Williams–Campbell syndrome: a case report

Introduction: Williams–Campbell syndrome, also known as bronchomalacia, is a rare disorder characterized by a deficiency of cartilage in subsegmental bronchi, leading to distal airway collapse and bronchiectasis. There have been few reports about patients affected by saccular bronchiectasis, paracicatricial emphysema, and diminished cartilage. These are all characteristic of Williams–Campbell syndrome.

Case presentation: This report presents a 57-year-old woman with progressive dyspnea, cough, sputum production, and fever. The clinical and laboratory examination revealed that the patient had a respiratory infection due to bronchiectasis caused by Williams–Campbell syndrome, which was undiagnosed in the patient until then.

Conclusion: Although a rare syndrome, when patients' signs and symptoms include recurrent respiratory infections, bronchiectasis, productive cough, and dyspnea, Williams–Campbell syndrome should be included in the differential diagnosis.

Keywords: bronchiectasis, Williams–Campbell syndrome, bronchomalacia

Introduction

Williams–Campbell syndrome is a rare disorder characterized by a deficiency of cartilage in subsegmental bronchi, leading to distal airway collapse and bronchiectasis. Williams–Campbell syndrome results from the absence of cartilage rings beyond the first and second bronchial divisions with resultant bronchiectasis, which typically affects the fourth- to sixth-order bronchi. Affected patients have normal caliber trachea and central bronchi. The symptoms and prognosis ultimately depend on the extent of cartilage maldevelopment of the bronchi. Although the syndrome has been best described in children with recurrent pneumonia and broncho-obstructive symptoms such as coughing and wheezing, there have been recent descriptions in adults as well.

Case presentation

A 57-year-old woman presented in the emergency department complaining of progressive dyspnea for the previous 5 days. On clinical examination she had productive cough with clear sputum and intermittent fever. She was a lifelong non-smoker but with multiple hospitalizations due to respiratory infections and asthma-like episodes from the age of 8 years. During the past 15 years the patient was treated symptomatically with antibiotics, nebulizer bronchodilators, and long-term oxygen therapy (2 L/min/24 hours). She had daily exposure to gases from a brazier used at home.
since childhood. Her family past medical history included her 30-year-old sister’s sudden death from unknown reasons and her mother’s infection from tuberculosis.

At the time of admission her axillary temperature was 38°C and blood pressure 120/70 mmHg. The patient was in respiratory insufficiency with an arterial pH of 7.46, PCO₂ of 38 mmHg, PO₂ of 72 mmHg, and HCO₃ of 30 mmol/L on air. Lung auscultation revealed widespread bilateral crepitating and cracks over the lower part of both lungs. No lymphadenopathy was found and examination of other systems was unremarkable. The electrocardiogram was normal with a heart rate of 69 beats per minute.

Laboratory data showed an elevated erythrocyte sedimentation rate of 100 mm/h and a white blood count of 9000/µL without peripheral blood eosinophilia, and C-reactive protein was 9.3 mg/dL. Biochemical values were normal. Her sputum sample for acid-fast bacilli was negative on multiple occasions. Sputum sent for fungal culture did not grow any organisms. Screening for Aspergillus in sputum cultivation was also negative. Pseudomonas aeruginosa was isolated from her sputum culture. Urine antigen for Legionella and Streptococcus was negative.

Her chest X-ray demonstrated increased lung volumes and revealed multiple ring shadows in lungs that were almost symmetrical in distribution and more apparent in the mid and lower zones. There was the sense of air-fluid levels in some of them corresponding to cystic bronchiectasis. Also, there was thickening of the interstitial network. The trachea and main bronchi were normal (Figure 1).

A laboratory investigation was performed for α-1 antitrypsin level and was normal. Total serum immunoglobulin (IgE) concentration was elevated at 2500 U/mL (normal range 0–165 U/mL). Immediate cutaneous reaction to Aspergillus fumigatus was not available at the time of the patient’s hospitalization; nevertheless, A. fumigatus-specific serum IgE levels were 1.37 U/mL. This is essential for the diagnosis of allergic bronchopulmonary aspergillosis.³

Total serum IgG, IgA, and IgM levels were quantified to rule out hypogammaglobulinemia and did not reveal any deficiencies. Antinuclear and antineutrophilic cytoplasmic antibody tests were performed to exclude Churg–Strauss syndrome, and the results were negative.

A pulmonary function test demonstrated a mixed obstructive and restrictive pattern. There was a reduced forced vital capacity of 1200 mL (43.35% pred), reduced expiratory volume in 1 second of 720 mL (30.7% pred) with Tiffeneau index, reduced expiratory volume in 1 second/forced vital capacity of 76.29, and carbon monoxide-diffusing capacity of 28%.

A contrast-enhanced computed tomography (CT) scan of the chest demonstrated central cylindrical/cystic bronchiectasis involving segmental/subsegmental bronchi, bronchial wall thickening with air-fluid levels, and mucus plugging with predominant localization in the lower lobes. There was ground glass and tree-in-bud scattered in all lobes, a finding consistent with pneumonitis and acute bronchiolitis. Chest CT also revealed a mosaic picture, which also emerges in all lobe findings consistent with bronchiolitis obliterans. Collapse of bronchi with distal air trapping, the result of an excessively compliant bronchial wall, particularly in the left and right lower lobe, was also present (Figure 2).

We also performed fiberbronchoscopy on the patient. The findings were unremarkable. The bronchoalveolar lavage fluid obtained from the patient for an acid-fast bacillus was negative on multiple occasions, but the same P aeruginosa was again isolated. The bronchoalveolar lavage fluid was negative for Aspergillus antibodies (IgG and IgM).

A diagnosis of saccular bronchiectasis due to Williams–Campbell syndrome was made based on clinicoradiological findings, laboratory testing, and past medical history. Genetic testing was planned, but the patient refused further investigations due to financial constraints. The patient was treated with intravenous antibiotics; third-generation cephalosporins, ciprofloxacin, and acetylcysteine; and chest physiotherapy, and was discharged after 1 month.

Written informed consent was obtained from the patient upon discharge for publication of this case report and all accompanying images.

**Discussion**

In 1960, Williams and Campbell first reported the unusual pattern of bronchiectasis in five patients whose disease presented in infancy with similar clinical features.
They noted markedly compliant, soft central bronchi that ballooned on inspiration and collapsed on expiration. In this syndrome, bronchiectasis due to deficiency in cartilage of the third- to sixth-order bronchi generation is observed.

The syndrome is usually presented in childhood, with recurrent pneumonia and broncho-obstructive symptoms such as coughing and wheezing. Nevertheless, adult cases have been reported without pathologic confirmation. The symptoms and prognosis depend on the extent of cartilage maldevelopment. CT imaging demonstrates bilateral cylindrical/cystic bronchiectasis distal to the third-generation bronchi (segmental/subsegmental) with hyperinflation of the lung. On expiration, collapse of the bronchi with distal air trapping is observed as the result of an excessively compliant bronchial wall. The trachea and central bronchi remain normal in caliber, a distinguished feature of this disease.

A genetic background and familial occurrence are also reported. The compliant bronchi collapse during coughing, leading to poor airway drainage. Subsequently, progressive obstructive disease develops, causing hyperinflation of the lung and segmental or lobar collapse. Recurrent destruction of the bronchial tree and inadequate clearance of mucus result in further damage to the lung parenchyma.

The long-term prognosis is variable, with rapid clinical deterioration and death in some children and prolonged survival in others. No specific treatment exists for Williams–Campbell syndrome. Prophylaxis from exacerbations remain the basis of treatment. Prophylaxis can be achieved if an oral or intravenous antibiotic is given for 7–10 days or until sputum production decreases. For severe cases, several different antibiotics may be used sequentially in a continuous regimen to minimize bacterial resistance. Transplantation has been reported in a patient with severe respiratory symptoms from Williams–Campbell syndrome, but the patient died 1 year later. Upon postmortem examination it was observed that the main bronchi had bronchomalacia, which was attributed to a respiratory infection during the postsurgery period. Moreover, when necessary, based on the patient’s symptoms (bronchospasm and thick, tenacious sputum), a bronchodilator, combined with postural drainage and chest percussion, can help remove secretions. Respiratory exercise with free flow and bronchoscopy may be used to help mobilize secretions. Hypoxia requires oxygen therapy. Noninvasive positive airway pressure can be used when respiratory acidosis exists. Other acquired and congenital conditions associated with bronchiectasis, including ciliary dyskinesia, cystic fibrosis, allergic bronchopulmonary aspergillosis, and immunoglobin deficiencies, must be excluded.

We were able to exclude cystic fibrosis, immunodeficiencies, and aspergillosis.

Conclusion
Differential diagnosis of saccular bronchiectasis includes cystic fibrosis, allergic bronchopulmonary aspergillosis, α-1 antitrypsin deficiency, Ig deficiency, radiation fibrosis, tuberculosis, immotile (dyskinetic) cilia syndrome, Mounier-Kuhn syndrome, and Williams–Campbell syndrome. When patients’ signs and symptoms include recurrent respiratory infections, bronchiectasis, productive cough, and dyspnea, Williams–Campbell syndrome should be included in the differential diagnosis.

Author contributions
PZ, MK, KP, and PB wrote the manuscript. AM, VZ, DP, NL, and TK diagnosed and treated the patient. SB performed the
microbiology tests. NC evaluated the radiographic images. NK and KZ provided useful insights.

Disclosure

The authors report no conflicts of interest in this work.

References