Antibody-induced secondary treatment failure in a patient treated with botulinum toxin type A for glabellar frown lines

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Abstract: Botulinum toxin type A (BTX-A) preparations are widely used nonsurgical treatments for facial wrinkles. Higher doses of BTX-A are also used for therapeutic purposes in the treatment of conditions involving increased muscle tone, such as cervical dystonia. The phenomenon of antibody-induced treatment failure is well known in the therapeutic setting, but reports are also emerging following cosmetic use of BTX-A. We describe the case of a 41-year-old female nurse who developed secondary treatment failure during 6 years of BTX-A treatment for glabellar lines. After a good response to the first BTX-A injection, the intensity and duration of effect decreased after subsequent treatments. Antibody tests revealed a high titer of neutralizing anti-BTX-A antibodies. This case shows secondary treatment failure due to the production of neutralizing antibodies following administration of BTX-A formulations for cosmetic purposes and demonstrates that immunogenicity of BTX-A preparations is an important consideration, even in the cosmetic setting.

Keywords: botulinum toxin type A, neutralizing antibodies, antibody-induced treatment failure

Introduction

Botulinum toxin type A (BTX-A) preparations are commonly used in therapeutic and cosmetic applications with great success. However, after initial good responses, therapy can subsequently fail either partially or completely (secondary therapy failure) due to a number of causes, including inadequate dosage, injection of inappropriate muscles, and development of BTX-A neutralizing antibodies. Antibody-induced treatment failure following treatment with BTX-A for therapeutic purposes has been reported to range from 4% to 10% of patients treated and to decrease to 1%–6% after the foreign protein load of the preparation used is reduced. The risk of developing antibody-induced treatment failure has been shown to increase with short injection intervals and high injected doses. Despite lower BTX-A dosages being used in cosmetic applications compared with therapeutics, there are now emerging reports of antibody-induced treatment failure in facial esthetics.

Case report

Here we report the case of a 41-year-old Caucasian woman who had been receiving BTX-A preparations for the treatment of glabellar lines for 6 years (Table 1). She was initially treated in 2004 with a commercially available BTX-A preparation, abobotulinumtoxinA (Dysport®, Ipsen Ltd, Slough, UK). The effects of treatment lasted for 3–4 months. However, following her next treatment with
abobotulinumtoxinA in the glabellar region, the duration of effect was reduced to 8 weeks. From 2005 to 2008, prior to presentation at our clinic, the patient received further injections of abobotulinumtoxinA in the glabellar area twice yearly and reported that the duration of effect subsequently diminished to a maximum effect of 3–4 weeks’ duration. From the beginning of 2009, we treated this patient with other BTX-A preparations, first with onabotulinumtoxinA (Botox®/Vistabel®, Allergan, Irvine, CA), and more recently with incobotulinumtoxinA (Xeomin®/Bocouture®, Merz Pharmaceuticals GmbH, Frankfurt, Germany).

The first treatment in our clinic was 28 U of onabotulinumtoxinA in the glabellar area, but the treatment was suboptimal and the patient returned approximately 2 weeks later, when she received an additional 9 U of onabotulinumtoxinA. For this second treatment and subsequent treatments, BTX-A was injected in the periorbital region as well as the glabellar region at the patient’s request. The patient reported the duration of effect to be 2–3 weeks. Three months later, the patient received one further treatment in the glabellar and periorbital areas, with 10 U onabotulinumtoxinA, a lower dose than usual, as requested by the patient. However, the patient was still dissatisfied with the treatment outcome and duration of effect. Therefore, we changed to administration of incobotulinumtoxinA at a higher dose (20 U) into the glabellar and periorbital regions, but the duration of effect was only 3–4 weeks. Indeed, two subsequent injections of incobotulinumtoxinA at higher doses (22 U and 44 U) also failed to elicit a response of longer duration. The clinical photograph taken approximately 1 month after the final injection shows no remaining effect of neurotoxin (Figure 1C). Therefore, we considered the possibility that the patient had neutralizing anti-BTX-A antibodies. This seemed likely since neutralizing anti-BTX-A antibodies would not be overcome by switching to another BTX-A formulation, and high antibody titers could prevent a response even to larger doses.

In December 2009, the patient’s serum was tested for the presence of neutralizing anti-BTX-A antibodies at a specialized laboratory (Toxogen GmbH, Hannover, Germany) using an in vitro mouse hemidiaphragm assay.10 The patient was positive for a high titer of neutralizing antibodies, suggesting that the cause of the secondary therapy failure experienced by this patient was neutralizing anti-BTX-A antibodies.

In this example, following treatment with a complexing protein-containing BTX-A formulation, the patient developed neutralizing antibodies and subsequently did not respond to any of the BTX-A formulations tested. Resistance that develops following the cosmetic use of BTX-A may impact on the success of any subsequent therapeutic BTX-A treatment (eg, for poststroke spasticity) that the patient may need in the future. It also limits further esthetic use of BTX-A. Some clostridial complexing proteins have been found to enhance antibody production,11 and may therefore increase the risk of neutralizing anti-BTX-A antibodies. Indeed, reducing the foreign protein load of the early preparation of onabotulinumtoxinA decreased the risk of neutralizing anti-BTX-A antibodies.4 A further reduction in the protein load of BTX-A injections, for instance by using highly purified formulations,12 may be beneficial in patients receiving several cycles of BTX-A injections.

For this patient, switching to a botulinum toxin type B (BTX-B) preparation was not considered because the data show that BTX-B has a greater diffusion potential13 and a

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**Table 1 Treatment history**

<table>
<thead>
<tr>
<th>Date</th>
<th>BTX-A preparation</th>
<th>Dosage</th>
<th>Area treated</th>
<th>Duration of effect</th>
<th>Approximate treatment interval</th>
</tr>
</thead>
<tbody>
<tr>
<td>2004</td>
<td>AbobotulinumtoxinA</td>
<td>N/A</td>
<td>Glabellar</td>
<td>First treatment: 12–16 weeks; Second treatment: maximum 8 weeks</td>
<td></td>
</tr>
<tr>
<td>2005</td>
<td>AbobotulinumtoxinA</td>
<td>N/A</td>
<td>Glabellar</td>
<td>4–8 months</td>
<td>6 months</td>
</tr>
<tr>
<td>2006</td>
<td>AbobotulinumtoxinA</td>
<td>N/A</td>
<td>Glabellar</td>
<td>3–4 weeks</td>
<td>6 months</td>
</tr>
<tr>
<td>2007</td>
<td>AbobotulinumtoxinA</td>
<td>N/A</td>
<td>Glabellar</td>
<td>3–4 weeks</td>
<td>6 months</td>
</tr>
<tr>
<td>2008</td>
<td>AbobotulinumtoxinA</td>
<td>N/A</td>
<td>Glabellar</td>
<td>3–4 weeks</td>
<td>6 months</td>
</tr>
<tr>
<td>February 11, 2009</td>
<td>OnabotulinumtoxinA</td>
<td>28 U</td>
<td>Glabellar</td>
<td>Patient complained of incomplete treatment</td>
<td>N/A</td>
</tr>
<tr>
<td>February 25, 2009</td>
<td>OnabotulinumtoxinA</td>
<td>9 U</td>
<td>Glabellar, periorbital</td>
<td>2–3 weeks</td>
<td>1.5 months</td>
</tr>
<tr>
<td>May 28, 2009</td>
<td>OnabotulinumtoxinA</td>
<td>10 U</td>
<td>Glabellar, periorbital</td>
<td>2–3 weeks</td>
<td>3 months</td>
</tr>
<tr>
<td>August 26, 2009</td>
<td>IncobotulinumtoxinA</td>
<td>20 U</td>
<td>Glabellar, periorbital</td>
<td>3–4 weeks</td>
<td>3 months</td>
</tr>
<tr>
<td>December 24, 2009</td>
<td>IncobotulinumtoxinA</td>
<td>22 U</td>
<td>Glabellar, periorbital</td>
<td>3–4 weeks</td>
<td>4 months</td>
</tr>
<tr>
<td>January 19, 2010</td>
<td>IncobotulinumtoxinA</td>
<td>44 U</td>
<td>Glabellar, periorbital</td>
<td>3–4 weeks</td>
<td>1 month</td>
</tr>
</tbody>
</table>

Abbreviations: N/A, not available; BTX-A, botulinum toxin type A.


