and may be associated with a poor prognosis for rapid healing.\textsuperscript{8,14} The presence of fungal microorganisms on cytological scrapings from a corneal lesion is a significant finding indicating advanced fungal disease.\textsuperscript{21}

Even in cases where culturing or histopathology cannot identify a fungal agent, fungal DNA can be present and identified by polymerase chain reaction.\textsuperscript{2,22} Laser scanning confocal microscopy is a new emerging technology useful as an in vivo, noninvasive diagnostic test for ocular surface diseases that can aid in the diagnosis of equine fungal keratitis and confirm the presence of fungi in deep corneal stromal lesions in horses.\textsuperscript{23–26}

Differential diagnoses for equine keratomycosis depend on the type of fungal infection but generally include keratoconjunctivitis sicca,\textsuperscript{16} bacterial keratitis, eosinophilic keratoconjunctivitis, immune-mediated keratitis, herpes virus keratitis, traumatic keratopathy, corneal degeneration, calcific band keratopathy, and corneal neoplasia.\textsuperscript{3}

Regardless of the clinical manifestation (epithelial keratitis, ulcerative keratitis, stromal abscess, melting ulcers, iris prolapse), some of the same treatment steps are utilized in all forms of equine keratomycosis, and others are used primarily in the more severe forms of equine keratomycosis. Medical therapy, even with adjunctive surgical procedures, remains the major source of disease control for equine keratomycosis. Subpalpebral lavage systems aid medication instillations in horses with keratomycosis because frequent topical instillations over prolonged periods of time are generally required.\textsuperscript{14}

Selection of specific and efficacious therapeutic topical and systemic medication regimen for equine keratomycosis requires recognition of the presence of fungal and bacterial infection, the degree of stromal destruction, the level of tear film proteinase activity, the presence and severity of intraocular inflammation, and the degree of ocular pain.\textsuperscript{3} Treatment must be directed against the fungi as well as against the iridocyclitis that occurs following fungal replication and death.\textsuperscript{3,16} The role of antiprotease therapy in corneal destruction caused by equine keratomycosis may ultimately be of greater importance than the fungal infection itself, and inability to control hyperproteinase activity is often the major reason for treatment failure.\textsuperscript{2,14} Therapy is often quite prolonged, scarring of the cornea can be prominent,\textsuperscript{16} and surgical therapy for equine keratomycosis may be necessary depending on the clinical manifestation of the disease.\textsuperscript{22,27–30} It is important to consider that horses with keratomycosis usually receive four or more topical medications, and that rarely the possible conflicting effects of one drug on another are considered. It is thus prudent to allow a minimum of 5 minutes between each medication using subpalpebral lavage tubes.\textsuperscript{31} Despite the increased awareness of owners and veterinarians, and despite aggressive therapy, some forms of equine keratomycosis still retain a guarded prognosis for sight.\textsuperscript{32}

**Medical therapy**

In addition to in vitro susceptibility data, selection of an ophthalmic antifungal should include consideration of drug toxicity, tissue penetration, ease of administration, and availability.\textsuperscript{13} Three major classes of antymycotic drugs are available, ie, polyenes (natamycin and amphotericin B), azoles, and nucleoside analogs.\textsuperscript{33} In clinical practice, equine
keratomycosis is most effectively treated with the azoles and/or natamycin. Polyenes have an excellent spectrum of activity, but penetrate the intact cornea poorly, whereas azoles have a good corneal penetration but vary in their fungal susceptibility. Amphoterocin B is more effective against yeasts, in particular Candida spp and Aspergillus spp, but has a limited effect on Fusarium. The combination of topical amphotericin B with subconjunctival injection of fluconazole in humans was more effective than the use of amphotericin B alone, suggesting that combination antifungal therapy can be a good choice.

Natamycin is generally effective against filamentous fungi, although frequent administration is necessary to achieve therapeutic levels in the cornea because of its reported poor penetration in this tissue. We would argue that natamycin is highly active in deep stromal abscess and penetrates corneas debrided of epithelium quite easily when normal permeability function is diminished, and thus remains a very important drug to use in equine keratomycosis.

Miconazole, natamycin, fluconazole, econazole, voriconazole, clotrimazole, and itraconazole have been successfully used topically to treat fungal ulcers in horses but their effects can vary according to geographic region and can change over time. In Florida in 1998, fungi such as Aspergillus and Fusarium were equally susceptible to natamycin and miconazole, but presently miconazole is most effective against Aspergillus and natamycin against Fusarium. Miconazole and ketoconazole were effective against Fusarium spp and itraconazole was particularly effective against Aspergillus spp. Recently, in vitro testing of the effects of itraconazole, miconazole, and natamycin at different concentrations on cellular morphology and cellular proliferation of equine keratocytes were evaluated. Itraconazole showed markedly fewer cytopathologic effects, and natamycin produced the most severe morphological changes in keratocytes at all concentrations and time points. The use of irritating drugs is often necessary to eliminate specific fungal species, but clinicians should exercise some caution in using drugs such as natamycin when fungi are not present.

In a recent in vitro study, voriconazole penetrated the intact horse cornea and appeared to be the most effective antifungal drug for initial treatment of equine keratomycosis. It is now our first choice for antifungal therapy because common equine fungal pathogens, including Fusarium, Aspergillus, and Candida are susceptible to it. Voriconazole concentrations were also established at therapeutic levels in horse aqueous humor after oral and topical administration, and were also detected in the plasma following topical administration. Subconjunctival voriconazole has also been used to treat equine keratomycosis cases, although its efficacy by this method has not been verified. Although reported as being less frequent than other fungi, Candida spp can also be treated with amphotericin B, natamycin, or flucytosine, a nucleoside analog. The 1% flucytosine parenteral solution is well tolerated for topical ophthalmic use.

Silver sulfadiazine and dilute (1:50) povidine iodine can also be used for equine keratomycosis. Silver sulfadiazine has both antibacterial and antifungal properties, and povidine iodine is effective against bacteria, fungi, viruses, and protozoa. Although the antifungal activity of silver sulfadiazine was not evaluated in vivo, the results of one in vitro study and many clinical anecdotes show that it could be useful in clinical cases of equine keratomycosis. We do not recommend silver sulfadiazine in equine keratomycosis in Florida because it does not appear to work well. We use dilute povidine iodine twice daily in our lavage tubing treatments. Chlorhexidine gluconate 0.2% has been used effectively to treat fungal keratitis resistant to antifungal drugs in humans and needs to be evaluated in horses.

Antifungal drug resistance is well recognized, and its prevention depends on maximizing the pharmacodynamic properties of the particular drug class, use of local rather than systemic treatment, and practicing good hygiene. We have noted resistance to miconazole to Fusarium, and natamycin resistance by Aspergillus in Florida in the past decade.

Prophylactic topical antibacterial drugs include triple antibiotic preparations (neomycin, polymyxin B, gramicidin, or bacitracin), tetracyclines, macrolides, and aminoglycosides. Gentamicin, ciprofloxacin, and tobramycin ophthalmic solutions may be used topically to treat Gram-negative bacteria. Amikacin 10 mg/L is another choice. Chloramphenicol may be useful, although it is bacteriostatic. Cefazolin 55 mg/mL is the best treatment for Gram-positive infections such as Streptococcus.

Iridocyclitis is present to some degree with the different forms of equine keratomycosis, and varies according to progression of the disease. Ocular pain and uveitis may increase for a time after starting antifungal medications due to fungus death. Uveitis must be controlled to prevent blindness, and is treated in horses by both the topical and systemic routes. Nonsteroidal anti-inflammatory agents such as flunixin meglumine (1 mg/kg orally, intravenously, or intramuscularly twice daily) and phenylbutazone (2 mg/kg orally twice daily) can be used. Both are effective in reducing
uveal exudation and relieving ocular discomfort, but flunixin meglumine is the most frequently used and efficacious nonsteroidal anti-inflammatory agent for systemic treatment of iridocyclitis in horses.\textsuperscript{14} Flunixin and all other systemically administered nonsteroidal anti-inflammatory agents should be used at the lowest dosage to control pain, because they appear to reduce the speed of corneal vascularization at higher doses.\textsuperscript{14} Atropine 1% solution is effective in causing pupillary dilation and stabilizing the blood-aqueous barrier. It also minimizes synchia formation and reduces ciliary muscle spasm, a factor contributing to ocular discomfort. Horses on atropine should be monitored carefully for signs of reduced intestinal motility and episodes of colic.\textsuperscript{41} Despite their potential to prevent rejection and minimize inflammation, corticosteroids are contraindicated in horses because of a high incidence of infectious and collagenolytic keratitis.\textsuperscript{2,42} Resolution of the corneal condition results in a gradual resolution of the signs of uveitis.\textsuperscript{32}

After corneal damage, elevated levels of matrix metalloproteinases from inflammatory cells, epithelial cells, and keratocytes can be detected in the tears, relative to that of the contralateral healthy eye.\textsuperscript{43} The metalloproteinases play a role in the degradation of matrix components, such as collagen.\textsuperscript{32,44} As levels of matrix metalloproteinases and serine proteases (like neutrophil elastase) become elevated in the tears of horses, prevention and control of collagenolysis is extremely important to speed corneal healing and reduce scarring. Metalloproteinases are inhibited by tissue inhibitor of metalloproteinase, serum and disodium ethylenediamine tetra-acetic acid, tetracyclines, and acetylcysteine. Serine proteases are inhibited by prealbumin inhibitor. Serum contains growth factors, fibronectin, and vitamins (especially vitamin A) that stimulate epithelial cell growth, migration, and differentiation.\textsuperscript{32,33,43} Serum was compared with fresh frozen plasma with regard to corneal epithelium healing in vitro and differences were found. Serum contains more concentrated levels of epithelial growth factor, platelet-derived growth factor, and vitamin A compared with plasma, which probably makes serum significantly superior with regard to stimulation of corneal healing.\textsuperscript{45} Serum or plasma can be administered topically as often as necessary, and is replaced by new serum every 8 days.\textsuperscript{14} Ten percent acetylcysteine or 0.17% ethylenediamine tetra-acetic acid and tetracyclines (oxytetracyclines, doxycycline) can also be administered until stromal liquefaction diminishes. In some eyes, both ethylenediamine tetra-acetic acid and serum are needed and used simultaneously to stop melting.\textsuperscript{14,44,46}

**Surgical treatment**

Fungal keratitis in horses has been described and grouped into three basic categories, ie, superficial keratomycoses (including tear film alterations and microerosions, superficial ulceration, and plaque formation), stromal ulcerative keratomycosis (including deep ulcers with corneal furrowing, melting ulcers, and corneal perforation with iris prolapse), and stromal abscesses.\textsuperscript{3} Aggressive medical treatment for all forms of equine keratomycosis may need to be combined with surgical therapy in eyes where there is a poor response to topical antifungal therapy, epithelial and fungal debris is pronounced, collagenolytic activity is extreme,\textsuperscript{9} stromal loss is greater than one third corneal thickness (including deep ulcers and descemetocoeles), corneal perforation occurs, and/or if the anterior uveitis cannot be controlled.\textsuperscript{3,14} The surgical procedure chosen depends on the depth and location of the offending lesion.\textsuperscript{22}

**Epithelial debridement/superficial keratectomy**

We have observed that dead fungi are difficult to remove by phagocytosis and enzymatic dissolution due to their large size, and their physical presence may inhibit corneal healing, and thus need to be removed surgically. Epithelial debridement with topical anesthetics and a blade or cotton swab removes devitalized corneal tissue and dead hyphae in superficial forms of equine keratomycosis,\textsuperscript{16} and can accompany medical therapy or precede more invasive surgeries to improve antifungal drug penetration. This modification of a superficial keratectomy can be realized with a cotton-tipped applicator, spatulas, scalpels, forceps,\textsuperscript{29} and/or a diamond burr.\textsuperscript{47} Debridement should be continued until only firmly adhered epithelium at the edge of the fungal lesion remains.\textsuperscript{29} Fungal removal by debridement speeds healing, minimizes scarring, and decreases the stimulus for iridocyclitis.\textsuperscript{16} Debridement is not suitable for deeper lesions.

**Amniotic membrane transplantation**

Amniotic membrane can be used successfully to preserve globe structure and vision in horses with equine keratomycosis and mild or severe keratomalacia,\textsuperscript{48} and has replaced the use of large diameter 180 degree or 360 degree conjunctival flaps.\textsuperscript{27} The amnion tissue is harvested from normal equine placenta but is not yet available commercially.\textsuperscript{41} This biomaterial acts as a self-sacrificing dressing because it attracts proteases and microbes that could attack the cornea, and allows the cornea to heal. It is believed that the amnion provides proteins like lumican and dermatopontin to speed corneal healing.
layers, and in combination with conjunctival grafts. Membrane can also be used alone, in single or multiple keratoplasty in horses with encouraging results. Amniotic University of Florida combined with penetrating and lamellar proliferation and migration, dermatopontin induces fibroblast cell adhesion. We have utilized amnion grafts at the Veterinary Medicine: Research and Reports 2012:3 in the horse. Lumican modulates cellular behavior like proliferation and migration, and dermatopontin induces fibroblast cell adhesion. We have utilized amnion grafts at the University of Florida combined with penetrating and lamellar keratoplasty in horses with encouraging results. Amniotic membrane can also be used alone, in single or multiple layers, and in combination with conjunctival grafts.

### Conjunctival grafts

Pedicle conjunctival grafts are frequently used for treatment of deep, melting, and large diameter corneal ulcers, descemetoceles, and perforated corneal ulcers with iris prolapse. They contain blood vessels and lymphatics thus offering antibacterial, antifungal, antiviral, antiprotease, and anticollegenase effects. When harvested from the limbal area, the transplanted conjunctival epithelium contains stem cells capable of producing corneal epithelium. Conjunctival grafts also provide mechanical support for a thin or weakened cornea, and a route for systemic antibiotic to be delivered to the corneal ulcer. Conjunctival grafts are used in conjunction with primary repair or penetrating keratoplasty in cases of iris prolapse or large diameter full thickness stromal abscesses.

### Penetrating and lamellar keratoplasties

Corneal transplantation for treatment of severe inflammatory keratopathies in horses with corneal fungal infection and melting has been used successfully at the University of Florida since 1993. Penetrating keratoplasty is a full thickness microsurgical transplantation of the corneal epithelium, stroma, and Descemet's membrane/endothelium. Penetrating keratoplasty is performed in horses for therapeutic and tectonic reasons in melting and fungal ulcers with extensive stromal loss, iris prolapse/descemetoceles, and full thickness stromal abscesses. It is a viable, routine surgical technique in horses with severe fungal keratitis, and has a very good visual outcome, although the penetrating keratoplasty site remains opaque to some degree due to graft rejection. Frozen donor cornea up to 13 mm in diameter can be sutured in place with 7-0 to 8-0 sutures to replace diseased or missing cornea.

**Table 1 Clinical signs from equine keratomycosis and their respective treatment**

<table>
<thead>
<tr>
<th>Clinical appearance</th>
<th>Treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tear film instability/</td>
<td>Topical antibiotics, serum, atropine, antifungals; systemic flunixin</td>
</tr>
<tr>
<td>microlesions</td>
<td>meglumine; fly masks</td>
</tr>
<tr>
<td>Epithelial keratopathy</td>
<td>Topical antibiotics, serum, atropine, antifungals; systemic flunixin</td>
</tr>
<tr>
<td></td>
<td>meglumine; fly masks</td>
</tr>
<tr>
<td>Subepithelial infiltrates</td>
<td>Topical antibiotics, serum, atropine, antifungals; systemic flunixin</td>
</tr>
<tr>
<td></td>
<td>meglumine; fly masks</td>
</tr>
<tr>
<td>Superficial ulcers</td>
<td>Topical antifungals, antibiotics, serum and/or ethylenediamine tetra-acetic acid, atropine; systemic flunixin meglumine; protective hood</td>
</tr>
<tr>
<td>Deep ulcers</td>
<td>Conjunctival pedicle graft after debridement or use of grafts; partial temporary tarsorrhaphy; clinical treatment as described; protective hood</td>
</tr>
<tr>
<td>Plaques</td>
<td>Excision of the plaque and underlying superficial stroma; keratoplasty (corneal conjunctival or amnion graft); partial temporary tarsorrhaphy clinical treatment as described; protective hood</td>
</tr>
<tr>
<td>Melting ulcers</td>
<td>Keratectomy and deep anterior lamellar keratoplasty; partial temporary tarsorrhaphy; clinical treatment as described; protective hood</td>
</tr>
<tr>
<td>Descemetoceles</td>
<td>Keratectomy and replacement with cornea (deep anterior lamellar keratoplasty) or Biosist® in combination with amnion or conjunctival graft; partial temporary tarsorrhaphy; clinical treatment as described; protective hood</td>
</tr>
<tr>
<td>Iris prolapse</td>
<td>Excision of iris tissue necrotic or contaminated; penetrating keratoplasty with conjunctival graft; partial temporary tarsorrhaphy; clinical treatment as described</td>
</tr>
<tr>
<td>Stromal abscess</td>
<td>Posterior lamellar keratoplasty for deep stromal abscess in the axial cornea or deep endothelial lamellar keratoplasty for limbal deep stromal abscess; partial temporary tarsorrhaphy; clinical treatment as described; protective hood</td>
</tr>
<tr>
<td>Failure of treatment</td>
<td>Enucleation</td>
</tr>
</tbody>
</table>

**Note:** The clinical therapy is generally similar, but the interval between instillations is reduced when the clinical manifestation becomes worse.
visual outcomes and more rapid healing when compared with penetrating keratoplasty. The deep anterior lamellar keratoplasty is another type of lamellar keratoplasty to replace the epithelium and stroma in melting ulcers where Descemet’s membrane is intact in the horse.

Posterior lamellar keratoplasty is indicated for deep stromal abscesses with a clear or slightly vascular overlying anterior stroma in the axial cornea. A deep lamellar endothelial keratoplasty has been used for limbal deep stromal abscesses in horses where the superficial cornea may be vascularized but is otherwise normal. Exposure of the abscess is accomplished by dissection of the cornea to the depth of the lesion, removal of the abscess with a corneal trephine, replacement and suturing of the split thickness donor corneal graft into the surgical site, and then surgical replacement of the overlying remaining normal corneal layers. The anterior chamber can be reformed with sodium hyaluronate, air, or lactate Ringer’s solution.

Deep anterior lamellar keratoplasty is a surgical technique to remove the entire corneal stroma down to bare normal Descemet’s membrane and endothelium, and is indicated in horses with rapidly progressive, large diameter, catastrophic, and melting corneal ulcers with and without exposure of Descemet’s membrane. Dissection and removal of superficial to deep layers of necrotic cornea is followed by suturing of donor corneal epithelium and anterior stroma over the surgical lesion.

Surgical lamellar and penetrating keratoplasties can also be used with biologic membranes like amniotic membrane and small intestinal porcine submucosa. The small intestinal submucosa graft provides a scaffold for corneal healing as well as additional strength to the overlying bulbar conjunctival graft. Grafts can be used to replace the missing stromal and epithelial cells.

Third eyelid flaps and tarsorrhaphy
A temporary tarsorrhaphy is recommended after most equine corneal surgery, such as amnion and conjunctival grafts, penetrating keratoplasty, deep endothelial lamellar keratoplasty, deep anterior lamellar keratoplasty, and posterior lamellar keratoplasty. It minimizes to the cornea surface and improves graft adherence. Third eyelid flaps can provide physical support to weakened corneas but are not indicated in most cases of deep equine keratomycosis. In the postoperative period, use of a protective hood with a plastic eyecup is important to minimize rubbing of the eye. Horses with surgical treatment of equine keratomycosis should be evaluated daily to confirm that there are no complications.

Therapeutic management of keratomycotic lesions (Table 1)

Tear film instability with and without microlesions
Horses with fungal-induced tear film instability display generalized and/or punctate haziness of the cornea. The lesions are rose Bengal-positive and may represent a form of qualitative keratoconjunctivitis sicca. Rose Bengal retention is also found with quantitative keratoconjunctivitis sicca, viral keratitis, corneal edema, and severe corneal scarring in the horse. Aspergillus is the most common cause of this in our experience.

Horses with ocular pain, a dry or hazy appearance to the cornea in a multifocal, punctate, or generalized pattern, and no uveitis, should be stained with rose Bengal and fluorescein stains. Keratomycosis, viral keratitis, keratoconjunctivitis sicca, immune-mediated keratitis, and conditions caused by environmental air pollution should be considered in the differential diagnosis. Superficial epithelial lesions of the cornea can be divided into microerosions, microcysts, and keratopathies. Epithelial and subepithelial lesions require a biomicroscopic lamp for differentiation. Punctate epithelial microerosions are fine lesions that stain with rose Bengal and stain faintly with fluorescein. They appear in areas of cornea in which there has been partial thickness loss of normal surface epithelial cells with exposure of underlying immature corneal epithelial cells, appearance of punctate microcysts in areas of epithelial healing with recurrent erosions, and cystic spaces in edematous corneal epithelium. Rose Bengal is able to stain epithelial cells and keratin, exposed stroma, and degenerative epithelial cells only when there is a generalized deficiency of the mucin layer of the precorneal tear film. Hyphae can be obtained from corneal scrapings and fungi cultured from the scraping site in fungal-induced microerosions. Medical therapy should include topical antibiotics, serum, atropine, and antifungals, and systemic flunixin meglumine. Fly masks are recommended to reduce tear film evaporation. This form of keratomycosis can progress to the ulcerative form of keratomycosis, but generally has a favorable prognosis.

Epithelial keratopathy
Fungi can invade the epithelium to cause a very fine punctate or generalized haze. The diagnosis of epithelial keratomycosis is made with negative rose Bengal staining, negative or weak fluorescein staining, epithelial opacities from slit-lamp examination, and cytologic evidence of hyphae,
fungal culture, and positive resolution of the condition with antifungal medication. Medical therapy should include topical antibiotics, serum, and antifungals, and systemic flunixin meglumine. Fly masks are recommended to reduce tear film evaporation. This form of keratomycosis can progress to the subepithelial form of keratomycosis, and generally has a favorable prognosis once diagnosed and treated. We suspect that this type and the subepithelial form of eosinophilic keratoconjunctivitis are common to horses in any geographic region. Horses with epithelial keratomycosis and subepithelial infiltrates have been reported anecdotally in the US, UK, Japan, France, Germany, Canada, Denmark, and Finland.

Subepithelial infiltrates
Subepithelial infiltrates can be caused by fungi, lymphocytes, neutrophils, or fibroblasts. Diagnosis of this condition requires the slit-lamp biomicroscope for localization. Fungi have been identified from subepithelial punctate opacities (Figure 1B) which retain neither fluorescein nor rose Bengal stains prior to scraping for cytology. Treatment is the same as described for epithelial form. We feel that equine subepithelial infiltrates represent a quite common and distinct keratopathy in the horse, and that many horses with equine subepithelial keratomycosis are able to resolve the condition spontaneously. Equine subepithelial keratomycosis may be more prevalent in temperate climates than the ulcerative forms noted in subtropical environments. We also suggest that equine subepithelial keratomycosis may be a preliminary event in the formation of corneal ulcers in some horses, and deep stromal abscesses in other horses.

Superficial ulcers
Superficial fungal ulcers arise from loss of the cornea epithelium and stain positively for fluorescein (Figure 1C–E).  

Figure 1 Clinical appearance of different forms of equine keratomycosis. (A) Rose Bengal-positive epithelial microerosions. (B) Subepithelial keratomycosis with microerosions. (C) Superficial ulcerative keratitis fluorescein stained. (D) Superficial keratomycosis fluorescein weakly positive. (E) Superficial ulcerative keratitis rose Bengal-positive showing tear film instability. (F) Superficial fungal plaque; note the brownish color, corneal edema, and neovascularization.
Due to the lack of epithelium, the cornea becomes more susceptible to opportunistic infections. Ulcers also result in increased tear film matrix metalloproteinase activity, and there is some degree of iridocyclitis. Medical therapy for a superficial fungal ulcer should include systemic flunixin meglumine, and topical antifungals, antibiotics, serum and/or ethylenediamine tetra-acetic acid, and atropine. The frequency is determined by the severity of the condition.

Deep ulcers

The loss of more than the initial third of the stroma is considered a deep ulcer. Some deep ulcers can heal with medical therapy alone, but deep ulcers and descemetoceles that are rapidly progressive with increasing signs of uveitis, and areas of marked stromal melting, stromal loss or marked cellular infiltrate are considered complicated, likely infected, and require surgical intervention. Rupture may be imminent. Surgical therapy for a deep fungal ulcer could be a conjunctival pedicle graft after debridement of the corneal lesion, penetrating keratoplasty, deep anterior lamellar keratoplasty, or use of biomaterial grafts such as amnion or Biosist™ to help the weakened cornea and prevent its rupture. Medical therapy for deep ulcers remains aggressive, and should include systemic flunixin meglumine, as well as topical therapy of antifungals, antibiotics, serum and ethylenediamine tetra-acetic acid, and atropine. The frequency should be increased to every hour in complicated and progressive cases.

Plaques

A dense, dark-colored plaque elevated off the corneal surface can be caused by fungal invasion of the corneal epithelium and stroma. The plaques appear film-like and/or fluffy in the early stages, and are typically discolored (Figures 1F and 2A). The lesions begin as a small opacity, but can enlarge rapidly and occupy more than half of the corneal surface. The specific reason for this fungal plaque and abscess formation is unknown. The presence of a deep stromal excavation (furrow) bordering the plaque is considered a serious development. The furrow precedes corneal vascularization, can progress rapidly over 24–48 hours, causes pain, and can lead to corneal rupture; like other forms of ulcerative keratomycosis in horses, corneas with fungal plaques heal very slowly. In most cases of fungal plaques, removal by keratectomy speeds healing time. Fungal plaques can benefit from excision of the plaque and underlying superficial stroma, and a keratoplasty with placement of a corneal conjunctival or amnion graft. Medical therapy should include systemic flunixin meglumine, as well as topical antifungals, antibiotics, serum and ethylenediamine tetra-acetic acid, and atropine.

Melting ulcers

Stromal malacia or melting occurs as a result of collagenolysis due to protease liberation from invading neutrophils, microorganisms, and corneal epithelial cells or keratoctyes (Figure 2B and C). The result is loss of rigidity and structure of the corneal collagen with development of a deep ulcer or descemetocele. Standard treatment for melting fungal ulcers includes topical antifungals and antibiotics, serum and ethylenediamine tetra-acetic acid, atropine, as well as systemic flunixin meglumine. It is imperative and often difficult to control collagenolysis medically.

Keratectomy and deep anterior lamellar keratoplasty may be indicated to speed healing by removing infected and necrotic tissue, encouraging vascularization, minimizing scarring, and decreasing the stimulus for anterior uveitis. A conjunctival or amnion graft can restore the anterior cornea, and a temporary partial tarsorrhaphy avoids additional trauma to the cornea. In melting ulcers, tear film protease/collagenase activity can digest absorbable sutures or the graft itself, resulting in possible corneal perforation such that antiprotease therapy should be rigorously maintained postoperatively.

Descemetoceles

A descemetocele is a deep corneal lesion in which the corneal epithelium and stroma are completely destroyed, leaving a corneal lesion lined only by Descemet’s membrane and corneal endothelium (Figure 2D). Once this barrier is breached, a full thickness corneal perforation occurs, the aqueous humor is lost, and the iris can prolapse. Descemetocele and iris prolapse can be a result of keratomalacia, in which collagenolysis is not controlled. It is not uncommon for stromal melting to progress rapidly to full-thickness corneal perforation within 48 hours in the most severe cases. Because of the fragile nature of descemetoceles, as well as the potential for intraocular inflammatory damage with perforation, descemetocele repairs should be considered as an emergency. Vision should always be assessed or attempted to determine the likely visual outcome. Evaluation of the consensual pupillary light reflex (if possible) and dazzle reflex may provide some information on visual capability, and ocular ultrasonography can also be used to assess the posterior segment.

Medical therapy should include systemic flunixin meglumine, and topical antifungals, antibiotics, serum and ethylenediamine tetra-acetic acid, and atropine. Surgical
treatment of descemetoceles consists of keratectomy to remove the necrotic and infected tissue,\textsuperscript{16} and replacement of the missing cornea with cornea (deep anterior lamellar keratoplasty), or replacement with collagen from Biosist or Acell\textsuperscript{TM} in combination with an amnion or conjunctival graft\textsuperscript{16,28} and partial temporary tarsorrhaphy.\textsuperscript{46} Sometimes, conjunctival grafts alone result in continuing leakage of aqueous humor through a corneal perforation, and the use of cornea or another tissue can provide the best results.\textsuperscript{29}

**Iris prolapse**

In keratomycosis, an iris prolapse can result from progression of ulcerative keratitis, or rupture from a stromal abscess.\textsuperscript{1} The prognosis is guarded for ocular survival and vision if surgery is not attempted. Surgical management requires corneal repair and stabilization. Prolapsed iris tissue that appears desiccated, necrotic or contaminated, or that has been prolapsed from more than 24 hours, should be excised rather than replaced in the anterior chamber. It is important to perform a keratectomy on the margins of the corneal defect to debride the devitalized and infected tissue. A penetrating keratoplasty with conjunctival graft can then be placed.\textsuperscript{44} Vision can be achieved in up to 68% of eyes with iris prolapse using a technique of penetrating keratoplasty/conjunctival flap.\textsuperscript{60} Medical therapy should include systemic flunixin meglumine, and topical antifungals, antibiotics, serum and ethylenediamine tetra-acetic acid, and atropine.\textsuperscript{8}

**Stromal abscess**

Fungal tropism for glycosaminoglycans has been suggested, based on the fact that hyphae are often found sequestered

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**Figure 2** Clinical appearance of equine keratomycosis. (A) Necrotic fungal plaque and intense corneal perilimbal vascularization. (B) Corneal melting and cellular infiltration. (C) Corneal melting and hypopyon into the anterior chamber. (D) Descemetocele, perilimbal neovascularization, hypopyon, and fibrin into the anterior chamber. (E) Deep stromal abscess in the axial cornea and intense corneal edema and vascularization. (F) Deep stromal abscess at 10 o’clock with intense perilimbal vascularization and hypopyon, fibrin, and hyphema in the anterior chamber.
in the posterior stroma near and/or infiltrating Descemet's membrane where glycosaminoglycans exist in abundance. Stromal abscesses vary widely in size, location in the cornea, border clarity, and stromal depth. Superficial stromal abscesses can be bacterial or fungal in origin, whereas deep stromal abscesses (Figure 2E and F) are thought to be more likely fungal in origin, and are thus often refractory to medical treatment compared with more superficial abscesses. The corneal epithelium may or may not be defective in the presence of a stromal abscess.2

Deep stromal abscesses are generally refractory to medical management due to poor penetration of medications, and the inability of the cornea to remove dead fungal hyphae.2,42 Penetrating keratoplasty is the treatment of choice for a full thickness corneal stromal abscess. The most effective surgical treatment for a deep stromal abscess is penetrating keratoplasty or deep endothelial lamellar keratoplasty because it removes infectious organisms, necrotic stroma, and metabolites of degenerating leukocytes, while preserving the overlying stroma and epithelium.42 Penetrating keratoplasty is indicated for deep stromal abscesses in the axial cornea and deep endothelial lamellar keratoplasty has been used for limbal deep stromal abscess.29,53 Our success rate using these procedures approaches 90%.28 If the donor button exceeds 8 mm in diameter, a pedicle stromal graft is also indicated over the surgical site if the donor button exceeds 8 mm in diameter.28,53 Our success rate using these procedures approaches 90%.28

Disclosure
The authors report no conflicts of interest in this work.

References


